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(54) **ATHEROSCLEROSIS-ASSOCIATED GENES**

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(57) **ABSTRACT**

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The present invention relates to a combination comprising a plurality of cDNAs which are differentially expressed in cardiovascular diseases. The combination and compositions can be used entirely or in part to diagnose, to stage, to treat, or to monitor the progression or treatment of disorders associated with atherosclerosis.

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5'  GCT GCC TGC CGG TGC TCT TCG TGG CTC TGG GCA TGG CCT CGG ACC CCA TCT TCA
    9      18      27      36      45      54
    ---
    CGC TGG CGC CCC CGC TGC ATT GCC ACT ACG GGG CCT TCC CCC CTA ATG CCT CTG
    63      72      81      90      99
    ---
    GCT GGG AGC AGC CTC CCA ATG CCA GCG GCG TCA GCG TCG CCA GCG CTG CCC TAG
    117     126     135     144     153
    ---
    CAG CCA GCG CCG CCA GCC GTG TCG CCA CCA GTA CCG ACC CCT CGT GCA GCG GCT
    171     180     189     198
    ---
    TCG CCC CGC CGG ACT TCA ACC ATT GCC CTC AAG GAT TGG GAC TAT AAT GGC CTT
    225     234     243     252     261
    ---
    CCT GTG CTC ACC ACC AAC GCC ATC GGC CAG TGG GAT CTG GTG TGT GAC CTG GGC
    279     288     306     315
    ---
    TGG CAG GTG ATC CTG GAG CAG ATC TTC ATC TTG GGC TTT GCC TCC GGC TAC
    333     342     351     360     369     378
    ---
    
```

FIGURE 1A

711	720	729	738	747	756
TGC ATC CTC	TTT TAT GGC TGG CCT GGT TTG TTC CTG GAG TCC GCA	CGG			
---	---	---	---	---	---
C I L	F L F Y G W P G L F L E S A R				
---	---	---	---	---	---
765	774	783	792	801	810
TGG CTG ATA	GTG AAG CGG CAG ATT GAG GAG GCT CAG TCT GTG CTG AGG ATC	CTG			
---	---	---	---	---	---
W L I	V K R Q I E E A Q S V L R I L				
---	---	---	---	---	---
819	828	837	846	855	864
GCT GAG CGA	AAC CGG CCC CAT GGG CAG ATG CTG GGG GAG GAG GCC CAG GAG	GCC			
---	---	---	---	---	---
A E R	N R P H G Q M L G E E A Q E A				
---	---	---	---	---	---
873	882	891	900	909	918
CTG CAG GAC	CTG GAG AAT ACC TGC CCT CTC CCT GCA ACA TCC TCC TTT TCC	TTT			
---	---	---	---	---	---
L Q D	L E N T C P L P A T S F S F				
---	---	---	---	---	---
927	936	945	954	963	972
GCT TCC CTC	CTC AAC TAC CGC AAC ATC TGG AAA AAT CTG CTT ATC	CTG			
---	---	---	---	---	---
A S L	L L N Y R N I W K N L L I L G F				
---	---	---	---	---	---

FIGURE 1C

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981      990      999      1008      1017      1026
ACC AAC TTC ATT GCC CAT GCC ATT CGC CAC TGC TAC CAG CCT GTG GGA GGA GGA
---
T   N   F   I   A   H   A   I   R   C   Y   Q   P   V   G   G   G
1035      1044      1053      1062      1071      1080
GGG AGC CCA TCG GAC TTC TAC CTG TGC TCT CTG CTG GCC AGC GGC ACC GCA GCC
---
G   S   P   S   D   F   Y   L   C   S   L   L   A   S   G   T   A   A
1089      1098      1107      1116      1125      1134
CTG GCC TGT GTC TTC CTG GGG GTC ACC GTG GAC CGA TTT GGC CGC CGG GGC ATC
---
L   A   C   V   F   L   G   V   T   V   D   R   F   G   R   R   G   I
1143      1152      1161      1170      1179      1188
CTT CTT CTC TCC ATG ACC CTT ACC GGC ATT GCT TCC CTG GTC CTG CTG GGC CTG
---
L   L   L   S   M   T   L   T   G   I   A   S   L   V   L   L   G   L
1197      1206      1215      1224      1233      1242
TGG GAT TAT CTG AAC GAG GCT GCC ATC ACC ACT TTC TCT GTC CTT GGC CTC TTC
---
W   D   Y   L   N   E   A   A   I   T   F   S   V   L   G   L   F

```

FIGURE 1D

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1251      1260      1269      1278      1287      1296
TCC TCC CAA GCT GCC GCC ATC CTC AGC ACC CTC CTT GCT GAG GTC ATC CCC
---
S S Q A A A I L S T L L A A E V I P
---

1305      1314      1323      1332      1341      1350
ACC ACT GTC CGG GGC CGT GGC CTG GGC CTG ATC ATG GCT CTA GGG GCG CTT GGA
---
T T V R G R G L G L I M A L G A L G
---

1359      1368      1377      1386      1395      1404
GGA CTG AGC GGC CCG GCC CAG CGC CTC CAC ATG GGC CAT GGA GCC TTC CTG CAG
---
G L S G P A Q R L L H M G H G A F L Q
---

1413      1422      1431      1440      1449      1458
CAC GTG GTG CTG GCG GCC TGC GCC CTC CTC TGC ATT CTC AGC ATT ATG CTG CTG
---
H V V L A A C A L L C I L S I M L L
---

1467      1476      1485      1494      1503      1512
CCG GAG ACC AAG CGC AAG CTC CTG CCC GAG GTG CTC CCG GAC GGG GAG CTG TGT
---
P E T K R K L L L P E V L R D G E L C

```

FIGURE 1E

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1521      1530      1539      1548      1557      1566
CGC CGG CCT TCC CTG CTG CGG CAG CCA CCC CCT ACC CGC TGT GAC CAC GTC CCG
---
R  R  P  S  L  L  R  Q  P  P  T  R  C  D  H  V  P
---

1575      1584      1593      1602      1611      1620
CTG CTT GCC ACC CCC AAC CCT GCC CTC TGA GCG GCC TCT GAG TAC CCT GGC GGG
---
L  L  A  T  P  N  P  A  L
---

1629      1638      1647      1656      1665      1674
AGG CTG GCC CAC ACA GAA AGG TGG CAA GAA GAT CGG GAA GAC TGA GTA GGG AAG
---

1683      1692      1701      1710      1719      1728
GCA GGG CTG CCC AGA AGT CTC AGA GGC ACC TCA CGC CAG CCA TCG CGG AGA GCT
---

1737      1746      1755      1764      1773      1782
CAG AGG GCC GTC CCC ACC CTG CCT CCT CCC TGC TGC TTT GCA TTC ACT TCC TTG
---

1791      1800      1809      1818      1827      1836
GCC AGA GTC AGG GGA CAG GGA GAG AGC TCC ACA CTG TAA CCA CTG GGT CTG GGC
---
    
```

FIGURE 1F

```

1845      1854      1863      1872      1881      1890
TCC ATC CTG CGC CCA AAG ACA TCC ACC CAG ACC TCA TTA TTT CTT GCT CTA TCA
-----
1899      1908      1917      1926
TTC TGT TTC AAT AAA GAC ATT TGG AAT AAA AAA AAA AAA 3'
-----

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FIGURE 1G

ATHEROSCLEROSIS-ASSOCIATED GENES

FIELD OF THE INVENTION

[0001] The invention relates to a combination of isolated cDNAs that are significantly co-expressed with one or more known atherosclerosis-associated genes. The invention also relates to the use of the combination in the diagnosis, prognosis, treatment, and evaluation of therapies for disorders associated with atherosclerosis.

BACKGROUND OF THE INVENTION

[0002] Atherosclerosis is a disorder characterized by cellular changes in the arterial intima and the formation of arterial plaques containing intracellular and extracellular deposits of lipids. The thickening of artery walls and the narrowing of the arterial lumen underlies the pathologic condition in most cases of coronary artery disease, aortic aneurysm, peripheral vascular disease, and stroke. A number of metabolic pathways and a cascade of molecular events is involved in the cellular morphogenesis, proliferation, and cellular migration that results in atherogenesis (Libby et al. (1997) *Int J Cardiol* 62 (S2):23-29).

[0003] The artery walls consist of three layers: the intima (innermost), the media, and the adventitia (outermost). The intima consists of a layer of endothelial cells lining the lumen of arteries and arterioles. Endothelial cells form a barrier against the indiscriminate entry of substances from the blood into the artery. Specific transporter proteins expressed by endothelial cells facilitate barrier function. Endothelial cells also secrete a number of substances which help regulate downstream vascular contractility blood coagulation, and other aspects of vascular biology. The medial layer of the arterial wall contains smooth muscle cells in a matrix of collagen and elastic fibers produced by the smooth muscle cells. Contraction and relaxation of the smooth muscle layer allows arteries and arterioles to modulate blood pressure and blood flow. The outermost layer of the arterial wall, the adventitia, is a mixture of collagen bundles, elastic fibers, some smooth muscle cells, fibroblasts and nerve cells. The adventitia provides structural integrity to the blood vessel and acts as a support matrix for the media and intima.

[0004] Initiation of an atherosclerotic lesion often occurs following vascular endothelial cell injury often attributable to hypertension, diabetes mellitus, hyperlipidemia, fluctuating shear stress, smoking, or transplant rejection. Nitric oxide and superoxide anions are released and react to form cytotoxic peroxynitrite radicals that cause injury to the endothelium and myocytes of the intima and lead to expression of a variety of molecules that produce local and systemic effects. These effects include the release of mediators of inflammation such as cytokines, complement components, prostaglandins, and downstream transcription factors. Such mediators promote monocyte infiltration of the vascular intima and lead to the upregulation of adhesion molecules which encourages attachment of the monocytes to the damaged endothelial cells. Simultaneously, components of the extracellular matrix including collagens, fibrinogens, and matrix Gla protein are induced and provide sites for monocyte attachment, and annexins, plasminogen activator inhibitor 1, and nitric oxide synthases are induced to counteract these effects.

[0005] Monocytes that infiltrate the lesion accumulate modified low density lipoprotein through scavenger receptors such as CD36 and macrophage scavenger receptor type I. The abundance of modified lipids is a factor in atherogenesis and is influenced by modifying enzymes such as lipoprotein lipase, carboxyl ester lipase, serum amyloid P component, LDL-receptor related protein, microsomal triglyceride transfer protein, and serum esterases such as paraoxonase. Lipid metabolism is governed by cholesterol biosynthesis enzymes such as 3-hydroxy-3-methylglutaryl coenzyme A synthase, and products of the apolipoprotein genes. Modified lipid stabilization and accumulation is aided by perilipin and alpha-2-macroglobulin.

[0006] As monocytes accumulate in the lesion, they can rupture and release free cholesterol, cytokines, and procoagulants into the surrounding environment. This is the process that leads to plaque development; plaque consists of a mass of lipid-engorged monocytes and a lipid-rich necrotic core covered by a fibrous cap. The gradual progression of plaque growth is punctuated by thrombus formation and leads to clinical symptoms such as unstable angina, myocardial infarction, or stroke. Thrombus formation is initiated by episodic plaque rupture which exposes flowing blood to tissue factors, which induce coagulation, and collagen, which activates platelets. After initiation of the atherosclerotic lesion, enzymes that degrade extracellular matrix components (ECM) such as matrix metalloproteinases and cathepsin K are up-regulated, and inhibitors of ECM are down-regulated. This results in destabilization of the atherosclerotic lesion and subsequent complications including myocardial infarction, angina, and stroke. Further arterial occlusion and infiltration increase with the expression of coagulation factors and down-regulation of their inhibitors, antithrombin III, and lipoprotein-associated coagulation inhibitor.

[0007] Smooth muscle cells build up in the arterial media and constitute one of the principal cell types in atherosclerotic and restenotic lesions. They show a high degree of plasticity and are able to shift between a differentiated, contractile phenotype and a less differentiated, synthetic phenotype. This modulation occurs as a response to factors secreted from cells at the site of vascular injury and results in structural reorganization with a loss of myofilaments and the formation of an extensive endoplasmic reticulum and a large Golgi complex. Genes encoding secreted protein, acidic and rich in cysteine (SPARC) and endothelin-1 contribute to these changes. At the same time, the expression of cytoskeletal proteins such as calponin, myosin, desmin, and other gene products in the cells is altered. As a result, the smooth muscle cells lose their contractility and become able to migrate from the media to the intima, to proliferate, and to secrete extracellular matrix components which contribute to arterial intimal thickening.

[0008] The initiation and progression of atherosclerotic lesion development requires the interplay of various molecular pathways. Many genes that participate in these processes are known, and some of them have been shown to have a direct role in atherosclerosis pathogenesis by animal model experiments, in vitro assays, and epidemiological studies (Krettek et al. (1997) *Arterioscler Thromb Vasc Biol* 17:2897-2903; Fisher et al. (1997) *Atherosclerosis* 135:145-159; Shih et al. (1998) *Circulation* 95:2684-2693; and Bocan et al. (1998) *Atherosclerosis* 139:21-30).

[0009] The present invention satisfies a need in the art by providing a combination comprising a plurality of cDNAs that are useful for diagnosis, prognosis, treatment, and evaluation of therapies for disorders associated with atherosclerosis.

SUMMARY OF THE INVENTION

[0010] The invention provides a combination of isolated cDNAs that are significantly co-expressed with one or more known atherosclerosis-associated genes. The combination comprises the isolated cDNAs having the nucleic acid sequences of SEQ ID NOs:1-25 and the complements of SEQ ID NOs:1-25. In one embodiment, the combination is placed on a substrate. In another embodiment, the substrate is a microarray.

[0011] The invention also provides a method for using the combination to detect gene expression in a sample containing nucleic acids, the method comprising hybridizing the substrate containing the combination to the nucleic acids of the sample under conditions for formation of one or more hybridization complexes and detecting hybridization complex formation, wherein complex formation indicates gene expression in the sample. In one embodiment, the sample is from artery or is obtained during microsurgery to open a blocked artery. In another embodiment, complex formation is compared to standards and is diagnostic of a disorder associated with atherosclerosis.

[0012] The invention further provides a method of using a combination to screen a plurality of molecules to identify at least one ligand which specifically binds a cDNA of the combination, the method comprising combining the substrate containing the combination with molecules under conditions to allow specific binding; and detecting specific binding, thereby identifying a ligand which specifically binds at least one cDNA of the combination. In one embodiment, the molecules are selected from DNA molecules, peptides, proteins, RNA molecules, and transcription factors.

[0013] The invention provides an isolated cDNA comprising a polynucleotide having the nucleic acid sequence of SEQ ID NO:8 and the complement of SEQ ID NO:8. In different embodiments, the cDNA is used as a probe, in an expression vector, and in assays for diagnosis, prognosis, and treatment of disorders associated with atherosclerosis. The invention also provides a composition comprising the cDNA and a labeling moiety. The invention further provides a method for using the cDNA to screen a plurality of molecules to identify a ligand which specifically binds the cDNA, the method comprising combining the cDNA with a sample under conditions to allow specific binding; recovering the bound cDNA; and separating the ligand from the bound cDNA, thereby obtaining purified ligand. In one embodiment, the molecules to be screened are selected from DNA molecules, peptides, proteins, RNA molecules, and transcription factors. The invention yet further provides a method for using a cDNA to detect gene expression in a sample containing nucleic acids, the method comprising hybridizing the cDNA to nucleic acids of a sample under conditions for formation of one or more hybridization complexes; and detecting hybridization complex formation, wherein complex formation indicates gene expression in the sample. In one embodiment, the cDNA is attached to a

substrate. In another embodiment, gene expression when compared to standards is diagnostic of a disorder associated with atherosclerosis.

[0014] The invention provides a vector containing the cDNA, and a host cell containing the vector. The invention also provides a method for producing a peptide or protein, the method comprising culturing the host cell under conditions for expression of the peptide or protein; and recovering the peptide or protein so produced from cell culture.

[0015] The invention provides a purified peptide or protein comprising an amino acid sequence expressed by a cDNA of the invention. In one embodiment, the protein comprises the amino acid sequence of SEQ ID NO:26. The invention additionally provides a composition comprising the protein and a pharmaceutical carrier. The invention also provides a method for using a peptide or protein to screen a plurality of molecules to identify at least one ligand which specifically binds the protein. In one embodiment, the molecules to be screened are selected from agonists, antagonists, antibodies, DNA molecules, transcription factors, RNA molecules, and small drug molecules or compounds. The invention further provides a method of using a peptide or protein to purify a ligand.

[0016] The invention provides a method for using the peptide or protein to produce an antibody which specifically binds the protein. A method for preparing polyclonal antibodies comprises immunizing a animal with peptide or protein under conditions to elicit an antibody response, isolating animal antibodies, attaching the peptide or protein to a substrate, contacting the substrate with isolated antibodies under conditions to allow specific binding to the peptide or protein, dissociating the antibodies from the peptide or protein, thereby obtaining purified polyclonal antibodies. A method for preparing monoclonal antibodies comprises immunizing a animal with a peptide or protein under conditions to elicit an antibody response, isolating antibody producing cells from the animal, fusing the antibody producing cells with immortalized cells in culture to form monoclonal antibody producing hybridoma cells, culturing the hybridoma cells, and isolating monoclonal antibodies from culture.

[0017] The invention provides purified antibodies which bind specifically to a peptide or protein. The invention also provides a method for using an antibody to detect expression of a peptide or protein in a sample, the method comprising combining the antibody with a sample under conditions for formation of antibody:peptide or protein complexes, and detecting complex formation, wherein complex formation indicates expression of the peptide or protein in the sample. In one aspect, the amount of complex formation when compared to standards is diagnostic of a disorder of the nervous system.

[0018] The invention provides a method for immunopurification of a protein comprising attaching an antibody to a substrate, exposing the antibody to a sample containing protein under conditions to allow antibody:protein complexes to form, dissociating the protein from the complex, and collecting purified protein. The invention also provides an array upon which a cDNA encoding a protein, the protein, or an antibody which specifically binds the protein are immobilized. The invention also provides a composition comprising a cDNA, a protein, an antibody, or a ligand which has agonistic or antagonistic activity.

[0019] The invention provides an antibody comprising an antigen binding site, wherein the antigen binding site specifically binds to the protein. The invention also provides a method for treating a disorder associated with the differential expression of a cDNA that is coexpressed with one or more known atherosclerosis-associated genes in a subject in need, the method comprising the step of administering to the subject in need the antibody in an amount effective for treating the disorder. The invention further provides an immunoconjugate comprising the antigen binding site of the antibody or joined to a therapeutic agent. The invention additionally provides a method for treating a disorder associated with the differential expression of a cDNA that is coexpressed with one or more known atherosclerosis-associated genes in a subject in need, the method comprising the step of administering to the subject in need the immunoconjugate in an amount effective for treating the disorder.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING, FIGURE AND TABLES

[0020] The Sequence Listing provides exemplary cDNAs associated with atherosclerosis including polynucleotide sequences SEQ ID NOs:1-25 and the polypeptide sequence, SEQ ID NO:26. Each sequence is identified by a sequence identification number (SEQ ID NO).

[0021] FIG. 1 shows the cDNA having the nucleic acid sequence of SEQ ID NO:8 which encodes the protein having the amino acid sequence of SEQ ID NO:26. The alignment was produced using MAdNASIS,PRO software (Hitachi Software Engineering, South San Francisco Calif.).

[0022] Table 4 shows the co-expression values (-log p) between the known atherosclerosis-associated genes (abbreviations as shown in Table 3) and the cDNAs of the invention (SEQ IDs).

[0023] Table 5 summarizes the highly significantly co-expression between each cDNA (SEQ ID) and two known atherosclerosis-associated genes (Gene 1 and Gene 2). P-value and the function or importance of the known atherosclerosis-associated gene are derived from Tables 3 and 4.

[0024] Table 6 shows transcript images for several of the cDNAs (SEQ ID) of the invention. Column 1 shows the SEQ ID; column 2, the library name; column 3, the number of cDNAs in the library; column 4, the description of the sample from which the library was constructed; column 5, transcript abundance; and column 6, percent transcript abundance. These sets demonstrate differential expression within experiments using cardiovascular tissues.

[0025] Table 7 shows microarray data for several of the cDNAs (SEQ ID) of the invention. Column 1 shows the SEQ ID; column 2, the name of the microarray (GEM) used for the experiment; column 3, the log₂ (C5/Cy3 ratio); column 4, the description of the Cy3 sample; and column 5, the description of the Cy5 sample. These data demonstrate differential expression of the cDNAs in experiments using cardiovascular samples.

DESCRIPTION OF THE INVENTION

[0026] It must be noted that as used herein and in the appended claims, the singular forms "a", "an", and "the" include the plural reference unless the context clearly dic-

tates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

[0027] Definitions

[0028] "Antibody" refers to intact immunoglobulin molecule, a polyclonal antibody, a monoclonal antibody, a chimeric antibody, a recombinant antibody, a humanized antibody, single chain antibodies, a Fab fragment, an F(ab')₂ fragment, an Fv fragment; and an antibody-peptide fusion protein.

[0029] "Antigenic determinant" refers to an antigenic or immunogenic epitope, structural feature, or region of an oligopeptide, peptide, or protein which is capable of inducing formation of an antibody which specifically binds the protein. Biological activity is not a prerequisite for immunogenicity.

[0030] "Array" refers to an ordered arrangement of at least two cDNAs, proteins, or antibodies on a substrate. At least one of the cDNAs, proteins, or antibodies represents a control or standard, and the other represents a cDNA, protein, or antibody of diagnostic or therapeutic interest. The arrangement of at least two and up to about 40,000 cDNAs, proteins, or antibodies on the substrate assures that the size and signal intensity of each labeled complex, formed between each cDNA and at least one nucleic acid, each protein and at least one ligand or antibody, or each antibody and at least one protein to which the antibody specifically binds, is individually distinguishable.

[0031] "Atherosclerosis-associated cDNA" refers to isolated polynucleotide that exhibits a statistically significant co-expression pattern with known atherosclerosis-associated genes. The cDNAs are represented by SEQ ID NOs:1-25 of the Sequence Listing and the complements of SEQ ID NOs:1-25. They may be of recombinant or synthetic origin, used in its double-stranded or single-stranded form, and combined with vitamins, minerals, carbohydrates, lipids, proteins, other nucleic acids, a pharmaceutical carrier or a labeling moiety to perform a particular activity or form a useful composition.

[0032] An "atherosclerotic sample or tissue" may be taken using needles, catheters, or scalpels and include vessels including the aorta, arteries, arterioles, endothelial cells, plaque, and blood. The sample may contain nucleic acids, proteins, antibodies, and the like. Additionally the sample may comprise the soluble fraction of a cell preparation; an aliquot of media in which cells were grown; a chromosome, an organelle, or membrane isolated or extracted from a cell; genomic DNA, RNA, or cDNA in solution or bound to a substrate.

[0033] A "combination" comprises at least two sequences selected from SEQ ID NOs:1-25 as presented in the Sequence Listing and the complements of SEQ ID NOs:1-25.

[0034] "Differential expression" refers to an increased or up-regulated or a decreased or down-regulated expression as detected by absence, presence, or at least two-fold change in the amount of transcribed messenger RNA or translated protein in a sample.

[0035] “Disorders associated with atherosclerosis” include angina pectoris, coronary artery disease, myocardial infarction, hypertension, transient cerebral ischemia, mesenteric ischemia, peripheral vascular disease, renal artery stenosis, and stroke.

[0036] An “expression profile” is a representation of gene expression in a sample. A nucleic acid expression profile is produced using sequencing, hybridization, or amplification technologies and mRNAs or cDNAs from a sample. A protein expression profile, although time delayed, mirrors the nucleic acid expression profile and uses two-dimensional polyacrylamide electrophoresis (2D-PAGE) and mass spectrophotometry (MS) or western analysis, enzyme-linked immunosorbent assays (ELISAs), fluorescence activated cell sorting (FACS), radioimmunoassays (RIAs), or arrays and labeling moieties or antibodies to detect expression in a sample. The nucleic acids, proteins, or antibodies may be used in solution or attached to a substrate, and their detection is based on methods and labeling moieties well known in the art.

[0037] A “hybridization complex” is formed between a cDNA of the invention and a nucleic acid of a sample when the purines of one molecule hydrogen bond with the pyrimidines of the complementary molecule, e.g., 5'-A-G-T-C-3' base pairs with its complete complement, 3'-T-C-A-G-5'. The degree of complementarity and the use of nucleotide analogs affect the efficiency and stringency of hybridization reactions.

[0038] “Identity” as applied to sequences, refers to the quantification (usually percentage) of nucleotide or residue matches between at least two sequences aligned using a standardized algorithm such as Smith-Waterman alignment (Smith and Waterman (1981) *J Mol Biol* 147:195-197), CLUSTALW (Thompson et al. (1994) *Nucleic Acids Res* 22:4673-4680), or BLAST2 (Altschul et al. (1997) *Nucleic Acids Res* 25:3389-340). BLAST2 may be used in a standardized and reproducible way to insert gaps in one of the sequences in order to optimize alignment and to achieve a more meaningful comparison between them.

[0039] “Similarity” as applied to proteins uses the same algorithms but takes into account conservative substitutions of nucleotides or residues.

[0040] “Isolated or purified” refers to a cDNA or protein that is removed from its natural environment and that is separated from other components with which it is naturally present.

[0041] “Genes known to be associated with atherosclerosis” include human 22 kDa smooth muscle protein, calponin (CNN1), pro alpha 1 (I) collagen (COL1A1), collagen alpha-2 type I (COL1A2), collagen alpha-6 type I (COL6A1), procollagen alpha 2(V) (COL5A2), collagen VI alpha-2 (COL6A2), type VI collagen alpha 3 (COL6A3), pro-alpha-i type 3 collagen (COL3A1), pro-alpha-1(V) collagen (COLSA1), matrix Gla protein (MGP), cathepsin K (CSTK), fibrinogen beta chain gene (FBG), pre-pro-von Willebrand factor (VWF), platelet endothelial cell adhesion molecule (PECAM-1), antithrombin III variant (AT3), lipoprotein lipase (LPL), alpha-2-macroglobulin (A2M), apolipoprotein AI (APOA1), apolipoprotein AII (APOA1)₂, apolipoprotein B-100 (APOB), lipoprotein apoCII (APOC2), pre-apolipoprotein CIII (APOC3), apolipoprotein apo C-IV

(APOC4), macrophage scavenger receptor type I (MSR1), human antigen CD36 gene (CD36), serum amyloid P component (SAP), carboxyl ester lipase gene (CEL), paraoxonase 1 (PONI), paraoxonase 2 (PON2), paraoxonase 3 (PON3), perilipin (PLIN), prostaglandin D2 synthase (PTGDS), annexin II/lipocortin II (ANX2), annexin I/lipocortin (ANX1), and secreted protein, acidic and rich in cysteine (SPARC).

[0042] “Labeling moiety” refers to any reporter molecule whether a visible or radioactive label, stain or dye that can be attached to or incorporated into a cDNA or protein. Visible labels and dyes include but are not limited to anthocyanins, B glucuronidase, BIODIPY, Coomassie blue, Cy3 and Cy5, digoxigenin, FITC, green fluorescent protein, luciferase, spiro red, silver, and the like. Radioactive markers include radioactive forms of hydrogen, iodine, phosphorous, sulfur, and the like.

[0043] “Ligand” refers to any agent, molecule, or compound which will bind specifically to a complementary site on a cDNA molecule, a polynucleotide, an epitope or a protein. Such ligands stabilize or modulate the activity of polynucleotides or proteins and may be composed of inorganic or organic substances including nucleic acids, proteins, carbohydrates, fats, and lipids.

[0044] “Markers for disorders associated with atherosclerosis” refers to cDNAs, peptides or proteins, and antibodies which are useful in the diagnosis, prognosis, treatment, selection or evaluation of therapies for disorders associated with atherosclerosis. These markers are differentially expressed in samples from subjects predisposed to or manifesting one of these disorders. The known atherosclerosis-associated genes and their contribution and/or function to disorders associated with atherosclerosis are listed in TABLE3.

[0045] “Probe” refers to a cDNA of the invention that hybridizes to at least one nucleic acid in a sample. Where targets are single stranded, probes are complementary single strands. Probes can be labeled for use in hybridization reactions including Southern, northern, in situ, dot blot, array, and like technologies or in screening assays.

[0046] “Protein” refers to a polypeptide or any portion thereof. An “oligopeptide” is an amino acid sequence from about five residues to about 15 residues that is used as part of a fusion protein to produce an antibody that specifically binds the protein.

[0047] “Specific binding” refers to a special and precise interaction between two molecules which is dependent upon their structure, particularly their molecular side groups. For example, the intercalation of a regulatory protein into the major groove of a DNA molecule, the hydrogen bonding along the backbone between two single stranded nucleic acids, or the binding between an epitope of a protein and an agonist, antagonist, or antibody.

[0048] “Substrate” refers to any rigid or semi-rigid support to which cDNAs or proteins are bound and includes membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, capillaries or other tubing, plates, polymers, and microparticles with a variety of surface forms including wells, trenches, pins, channels and pores.

[0049] A “transcript image” (TI) is a profile of gene transcription activity in a particular tissue at a particular

time. TI provides assessment of the relative abundance of expressed transcripts in the cDNA libraries of an EST database as described in U.S. Pat. No. 5,840,484, incorporated herein by reference.

[0050] "Variant" refers to molecules that are recognized variations of a polynucleotide or a protein. Splice variants may be determined by BLAST score, wherein the score is at least 100, and most preferably at least 400. Allelic variants have a high percent identity to a polynucleotide and may differ by about three bases per hundred bases. "Single nucleotide polymorphism" (SNP) refers to a change in a single base as a result of a substitution, insertion or deletion. The change may be conservative (purine for purine) or non-conservative (purine to pyrimidine) and may or may not result in a change in an encoded amino acid.

[0051] The Method

[0052] The present invention encompasses a method for identifying cDNAs that are significantly co-expressed with known atherosclerosis-associated genes. In particular, the method identifies a combination of cDNAs useful in diagnosis, prognosis, treatment, and evaluation of therapies for disorders associated with atherosclerosis.

[0053] The method involves identifying cDNAs that are expressed in a plurality of cDNA libraries. These cDNAs include genes of known or unknown function whose expression patterns are compared with genes having a known function and disease association to determine whether a specified coexpression probability threshold is met. Through this comparison, a subset of the cDNAs having a high coexpression probability with the known genes can be identified.

[0054] The cDNAs may originate from cDNA libraries derived from a variety of sources including, but not limited to, eukaryotes such as human, mouse, rat, dog, monkey, plant, and yeast; prokaryotes such as bacteria; and viruses. The cDNAs can also be selected from a variety of sequence types including, but not limited to, expressed sequence tags (ESTs), assembled polynucleotide sequences, full length gene coding regions, promoters, introns, enhancers, 5' untranslated regions, and 3' untranslated regions. To have statistically significant analytical results, the cDNAs need to be expressed in at least three cDNA libraries.

[0055] The cDNA libraries used in the coexpression analysis of the present invention can be obtained from adrenal gland, biliary tract, bladder, blood cells, blood vessels, bone marrow, brain, bronchus, cartilage, chromaffin system, colon, connective tissue, cultured cells, embryonic stem cells, endocrine glands, epithelium, esophagus, fetus, ganglia, heart, hypothalamus, immune system, intestine, islets of Langerhans, kidney, larynx, liver, lung, lymph, muscles, neurons, ovary, pancreas, penis, peripheral nervous system, phagocytes, pituitary, placenta, pleurus, prostate, salivary glands, seminal vesicles, skeleton, spleen, stomach, testis, thymus, tongue, ureter, uterus, and the like. The number of cDNA libraries selected can range from as few as 3 to greater than 10,000. Preferably, the number of the cDNA libraries is greater than 500.

[0056] In a preferred embodiment, the cDNAs are assembled from related sequences, such as assembled sequence fragments derived from a single transcript. Assembly of the sequences can be performed using sequences of

various types including, but not limited to, ESTs, extensions, or shotgun sequences. In a most preferred embodiment, the cDNAs are derived from human sequences that have been assembled using the algorithm disclosed in U.S. Ser. No. 09/276,534, filed Mar. 25, 1999, incorporated herein by reference.

[0057] Experimentally, differential expression of the cDNAs can be evaluated by methods including, but not limited to, differential display by spatial immobilization or by gel electrophoresis, genome mismatch scanning, representational difference analysis, and transcript imaging. Additionally, differential expression can be assessed by microarray technology. These methods may be used alone or in combination.

[0058] The known atherosclerosis-associated genes were selected based on their function in pathways associated with atherogenesis, their use as diagnostic or prognostic markers, their behavior in model systems or their use as therapeutic targets.

[0059] The procedure for identifying a cDNA that exhibits a statistically significant coexpression pattern with known atherosclerosis-associated genes is as follows. First, the presence or absence of a gene in a cDNA library is defined: a gene is present in a cDNA library when at least one cDNA fragment corresponding to that gene is detected in a cDNA sample taken from the library, and a gene is absent from a library when no corresponding cDNA fragment is detected in the sample.

[0060] Second, the significance of gene coexpression is evaluated using a probability method to measure a due-to-chance probability of the coexpression. The probability method can be the Fisher exact test, the chi-squared test, or the kappa test. These tests and examples of their applications are well known in the art and can be found in standard statistics texts (Agresti (1990) *Categorical Data Analysis*, John Wiley & Sons, New York N.Y.; Rice (1988) *Mathematical Statistics and Data Analysis*, Duxbury Press, Pacific Grove Calif.). A Bonferroni correction (Rice, supra, p. 384) can also be applied in combination with one of the probability methods for correcting statistical results of one gene versus multiple other genes. In a preferred embodiment, the due-to-chance probability is measured by a Fisher exact test, and the threshold of the due-to-chance probability is set preferably to less than 0.001, more preferably to less than 0.00001.

[0061] To determine whether two genes, A and B, have similar coexpression patterns, occurrence data vectors can be generated as illustrated in Table 1. The presence of a gene occurring at least once in a library is indicated by a one, and its absence from the library, by a zero.

TABLE 1

Occurrence data for genes A and B					
	Library 1	Library 2	Library 3	...	Library N
gene A	1	1	0	...	0
gene B	1	0	1	...	0

[0062] For a given pair of genes, the occurrence data in Table 1 can be summarized in a 2x2 contingency table.

TABLE 2

Contingency table for co-occurrences of genes A and B			
	Gene A present	Gene A absent	Total
Gene B present	8	2	10
Gene B absent	<u>2</u>	<u>18</u>	<u>20</u>
Total	10	20	30

[0063] Table 2 presents co-occurrence data for gene A and gene B in a total of 30 libraries. Both gene A and gene B occur 10 times in the libraries. Table 2 summarizes and presents: 1) the number of times gene A and B are both present in a library; 2) the number of times gene A and B are both absent in a library; 3) the number of times gene A is present, and gene B is absent; and 4) the number of times gene B is present, and gene A is absent. The upper left entry is the number of times the two genes co-occur in a library, and the middle right entry is the number of times neither gene occurs in a library. The off diagonal entries are the number of times one gene occurs, and the other does not. Both A and B are present eight times and absent 18 times. Gene A is present, and gene B is absent, two times; and gene B is present, and gene A is absent, two times. The probability ("p-value") that the above association occurs due to chance as calculated using a Fisher exact test is 0.0003. Associations are generally considered significant if a p-value is less than 0.01 (Agresti, supra; Rice, supra).

[0064] This method of estimating the probability for co-expression of two genes makes several assumptions. The method assumes that the libraries are independent and are identically sampled. However, in practical situations, the selected cDNA libraries are not entirely independent, because more than one library may be obtained from a single subject or tissue. Nor are they entirely identically sampled, because different numbers of cDNAs may be sequenced from each library. The number of cDNAs sequenced typically ranges from 5,000 to 10,000 cDNAs per library. In addition, because a Fisher exact co-expression probability is calculated for each gene versus 45,233 other assembled genes, a Bonferroni correction for multiple statistical tests is used.

[0065] The Invention

[0066] The present invention identifies 25 atherosclerosis-associated cDNAs that exhibit strong association with genes known to be specifically expressed in atherosclerosis. The results presented in Tables 4 and 5 show that the expression of the 25 novel atherosclerosis-associated cDNAs have direct association with the expression of known atherosclerosis-associated genes as described in Table 3 and in the background of the invention. Therefore, the novel atherosclerosis-associated cDNAs can potentially be used in diagnosis, prognosis, treatment or evaluation of therapies for disorders associated with atherosclerosis. Further, the gene products of the 25 novel atherosclerosis-associated cDNAs are either potential therapeutics or targets for the development of therapeutics against disorders associated with atherosclerosis.

[0067] Therefore, in one embodiment, the present invention encompasses a combination comprising a plurality of cDNAs having the nucleic acid sequences of SEQ ID NOs:1-25 or the complements of SEQ ID NOs:1-25. These 25 cDNAs have been shown by the method of the present invention to have statistically significant co-expression with known atherosclerosis-associated genes and with each other. The invention also encompasses a cDNA comprising a polynucleotide having the nucleic acid sequence of SEQ ID NO:8 and the complement thereof. As shown in FIG. 1, SEQ ID NO:8 encodes the protein of SEQ ID NO:26. The invention further encompasses a protein comprising the polypeptide having the amino acid sequence of SEQ ID NO:26.

[0068] The protein encoded by SEQ ID NO:8 has 366 amino acids. Motif analyses of SEQ ID NO:26 shows one potential cAMP- and cGMP-dependent protein kinase phosphorylation site at residue S343, two potential casein kinase II phosphorylation sites at residues S179 and T351, and four potential protein kinase C phosphorylation sites at residues T29, S85, T269, and T324. Additionally, SEQ ID NO:26 contains a potential sugar transport protein signature sequence from residues L201 to S217.

[0069] cDNAs and Their Uses

[0070] cDNAs can be prepared by a variety of synthetic or enzymatic methods well known in the art. cDNAs can be synthesized, in whole or in part, using chemical methods well known in the art (Caruthers et al. (1980) *Nucleic Acids Symp Ser* (7):215-233). Alternatively, cDNAs can be produced enzymatically or recombinantly, by in vitro or in vivo transcription.

[0071] Nucleotide analogs can be incorporated into cDNAs by methods well known in the art. The only requirement is that the incorporated analog must base pair with native purines or pyrimidines. For example, 2,6-diaminopurine can substitute for adenine and form stronger bonds with thymidine than those between adenine and thymidine. A weaker pair is formed when hypoxanthine is substituted for guanine and base pairs with cytosine. Additionally, cDNAs can include nucleotides that have been derivatized chemically or enzymatically.

[0072] cDNAs can be synthesized on a substrate. Synthesis on the surface of a substrate may be accomplished using a chemical coupling procedure and a piezoelectric printing apparatus as described by Baldeschweiler et al. (PCT publication WO95/251116). Alternatively, the cDNAs can be synthesized on a substrate surface using a self-addressable electronic device that controls when reagents are added as described by Heller et al. (U.S. Pat. No. 5,605,662). cDNAs can be synthesized directly on a substrate by sequentially dispensing reagents for their synthesis on the substrate surface or by dispensing preformed DNA fragments to the substrate surface. Typical dispensers include a micropipette delivering solution to the substrate with a robotic system to control the position of the micropipette with respect to the substrate. There can be a multiplicity of dispensers so that reagents can be delivered to the reaction regions efficiently.

[0073] cDNAs can be immobilized on a substrate by covalent means such as by chemical bonding procedures or UV irradiation. In one method, a cDNA is bound to a glass surface which has been modified to contain epoxide or

aldehyde groups. In another method, a cDNA is placed on a polylysine coated surface and UV cross-linked to it as described by Shalon et al. (WO95/35505). In yet another method, a cDNA is actively transported from a solution to a given position on a substrate by electrical means (Heller, supra). cDNAs do not have to be directly bound to the substrate, but rather can be bound to the substrate through a linker group. The linker groups are typically about 6 to 50 atoms long to provide exposure of the attached cDNA. Preferred linker groups include ethylene glycol oligomers, diamines, diacids and the like. Reactive groups on the substrate surface react with a terminal group of the linker to bind the linker to the substrate. The other terminus of the linker is then bound to the cDNA. Alternatively, polynucleotides, plasmids or cells can be arranged on a filter. In the latter case, cells are lysed, proteins and cellular components degraded, and the DNA is coupled to the filter by UV cross-linking.

[0074] The cDNAs may be used for a variety of purposes. For example, the combination of the invention may be used on an array. The array, in turn, can be used in high-throughput methods for detecting a related polynucleotide in a sample, screening a plurality of molecules or compounds to identify a ligand, diagnosing a disorder such as diabetes, or inhibiting or inactivating a therapeutically relevant gene related to the cDNA.

[0075] When the cDNAs of the invention are employed on an array, the cDNAs are arranged in an ordered fashion so that each cDNA is present at a specified location. Because the cDNAs are at specified locations on the substrate, the hybridization patterns and intensities, which together create a unique, can be interpreted in terms of expression levels of particular genes and can be correlated with a particular metabolic process, condition, disorder, disease, stage of disease, or treatment.

[0076] Hybridization

[0077] The cDNAs or fragments or complements thereof may be used in various hybridization technologies. The cDNAs may be labeled using a variety of reporter molecules by either PCR, recombinant, or enzymatic techniques. For example, a commercially available vector containing the cDNA is transcribed in the presence of an appropriate polymerase, such as T7 or SP6 polymerase, and at least one labeled nucleotide. Commercial kits are available for labeling and cleanup of such cDNAs. Radioactive (Amersham Biosciences (APB), Piscataway N.J.), fluorescent (Qiagen-Operon, Alameda Calif.), and chemiluminescent labeling (Promega, Madison Wis.) are well known in the art.

[0078] A cDNA may represent the complete coding region of an mRNA or be designed or derived from unique regions of the mRNA or genomic molecule, an intron, a 3' untranslated region, or from a conserved motif. The cDNA is at least 18 contiguous nucleotides in length and is usually single stranded. Such a cDNA may be used under hybridization conditions that allow binding only to an identical sequence, a naturally occurring molecule encoding the same protein, or an allelic variant. Discovery of related human and mammalian sequences may also be accomplished using a pool of degenerate cDNAs and appropriate hybridization conditions. Generally, a cDNA for use in Southern or northern hybridizations may be from about 400 to about 6000 nucleotides long. Such cDNAs have high binding specificity in

solution-based or substrate-based hybridizations. An oligonucleotide, a fragment of the cDNA, may be used to detect a polynucleotide in a sample using PCR.

[0079] The stringency of hybridization is determined by G+C content of the cDNA, salt concentration, and temperature. In particular, stringency is increased by reducing the concentration of salt or raising the hybridization temperature. In solutions used for some membrane based hybridizations, addition of an organic solvent such as formamide allows the reaction to occur at a lower temperature. Hybridization may be performed with buffers, such as 5x saline sodium citrate (SSC) with 1% sodium dodecyl sulfate (SDS) at 60° C., that permit the formation of a hybridization complex between nucleic acid sequences that contain some mismatches. Subsequent washes are performed with buffers such as 0.2xSSC with 0.1% SDS at either 45° C. (medium stringency) or 65°-68° C. (high stringency). At high stringency, hybridization complexes will remain stable only where the nucleic acids are completely complementary. In some membrane-based hybridizations, preferably 35% or most preferably 50%, formamide may be added to the hybridization solution to reduce the temperature at which hybridization is performed. Background signals may be reduced by the use of detergents such as Sarkosyl or TRITON X-100 (Sigma-Aldrich, St. Louis Mo.) and a blocking agent such as denatured salmon sperm DNA. Selection of components and conditions for hybridization are well known to those skilled in the art and are reviewed in Ausubel et al. (1997, *Short Protocols in Molecular Biology*, John Wiley & Sons, New York N.Y., Units 2.8-2.11, 3.18-3.19 and 4.6-4.9).

[0080] Dot-blot, slot-blot, low density and high density arrays are prepared and analyzed using methods known in the art. cDNAs from about 18 consecutive nucleotides to about 5000 consecutive nucleotides in length are contemplated by the invention and used in array technologies. The preferred number of cDNAs on an array is at least about 100,000, a more preferred number is at least about 40,000, an even more preferred number is at least about 10,000, and a most preferred number is at least about 600 to about 800. The array may be used to monitor the expression level of large numbers of genes simultaneously and to identify genetic variants, mutations, and SNPs. Such information may be used to determine gene function; to understand the genetic basis of a disorder; to diagnose a disorder; and to develop and monitor the activities of therapeutic agents being used to control or cure a disorder. (See, e.g., U.S. Pat. No. 5,474,796; WO95/11995; WO95/35505; U.S. Pat. No. 5,605,662; and U.S. Pat. No. 5,958,342.)

[0081] Screening and Purification Assays Using cDNAs

[0082] A cDNA may be used to screen a library or a plurality of molecules or compounds for a ligand which specifically binds the cDNA. Ligands may be DNA molecules, RNA molecules, peptide nucleic acid molecules, peptides, proteins such as transcription factors, promoters, enhancers, repressors, and other proteins that regulate replication, transcription, or translation of the polynucleotide in the biological system. The assay involves combining the cDNA or a fragment thereof with the molecules or compounds under conditions that allow specific binding and detecting the bound cDNA to identify at least one ligand that specifically binds the cDNA.

[0083] In one embodiment, the cDNA may be incubated with a library of isolated and purified molecules or compounds and binding activity determined by methods such as a gel-retardation assay (U.S. Pat. No. 6,010,849) or a reticulocyte lysate transcriptional assay. In another embodiment, the cDNA may be incubated with nuclear extracts from biopsied and/or cultured cells and tissues. Specific binding between the cDNA and a molecule or compound in the nuclear extract is initially determined by gel shift assay. Protein binding may be confirmed by raising antibodies against the protein and adding the antibodies to the gel-retardation assay where specific binding will cause a supershift in the assay.

[0084] In another embodiment, the cDNA may be used to purify a molecule or compound using affinity chromatography methods well known in the art. In one embodiment, the cDNA is chemically reacted with cyanogen bromide groups on a polymeric resin or gel. Then a sample is passed over and reacts with or binds to the cDNA. The molecule or compound which is bound to the cDNA may be released from the cDNA by increasing the salt concentration of the flow-through medium and collected.

[0085] The cDNA may be used to purify a ligand from a sample. A method for using a cDNA to purify a ligand would involve combining the cDNA or a fragment thereof with a sample under conditions to allow specific binding, recovering the bound cDNA, and using an appropriate agent to separate the cDNA from the purified ligand.

[0086] Protein Production and Uses

[0087] The full length cDNAs or fragments thereof may be used to produce purified proteins using recombinant DNA technologies described herein and taught in Ausubel (supra; Units 16.1-16.62). One of the advantages of producing proteins by these procedures is the ability to obtain highly-enriched sources of the proteins thereby simplifying purification procedures.

[0088] The proteins may contain amino acid substitutions, deletions or insertions made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. Such substitutions may be conservative in nature when the substituted residue has structural or chemical properties similar to the original residue (e.g., replacement of leucine with isoleucine or valine) or they may be nonconservative when the replacement residue is radically different (e.g., a glycine replaced by a tryptophan). Computer programs included in LASERGENE software (DNASTAR, Madison Wis.) and algorithms included in RasMol software (University of Massachusetts, Amherst Mass.) may be used to help determine which and how many amino acid residues in a particular portion of the protein may be substituted, inserted, or deleted without abolishing biological or immunological activity.

[0089] Expression of Encoded Proteins

[0090] Expression of a particular cDNA may be accomplished by cloning the cDNA into a vector and transforming this vector into a host cell. The cloning vector used for the construction of cDNA libraries in the LIFESEQ databases (Incyte Genomics, Palo Alto Calif.) may also be used for expression. Such vectors usually contain a promoter and a polylinker useful for cloning, priming, and transcription. An

exemplary vector may also contain the promoter for β -galactosidase, an amino-terminal methionine and the subsequent seven amino acid residues of β -galactosidase. The vector may be transformed into competent *E. coli* cells. Induction of the isolated bacterial strain with isopropylthiogalactoside (IPTG) using standard methods will produce a fusion protein that contains an N terminal methionine, the first seven residues of β -galactosidase, about 15 residues of linker, and the protein encoded by the cDNA.

[0091] The cDNA may be shuttled into other vectors known to be useful for expression of protein in specific hosts. Oligonucleotides containing cloning sites and fragments of DNA sufficient to hybridize to stretches at both ends of the cDNA may be chemically synthesized by standard methods. These primers may then be used to amplify the desired fragments by PCR. The fragments may be digested with appropriate restriction enzymes under standard conditions and isolated using gel electrophoresis. Alternatively, similar fragments are produced by digestion of the cDNA with appropriate restriction enzymes and filled in with chemically synthesized oligonucleotides. Fragments of the coding sequence from more than one gene may be ligated together and expressed.

[0092] Signal sequences that dictate secretion of soluble proteins are particularly desirable as component parts of a recombinant sequence. For example, a chimeric protein may be expressed that includes one or more additional purification-facilitating domains. Such domains include, but are not limited to, metal-chelating domains that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex, Seattle Wash.). The inclusion of a cleavable-linker sequence such as ENTEROKINASEMAX (Invitrogen, San Diego Calif.) between the protein and the purification domain may also be used to recover the protein.

[0093] Suitable host cells may include, but are not limited to, mammalian cells such as Chinese Hamster Ovary (CHO) and human 293 cells, insect cells such as Sf9 cells, plant cells such as *Nicotiana tabacum*, yeast cells such as *Saccharomyces cerevisiae*, and bacteria such as *E. coli*. For each of these cell systems, a useful vector may also include an origin of replication and one or two selectable markers to allow selection in bacteria as well as in a transformed eukaryotic host. Vectors for use in eukaryotic host cells may require the addition of 3' poly(A) tail if the cDNA lacks poly(A).

[0094] Additionally, the vector may contain promoters or enhancers that increase gene expression. Many promoters are known and used in the art. Most promoters are host specific and exemplary promoters includes SV40 promoters for CHO cells; T7 promoters for bacterial hosts; viral promoters and enhancers for plant cells; and PGH promoters for yeast. Adenoviral vectors with the rous sarcoma virus enhancer or retroviral vectors with long terminal repeat promoters may be used to drive protein expression in mammalian cell lines. Once homogeneous cultures of recombinant cells are obtained, large quantities of secreted soluble protein may be recovered from the conditioned medium and analyzed using chromatographic methods well known in the art. An alternative method for the production of large amounts of secreted protein involves the transfor-

mation of mammalian embryos and the recovery of the recombinant protein from milk produced by transgenic cows, goats, sheep, and the like.

[0095] In addition to recombinant production, proteins or portions thereof may be produced manually, using solid-phase techniques (Stewart et al. (1969) *Solid-Phase Peptide Synthesis*, W H Freeman, San Francisco Calif.; Merrifield (1963) *J Am Chem Soc* 5:2149-2154), or using machines such as the 431A peptide synthesizer (Applied Biosystems (ABI), Foster City Calif.). Proteins produced by any of the above methods may be used as pharmaceutical compositions to treat disorders associated with null or inadequate expression of the genomic sequence.

[0096] Screening and Purification Assays Using Proteins

[0097] A protein or a portion thereof encoded by the cDNA may be used to screen a library or a plurality of molecules or compounds for a ligand with specific binding affinity or to purify a molecule or compound from a sample. The protein or portion thereof employed in such screening may be free in solution, affixed to an abiotic or biotic substrate, or located intracellularly. For example, viable or fixed prokaryotic host cells that are stably transformed with recombinant nucleic acids that have expressed and positioned a protein on their cell surface can be used in screening assays. The cells are screened against a library or a plurality of ligands and the specificity of binding or formation of complexes between the expressed protein and the ligand may be measured. The ligands may be agonists, antagonists, antibodies, DNA molecules, enhancers, small drug molecules, immunoglobulins, inhibitors, mimetics, peptide nucleic acid molecules, peptides, pharmaceutical agents, proteins, and regulatory proteins, repressors, RNA molecules, ribozymes, transcription factors, or any other test molecule or compound that specifically binds the protein. An exemplary assay involves combining the mammalian protein or a portion thereof with the molecules or compounds under conditions that allow specific binding and detecting the bound protein to identify at least one ligand that specifically binds the protein.

[0098] This invention also contemplates the use of competitive drug screening assays in which neutralizing antibodies capable of binding the protein specifically compete with a test compound capable of binding to the protein or oligopeptide or fragment thereof. One method for high throughput screening using very small assay volumes and very small amounts of test compound is described in U.S. Pat. No. 5,876,946. Molecules or compounds identified by screening may be used in a model system to evaluate their toxicity, diagnostic, or therapeutic potential.

[0099] The protein may be used to purify a ligand from a sample. A method for using a protein to purify a ligand would involve combining the protein or a portion thereof with a sample under conditions to allow specific binding, recovering the bound protein, and using an appropriate chaotropic agent to separate the protein from the purified ligand.

[0100] Production of Antibodies

[0101] A protein encoded by a cDNA of the invention may be used to produce specific antibodies. Antibodies may be produced using an oligopeptide or a portion of the protein with inherent immunological activity. Methods for produc-

ing antibodies include: 1) injecting an animal, usually goats, rabbits, or mice, with the protein, or an antigenically-effective portion or an oligopeptide thereof, to induce an immune response; 2) engineering hybridomas to produce monoclonal antibodies; 3) inducing in vivo production in the lymphocyte population; or 4) screening libraries of recombinant immunoglobulins. Recombinant immunoglobulins may be produced as taught in U.S. Pat. No. 4,816,567.

[0102] Antibodies produced using the proteins of the invention are useful for the diagnosis of prepathologic disorders as well as the diagnosis of chronic or acute diseases characterized by abnormalities in the expression, amount, or distribution of the protein. A variety of protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies specific for proteins are well known in the art. Immunoassays typically involve the formation of complexes between a protein and its specific binding molecule or compound and the measurement of complex formation. Immunoassays may employ a two-site, monoclonal-based assay that utilizes monoclonal antibodies reactive to two noninterfering epitopes on a specific protein or a competitive binding assay (Pound (1998) *Immunochemical Protocols*, Humana Press, Totowa N.J.).

[0103] Immunoassay procedures may be used to quantify expression of the protein in cell cultures, in subjects with a particular disorder or in model animal systems under various conditions. Increased or decreased production of proteins as monitored by immunoassay may contribute to knowledge of the cellular activities associated with developmental pathways, engineered conditions or diseases, or treatment efficacy. The quantity of a given protein in a given tissue may be determined by performing immunoassays on freeze-thawed detergent extracts of biological samples and comparing the slope of the binding curves to binding curves generated by purified protein.

[0104] Antibody Arrays

[0105] In an alternative to yeast two hybrid system analysis of proteins, an antibody array can be used to study protein-protein interactions and phosphorylation. A variety of protein ligands are immobilized on a membrane using methods well known in the art. The array is incubated in the presence of cell lysate until protein:antibody complexes are formed. Proteins of interest are identified by exposing the membrane to an antibody specific to the protein of interest. In the alternative, a protein of interest is labeled with digoxigenin (DIG) and exposed to the membrane; then the membrane is exposed to anti-DIG antibody which reveals where the protein of interest forms a complex. The identity of the proteins with which the protein of interest interacts is determined by the position of the protein of interest on the membrane.

[0106] Antibody arrays can also be used for high-throughput screening of recombinant antibodies. Bacteria containing antibody genes are robotically-picked and gridded at high density (up to 18,342 different double-spotted clones) on a filter. Up to 15 antigens at a time are used to screen for clones to identify those that express binding antibody fragments. These antibody arrays can also be used to identify proteins which are differentially expressed in samples (de Wildt et al. (2000) *Nature Biotechnol* 18:989-94).

[0107] Labeling of Molecules for Assay

[0108] A wide variety of reporter molecules and conjugation techniques are known by those skilled in the art and may be used in various cDNA, polynucleotide, protein, peptide or antibody assays. Synthesis of labeled molecules may be achieved using commercial kits for incorporation of a labeled nucleotide such as ³²P-dCTP, Cy3-dCTP or Cy5-dCTP or amino acid such as ³⁵S-methionine. Polynucleotides, cDNAs, proteins, or antibodies may be directly labeled with a reporter molecule by chemical conjugation to amines, thiols and other groups present in the molecules using reagents such as BIODIPY or FITC (Molecular Probes, Eugene Oreg.).

[0109] The proteins and antibodies may be labeled for purposes of assay by joining them, either covalently or noncovalently, with a reporter molecule that provides for a detectable signal. A wide variety of labels and conjugation techniques are known and have been reported in the scientific and patent literature including, but not limited to U.S. Pat. No. 3,817,837; U.S. Pat. No. 3,850,752; U.S. Pat. No. 3,939,350; U.S. Pat. No. 3,996,345; U.S. Pat. No. 4,277,437; U.S. Pat. No. 4,275,149; and U.S. Pat. No. 4,366,241.

[0110] Diagnostics

[0111] The cDNAs, or fragments thereof, may be used to detect and quantify differential gene expression; absence, presence, or excess expression of mRNAs; or to monitor mRNA levels during therapeutic intervention. Disorders associated with atherosclerosis include angina pectoris, coronary artery disease, myocardial infarction, hypertension, transient cerebral ischemia, mesenteric ischemia, peripheral vascular disease, renal artery stenosis, and stroke. These cDNAs can also be utilized as markers of treatment efficacy against the disorders noted above and other disorders, conditions, and diseases over a period ranging from several days to months. The diagnostic assay may use hybridization or amplification technology to compare gene expression in a biological sample from a patient to standard samples in order to detect altered gene expression. Qualitative or quantitative methods for this comparison are well known in the art.

[0112] For example, the cDNA may be labeled by standard methods and added to a biological sample from a patient under conditions for hybridization complex formation. After an incubation period, the sample is washed and the amount of label (or signal) associated with hybridization complexes is quantified and compared with a standard value. If the amount of label in the patient sample is significantly altered in comparison to the standard value, then the presence of the associated condition, disease or disorder is indicated.

[0113] In order to provide a basis for the diagnosis of a condition, disease or disorder associated with gene expression, a normal or standard expression profile is established. This may be accomplished by combining a biological sample taken from normal subjects, either animal or human, with a probe under conditions for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained using normal subjects with values from an experiment in which a known amount of a purified target sequence is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a particular

condition, disease, or disorder. Deviation from standard values toward those associated with a particular condition is used to diagnose that condition.

[0114] Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies and in clinical trial or to monitor the treatment of an individual patient. Once the presence of a condition is established and a treatment protocol is initiated, diagnostic assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in a normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

[0115] Gene Expression Profiles

[0116] A gene expression profile comprises a plurality of cDNAs and a plurality of detectable hybridization complexes, wherein each complex is formed by hybridization of one or more probes to one or more complementary nucleic acids in a sample. The cDNAs of the invention are used as elements on an array to analyze gene expression profiles. In one embodiment, the array is used to monitor the progression of disease. Researchers or clinicians can catalog the differences in gene expression between healthy and diseased tissues or cells. By analyzing changes in patterns of gene expression, disease can be diagnosed at earlier stages before the patient is symptomatic. The invention can be used to formulate a prognosis and to design a treatment regimen. The invention can also be used to monitor the efficacy of treatment. For treatments with known side effects, the array is employed to improve the treatment regimen. A dosage is established that causes a change in genetic expression patterns indicative of successful treatment. Expression patterns associated with the onset of undesirable side effects are avoided. This approach may be more sensitive and rapid than waiting for the patient to show inadequate improvement, or to manifest side effects, before altering the course of treatment.

[0117] Experimentally, expression profiles can also be evaluated by methods including, but not limited to, differential display by spatial immobilization or by gel electrophoresis, labeling with radionuclide and quantification using a scintillation counter, genome mismatch scanning, representational difference analysis, transcript imaging, quantitative PCR, and by protein or antibody arrays. Expression profiles produced by these methods may be contrasted with expression profiles produced using normal or diseased tissues. Of note is the correspondence between mRNA and protein expression has been discussed by Zweiger (2001, *Transducing the Genome*. McGraw-Hill, San Francisco, Calif.) and Glavas et al. (2001; T cell activation upregulates cyclic nucleotide phosphodiesterases 8A1 and 7A3, *Proc Natl Acad Sci* 98:6319-6342) among others.

[0118] In another embodiment, animal models which mimic a human disease can be used to produce expression profiles associated with a particular condition, disorder or disease; or treatment of the condition, disorder or disease. Novel treatment regimens may be tested in these animal models using arrays to establish and then follow expression profiles over time. In addition, arrays may be used with cell cultures or tissues removed from animal models to rapidly screen large numbers of candidate drug molecules, looking

for ones that produce an expression profile similar to those of known therapeutic drugs, with the expectation that molecules with the same expression profile will likely have similar therapeutic effects. Thus, the invention provides the means to rapidly determine the molecular mode of action of a drug.

[0119] Assays Using Antibodies

[0120] Antibodies directed against antigenic determinants of a protein encoded by a cDNA of the invention may be used in assays to quantify the amount of protein found in a particular human cell. Such assays include methods utilizing the antibody and a label to detect expression level under normal or disease conditions. The antibodies may be used with or without modification, and labeled by joining them, either covalently or noncovalently, with a labeling moiety.

[0121] Protocols for detecting and measuring protein expression using either polyclonal or monoclonal antibodies are well known in the art. Examples include, but are not limited to, western analysis, ELISA, RIA, FACS, and arrays. Such immunoassays typically involve the formation of complexes between the protein and its specific antibody and the measurement of such complexes. These assays are specifically described in Pound (supra).

[0122] Therapeutics

[0123] The cDNAs and fragments thereof can be used in gene therapy. cDNAs can be delivered ex vivo to target cells, such as cells of bone marrow. Once stable integration and transcription and or translation are confirmed, the bone marrow may be reintroduced into the subject. Expression of the protein encoded by the cDNA may correct a disorder associated with mutation of a normal sequence, reduction or loss of an endogenous target protein, or overexpression of an endogenous or mutant protein. Alternatively, cDNAs may be delivered in vivo using vectors such as retrovirus, adenovirus, adeno-associated virus, herpes simplex virus, and bacterial plasmids. Non-viral methods of gene delivery include cationic liposomes, polylysine conjugates, artificial viral envelopes, and direct injection of DNA (Anderson (1998) *Nature* 392:25-30; Dachs et al. (1997) *Oncol Res* 9:313-325; Chu et al. (1998) *J Mol Med* 76(3-4):184-192; Weiss et al. (1999) *Cell Mol Life Sci* 55(3):334-358; Agrawal (1996) *Antisense Therapeutics*, Humana Press, Totowa N.J.; and August et al. (1997) *Gene Therapy (Advances in Pharmacology*, Vol. 40), Academic Press, San Diego Calif.).

[0124] In addition, expression of a particular protein can be regulated through the specific binding of a fragment of a cDNA to a genomic sequence or an mRNA which encodes the protein or directs its transcription or translation. The cDNA can be modified or derivatized to any RNA-like or DNA-like material including peptide nucleic acids, branched nucleic acids, and the like. These sequences can be produced biologically by transforming an appropriate host cell with a vector containing the sequence of interest.

[0125] Molecules which regulate the activity of the cDNA or encoded protein are useful as therapeutics for diabetes mellitus, obesity, hypertension, atherosclerosis, polycystic ovarian syndrome, and cancers including breast, prostate, and colon. Such molecules include agonists which increase the expression or activity of the polynucleotide or encoded protein, respectively; or antagonists which decrease expression or activity of the polynucleotide or encoded protein,

respectively. In one aspect, an antibody which specifically binds the protein may be used directly as an antagonist or indirectly as a delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express the protein.

[0126] Additionally, any of the proteins, or their ligands, or complementary nucleic acid sequences may be administered as pharmaceutical compositions or in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to affect the treatment or prevention of the conditions and disorders associated with an immune response. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects. Further, the therapeutic agents may be combined with pharmaceutically-acceptable carriers including excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration used by doctors and pharmacists may be found in the latest edition of Remington's *Pharmaceutical Sciences* (Mack Publishing, Easton Pa.).

[0127] Model Systems

[0128] Animal models may be used as bioassays where they exhibit a phenotypic response similar to that of humans and where exposure conditions are relevant to human exposures. Mammals are the most common models, and most infectious agent, cancer, drug, and toxicity studies are performed on rodents such as rats or mice because of low cost, availability, lifespan, reproductive potential, and abundant reference literature. Inbred and outbred rodent strains provide a convenient model for investigation of the physiological consequences of underexpression or overexpression of genes of interest and for the development of methods for diagnosis and treatment of diseases. A mammal inbred to overexpress a particular gene (for example, secreted in milk) may also serve as a convenient source of the protein expressed by that gene.

[0129] Transgenic Animal Models

[0130] Transgenic rodents that overexpress or underexpress a gene of interest may be inbred and used to model human diseases or to test therapeutic or toxic agents. (See, e.g., U.S. Pat. No. 5,175,383 and U.S. Pat. No. 5,767,337.) In some cases, the introduced gene may be activated at a specific time in a specific tissue type during fetal or postnatal development. Expression of the transgene is monitored by analysis of phenotype, of tissue-specific mRNA expression, or of serum and tissue protein levels in transgenic animals before, during, and after challenge with experimental drug therapies.

[0131] Embryonic Stem Cells

[0132] Embryonic (ES) stem cells isolated from rodent embryos retain the potential to form embryonic tissues. When ES cells such as the mouse 129/SvJ cell line are placed in a blastocyst from the C57BL/6 mouse strain, they resume normal development and contribute to tissues of the live-born animal. ES cells are preferred for use in the creation of experimental knockout and knockin animals. The method for this process is well known in the art and the steps

are: the cDNA is introduced into a vector, the vector is transformed into ES cells, transformed cells are identified and microinjected into mouse cell blastocysts, blastocysts are surgically transferred to pseudopregnant dams. The resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains.

[0133] Knockout Analysis

[0134] In gene knockout analysis, a region of a gene is enzymatically modified to include a non-natural intervening sequence such as the neomycin phosphotransferase gene (*neo*; Capecchi (1989) *Science* 244:1288-1292). The modified gene is transformed into cultured ES cells and integrates into the endogenous genome by homologous recombination. The inserted sequence disrupts transcription and translation of the endogenous gene.

[0135] Knockin Analysis

[0136] ES cells can be used to create knockin humanized animals or transgenic animal models of human diseases. With knockin technology, a region of a human gene is injected into animal ES cells, and the human sequence integrates into the animal cell genome. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on the progression and treatment of the analogous human condition.

[0137] As described herein, the uses of the cDNAs, provided in the Sequence Listing of this application, and their encoded proteins are exemplary of known techniques and are not intended to reflect any limitation on their use in any technique that would be known to the person of average skill in the art. Furthermore, the cDNAs provided in this application may be used in molecular biology techniques that have not yet been developed, provided the new techniques rely on properties of nucleotide sequences that are currently known to the person of ordinary skill in the art, e.g., the triplet genetic code, specific base pair interactions, and the like. Likewise, reference to a method may include combining more than one method for obtaining or assembling full length cDNA sequences that will be known to those skilled in the art. It is also to be understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provided to illustrate the subject invention and are not included for the purpose of limiting the invention.

EXAMPLES

[0138] It is to be understood that this invention is not limited to the particular devices, machines, materials and methods described. Although particular embodiments are described, equivalent embodiments may be used to practice the invention. The described embodiments are provided to illustrate the invention and are not intended to limit the scope of the invention which is limited only by the appended claims.

[0139] I cDNA Library Construction

[0140] The cDNA library SMCCNOS01 was selected as an example to demonstrate the construction of cDNA libraries

from which the cDNAs co-expressed with known atherosclerosis-associated genes were derived. The SMC-CNOS01 subtracted coronary artery smooth muscle cell library was constructed using 7.56×10^6 clones from the SMCCNOT02 library and was subjected to two rounds of subtraction hybridization for 48 hours with 6.12×10^6 clones from SMCCNOT01.

[0141] The SMCCNOT02 library was constructed using RNA isolated from coronary artery smooth muscle cells removed from a 3-year-old Caucasian male. The cells were treated for 20 hours with TNF α and IL-1 β at 10 ng/ml each. The SMCCNOT01 was constructed using RNA isolated from untreated coronary artery smooth muscle cells from the same donor. Subtractive hybridization conditions were based on the methodologies of Swaroop et al. (1991; *Nucleic Acids Res* 19:1954) and Bonaldo et al. (1996; *Genome Research* 6:791).

[0142] For both cDNA libraries, SMCCNOT01 and SMC-CNOT02, the frozen coronary artery smooth muscle cells (50-100 mg) were homogenized in GTC buffer (4.0M guanidine thiocyanate, 0.1M Tris-HCl pH 7.5, 1% 2-mercaptoethanol). Two volumes of binding buffer (0.4M LiCl, 0.1M Tris-HCl pH 7.5, 0.02M EDTA) were added, and the resulting mixture was vortexed at 13,000 rpm. The supernatant was removed and combined with oligo d(T)₂₅ bound streptavidin particles (MPG). After rotation at room temperature, the mRNA-oligo d(T)₂₅ bound streptavidin particles were separated from the supernatant, washed twice with hybridization buffer 1 (0.15M NaCl, 0.01M Tris-HCl pH 8.0, 1 mM EDTA, 0.1% lauryl sarcosinate) using magnetic separation at each step to remove the supernatant from the particles. Bound mRNA was eluted from the particles with release solution and heated to 65° C. The supernatant containing eluted mRNA was magnetically separated from the particles and used to construct the cDNA libraries.

[0143] The RNA was used according to the recommended protocols in the SUPERScript plasmid system (Invitrogen). The cDNAs were fractionated on a SEPHAROSE CLIB column (APB), and those cDNAs exceeding 400 bp were ligated into pINCY plasmid (Incyte Genomics, Palo Alto Calif.). Recombinant plasmids were transformed into DH5 α competent cells or ELECTROMAX cells (Invitrogen).

[0144] II Isolation and Sequencing of cDNA Clones

[0145] Plasmid DNA was released from the cells and purified using the REAL Prep 96 plasmid kit (Qiagen, Valencia Calif.). The recommended protocol was employed except for the following changes: 1) the bacteria were inoculated into 1 ml of sterile TERRIFIC BROTH (BD Biosciences, San Jose Calif.) with carbenicillin at 25 mg/l and glycerol at 0.4%; 2) the cells were cultured for 19 hours and then lysed with 0.3 ml of lysis buffer; 3) following isopropanol precipitation, the plasmid DNA pellet was resuspended in 0.1 ml distilled water, and 4) the samples were transferred to a 96-well block for storage at 4° C.

[0146] The cDNAs were prepared using a MICROLAB 2200 system (Hamilton, Reno Nev.) in combination with the DNA ENGINE thermal cycler (MJ Research, Watertown Mass.). cDNAs were sequenced by the method of Sanger et al. (1975, *J Mol Biol* 94:441-446) using PRISM 377 (ABI) or MEGABACE 1000 sequencing systems (APB).

[0147] Most of the sequences were sequenced using standard protocols and kits (ABI) at solution volumes of 0.25x-1.0x concentrations. In the alternative, some of the sequences disclosed herein were sequenced using solutions and dyes from APB.

[0148] III Selection, Assembly, and Characterization of Sequences

[0149] The sequences used for co-expression analysis were assembled from EST sequences, 5' and 3' longread sequences, and full length coding sequences. The cDNAs claimed herein were expressed in at least three libraries.

[0150] The assembly process is described as follows. EST sequence chromatograms were processed and verified. Quality scores were obtained using PHRED (Ewing et al. (1998) Genome Res 8:175-185; Ewing and Green (1998) Genome Res 8:186-194), and edited sequences were loaded into a relational database management system (RDBMS). The sequences were clustered using BLAST with a product score of 50. All clusters of two or more sequences created a bin which represents one transcribed gene.

[0151] Assembly of the component sequences within each bin was performed using a modification of Phrap, a publicly available program for assembling DNA fragments (Green, P. University of Washington, Seattle Wash.). Bins that showed 82% identity from a local pair-wise alignment between any of the consensus sequences were merged.

[0152] Bins were annotated by screening the consensus sequence in each bin against public databases, such as GBpri

and GenPept from NCBI. The annotation process involved a FASTn screen against the GBpri database in GenBank. Those hits with a percent identity of greater than or equal to 75% and an alignment length of greater than or equal to 100 base pairs were recorded as homolog hits. The residual unannotated sequences were screened by FASTx against GenPept. Those alignments with an E value of less than or equal to 10^{-8} were recorded as homologs.

[0153] Sequences were then reclustered using BLASTn and Cross-Match, a program for rapid amino acid and nucleic acid sequence comparison and database search (Green, supra), sequentially. Any BLAST alignment between a sequence and a consensus sequence with a score greater than 150 was realigned using cross-match. The sequence was added to the bin whose consensus sequence gave the highest Smith-Waterman score (Smith et al. (1992) Protein Engineering 5:35-51) amongst local alignments with at least 82% identity. Non-matching sequences were moved into new bins, and assembly processes were repeated.

[0154] IV Coexpression Analyses of Atherosclerosis-Associated Genes

[0155] Known atherosclerosis-associated genes were selected to identify the cDNAs that are closely associated with atherosclerosis. The known atherosclerosis-associated genes which were used in this analysis all occur within the LIFESEQ Gold database (Incyte Genomics), and brief descriptions of their functions as they have been reported in the literature are listed in Table 3.

TABLE 3

<u>Descriptions of Known Atherosclerosis-Associated Genes</u>	
GENE	DESCRIPTION AND REFERENCES
Human 22kDa smooth muscle protein (SM22) calponin (CNN1)	Smooth muscle cell-specific gene which is down-regulated during smooth muscle cell dedifferentiation as part of atherogenic process (Sobue et al. (1998) Horm Res 50(S2):15-24; Sobue et al. (1999) Mol Cell Biochem 190:105-18) Calponin is smooth muscle-specific and may mediate smooth muscle contractility through binding of the amino-terminal end of the myosin regulatory light chain. Involved in phenotypic modulation of smooth muscle cells, a feature of atherosclerosis (Szymanski et al.(1999) Biochemistry 38:3778-84)
pro alpha 1(I) collagen (COL1A1)	Member of family of fibrous structural proteins. Most abundant structural component of the extracellular matrix. Secreted as procollagen and converted to collagen by matrix metalloproteinases. Collagens are important in atherosclerosis for promoting platelet aggregation and for providing sites for platelet adhesion to the vessel wall (Wen et al. (1999) Arterioscler Thromb Vasc Biol 19:519-24)
collagen alpha-2 type I (COL1A2)	see COL1A1 above
COL6A1	see COL1A1 above
procollagen alpha 2(V) (COL5A2)	see COL1A1 above
collagen VI alpha-2 (COL6A2)	see COL1A1 above
type VI collagen alpha3 (COL6A3)	see COL1A1 above
pro-alpha-1 type 3 collagen (COL3A1)	see COL1A1 above
pro-alpha-1 (V) collagen (COL3A1)	see COL1A1 above
matrix Gla protein (MGP)	Role in active calcification of vascular smooth muscle cells, suggested by expression study on VSMC in vitro differentiation study. Calcifying phenotype associated with high MGP levels. MGP knockout mice develop to term, but die up to 2 months after birth due to extensive calcification of the arteries, causing blood vessel rupture (Luo et al. (1997) Nature 386:78-81; Mori et al. (1998) FEES Lett 433:19-22)

TABLE 3-continued

Descriptions of Known Atherosclerosis-Associated Genes	
GENE	DESCRIPTION AND REFERENCES
cathepsin K (CTSK)	Nonmetalloenzyme, potent elastase present in advanced atherosclerotic plaques. Contributes to the breakdown of components of vascular extracellular matrix, reducing tensile strength, increasing plaque vulnerability (Sukhova et al.(1998) <i>J Clin Invest</i> 102:576-83)
fibrinogen beta chain gene (FGB)	Component of fibrin in the extracellular matrix. Fibrin deposition is an integral part of advanced atherosclerotic lesion development. Variation at the beta fibrinogen locus associated with peripheral atherosclerosis (Sueishi et al. (1998) <i>Semin Thromb Hemost</i> 24:255-260; Fowkes et al.(1992) <i>Lancet</i> 339:693-696)
pre-pro-von Willebrand factor (VWF)	Blood glycoprotein involved in normal hemostasis. Mediates adhesion of platelets to sites of vascular damage. Also acts as a cofactor in factor VIII activity in blood coagulation. Increased levels of VWF are found in atherosclerosis and in several of its major risk factors, including hypercholesterolemia, diabetes, obesity, hypertension. Levels serve as a predictor of adverse clinical outcome following vascular surgery, possibly as an indicator of thrombus formation (Sadler (1998) <i>Annu Rev Biochem</i> 67:395-424 Blann et al. (1994) <i>Eur J Vase Surg</i> 8:10-15; Kessler et al. (1998) <i>Diabetes Metab</i> 24:327-36; Folsom et al. (1997) <i>Circulation</i> 96:1102-1108)
platelet endothelial cell adhesion molecule (PECAM-1)	Signaling molecule in the migration of cells as part of the pathophysiology of vascular occlusive diseases such as atherosclerosis. Analysis of endothelial/monocyte co-cultures indicates oxidative stress induces transendothelial migration of monocytes as a result of phosphorylation of PECAM-1 (Rattan et al. (1997) <i>Am J Physiol</i> 273:E453-61)
antithrombin III variant (AT3)	ATIII is the sole blood component through which heparin exerts its anti-coagulation effect. Deficiency in ATIII causes recurrent venous thrombosis and pulmonary embolism and can be inherited in autosomal dominant fashion (Hultin et al. (1988) <i>Thromb Haemost</i> 59:468-73; Lane et al. (1996) <i>Blood Rev</i> 10:59-74)
lipoprotein lipase (LPL)	Hydrolyses triglyceride in chylomicrons and therefore regulates metabolism of circulating lipoproteins. Appears to have an atherogenic effect on the arterial wall due to its ability to alter the properties of LDL. Increased activity of LPL is found in atherosclerotic arteries when compared to normal. Expressed by macrophages in atherosclerotic lesions. Mutations in LPL responsible for familial hypercholesterolemia and premature atherosclerosis (Fisher et al. (1997) <i>Atherosclerosis</i> 135:145-159; Goldberg (1996) <i>J Lipid Res</i> 37:693-707; Gerdes et al. (1997) <i>Circulation</i> 96:733-740)
alpha-2-macroglobulin (A2M)	Foam cell formation—retains LDL cholesterol in the lipid core of atherosclerotic plaque (Llorente et al. (1998) <i>Rev Esp Cardiol</i> 51:633-641)
apolipoprotein AI (APOA1)	Participates in reverse cholesterol transport from tissues to the liver. Promotes cholesterol efflux from tissues and acts as a cofactor for lecithin cholesterol acyltransferase (LCAT). Mutations in ApoA1 and of ApoAI/CIII/AIV gene cluster assoc with atherosclerosis. Transgenic mice expressing high plasma APOA1 levels are protected from fatty streak development with a high atherogenic diet (Gordon et al. (1989) <i>Circulation</i> 79:8-15; Rubin et al. (1991) <i>Nature</i> 353:265-7; Karathanasis et al. (1987) <i>Proc Natl Acad Sci</i> 84:7198-7202)
apolipoprotein AII (APOA2)	Major component of HDL. Appears to have an opposite effect to that of APOAI, though exact function unknown. APOAII may have ability to convert HDL from an anti- to a pro-inflammatory particle, with paraoxonase having a role in this transformation process. Plasma APOAII levels significantly associated with plasma free fatty acid levels. Transgenic mice expressing varying levels of APOAII show increased atherosclerotic lesions than wt when fed an atherogenic diet. Possible interaction between diet/genotype and atherogenic potential (Escola-Gil et al. (1998) <i>J Lipid Res</i> 39:457-462; Warden et al. (1993) <i>Proc Natl Acad Sci</i> 90:10886-10890)
apolipoprotein B-100 (APOB)	Main apolipoprotein of chylomicrons and low density lipoproteins. Mutations in APOB100 underly familial defective apolipoprotein B-100 in which patients suffer from premature atherosclerosis. Mutations result in defect in binding of LDL to LDL receptor, and accumulation of plasma LDL. High-expressing APOB transgenic mice exhibit elevated VLDL-LDL cholesterol and atherogenic lesions (Callow et al. (1995) <i>J Clin Invest</i> 96:1639-1646; Brasaemle et al.(1997) <i>J Biol Chem</i> 272:9378-9387)
lipoprotein apoCII (APOC2)	Role in lipoprotein metabolism. Cofactor in the activity of lipoprotein lipase the enzyme that hydrolyzes triglycerides in plasma and transfers the fatty acids to tissues. Mutations in APOC2 responsible for hyperlipoproteinemia 1B, similar to lipoprotein lipase deficiency (Cox et al. (1978) <i>N Engl J Med</i> 299:1421-1424; Arimoto et al. (1998) <i>J Lipid Res</i> 39:143-151)
pre-apolipoprotein CIII (APOC3)	Inhibits lipoprotein lipase and hepatic lipase, decreases uptake of lymph chylomicrons by hepatic cells. APOA3 possibly delays breakdown of triglyceride rich particles. SstI RFLP in apoCIII is associated with plasma triglyceride and apoCIII levels and hyperlipidemic phenotypes (Henderson et al.(1987) <i>Hum Genet</i> 75:62-65)

TABLE 3-continued

Descriptions of Known Atherosclerosis-Associated Genes	
GENE	DESCRIPTION AND REFERENCES
apolipoprotein apoC-IV (APOC4)	APOC4 is a lipid-binding protein that has the potential to alter lipid metabolism. Human APOC4 transgenic mice are hypertriglyceridaemic compared to normal controls (Allan et al. (1996) <i>J Lipid Res</i> 37:1510-1518)
macrophage scavenger receptor type I (MSR1) Human antigen CD36 gene (CD36)	Mediates binding, internalisation and processing of negatively-charged macromolecules. Implicated in the pathological deposition of cholesterol in arterial walls during atherogenesis (Han et al. (1998) <i>Hum Mol Genet</i> 7:1039-1046) Acts as a scavenger receptor for oxidised LDL. Transient regulation under control of M-CSF during monocyte-macrophage differentiation increases foam cell accumulation, Possible role in atherogenesis: increased M-CSF levels detected in atherosclerotic lesions in rabbits and humans (Huh et al. (1996) <i>Blood</i> 87:2020-2028; Aitman et al. (1999) <i>Nat Genet</i> 21:76-83)
serum amyloid P component (SAP)	Plasma glycoprotein expressed in atherosclerotic lesions. Interacts with lipoproteins in specific manner (Li et al. (1995) <i>Arterioscler Thromb Vasc Biol</i> 15:252-257; Li et al. (1998) <i>Biochem Biophys Res Commun</i> 244:249-252)
carboxyl ester lipase gene (CEL)	CEL gene expression increases in presence of oxidised and native LDL in vitro. It is expressed in the vessel wall and in aortic extracts - may interact with cholesterol to modulate progression of atherosclerosis (Li et al. (1998) <i>Biochem J</i> 329:675-679)
paraoxonase 1 (PON1)	Serum esterase exclusively associated with high-density lipoproteins; it might confer protection against coronary artery disease by destroying pro-inflammatory oxidized lipids in oxidized low-density lipoproteins. PON1 gln192-to-arg polymorphism associated with coronary artery disease. Association between PON1 genetic variation and plasma LDL, HDL and non-HDL and apoB levels in genetically isolated Alberta Hutterite population. When fed on a high-fat, high-cholesterol diet, PON1-null mice were more susceptible to atherosclerosis than wild-type (Serrato et al. (1995) <i>J Clin Invest</i> 96:3005-3008; Boright et al. (1998) <i>Atherosclerosis</i> 139:131-136; Shih et al. (1998) <i>Nature</i> 394:284-287)
paraoxonase 2 (PON2)	Serum esterase exclusively associated with high-density lipoproteins; it might confer protection against coronary artery disease by destroying pro-inflammatory oxidized lipids in oxidized low-density lipoproteins. Common polymorphism at codon 311 (cys-ser) in PON2 associated with CHD alone and synergistically with the 192 polymorphism in PON1 in Asian Indians. Association between genetic variation in PON2 and plasma cholesterol and apolipoprotein A1 in genetically isolated Alberta Hutterite population (Sanghera et al. (1998) <i>Am J Hum Genet</i> 62:36-44; Boright supra)
paraoxonase 3 (PON3)	Serum esterase exclusively associated with high-density lipoproteins; it might confer protection against coronary artery disease by destroying pro-inflammatory oxidized lipids in oxidized low-density lipoproteins. Other members PON2, 3 associated with CHD and cholesterol levels (Laplaud et al. (1998) <i>Clin Chem Lab Med</i> 36:431-441)
perilipin (PLIN)	Lipid storage droplets of steroidogenic cells are surrounded by perilipins, family of phosphorylated proteins encoded by a single gene, detected in adipocytes and steroidogenic cells. Possible role in lipid metabolism (Brasaemle et al. (1997) <i>J Biol Chem</i> 272:9378-9387)
Prostaglandin D2 synthase (PTGDS)	Catalyses conversion of PGH2 to PGD2, a prostaglandin important in smooth muscle contraction/relaxation and potent inhibitor of platelet aggregation. Northern analysis shows strong specific expression in heart. Immunocytochemical localization to myocardial and atrio endocardial cells, and accumulates in end-stage atherosclerotic plaques. High plasma levels detected in severe angina patients (Eguchi et al. (1997) <i>Proc Natl Acad Sci</i> 94:14689-14694)
Annexin II/lipocortinII(ANX2)	Inhibits phospholipase A2 activity and the production of arachidonic acid, the precursor of the inflammatory mediators prostaglandins and leukotrienes. ANX2 is an important anti-inflammatory molecule that binds plasminogen and t-PA and is suspected of having a role in atherogenesis. Binding of plasminogen to ANX2 is specifically inhibited by the excess atherogenic Lp(a) (Hajjar et al. (1998) <i>J Invest Med</i> 46:364-369)
Annexin I/lipocortin(ANX1)	Inhibits phospholipase A2 activity and production of arachidonic acid, the precursor of the inflammatory mediators prostaglandins and leukotrienes. ANX1 is an important anti-inflammatory molecule (Wallner et al. (1986) <i>Nature</i> 320:77-81)
Secreted protein, acidic and rich in cysteine (SPARC)	Extracellular glycoprotein secreted by endothelial cells which has a suspected role in calcification of atherosclerotic plaques. Interacts with PDGF-B containing dimers and inhibits binding to its receptors. Expression of SPARC and PDGF is minimal in most adult tissues, but is enhanced following injury and advanced atherosclerotic lesions. Selective expression of SPARC causes rounding of adherent endothelial cells and influences extravasation of macromolecules (Raines et al. (1992) <i>Proc Natl Acad Sci</i> 89:1281-1285; Goldblum et al. (1994) <i>Proc Natl Acad Sci</i> 91:3448-3452)

[0156] From a total of 45,233 assembled gene sequences, 25 cDNAs were identified (SEQ ID NOs:1-25 of the Sequence Listing) that show strong association with the known atherosclerosis-associated genes.

[0157] Initially, the degree of association was measured by probability values using a cutoff p-value less than 0.00001. The sequences were further examined to ensure that the genes that passed the probability test had strong association with known atherosclerosis-associated genes. Details of the co-expression patterns for the known genes and co-expressed cDNAs are presented in Table 4. The entries in Table 4 are the negative log of the p-value ($-\log p$) for the coexpression of the two genes. Table 5 summarizes the highly significant co-expression relationships between each cDNA, two marker genes and their functions.

[0158] V Atherosclerosis-Associated cDNAs

[0159] Using the co-expression analysis method, cDNAs comprising the polynucleotides of SEQ ID NOs:1-25 and their complements, were identified by their highly significant co-expression with known atherosclerosis-associated genes.

[0160] BLAST and other motif searches were performed for SEQ ID NOs:1-25 according to Example VII. SEQ ID NO:8 was determined to be full length and translated as shown in FIG. 1.

[0161] VI Transcript Imaging

[0162] Transcript images were performed for several of the cDNAs of the invention using the LIFESEQ GOLD database (July 02 release, Incyte Genomics). This process allowed assessment of the relative abundance of the expressed polynucleotides in all of the cDNA libraries, but those in the cardiovascular category are specifically emphasized. Criteria for transcript imaging can be selected from category, number of cDNAs per library, library description, disease indication, clinical relevance of sample, and the like.

[0163] All sequences and cDNA libraries in the LIFESEQ database have been categorized by system, organ/tissue and cell type. For each category, the number of libraries in which the sequence was expressed were counted and shown over the total number of libraries in that category. For each library, the number of cDNAs were counted and shown over the total number of cDNAs in that library. In some transcript images, all normalized or subtracted libraries, which have high copy number sequences removed prior to processing, and all mixed or pooled tissues, which are considered non-specific in that they contain more than one tissue type or more than one subject's tissue, can be excluded from the analysis. Treated and untreated cell lines and/or fetal tissue data can also be excluded where clinical relevance is emphasized. Conversely, fetal tissue can be emphasized wherever elucidation of inherited disorders or differentiation of particular adult or embryonic stem cells into tissues or organs such as heart, kidney, nerves or pancreas would be aided by removing clinical samples from the analysis. Transcript imaging can also be used to support data from other methodologies such as guilt-by-association and hybridization analyses.

[0164] The transcript images for SEQ ID NOs:3-6, 8,13, 15-20, and 22 are shown in Table 6. The first column shows library name; the second column, the number of cDNAs

sequenced in that library; the third column, the description of the library; the fourth column, absolute abundance of the transcript in the library; and the fifth column, percentage abundance of the transcript in the library. In some cases, the normal library shows differential expression of the cDNA; in others, the induced or diseased library shows differential expression. For example, for SEQ ID NO:8, the endothelial cells treated with growth factors (VEGF and EF) show 2-8x higher expression than untreated endothelial cells in the same experiment.

[0165] These data confirm the differential expression of SEQ ID NOs:3-6, 8,13, 15-20, and 22 under conditions that correlate with disorders associated with atherosclerosis. VII Homology Searching for Atherosclerosis-Associated cDNAs and Polypeptides The polynucleotide sequences, SEQ ID NO:1-25, and polypeptide sequence, SEQ ID NO:26, were queried against databases derived from sources such as GenBank and SwissProt. These databases, which contain previously identified and annotated sequences, were searched for regions of similarity using BLAST (Altschul, supra). BLAST searched for matches and reported only those that satisfied the probability thresholds of 10⁻²⁵ or less for nucleotide sequences and 10⁻⁸ or less for polypeptide sequences.

[0166] The polypeptide sequence was also analyzed for known motif patterns using MOTIFS, SPSCAN, BLIMPS, and HMM-based protocols. MOTIFS (Genetics Computer Group, Madison Wis.) searches polypeptide sequences for patterns that match those defined in the Prosite Dictionary of Protein Sites and Patterns (Bairoch, supra) and displays the patterns found and their corresponding literature abstracts. SPSCAN (Genetics Computer Group) searches for potential signal peptide sequences using a weighted matrix method (Nielsen et al. (1997) Protein Engineering 10:1-6). Hits with a score of 5 or greater were considered. BLIMPS uses a weighted matrix analysis algorithm to search for sequence similarity between the polypeptide sequences and those contained in BLOCKS, a database consisting of short amino acid segments, or blocks of 3-60 amino acids in length, compiled from the PROSITE database (Henikoff; supra; Bairoch, supra), and those in PRINTS, a protein fingerprint database based on non-redundant sequences obtained from sources such as SwissProt, GenBank, PIR, and NRL-3D (Attwood et al. (1997) J Chem Inf Comput Sci 37:417-424). For the purposes of the present invention, the BLIMPS searches reported matches with a cutoff score of 1000 or greater and a cutoff probability value of 1.0x10⁻³. HMM-based protocols were based on a probabilistic approach and searched for consensus primary structures of gene families in the protein sequences (Eddy, supra; Sonnhammer, supra). More than 500 known protein families with cutoff scores ranging from 10 to 50 bits were selected for use in this invention.

[0167] VIII Hybridization Technologies: Selection of Sequences, Microarray Preparation and Use

[0168] SEQ ID NO:1-25 are represented among the template sequences in the LIFESEQ GOLD database (Incyte Genomics). Several of these sequences, specifically SEQ ID NOs:1, 2, 10, 12, 18, and 32-34 have been used on microarrays in experiments investigating differential gene expression in cardiovascular samples. Table 7 presents the results

of these experiments; results were significant if the log₂ Cy/Cy5 ratio exceeded ± 1.00 in either the normal or the induced or disease state.

[0169] Exemplary Experimental Materials and Protocols

[0170] Activation of the vascular endothelium is considered to be a central event in a wide range of both physiological and pathophysiological processes, such as vascular tone regulation, coagulation and thrombosis, atherosclerosis, and inflammation.

[0171] HAEC, human aortic endothelial cells, are primary cells derived from the endothelium of a human aorta. They have been used as an experimental model for investigating the role of the endothelium in human vascular biology in vitro. HAECs were grown to 85% confluency, split into two samples, one of which was then treated with growth factors, LDL, cytokines, O₂, and the like for variable time periods.

[0172] ECV304, HUAEC, and HUVEC are endothelial cell lines derived from the endothelium of the human umbilical artery or vein. This cell model has been extensively used to study the functional biology of human endothelial cells. These cells were also grown to about 85% confluency, split into samples, one of which was then treated with growth factors, LDL, cytokines, O₂, and the like for variable time periods.

[0173] The experimental treatments and time of exposure are shown by the tissue description in Table 7.

[0174] Exemplary Activators and Inducers

[0175] TNF- α is a pleiotropic cytokine that is known to play a central role in the mediation of inflammatory responses through activation of multiple signal transduction pathways. TNF- α is produced by activated lymphocytes, macrophages, and other white blood cells, and is known to activate endothelial cells. Monitoring the endothelial cells' response to TNF- α at the level of the mRNA expression can provide information necessary for better understanding of both TNF- α signaling pathways and endothelial cell biology.

[0176] PMA is a broad activator of the protein kinase C-dependent pathways. Ionomycin is a calcium ionophore that permits the entry of calcium in the cell, hence increasing the cytosolic calcium concentration. The combination of PMA and ionomycin activates two of the major signaling pathways used by mammalian cells to interact with their environment. In T cells, the combination of PMA and ionomycin mimics the type of secondary signaling events elicited during optimal B cell activation.

[0177] Microarrays

[0178] The HUMAN GENOME GEM series 1-5 microarrays (Incyte Genomics) contain 45,320 array elements which represent 22,632 annotated clusters and 22,688 unannotated clusters. For the UNIGEM series microarrays (Incyte Genomics), Incyte clones were mapped to non-redundant Unigene clusters (Unigene database (build 46), NCBI; Shuler (1997) *J Mol Med* 75:694-698), and the 5' clone with the strongest BLAST alignment (at least 90% identity and 100 bp overlap) was chosen, verified, and used in the construction of the microarray. The UNIGEM V 2.0 microarray (Incyte Genomics) contains 8,502 array elements which represent 8,372 annotated genes and 130 unannotated clusters.

[0179] To construct microarrays, cDNAs were amplified from bacterial cells using primers complementary to vector sequences flanking the cDNA insert. Thirty cycles of PCR increased the initial quantity of cDNAs from 1-2 ng to a final quantity greater than 5 μ g. Amplified cDNAs were then purified using SEPHACRYL-400 columns (APB). Purified cDNAs were immobilized on polymer-coated glass slides. Glass microscope slides (Corning, Corning N.Y.) were cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides were etched in 4% hydrofluoric acid (VWR Scientific Products, West Chester Pa.), washed thoroughly in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich) in 95% ethanol. Coated slides were cured in a 110° C. oven. cDNAs were applied to the coated glass substrate using a procedure described in U.S. Pat. No. 5,807,522. One microliter of the cDNA at an average concentration of 100 ng/ μ l was loaded into the open capillary printing element by a high-speed robotic apparatus which then deposited about 5 nl of cDNA per slide.

[0180] Microarrays were UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene), and then washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites were blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (Tropix, Bedford Mass.) for 30 minutes at 60° C. followed by washes in 0.2% SDS and distilled water as before.

[0181] Isolation and Labeling of Sample cDNAs

[0182] Cells were harvested and lysed in 1 ml of TRIZOL reagent (5×10^6 cells/ml; Invitrogen). The lysates were vortexed thoroughly and incubated at room temperature for 2-3 minutes and extracted with 0.5 ml chloroform. The extract was mixed, incubated at room temperature for 5 minutes, and centrifuged at 16,000 \times g for 15 minutes at 4° C. The aqueous layer was collected, and an equal volume of isopropanol was added. Samples were mixed, incubated at room temperature for 10 minutes, and centrifuged at 16,000 \times g for 20 minutes at 4° C. The supernatant was removed, and the RNA pellet was washed with 1 ml of 70% ethanol, centrifuged at 16,000 \times g at 4° C., and resuspended in RNase-free water. The concentration of the RNA was determined by measuring the optical density at 260 nm.

[0183] Poly(A) RNA was prepared using an OLIGOTEX mRNA kit (Qiagen) with the following modifications: OLIGOTEX beads were washed in tubes rather than spin columns, resuspended in elution buffer, and then loaded onto spin columns to recover the mRNA. To obtain maximum yield, the mRNA was eluted twice.

[0184] Each poly(A) RNA sample was reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/ μ l oligo-d(T) primer (21mer), 1 \times first strand buffer, 0.03 units/ μ l RNase inhibitor, 500 μ M dATP, 500 μ M dGTP, 500 μ M dTTP, 40 μ M dCTP, and 40 μ M either dCTP-Cy3 or dCTP-Cy5 (APB). The reverse transcription reaction was performed in a 25 ml volume containing 200 ng poly(A) RNA using the GEMBRIGHT kit (Incyte Genomics). Specific control poly(A) RNAs (YCFR06, YCFR45, YCFR67, YCFR85, YCFR43, YCFR22, YCFR23, YCFR25, YCFR44, YCFR26) were synthesized by in vitro transcription from non-coding yeast genomic DNA (W. Lei, unpublished). As quantitative controls, control mRNAs (YCFR06, YCFR45,

YCFR67, and YCFR85) at 0.002 ng, 0.02 ng, 0.2 ng, and 2 ng were diluted into reverse transcription reaction at ratios of 1:100,000, 1:10,000, 1:1,000, 1:100 (w/w) to sample mRNA, respectively. To sample differential expression patterns, control mRNAs (YCFR43, YCFR22, YCFR23, YCFR25, YCFR44, YCFR26) were diluted into reverse transcription reaction at ratios of 1:3, 3:1, 1:10, 10:1, 1:25, 25:1 (w/w) to sample mRNA. Reactions were incubated at 37° C. for 2 hr, treated with 2.5 ml of 0.5M sodium hydroxide, and incubated for 20 minutes at 85° C. to stop the reaction and degrade the RNA.

[0185] cDNAs were purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech). Cy3- and Cy5-labeled reaction samples were combined as described below and ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The cDNAs were then dried to completion using a SpeedVAC system (Savant Instruments, Holbrook N.Y.) and resuspended in 14 μ l 5 \times SSC, 0.2% SDS.

[0186] Hybridization and Detection

[0187] Hybridization reactions contained 9 μ l of sample mixture containing 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5 \times SSC, 0.2% SDS hybridization buffer. The mixture was heated to 65° C. for 5 minutes and was aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The microarrays were transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber was kept at 100% humidity internally by the addition of 140 μ l of 5 \times SSC in a corner of the chamber. The chamber containing the microarrays was incubated for about 6.5 hours at 60° C. The microarrays were washed for 10 min at 45° C. in low stringency wash buffer (1 \times SSC, 0.1% SDS), three times for 10 minutes each at 45° C. in high stringency wash buffer (0.1 \times SSC), and dried.

[0188] Reporter-labeled hybridization complexes were detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Santa Clara Calif.) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light was focused on the microarray using a 20 \times microscope objective (Nikon, Melville N.Y.). The slide containing the microarray was placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm \times 1.8 cm microarray used in the present example was scanned with a resolution of 20 micrometers.

[0189] In two separate scans, the mixed gas multiline laser excited the two fluorophores sequentially. Emitted light was split, based on wavelength, into two photomultiplier tube detectors (PMT R1477; Hamamatsu Photonics Systems, Bridgewater N.J.) corresponding to the two fluorophores. Appropriate filters positioned between the microarray and the photomultiplier tubes were used to filter the signals. The emission maxima of the fluorophores used were 565 nm for Cy3 and 650 nm for Cy5. Each microarray was typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus was capable of recording the spectra from both fluorophores simultaneously.

[0190] The sensitivity of the scans was calibrated using the signal intensity generated by a cDNA control species.

Samples of the calibrating cDNA were separately labeled with the two fluorophores and identical amounts of each were added to the hybridization mixture. A specific location on the microarray contained a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000.

[0191] The output of the photomultiplier tube was digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Norwood, Mass.) installed in an IBM-compatible PC computer. The digitized data were displayed as an image where the signal intensity was mapped using a linear 20-color transformation to a pseudo-color scale ranging from blue (low signal) to red (high signal). The data was also analyzed quantitatively. Where two different fluorophores were excited and measured simultaneously, the data were first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

[0192] A grid was superimposed over the fluorescence signal image such that the signal from each spot was centered in each element of the grid. The fluorescence signal within each element was then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis was the GEMTOOLS gene expression analysis program (Incyte Genomics). Significance was defined as signal to background ratio exceeding 2 \times and area hybridization exceeding 40%.

[0193] IX Further Characterization of Differentially Expressed cDNAs and Proteins

[0194] Clones were aligned against the LIFESEQ Gold 5.1 database (Incyte Genomics) and an Incyte template and its sequence variants were chosen for each clone. The template and variant sequences were aligned against the GenBank nucleotide sequence databases using BLASTn (vers. 2.0, NCBI) to acquire annotation. The template and variant sequences were translated into amino acid sequences which were aligned against GenPept and other protein databases using BLASTp (vers. 2.0, NCBI) to acquire annotation and characterization, i.e., structural motifs. Table 3 shows the GenBank annotations (where available) for SEQ ID NOs:1-25 of this invention as produced by BLAST analysis.

[0195] Percent sequence identity can be determined electronically for two or more amino acid or nucleic acid sequences using the MEGALIGN program, a component of LASERGENE software (DNASTAR). The percent identity between two amino acid sequences is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one hundred. Gaps of low or of no homology between the two amino acid sequences are not included in determining percentage identity.

[0196] Sequences with conserved protein motifs may be searched using the BLOCKS search program. This program analyses sequence information contained in the Swiss-Prot and PROSITE databases and is useful for determining the classification of uncharacterized proteins translated from genomic or cDNA sequences (Bairoch, supra; Attwood, supra). PROSITE database is a useful source for identifying

functional or structural domains that are not detected using motifs due to extreme sequence divergence. Using weight matrices, these domains are calibrated against the SWISS-PROT database to obtain a measure of the chance distribution of the matches.

[0197] The PRINTS database can be searched using the BLIMPS search program to obtain protein family "fingerprints". The PRINTS database complements the PROSITE database by exploiting groups of conserved motifs within sequence alignments to build characteristic signatures of different protein families. For both BLOCKS and PRINTS analyses, the cutoff scores for local similarity were:

[0198] >1300=strong, 1000-1300=suggestive; for global similarity were: $p < \exp^{-3}$; and for strength (degree of correlation) were: >1300=strong, 1000-1300=weak.

[0199] X Other Hybridization Technologies and Analyses

[0200] Other hybridization technologies utilize a variety of substrates such as nylon membranes, capillary tubes, etc. Arranging cDNAs on polymer coated slides is described in Example V; sample cDNA preparation and hybridization and analysis using polymer coated slides is described in examples VI and VII, respectively.

[0201] The cDNAs are applied to a membrane substrate by one of the following methods. A mixture of cDNAs is fractionated by gel electrophoresis and transferred to a nylon membrane by capillary transfer. Alternatively, the cDNAs are individually ligated to a vector and inserted into bacterial host cells to form a library. The cDNAs are then arranged on a substrate by one of the following methods. In the first method, bacterial cells containing individual clones are robotically picked and arranged on a nylon membrane. The membrane is placed on LB agar containing selective agent (carbenicillin, kanamycin, ampicillin, or chloramphenicol depending on the vector used) and incubated at 37° C. for 16 hr. The membrane is removed from the agar and consecutively placed colony side up in 10% SDS, denaturing solution (1.5 M NaCl, 0.5 M NaOH), neutralizing solution (1.5 M NaCl, 1 M Tris, pH 8.0), and twice in 2×SSC for 10 min each. The membrane is then UV irradiated in a STRATALINKER UV-crosslinker (Stratagene).

[0202] In the second method, cDNAs are amplified from bacterial vectors by thirty cycles of PCR using primers complementary to vector sequences flanking the insert. PCR amplification increases a starting concentration of 1-2 ng nucleic acid to a final quantity greater than 5 μ g. Amplified nucleic acids from about 400 bp to about 5000 bp in length are purified using SEPHACRYL-400 beads (APB). Purified nucleic acids are arranged on a nylon membrane manually or using a dot/slot blotting manifold and suction device and are immobilized by denaturation, neutralization, and UV irradiation as described above.

[0203] Hybridization probes derived from cDNAs of the Sequence Listing are employed for screening cDNAs, mRNAs, or genomic DNA in membrane-based hybridizations. Probes are prepared by diluting the cDNAs to a concentration of 40-50 ng in 45 μ l TE buffer, denaturing by heating to 100° C. for five min and briefly centrifuging. The denatured cDNA is then added to a REDIPRIME tube (APB), gently mixed until blue color is evenly distributed, and briefly centrifuged. Five microliters of [³²P]dCTP is added to the tube, and the contents are incubated at 37° C.

for 10 min. The labeling reaction is stopped by adding 5 μ l of 0.2M EDTA, and probe is purified from unincorporated nucleotides using a PROBEQUANT G-50 microcolumn (APB). The purified probe is heated to 100° C. for five min and then snap cooled for two min on ice.

[0204] Membranes are pre-hybridized in hybridization solution containing 1% Sarkosyl and 1x high phosphate buffer (0.5 M NaCl, 0.1 M Na₂HPO₄, 5 mM EDTA, pH 7) at 55° C. for two hr. The probe, diluted in 15 ml fresh hybridization solution, is then added to the membrane. The membrane is hybridized with the probe at 55° C. for 16 hr. Following hybridization, the membrane is washed for 15 min at 25° C. in 1mM Tris (pH 8.0), 1% Sarkosyl, and four times for 15 min each at 25° C. in 1 mM Tris (pH 8.0). To detect hybridization complexes, XOMAT-AR film (Eastman Kodak, Rochester N.Y.) is exposed to the membrane overnight at -70° C., developed, and examined.

[0205] XI Expression of the Encoded Protein

[0206] Expression and purification of a protein encoded by a cDNA of the invention is achieved using bacterial or virus-based expression systems. For expression in bacteria, cDNA is subcloned into a vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into bacterial hosts, such as BL21(DE3). Antibiotic resistant bacteria express the protein upon induction with IPTG. Expression in eukaryotic cells is achieved by infecting *Spodoptera frugiperda* (Sf9) insect cells with recombinant baculovirus, *Autographica californica* nuclear polyhedrosis virus. The polyhedrin gene of baculovirus is replaced with the cDNA by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of transcription.

[0207] For ease of purification, the protein is synthesized as a fusion protein with glutathione-S-transferase (GST; APB) or a similar alternative such as FLAG. The fusion protein is purified on immobilized glutathione under conditions that maintain protein activity and antigenicity. After purification, the GST moiety is proteolytically cleaved from the protein with thrombin. A fusion protein with FLAG, an 8-amino acid peptide, is purified using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak, Rochester N.Y.).

[0208] XII Production of Antibodies

[0209] A denatured protein from a reverse phase HPLC separation is obtained in quantities up to 75 mg. This denatured protein is used to immunize mice or rabbits following standard protocols. About 100 μ g is used to immunize a mouse, while up to 1 mg is used to immunize a rabbit. The denatured protein is radioiodinated and incubated with murine B-cell hybridomas to screen for monoclonal antibodies. About 20 mg of protein is sufficient for labeling and screening several thousand clones.

[0210] In another approach, the amino acid sequence translated from a cDNA of the invention is analyzed using PROTEAN software (DNASTAR) to determine antigenic

determinants of the protein. The optimal sequences for immunization are usually at the C-terminus, the N-terminus, and those intervening, hydrophilic regions of the protein that are likely to be exposed to the external environment when the protein is in its natural conformation. Typically, oligopeptides about 15 residues in length are synthesized using an 431 peptide synthesizer (ABI) using Fmoc-chemistry and then coupled to keyhole limpet hemocyanin (KLH; Sigma-Aldrich) by reaction with M-maleimidobenzoyl-N-hydroxysuccinimide ester. If necessary, a cysteine may be introduced at the N-terminus of the peptide to permit coupling to KLH. Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. The resulting antisera are tested for antipeptide activity by binding the peptide to plastic, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radioiodinated goat anti-rabbit IgG.

[0211] Hybridomas are prepared and screened using standard techniques. Hybridomas of interest are detected by screening with radioiodinated protein to identify those fusions producing a monoclonal antibody specific for the protein. In a typical protocol, wells of 96 well plates (FAST, Becton-Dickinson, Palo Alto Calif.) are coated with affinity-purified, specific rabbit-anti-mouse (or suitable anti-species Ig) antibodies at 10 mg/ml. The coated wells are blocked with 1% BSA and washed and exposed to supernatants from hybridomas. After incubation, the wells are exposed to radiolabeled protein at 1 mg/ml. Clones producing antibodies bind a quantity of labeled protein that is detectable above background.

[0212] Such clones are expanded and subjected to 2 cycles of cloning at 1 cell/3 wells. Cloned hybridomas are injected into pristane-treated mice to produce ascites, and monoclonal antibody is purified from the ascitic fluid by affinity chromatography on protein A (APB). Monoclonal antibodies with affinities of at least $10^8 M^{-1}$, preferably 10^9 to $10^{10} M^{-1}$ or stronger, are made by procedures well known in the art.

[0213] XIII Purification of Naturally Occurring Protein Using Antibodies

[0214] Naturally occurring or recombinant protein is purified by immunoaffinity chromatography using antibodies

specific for the protein. An immunoaffinity column is constructed by covalently coupling the antibody to CNBr-activated SEPHAROSE resin (APB). Media containing the protein is passed over the immunoaffinity column, and the column is washed using high ionic strength buffers in the presence of detergent to allow preferential absorbance of the protein. After coupling, the protein is eluted from the column using a buffer of pH 2-3 or a high concentration of urea or thiocyanate ion to disrupt antibody/protein binding, and the protein is collected.

[0215] XIV Screening for Molecules That Specifically Bind the cDNA or Protein

[0216] The cDNA or fragments thereof and the protein or portions thereof are labeled with ^{32}P -dCTP, Cy3-dCTP, Cy5-dCTP (APB), or BIODIPY or FITC (Molecular Probes), respectively. Candidate molecules or compounds previously arranged on a substrate are incubated in the presence of labeled nucleic or amino acid. After incubation under conditions for either a cDNA or a protein, the substrate is washed, and any position on the substrate retaining label, which indicates specific binding or complex formation, is assayed. The binding molecule is identified by its arrayed position on the substrate. Data obtained using different concentrations of the nucleic acid or protein are used to calculate affinity between the labeled nucleic acid or protein and the bound molecule. High throughput screening using very small assay volumes and very small amounts of test compound is fully described in U.S. Pat. No. 5,876,946.

[0217] All patents and publications mentioned in the specification are incorporated herein by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention that are obvious to those skilled in the field of molecular biology or related fields are intended to be within the scope of the following claims.

TABLE 4

Co-expression of 25 cDNAs with known atherosclerosis genes (- log p).													
SEQ ID	CNN1	COL1A1	COL1A2	COL6A1	COL5A2	COL6A2	COL6A3	COL3A1	COL5A1	MGP	CTSK	FGF	VWF
1	0	1	0	0	0	0	0	0	0	0	0	4	1
2	1	1	3	2	2	2	4	3	4	5	3	1	6
3	0	1	0	1	1	1	0	0	1	1	1	4	0
4	1	0	1	1	1	0	1	1	1	0	1	7	0
5	5	9	12	13	3	14	11	12	4	10	13	1	3
6	6	4	7	9	4	13	5	6	8	14	6	1	12
7	5	4	5	5	2	7	5	6	5	10	1	2	1
8	1	1	0	2	1	1	0	1	0	1	0	1	1
9	1	1	4	2	1	6	4	4	5	6	1	0	10
10	3	4	3	3	3	4	4	3	1	4	1	0	5
11	1	1	1	1	1	1	1	0	1	1	2	1	1
12	11	4	7	9	2	9	11	10	7	11	3	0	5
13	2	6	5	6	9	8	4	6	7	1	3	1	1
14	0	0	0	0	0	0	0	1	0	1	1	4	1
15	17	7	13	17	2	25	12	11	6	21	6	3	7
16	13	22	27	27	15	33	24	34	15	20	18	0	8
17	2	1	3	3	0	2	6	3	1	9	1	0	7

TABLE 4-continued

Co-expression of 25 cDNAs with known atherosclerosis genes (- log p).													
18	6	5	6	7	3	7	10	6	2	14	5	0	5
19	0	1	1	2	1	1	0	1	0	0	1	7	0
20	8	7	8	12	3	9	11	10	3	11	6	1	8
21	0	2	4	2	4	2	5	5	3	2	1	0	4
22	1	1	0	1	3	1	2	1	1	1	1	4	1
23	1	1	0	1	0	1	0	1	0	0	0	7	0
24	0	1	2	0	0	0	0	1	0	1	0	8	1
25	48	11	17	24	3	26	18	17	13	20	6	1	8

SEQ ID	PECAM1	AT3	LPL	A2M	APOA1	APOA2	APOB	APOC2	APOC3	APOC4	MSR1	CD36	SAP	CEL
1	0	5	0	1	3	4	2	2	5	4	0	0	6	0
2	7	0	1	4	0	1	0	0	0	0	2	3	0	1
3	0	8	1	1	5	4	3	3	5	7	0	0	8	1
4	0	4	1	2	4	5	4	4	6	2	0	1	3	0
5	2	0	4	9	1	0	0	1	0	0	4	2	0	2
6	6	1	5	7	0	1	1	0	1	0	7	3	0	2
7	4	3	3	8	1	1	2	0	2	1	2	7	2	0
8	0	0	3	3	0	0	0	2	0	0	1	1	1	0
9	8	0	5	6	1	1	1	2	0	0	5	6	0	1
10	3	0	4	4	0	0	0	0	0	0	9	5	0	0
11	1	2	1	0	3	1	2	2	1	0	0	2	3	6
12	2	1	3	6	2	0	1	0	0	0	4	3	0	3
13	0	0	1	1	3	2	1	2	1	0	1	2	1	3
14	0	5	1	1	4	5	5	5	3	1	0	0	2	1
15	5	1	1	13	1	1	1	0	1	0	10	1	1	0
16	7	0	3	12	1	0	0	1	1	0	9	5	0	1
17	5	0	6	7	0	0	0	1	0	0	5	9	0	1
18	4	0	6	10	1	0	1	0	0	0	8	5	1	1
19	1	8	1	3	5	8	4	6	8	9	1	1	11	0
20	2	0	12	10	1	1	0	1	0	0	9	14	0	0
21	4	0	8	5	0	1	0	1	1	0	5	11	1	1
22	4	9	0	2	6	8	7	6	4	3	1	1	7	0
23	0	6	0	1	6	7	7	4	8	1	0	1	7	1
24	0	7	0	2	8	10	10	7	8	4	0	0	5	1
25	2	0	2	20	1	0	3	0	0	0	10	4	0	1

SEQ ID	PON1	PON2	PON3	PLIN	PTGDS	ANX2	ANX1	SPARC	SM22
1	5	1	2	0	0	2	0	1	0
2	1	6	1	1	2	3	4	8	3
3	5	2	2	0	0	1	0	1	0
4	2	0	1	0	0	1	0	1	1
5	1	1	1	6	4	6	3	9	9
6	1	4	0	11	7	6	4	11	14
7	1	4	5	2	1	11	7	7	8
8	1	5	0	1	23	1	1	4	2
9	0	7	1	6	3	3	2	7	4
10	0	1	0	15	2	2	1	2	4
11	6	2	7	4	0	0	2	1	0
12	1	3	1	3	3	6	3	7	9
13	2	3	2	2	1	1	3	4	2
14	2	2	1	1	0	0	0	1	0
15	0	2	1	1	12	7	9	14	27
16	1	2	2	7	6	12	9	22	20
17	0	1	0	6	2	3	1	6	3
18	1	2	3	8	6	3	10	6	11
19	5	1	2	0	1	0	1	0	1
20	0	0	0	19	4	4	2	7	10
21	0	1	0	8	0	2	2	3	3
22	4	2	2	1	2	1	3	1	1
23	4	1	3	1	0	0	0	1	0
24	3	1	1	0	2	1	1	1	0
25	1	1	1	3	11	9	6	9	30

[0218]

TABLE 5

Summary of coexpressed genes and their function in atherosclerosis-associated disorders						
SEQ ID	Pvalue	Gene 1	Gene function from Table 3	Pvalue	Gene 2	Gene function from Table 3
1	6	SAP	associated with amyloid P in lesions	5	APOC3	associated with plasma triglyceride and hyperlipidemia
2	8	SPARC	calcification of plaques	7	PECAM-	implicated in migration of cells under O2 stress
3	8	AT3	deficiency causes recurrent venous thrombosis	8	SAP	associated with amyloid P in lesions
4	7	FGB	fibrin deposition in plaque	6	APOC3	apolipoprotein CIII
5	14	COL6A2	promoting platelet aggregation	12	CTSK	present in advanced plaques
6	14	SM22	downregulated during atherogenesis	14	MGP	calcification associated with high expression
7	11	ANX2	role in atherogenesis	10	MGP	calcification associated with high expression
8	23	PTDGS	accumulates in end-stage atherosclerotic plaques	15	AGT	associated with higher total cholesterol levels
9	10	VWF	Increased levels found in atherosclerosis	8	PECAM-	implicated in migration of cells under O2 stress
10	15	PLIN	functions in lipid metabolism	9	MSR1	deposits cholesterol during atherogenesis
11	7	PON3	associated with coronary heart disease and cholesterol levels	6	CEL	implicated in the progression of atherosclerosis
12	11	COL6A3	promoting platelet aggregation	11	MGP	calcification associated with high expression
13	9	COL5A2	promoting platelet aggregation	8	COL6A2	promoting platelet aggregation
14	5	APOA/B	accumulation of plasma LDL in atherogenesis	5	AT3	deficiency causes recurrent venous thrombosis
15	27	SM22	downregulated during atherogenesis	25	COL6A2	promoting platelet aggregation
16	34	COL3A1	promoting platelet aggregation	22	SPARC	calcification of plaques
17	9	MGP	calcification associated with high expression	9	CD36	receptor for oxidised LDL
18	11	SM22	downregulated during atherogenesis	10	A2M	retains LDL in core of plaque
19	11	SAP	associated with amyloid P in lesions	9	APOC4	alters lipid metabolism toward hypertriglyceridaemia
20	19	PLIN	functions in lipid metabolism	14	CD36	receptor for oxidised LDL
21	11	CD36	receptor for oxidised LDL	8	PLIN	functions in lipid metabolism
22	9	AT3	deficiency causes recurrent venous thrombosis	8	APOA2	associated with increased atherosclerotic lesions
23	8	APOC3	associated with plasma triglyceride and hyperlipidemia	7	SAP	associated with amyloid P in lesions
24	10	APOA/B	accumulation of plasma LDL in atherogenesis	8	FGB	fibrin deposition in plaque
25	48	CNN1	modulation of SMC, a feature of atherosclerosis	26	COL6A2	promoting platelet aggregation

[0219]

TABLE 6

Cardiovascular transcript images for the coexpressed cDNAs					
SEQ ID	Library	cDNAs	Description of Sample	Abundance	% Abundance
3	SMCRUNT01	3472	renal vein, smooth muscle cells, 57M, Untx	1	0.0288
	ENDVUNT01	5215	microvascular, dermal, endothelial cells, 22F, Untx	1	0.0192
	HEAANOT01	12578	heart, coronary artery, CAD, 46M	1	0.0080
4	MONOTXN05	2709	periph blood, monocytes, 42F, t/IL-10, LPS, NORM	1	0.0369
	MONOTXT02	3554	periph blood, monocytes, 42F, t/IL-10, LPS	1	0.0281
	MCLRUNT01	6149	periph blood mononuclear cells, 60M, untreated	1	0.0163
5	ARTANOT06	6311	aorta, adventitia, 48M	2	0.0317
	HEAONOT02	3482	heart, aorta, 10M	1	0.0287
	HEAONOT04	4002	heart, aorta, 12F	1	0.0250
	ARTANOT07	5716	aorta, adventitia, 65F	1	0.0175
6	HEAONOE01	3639	heart, aorta, 39M, 5RP	10	0.2748
	HEAONOT03	3720	heart, aorta, aw/cerebral agenesis, 27F	3	0.0806
	HEAPNOT01	3502	heart, coronary artery, plaque, pool	2	0.0571

TABLE 6-continued

Cardiovascular transcript images for the coexpressed cDNAs				
SEQ ID Library	cDNAs	Description of Sample	Abundance	% Abundance
8	ENDVXT01	1882 microvascular, dermal, endothelial cells, 22F, t/bFGF, EF	3	0.1594
	ENDVXT02	1876 microvascular, dermal, endothelial cells, 22F, t/VEGE, EF	1	0.0533
	ENDVUNT01	5215 microvascular, dermal, endothelial cells, 22F, Untx	1	0.0192
13	SMCCNOS01	3494 coronary artery, smooth muscle cells, 3M, t/TNF, IL-1, SUB	1	0.0286
	SMCANOT01	7327 aortic smooth muscle line, M	1	0.0136
15	SMCRXT01	3453 renal vein, smooth muscle cells, 57M t/TNF, IL1	2	0.0579
	SMCRUNT01	3472 renal vein, smooth muscle cells, 57M, Untx	1	0.0288
16	SMCCNOT01	4266 coronary artery, smooth muscle cells, 3M	5	0.1172
	SMCCNOT02	3980 coronary artery, smooth muscle cells, 3M, t/TNF, IL-1	2	0.0503
	SMCCNOS01	3494 coronary artery, smooth muscle cells, 3M, t/TNF, IL-1, SUB	1	0.0286
17	ENDIUNT01	3582 iliac artery, endothelial cells, F, control, untreated	26	0.7259
	ENDITXT01	3464 iliac artery, endothelial cells, F, t/1% oxygen 24 hr	21	0.6062
	ENDINOT02	3208 iliac artery, endothelial cells, F, t/TNF, IL-1 20 hr	10	0.3117
18	ENDVXT01	1882 microvascular, dermal, endothelial cells, 22F, t/bFGF, EF	1	0.0531
	ENDVNOT01	4955 microvascular, dermal, endothelial cells, 18F, untreated	1	0.0202
19	ENDATXP01	1877 aorta, endothelial cells, t/TNF, TIGR	1	0.0533
	ENDVXT02	1876 microvascular, dermal, endothelial cells, 22F, t/VEGF, EF	1	0.0533
	ENDVXT01	1882 microvascular, dermal, endothelial cells, 22F, t/bFGF, EF	1	0.0531
	ENDVNOT01	4955 microvascular, dermal, endothelial cells, 18F, untreated	1	0.0202
20	HEAANOT01	12578 heart, coronary artery, CAD, 46M	13	0.1034
	HEAONOT05	3959 heart, aorta, 17F	3	0.0758
	HEAONOT04	4002 heart, aorta, 12F	1	0.0250
22	ENDITXT01	3464 iliac artery, endothelial cells, F, t/1% oxygen 24 hr	1	0.0289
	HEAONOE01	3639 heart, aorta, 39M, 5RP	1	0.0275
	HEAANOT01	12578 heart, coronary artery, CAD, 46M	1	0.0080

[0220]

TABLE 7

Microarray data from cardiovascular experiments				
SEQ ID	GEM	Log2 (Cy5/Cy3)	Cy3 Sample	Cy5 Sample
1	LG1	-1.01	ECV304 Line, Untx	ECV304 Line, t/PMA + Iono
2	HG1	-1.03	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 1 hr, Nrm1
2	HG1	-1.03	HUAEC Cells, Untx, Nrm1	HUAEC Cells, t/TNFa 10 ng/mL 24 hr, Nrm1
2	HG1	-1.03	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/IL1b + TNFa 10 ng/mL, 10 ng/mL 2 4hr, 4 hr, Nrm1
2	HG1	-1.07	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/PMA + TNFa 10 nM, 10 ng/mL 24 hr, 1 hr, Nrm1
2	HG1	-1.07	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 48 hr, Nrm1
2	HG1	-1.10	HUVEC Cells, Untx	HUVEC Cells, t/IL4 10 ng/mL 24 hr
2	HG1	-1.10	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 2 hr, Nrm1
2	HG1	-1.12	HUVEC Cells, Untx, 48 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 1 hr
2	HG1	-1.12	HUVEC Cells, Untx	HUVEC Cells, t/IL10 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 4 hr
2	HG1	-1.13	HUAEC Cells, Untx, Nrm1	HUAEC Cells, t/TNFa 10 ng/mL 4 hr, Nrm1
2	HG1	-1.14	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/CHX + TNFa 10 mcg/mL, 10 ng/mL 30 min, 24 hr, Nrm1
2	HG1	-1.14	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/PMA 10 nM 24 hr, Nrm1
2	HG1	-1.15	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 24 hr, Nrm1
2	HG1	-1.17	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 24 hr, Nrm1
2	HG1	-1.18	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/8ClcAMP + TNFa 7.5 microM, 10 ng/mL 24 hr, 4 hr, Nrm1
2	HG1	-1.19	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 24 hr, Nrm1
2	HG1	-1.21	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TL1b + TNFa 10 ng/mL, 10 ng/mL 2 4hr, 24 hr, Nrm1

TABLE 7-continued

Microarray data from cardiovascular experiments				
SEQ ID	GEM	Log2 (Cy5/Cy3)	Cy3 Sample	Cy5 Sample
2	HG1	-1.22	HIAEC Cells, Untx, Nrm1	HIAEC Cells, t/TNFa 10 ng/mL 24 hr, Nrm1
2	HG1	-1.22	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/TNFa + TNFa .1 ng/ml, 10 ng/ml 24 hr, 24 hr, Nrm1
2	HG1	-1.23	HUVEC Cells, Untx	HUVEC Cells, t/TNFa 10 ng/mL 4 hr
2	HG1	-1.26	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/8ClcAMP + TNFa 7.5 microM, 10 ng/mL 24 hr, 24 hr, Nrm1
2	HG1	-1.28	HUVEC Cells, Untx	HUVEC Cells, t/TNFa .1 ng/mL 4 hr
2	HG1	-1.29	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 4 hr, Nrm1
2	HG1	-1.29	HIAEC Cells, Untx, Nrm1	HIAEC Cells, t/TNFa 10 ng/mL 4 hr, Nrm1
2	HG1	-1.30	HUVEC Cells, Untx	HUVEC Cells, t/IL10 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr
2	HG1	-1.31	HAEC Cells, Untx, Nrm1	HAEC Cells, t/TNFa 10 ng/ml 10 hr, Nrm1
2	HG1	-1.31	HAEC Cells, Untx, Nrm1	HAEC Cells, t/TNFa 10 ng/ml 24 hr, Nrm1
2	HG1	-1.34	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/ml, 10 ng/ml 24 hr, 4 hr, Nrm1
2	HG1	-1.36	HAEC Cells, Untx, Nrm1	HAEC Cells, t/TNFa 10 ng/ml 4 hr, Nrm1
2	HG1	-1.40	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/ml, 10 ng/ml 24 hr, 24 hr, Nrm1
2	HG1	-1.43	HUVEC Cells, Untx	HUVEC Cells, t/TNFa 10 ng/mL 24 hr
2	HG1	-1.44	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/IL1b 10 ng/mL 24 hr, Nrm1
2	HG1	-1.48	HUVEC Cells, Untx	HUVEC Cells, t/TNFa 10 ng/mL 24 hr
2	HG1	-1.52	HUAEC Cells, Untx, Nrm1	HUAEC Cells, t/TNFa 10 ng/mL 8 hr, Nrm1
2	HG1	-1.53	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 4 hr, Nrm1
2	HG1	-1.53	HUVEC Cells, Untx	HUVEC Cells, t/CHX + TNFa 10 mcg/ml, 10 ng/ml 30 min, 23.5 hr
2	HG1	-1.53	HUVEC Cells, Untx	HUVEC Cells, t/TNFa 1 ng/mL 24 hr
2	HG1	-1.55	HAEC Cells, Untx, Nrm1	HAEC Cells, t/TNFa 10 ng/ml 8 hr, Nrm1
2	HG1	-1.58	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/PD98059 + TNFa 50 microM, 10 ng/mL 24 hr, 24 hr, Nrm1
2	HG1	-1.59	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 3 d, Nrm1
2	HG1	-1.59	HAEC Cells, Untx, Nrm1	HAEC Cells, t/TNFa 10 ng/ml 6 hr, Nrm1
2	HG1	-1.61	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/IL1b + TNFa 10 ng/mL, 10 ng/mL 24 hr, 1 hr, Nrm1
2	HG1	-1.62	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/TNFa 10 ng/ml 24 hr, Nrm1
2	HG1	-1.72	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 8 hr, Nrm1
2	HG1	-1.80	HUVEC Cells, Untx, Nrm1, 24 hr	HUVEC Cells, t/PD98059 + TNFa 50 microM, 10 ng/mL 24 hr, 4 hr, Nrm1
2	HG1	-1.83	HIAEC Cells, Untx, Nrm1	HIAEC Cells, t/TNFa 10 ng/mL 8 hr, Nrm1
2	HG1	-2.07	HUVEC Cells, Untx, 48 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr
2	HG1	-2.15	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/TNFa 10 ng/mL 8 hr, Nrm1
2	HG1	-2.18	HUVEC Cells, Untx, 48 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 4 hr
10	UG1	3.38	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/CHX + TNFa 10 mcg/mL, 10 ng/mL 30 min, 4 hr, Nrm1
10	UG1	3.16	HUVEC Cells, Untx, Nrm1	HUVEC Cells, t/CHX + TNFa 10 mcg/mL, 10 ng/mL 30 min, 1 hr, Nrm1
10	UG1	2.85	HUAEC Cells, Untx, Nrm1	HUAEC Cells, t/TNFa 10 ng/mL 2 hr, Nrm1
10	UG1	2.68	HPAEC Cells, Untx, Nrm1	HPAEC Cells, t/TNFa 10 ng/mL 1 hr, Nrm1
10	UG1	2.64	HPAEC Cells, Untx, Nrm1	HPAEC Cells, t/TNFa 10 ng/mL 2 hr, Nrm1

TABLE 7-continued

Microarray data from cardiovascular experiments				
SEQ ID	GEM	Log2 (Cy5/Cy3)	Cy3 Sample	Cy5 Sample
10	UG1	2.60	HUVEC Cells, Untx, Nrml, 24 hr	HUVEC Cells, t/PMA + TNFa 100 nM, 10 ng/mL 24 hr, 1 hr, Nrml
10	UG1	2.55	HUVEC Cells, Untx, Nrml, 24 hr	HUVEC Cells, t/PMA + TNFa 100 nM, 10 ng/mL 24 hr, 24 hr, Nrml
10	UG1	2.47	HUVEC Cells, Untx, 0 hr, Nrml	HUVEC Cells, t/Dex + TNFa 100 nM, 10 ng/mL 24 hr, 1 hr Nrml
10	UG1	2.40	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/TNFa 10 ng/mL 48 hr, Nrml
10	UG1	2.39	HUVEC Cells, Unix, Nrml, 0 hr	HUVEC Cells, t/Dex + TNFa 100 nM, 10 ng/mL 24 hr, 24 hr Nrml
10	UG1	2.27	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/IL10 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr, Nrml
10	UG1	2.24	HUAEC Cells, Untx, Nrml	HUAEC Cells, t/TNFa 10 ng/mL 4 hr, Nrml
10	UG1	2.18	HUAEC Cells, Untx, Nrml	HUAEC Cells, t/TNFa 10 ng/mL 1 hr, Nrml
10	UG1	2.17	HUVEC Cells, Untx, Nrml, 0 hr	HUVEC Cells, t/Dex + TNFa 10 nM, 10 ng/mL 24 hr, 24 hr Nrml
10	UG1	2.14	HUVEC Cells, Untx, 24 hr	HUVEC Cells, t/Lipom + TNFa 8 mcg/mL, 10 ng/ml 4 hr, 4 hr, 1 hr
10	UG1	2.11	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/IL1b + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr, Nrml
10	UG1	2.04	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/IFNg + TNFa 200 ng/mL, 10 ng/mL 24 hr, 24 hr, Nrml
10	UG1	1.95	HUVEC Cells, Untx, Nrml, 0 hr	HUVEC Cells, t/Dex + TNFa 10 nM, 10 ng/mL 24 hr, 1 hr Nrml
10	UG1	1.91	HUVEC Cells, Untx, 24 hr	HUVEC Cells, t/Lipom + asOligo + TNFa 8 mcg/mL, 100 nM, 50 nM, 10 ng/ml 4 hr, 4 hr, 1 hr
10	UG1	1.91	HUVEC Cells, Untx, Nrml, 24 hr	HUVEC Cells, t/PD98059 + TNFa 50 microM, 10 ng/mL 24 hr, 1 hr, Nrml
10	UG1	1.88	HUVEC Cells, Untx, Nrml, 0 hr	HUVEC Cells, t/Dex + TNFa 100 nM, 10 ng/mL 24 hr, 4 hr Nrml
10	UG1	1.88	HUVEC Cells, Untx, Nrml, 0 hr	HUVEC Cells, t/Dex + TNFa 10 nM, 10 ng/mL 24 hr, 4 hr Nrml
10	UG1	1.86	HUVEC Cells, Untx, 48 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr
10	UG1	1.82	HUVEC Cells, Untx, 24 hr	HUVEC Cells, t/Lipom + TNFa 8 mcg/mL, 10 ng/ml 4 hr, 4 hr, 24 hr
10	UG1	1.82	HUVEC Cells, Untx, 48 hr	HUVEC Cells, t/IL4 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 1 hr
10	UG1	1.76	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/IL1b 10 ng/mL 24 hr, Nrml
10	UG1	1.74	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/TNFa 10 ng/mL 24 hr, Nrml
10	HG3	1.20	HUAEC Cells, Untx, Nrml	HUAEC Cells, t/TNFa 10 ng/mL 1 hr, Nrml
10	HG3	1.04	HUAEC Cells, Untx, Nrml	HUAEC Cells, t/TNFa 10 ng/mL 2 hr, Nrml
12	HG5	1.02	ECV304 Line, t/TNFa 10 ng/mL 3 d, Nrml	ECV304 Line, Untx, Nrml
12	HG5	-1.07	HUVEC Cells, Untx	HUVEC Cells, t/IL10 + TNFa 10 ng/mL, 10 ng/mL 24 hr, 4 hr
12	HG5	-1.16	HUAEC Cells, Untx, Nrml	HUAEC Cells, t/TNFa 10 ng/mL 8 hr, Nrml
12	HG5	-1.18	HMVEC Cells, Untx, Nrml	HMVEC Cells, t/TNFa 10 ng/mL 24 hr, Nrml
12	HG5	-1.24	HMVEC Cells, Untx, Nrml	HMVEC Cells, t/TNFa 10 ng/mL 8 hr, Nrml
12	HG5	-1.50	HIAEC Cells, Untx, Nrml	HIAEC Cells, t/TNFa 10 ng/mL 8 hr, Nrml
18	HG2	-2.38	HUVEC Cells, Untx, Nrml	HUVEC Cells, t/IFNg + TNFa 10 ng/mL, 10 ng/mL 24 hr, 24 hr, Nrml
32	HG3	-1.13	HUVEC Cells, Untx 24 hr, Nrml	HUVEC Cells, t/PMA + TNFa 10 nM, 10 ng/mL 24 hr, 24 hr, Nrml
33	LG1	-1.17	Tangier	Fibroblast Cells, t/LDL Cholesterol, Nrml
34	UG1	-1.04	ECV304 Line, Untx	ECV304 Line, t/PMA + Iono 1 microM, 1 meg/ml 4 hr
34	UG1	-1.11	ECV304 Line, Untx	ECV304 Line, t/PMA + Iono 1 microM, 1 meg/ml 5 hr

[0221]

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<210> SEQ ID NO 3
<211> LENGTH: 586
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 48, 66, 560, 574, 577, 580
<223> OTHER INFORMATION: a, t, c, g, or other

<400> SEQUENCE: 3

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gggcgacgtg gagtggggcg gtctggcg gccgagatgc cgtcctgcgc tgcgagggac 180
ccatccccga cgtcaccttc gagctgctgc gcgagggcga gacgaaggcc gtgaagacgg 240
tccgcacccc cggggcccg gcgaacctcg agctgatctt cgtggggccc cagcacgccg 300
gcaactacag gtgccgctac cgtcctggg tgccccacac cttogaatcg gagctcagcg 360
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tgtcctcaga agtgccggg attctggact ggctccctcc cctcctgttg cagcacaagg 480
ccgggggtctc tggggggctg gagaagcctc cctcattcct cccaggaatt aataaatgtg 540
aagagagctc tgtttaaan aaaaaaaaaa aaanaanan aaccaa 586
    
```

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<210> SEQ ID NO 4
<211> LENGTH: 433
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
    
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<400> SEQUENCE: 4
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ccacaacaga cgggacaact tgcaagctg caaccccagg acagagctgg agccagggcc 180
agctggatgc ccatgttcca gaggcgaagg aggcgagaca cccacttccc catctgcatt 240
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cctgcctcgc ccccgctccc tcccttctt atttattct gctgccccag aacataggtc 360
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aaaaaaaaaa aaa 433
    
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```

<210> SEQ ID NO 5
<211> LENGTH: 3111
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 44
<223> OTHER INFORMATION: a, t, c, g, or other
    
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<400> SEQUENCE: 5
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cccgcttcgc ccaactgggt ctcccggctg cagtgccagg gcgcaggac cgcccgatct 180
cccgcctccg ccacctccgc caccatgctg ctccccagc tctgctggct gccgctgctc 240
gctgggctgc tcccgcgggt gcccgctcag aagttctcgg cgtcacggt tttgagagtg 300
gatcaagata aagacaagga ttgtagcttg gactgtcgg gttcgccca gaaacctctc 360
tgcgcactg acggaaggac ctctctttcc cgttgtgaat ttcaactgac caagtgaaa 420
gatccccagc tagagattgc atatcgagga aactgcaaag acgtgtccag gtgtgtggcc 480
gaaaggaagt ataccagga gcaagcccg aaggagtctc agcaagtgtt cattcctgag 540
tgcaatgacg acggcaccta cagttaggtc cagtgtcaca gctacacggg atactgctgg 600
    
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tgcgtcacgc	ccaacgggag	gccatcagc	ggcactgccg	tggcccacaa	gacgccccgg	660
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gccgcagctc	cagcgttga	gactcagcct	caaggagatg	aagaagatat	tgcatcacgt	780
taccctacco	tttgactga	acaggttaa	agtcggcaga	acaaaaccaa	taagaattca	840
gtgtcatcct	gtgaccaaga	gcaccagtct	gccctggagg	aagccaagca	gcccagaac	900
gacaatgtgg	tgatccctga	gtgtgcgcac	ggcgcctct	acaagccagt	gcagtgccac	960
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ccgggacctg	tacaaggccc	gccagctaca	aggttgtccg	ggtgccaaaa	agcatgagtt	1140
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aaccagctgc	caccctcagg	cctgggcccc	agagctcagg	gcaccagtg	tcttaaggaa	2760
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ttaagaaaca gttatgatcc taaacttttt ggataatcct ttatatttct gacctttgaa 3060
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<210> SEQ ID NO 6
<211> LENGTH: 2311
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 474-544, 2288, 2295-2299
<223> OTHER INFORMATION: a, t, c, g, or other

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<400> SEQUENCE: 6
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gtggcagcct ccccgccgag gcctcctgca ccacctagag ccccccaccc gaccccaccc 180
cgggagggca gagccagaag aaggctcatt agacctgggg gaccocaaag gtctggcctc 240
tttgggcagc cccagagatg aggggtcagc agaggagagc tctgggggtt gggatgggtt 300
agggacgcaa gcttgagttc tagcccttgc tctcattcag ctgtttgtgt accctgggta 360
agacccttcc ttgtttgacc ctacagtttc ccatctgttt aatgggtggc ttggccaagg 420
caatccacaa acgtcaaaa tccccttccc atcagtacac acaccgatgc acannnnnnn 480
nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn 540
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tggtgcccag tgacctgggg gagcctggct gcaggccctc actggttccc taaaccttgg 1560
tggtctgatg tcaggctccc aggggggact caggaggaga tatggctgag ttctgtatgt 1620

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taaaaaanaa aaaannnna aaaaaaaag g 2311

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<210> SEQ ID NO 7
<211> LENGTH: 1866
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

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<400> SEQUENCE: 7

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gccatcatca tgtccacgtc gctacgagtc agcccatcca tccatggcta ccaactcgac 180
acagcctcto gtaagaaagc cgtgggcaac atctttgaaa acacagacca agaatcacta 240
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gccatagatc aagatgtgga ggagaaaacg cgtgccctga tggcctttaa gaagaggaca 360
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gaagaggatg gataaggacg ttatccaaga atggacattc aaagaccaag tgagtttgtg 480
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accaaacttt ttcaaaacaa attcttacgt caaatatctg ggaagtttct ctgtccaat	1560
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tgtatccatt aaggatgagt tttaaaaggc tttctctca tacttttgaa aaatttcttc	1680
tatgattaca gtagctatgt acatgtgtac atctattttt cccaagcaat atgttttggg	1740
tttagagtct gagtgtgac caagattctg tgtgttacta ctgtttgttt aataggaaca	1800
aatatagaaa taatattatc tctttgotta tttcccgta aaactataat aaaatgttc	1860
taggaa	1866

<210> SEQ ID NO 8
 <211> LENGTH: 1929
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature

<400> SEQUENCE: 8

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ctcccaatgc cagcggcgtc agcgtgcca gcgtgcctt agcagccagc gccgccagcc	180
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aaaaaaaaa 1929

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<210> SEQ ID NO 9
<211> LENGTH: 1831
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

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<400> SEQUENCE: 9

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gggagggcca tgatttccct cccggggccc ctggtgacca acttgcctgc gttttgttc 180
ctggggctga gtgccctcgc gcccccctcg cgggccacgc tgcaactgca cttgcccgcc 240
aaccggttgc aggcggtgga gggaggggaa gtggtgcttc cagcgtggtc caccttgcac 300
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aaagaaaagg aggatcaggt gttgtcctac atcaatgggg tcacaacaag caaacctgga 420
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gagaaagact ctggccccta cagctgctcc gtgaatgtgc aagacaaaca aggcaaatct 540
aggggccaca gcatcaaac cttagaactc aatgtactgg ttccctcagc tctccatcc 600
tgccgtctcc aggggtgccc ccatgtgggg gcaaactgca ccctgagctg ccagtctcca 660
aggagtaagc ccgctgtcca ataccagtgg gatcggcagc ttccatcctt ccagactttc 720
tttgaccag cattagatgt catccgtggg tctttaagcc tcaccaacct ttcgtcttc 780
atggctggag tctatgtctg caaggccac aatgaggtgg gcaactgcca atgtaatgtg 840
acgctggaag tgagcacagg tcagtgagg ggccctggagc tgcagtggtt gctggagctg 900
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cctccgcacg agccctccgg ccaccccctg gccctcccag gcctggtgca ttgacccca 1140
cgcccagtct ctccagccag gccctgccct caccaagact gccacgaca gatggggccc 1200
accctcaacc aatatcccc atccctggtg gggtttcttc ctctggcttg agccgatgg 1260
gtgctgtgctg tgtgatggtg cctgcccaga gtcaagctgg ctctctggtc tgatgacccc 1320

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accactcatt ggctaaaagga tttgggggtct ctccttccta taagggtcac ctctagcaca 1380
gaggcctgag tcatgggaaa gagtcacact cctgaccctt agtactctgc ccccacctct 1440
ctttactgtg ggaaaacct ctcagtaaga cctaagtgtc caggagacag aaggagaaga 1500
ggaagtggat ctggaattgg gaggagocct caccaccccc tgactcctcc ttatgaagcc 1560
agctgctgaa attagctact caccaagagt gaggggcaga gacttccagt cactgagtct 1620
cccaggcccc cttgatctgt accccacccc tatctaacac cacccttggc tcccactcca 1680
gctccctgta ttgataaac ctgtcaggct ggcttggtta ggttttactg gggcagagga 1740
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taaagataca taatgtttgt atgagataag a 1831

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<210> SEQ ID NO 10
<211> LENGTH: 1453
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 903-935
<223> OTHER INFORMATION: a, t, c, g, or other

<400> SEQUENCE: 10

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tctccctggc ccaggctcca gccacaggca cctctcctgc ccccgccac cctcctgacc 120
gcagctccca ggccttgagg acctccaggc tttcctgccc tgggcagccc cacctcacag 180
ccagagtcaa tgcttcatg ggaagggctc ccagccacac ccagagtggc ccaaagctgt 240
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cctccccacc ctctcagag gcccggggaa ggaagagca ggtcagtaca gaggttctgt 360
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tgactgctag agggcagggg tgcagggtc aggggggccc ggtggtcct ttggggctgg 480
tgttctacg tcagtcccca cctggggaat aaactccagc ctctcctgct catacagaag 540
gaactggttg ggtttgcttt atgggatctt tgagacaaa acagatgctc ctggttgctg 600
ggggagggtg tgagcacgga gtatttctgt ccctcgtgaa gtcacgtcac acaggggaga 660
ggcgaggtcg atggaactcg ccacgcacag gctctggctc tgggaaggag gatgatgagt 720
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aatgaaccga tggttgagg attgtcacgg gaggaacatg acaccgaag ggaacttagg 840
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tgnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnttggt cactcaga acccaggaca 960
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agctcagagg ccccagagg gtgtgtagga ggcgaggcc cgcagagcac agagcaggag 1140
aagggcttgg gccctgagg aaaaagccat tctggacacc aggggacctg gacggagggg 1200
ccccacagcc cgtgccccac gccgcctgga ggccagaggg gtcagtggcc ctgctgtccc 1260
ggctccatct tggttctagc cgccacctgt atgaacacag tggcccggct taacgcacta 1320

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accagcctc tccctgtgc ccacaggag tagcaagacc cacccacac tgccttcacc 1380
atctacacca gtgacgocgc tgtgtgtctt agcatggaaa taaataaacc tgaatgcaaa 1440
aaaaaaaaa agg 1453

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<210> SEQ ID NO 11
<211> LENGTH: 443
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

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<400> SEQUENCE: 11

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gatatagaca acttccagag tcaccagtgt gcaaatggag ccacctgcat tagtcatact 60
aatggctatt cttgcctctg ttttgaaat tttacaggaa aatthttgag acagagcaga 120
ttaccctcaa cagtctgtgg gaatgagaag acaaatctca cttgtacaa tggaggcaac 180
tgcacagagt tccagactga attaaaatgt atgtgccggc caggthttac tggagaatgg 240
tgtgaaaagg acattgatga gtgtgcctct gatccgtgtg tcaatggagg tctgtgccag 300
gacttactca acaaatccca gtgcctctgt gatgttgctt ttgctggcga gcgctgcgag 360
gtggacttgg cagatgactt gatctcogac atthttacca ctattggctc agtgactgtc 420
gccttgttac tgatcctctt gct 443

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<210> SEQ ID NO 12
<211> LENGTH: 1537
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 284-285, 287
<223> OTHER INFORMATION: a, t, c, g, or other

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<400> SEQUENCE: 12

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aaaaaaaaa acccggtagc attgtccctt cccactgac aaacttatca aatccagaag 60
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atgtatthtt agaattacgg cagcatcaga cctgaatthtt gtgagthttcg tggctthtct 180
ttaaatacacc atthccctaa aaacggthttc thttccctta gaaatgctgg tggcaacttg 240
atgaaacagc caaatgcacc agggcaggtc actthcccaa aaananaag aaaaaaact 300
cattgagata gctacagttc tataggthaa thtaaacctt cththttcta ctcatthttg 360
aaagcaaaat tacatthttac ththttacat aaccagtga aagacgttg aagcctacag 420
ctcactgthtt thgtgctctt ggaatgthtt aggtggthtt thtaaccagt gatththtaac 480
gtgcagtga aatthttgag ththtaaacac cagctaaagt agtcaactt gatccccatt 540
aaaaatcaag gaatthgggg tcgggggagg ththtaggag gatccagaat gacctcccag 600
aattactgtg cgtacaactt thththttcag agththttcatt ggaatgthta gagththttatg 660
aaagacagth thtaaaacta thctgagthta aatthtaata ththtaaaaa ththttgtact 720
agactthttg cagctthttg aaagtagcag agththttcatca thccacatat ataacagagc 780
ataaaththtt tataatcagc cacctthttgc thgtthttgag thagactgthtt thctgtthta 840
ggtgthtaagc atcgcacagc aaaaaaatct thttctctct ctcgattgta gcatagcctg 900

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acagctctag atacagcatt tctatgatga aaaatgagta tccatcagga aatctagaag 960
actagccgtg ttttctcaga ctccaccttt gtttgcactc tgttgctgt gaggagcttt 1020
ctggcatgtg attatttact tcaaaactag agttccaagc acctacatta attattttat 1080
attgtgtgca gaatagtata tcttttaatg tcagatatga tacactgcac atattgcttt 1140
tgcactctta aaatTTTTgt actaaataat agaaaatatt tatattcttt gagtgtgagc 1200
tttgaataga tggcattatc actttattgt tttttaaca aaaactTTTT ctcaattatt 1260
ctattgcaat gttattctga gcaagtccta tgccaaatat cttgtataat gtttztatgg 1320
aagattaaat tttactcttg tgtgtaaga ctatttcagt tactgatttt atagtggaa 1380
tttgatattc cagcacaag tccacagtgt attcagaaat ccaagttggt gtcatacatt 1440
tcattttgat gtgaactttt ctttgcttcc ctttgttcta agaotccatt ttgcaataaa 1500
cgttttgaca gtaaaaaaaaa taaaaagga aaaaaaa 1537

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<210> SEQ ID NO 13
<211> LENGTH: 972
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

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<400> SEQUENCE: 13

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attaaacaca ccacgagaga acaatattcc aggagactta atagtgatta ctttcttcaa 180
tcaggaaatc gtttcagtgc ctcccttgta ggaatgcttt gttttgtgat gggttttctt 240
aaagaagagc acacctccgt ccaatctcct gagacagcca cgtotccgct gacatcccac 300
tgtgatgctt tcagatagtc agtgaatggt tctgataacc ttcattccagt atctgaaaca 360
caatgtgaga gattatattg ttttagataa taacatccca tttagttgac taaaatcttc 420
caaaactctga aagctgcaca ctgctactcc agagagtgcg ggtottagct cttctccttt 480
ctgacttcaa gatgaatctt tgggacgatg tttctggtgc ttggtccaca gtgattcact 540
tttgaaggag aggccacatg acatgaactg cctgggtgta caacctagct aacatatttg 600
atgctactcc tgttgtctgt actgcttatt caagtagtat tctaagtatt gttactaaaa 660
aacatggtgg gtaaagcaca atcctaccca tcattgtcct caaaataat tgotgacat 720
acacggccca gccattgcc ctccctgcat ctctgtgctg ctttgccatt tccccttcta 780
cccagcctcc tcaaggggta ccttggtgga tatttcagta cttaaaacca gactgtaatc 840
ataacctccc tctgtgtggc atcaataaat agccaaactc aaaaaaaaaa aaaaaaaaaa 900
aaaaaaaaaa aaaaatatac ggtcgcaagc ttattccctt tagtgagggt taatttttagc 960
ttgcaactgcc ta 972

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<210> SEQ ID NO 14
<211> LENGTH: 1544
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

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<400> SEQUENCE: 14

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agagccacaa gaccacacac acagccotta cctcctcag gactaccgaa ccttctggca	120
caccttgtag agagttttg ggttcacacc ccaaaatgac ccaacgatgt ccacacacca	180
ccaaaacca gccaatgggc cacctcttcc tccaagccca gatgcagaga tggacatggg	240
cagctggagg gtaggctcag aaatgaagg aaccocctcag tgggctgctg gacctatctt	300
tcccaagcct tgccattatc tctgtgagg aggccaggta gccgagggat caggatgcag	360
gctgctgtac ccgctctgcc tcaagatcc cccacacagg gctctggtt tcaactgctt	420
cgctctagat agtttaaatg ggaatcagat cccctggtg agagctaaga caaccaccta	480
ccagtgccca tgtcccttcc agctcacctt gagcagcctc agatcatctc tgtcactctg	540
gaagggacac cccagccagg gacggaatgc ctggtcttga gcaacctccc actgctggag	600
tgcgagtggg aatcagagcc tcctgaagcc tctgggaact cctcctgtgg ccaccaccaa	660
aggatgagga atctgagttg ccaacttcag gacgacacct ggcttgccac ccacagtga	720
ccacaggcca acctacgccc ttcactactt ggttctgtt taatcgactg gccocctgtc	780
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ccgagacgcc ttggctggca cctctgggg tcccccttt ccccaggca ggtcatcttt	900
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gtggacacag ctgtaattt ccaactggta tgccacttca gactcttca tggcagcgtt	1080
tgagctctc tgggtaaaa cttcccttg ttgactggcc ttcacagcca tggctgtgta	1140
caacagagga tcggtgagat tgagcagcgc ttggtgatct ctacagaaac aaccctgcc	1200
cgtagggcaa tctactttaa gttactogga caaagacccc aaagtggggc aacaactcca	1260
gagaggctgt gggaaatctc agaagcccc ctgtaagaga cagacatgag agacaagcat	1320
cttctttccc ccgcaagtc attttatttc cttcttctgc tgcctggaa gagaggcagt	1380
agcaaagaga tgagctcctg gatggcattt tccagggcag gagaaagtat gagagcctca	1440
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<210> SEQ ID NO 15
 <211> LENGTH: 1109
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <220> FEATURE:
 <221> NAME/KEY: unsure
 <222> LOCATION: 751-806, 884-885
 <223> OTHER INFORMATION: a, t, c, g, or other
 <400> SEQUENCE: 15

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ccagcttgcg catcatctgc ggcgggtcc cgatgagcct cctgttgccct ccgctggcgc	120
tgtctgtct tctcggggc cttgtggccc cagccacagc cgccaactgcc taccggccgg	180
actggaaccg tctgagcggc ctaaccgcg cccgggtaga gacctgcggg ggatgacagc	240
tgaaccgcct aaaggagggtg aaggctttcg tcacgcagga cattccattc tatcacaacc	300

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tggtgatgaa acacctccct ggggcccgacc ctgagctcgt gctgctgggc cgccgctacg	360
aggaactaga gcgcatocca ctcagtgaat tgaccgcga agagatcaat gcgctagtgc	420
aggagctcgg cttctaccgc aaggcggcgc ccgacgcgca ggtgcccccc gagtacgtgt	480
ggcgccccgc gaagccccca gaggaaactt cggaccacgc tgacctgtag gtccgggggc	540
gcggcggagc tgggacctac ctgcctgagt cctggagaca gaatgaagcg ctcagcatcc	600
cgggaatact tctcttgctg agagccgatg cccgtccccg ggccagcagc gatgggggtg	660
gggaggttct cccaacccca ctttcttctt tccccagctc cactaaattc ctcctgcct	720
taaaaaaaa aagaaaaacc aaacaacaa nnnnnnnnn nnnnnnnnn nnnnnnnnn	780
nnnnnnnnn nnnnnnnnn nnnnnntct ctatagtgc acctaaattc aattcactgg	840
ccgtcgtttt acaacgtcgt gactggcgac ggacaaagt atcnnttaa tcgccttgoa	900
gcacataccc ctttgccagc ctgggtaat aggggaagcg ggccggacc gatcggcctt	960
cccaaacagt tggggaagct tgaatgcgc gacattgggc cgacggcctt ctatacggga	1020
ggatctctaa acgcggcccg ggtgttggtt gggttaagc ggagtgtgac cccgcataat	1080
aacttttgca caggggccct ataggggccc	1109

<210> SEQ ID NO 16
 <211> LENGTH: 1740
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature

<400> SEQUENCE: 16

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ctctcactcc tcaactgagtc cactctgaac gtgctaaat gggaaaggag cgggtgtttg	180
ctgatctgtt aaattcttag tgaagtttcc ttgatttcca gtggctgctg ttgtttgagt	240
ttggtttgga gcaaaaactg ggtagtcta acatttctgg gactgaatcc aggcaagaga	300
aagaagaaaa agaagaagaa aaagaggag aaaaaggtag ggagaaata agggaggaga	360
gaagcacagt gaaagaaaa aaaagtcctt ttgcgacat cacattctg tgtttccct	420
cagcctggaa aacatattaa tcccagtgtt ttacgcccg gaaacaaaga gactaagcca	480
gactatgggg gaaagggaga taagaaggat cctggaactt taaagggga aagagtgaga	540
ttcagaaatc gccaggactg gactttaagg gacgtcctgt gtcagcaca gggactggca	600
cacacagaca cacgagaccg aggagaaact gcagacaaat ggagatacaa agacttagaa	660
ggacagctcc ttccacctca tcctacttgt ccagaaggta aaaagacaca gccagaaaga	720
aaaggcatcg gctcagctct cagatcagga caggctgtgg atctgtggcg gtactctgaa	780
agctggagct gcagcacacc ccttttgat tgctcacct cggtaaagag agagagggt	840
gggaggaaaa gtagtccatc taggaaactg tcctgggaac caaacttctg atttctttg	900
caaccctctg cattocatct ctatgagcca ccattggatt acacaatgac atggagaatg	960
ggacccggtt tcactatgct gttggccatg tggtagtgt gtggatcaga accccacccc	1020
catgccacta ttagaggcag ccacggagga cggaaagtgc ctttggttc tccggacagc	1080
agtaggccag ctcggtttct gaggcacact gggaggtctc gcggaattga gagatccact	1140

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ctggaggaac caaaccttca gcctctocag agaaggagga gtgtgcccgt gttgagacta 1200
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caaagaccag cagccagggg ctctccgcgt gagatgatca gagatgaggg gtcctcagct 1320
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cctcatgatg agcctgtga aggacgatgt gtactgtgag ctggcggaga ggcacatcca 1500
acagattgtg ctctccacc aggcaggtga ggaaggaggc aaggtagaa ggatcaccag 1560
cgagggccag atcctggagc agcccctggg accctagcct catocctaag ctgatgagct 1620
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<210> SEQ ID NO 17
<211> LENGTH: 4467
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 971, 978, 1295-1522
<223> OTHER INFORMATION: a, t, c, g, or other

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<400> SEQUENCE: 17

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ccttctcggg cgcgctctgt gcgtacttca ggcccagggc ccacgacggc ggcgaacatc 180
cggatcccct gaccatctgc ccgctgccg ttagagggac gcgctcccc gaagagaacg 240
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gcgccctcgt ggcgcgggga ctggccccg gggggcccc ttgatcccag gccagcgtt 360
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aaagagaggg acctttgcct acgtagatgt gtatgtgtag tgcgattttc ttcaaggaaa 480
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tattttttgt atatgtttg tagctctaaa aaggtcgaga tgcaataaca ctctgtaagc 720
aacgagttoa cctaagtaag gtcagatcc tagttttaaa aaccatttcc cattaataatg 780
aagttggagg aacagctgct tctggagccg gggcaaaaaa tttcaaggtg agcctggagc 840
attgtgtgtg gtgaagtaaa ataaaggctc aaaacgtgac ggcaaccocg caaaagggtg 900
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tattgtgtag agtcttgag aacaggtgtt ttccagtctc aaagcagtaa cttatacac 1140
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tggggaaga	agtggtcttc	accccctacc	cccannnnnn	nnnnnnnnnn	nnnnnnnnnn	1320
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gtctgccact	gaatttcag	tgactaagtg	gaaaaatata	aaacatatga	atataaagaa	1680
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tcccgaatca	aagatatgga	gaagcagaag	aaggagggca	ttgtttgcaa	agaggacaaa	3480
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gttgggggag tgtagaact gttccaatt aatgggagct ctgttgttga gcgagaagac 3720
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<210> SEQ ID NO 18
<211> LENGTH: 2965
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 1473-1777, 2948, 2951, 2961
<223> OTHER INFORMATION: a, t, c, g, or other

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<400> SEQUENCE: 18

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ctttgagaaa ttgactgaca tactggaaga gaacaccatt ttatctcagc ttagtgaaga 180
atcagtgcag gtccctgact cttattttcc cagaggccat ggagctgaga ttgagactag 240
ccttgtgggt ttcacactaa agagtttcct tgttatgggc aacatgcatg acctaatgtc 300
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aaaggaaaaa cagcacaatg gtgaccactg ataaaggctt tattaggtat atctgaggaa 420
gtgggtcaca tgaaatgtaa aaaggaatg aggtttttgt tgttttttg aagtaaaggc 480
aaacataaat attaccatga tgaattotag tgaatgacc ccttgacttt gcttttctta 540
atacagatat ttactgagag gaactatttt tataacacaa gaaaaattha caattgatta 600
aaagtatcca tgtcttggat acatacgtat ctatagagct ggcatgtaat tcttctctta 660
taaagaatag gtataggaaa gactgaataa aaatggaggg atatccctt ggatttcaat 720
tgcatgtgc aataagcaaa gaaggttga taaaagttct tgatcaaaaa gttcaaagaa 780
accagaattt tagacagcaa gctaaataaa tattgtaaaa ttgcaactata ttaggttaag 840
tattatttag gtattataat atgctttgta aattttatat tccaaatatt gctcaatatt 900

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ccaggatatt tttctataaa acttttttaa aataattgta tctatatatt caattttaca 1440
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gctgatatgt ttctgtatt ctagaaaaat ttttacctt tcacattatt tttgtacct 2880
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<210> SEQ ID NO 19

<211> LENGTH: 1734

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

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<221> NAME/KEY: misc_feature
<220> FEATURE:
<221> NAME/KEY: unsure
<222> LOCATION: 571-899
<223> OTHER INFORMATION: a, t, c, g, or other

<400> SEQUENCE: 19

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gaacggctac atctagaagg caacaaattg caagtactgg gaaaagatct cctcttgccg    180
cagccggacc tgcgctacct ctctcgtaac ggcaacaagc tggccagggt ggcagccggt    240
gccttcagg gctcggcgca gctggacatg ctggacctct ccaataactc actggccagc    300
gtgcccagg ggtctggtgc atccctaggg cagccaaact gggacatgcg ggatggcttc    360
gacatctccg gcaaccctgt gatctgtgac cagaacctga gcgacctcta tcgttggtt    420
caggcccaaa aagacaagat gttttccagc aatgacacgc gctgtgctgg gcctgaagcc    480
gtgaagggcc agacgctctc gggcagtgcc caagtcccag tgagaccagg ggcttggtt    540
gagggtgggg ggtctggtag aacactgcaa nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn    600
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ccttgactgt tgcccgggtg tcattttacc agtggttttc tgccagggtc tgtgtcctag   1500
gagaagggtt aagttaaac agattgocca ggtctccaaa cgatttgtca tgetgacctg   1560
agatcatcga agggggcacc tgccccggg caaggttgca ggggcaggat ggggctgaag   1620
ggatgagcag ggtcccgggc ccacctgctg atacagcatt ggccatgtgg gggctgcaat   1680
cggatttggg agaccctggg gcttgggggc atgtccattt tcccagttcc taaa       1734

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<210> SEQ ID NO 20
<211> LENGTH: 4005
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature

<400> SEQUENCE: 20

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tcatgggggc agtgaatgtc gccaaagga ccgtccagac cagtgtggac accaccaaga	180
ctgtcctaao tggtagcaag gacaccgtct gcagtggggt gaccgggtgct gcgaatgtgg	240
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acaccgtatc cactgggctc acaggggctg tgaacttggc caaagggact gtccagaccg	360
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ctggtgccgt aaatgtggcc aaaggcaccg tccagacagc tgtggacaca gccaaagcgg	480
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ggtcagggaa cgtggcgaca ggggccacc acactggcct cagcaccttc cagaactggt	780
tacctagtac ccccgcacc tcttggggtg gactcaccag tccaggacc acagacaatg	840
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tgcaggcagt gctgtggcat ccaggtcc gggcagctcc gttctcatgc tgaagtggg	2220
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<210> SEQ ID NO 21

<211> LENGTH: 846

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<400> SEQUENCE: 21

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tggtagcaga gcgagattcc atctcaaaaa aaaaaacag tatgcagta caaatttctt 180
aacctgttat caatgtctga gctacataat tatctttcta gttggagttt gttttaggtg 240

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tgtaccaact gacatttcag tttttctggt tgaagtccaa tgtattagtg actctgtggc	300
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gtcgtcgtct ctggtttcct gcatatacca atagcattac ctatgacttt ttttttcctg	420
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aaatacataa aaggaatact gctttttcct tttgtggctc aaaggtagct gcattttaaa	540
atatttgtga aaataaaaac ttttgttatt agaaaaaaaa aaaaaaaaaa aaaaaaaaaa	600
aaaaaaaaaa agaaagacca aaaaggaaga gaaggaaaa agaagaagag aaacggaggg	660
acaacgggaa acacagagag cgagccgggt acgaaaaagc ggaaggccaa cgcaggagaa	720
gaaagagagg ggggcccgtt cgctcattgt gggagtgtcc tcagagtatt gcgagtgggg	780
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gccggg	846

<210> SEQ ID NO 22
 <211> LENGTH: 1740
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <220> FEATURE:
 <221> NAME/KEY: unsure
 <222> LOCATION: 216-390, 846-889
 <223> OTHER INFORMATION: a, t, c, g, or other

<400> SEQUENCE: 22

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gagatnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnt gatatgaatg	900
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<210> SEQ ID NO 23
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<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
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<400> SEQUENCE: 23

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gatgaagaaa ctttggctga taatattgca gcattttcca gatggaagct gtatccaagg 180
atgctccgga atgttgctga aacagatctg tcgacttctg ttttaggaca gaggtcagc 240
atgccaatat gtgtgggggc tacggccatg cagcgcattg ctcattgtga cggcgagctt 300
gccactgtga gagcctgtca gtccctggga acgggcatga tgttgagttc ctgggccacc 360
tcctcaattg aagaagtggc ggaagctggc cctgaggcac ttcggttgct gctactgtat 420
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<210> SEQ ID NO 24
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<212> TYPE: DNA
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1714, 1716, 1731, 1740, 1744
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<400> SEQUENCE: 24

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<223> OTHER INFORMATION: a, t, c, g, or other
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<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<400> SEQUENCE: 26

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             20             25            30
Arg Leu Arg Val Ala Leu Ala Gly Glu Leu Val Gly Val Gly Gly
             35             40            45
His Phe Leu Phe Leu Gly Leu Ala Leu Val Ser Lys Asp Trp Arg
             50             55            60
Phe Leu Gln Arg Met Ile Thr Ala Pro Cys Ile Leu Phe Leu Phe
             65             70            75
Tyr Gly Trp Pro Gly Leu Phe Leu Glu Ser Ala Arg Trp Leu Ile
             80             85            90
Val Lys Arg Gln Ile Glu Glu Ala Gln Ser Val Leu Arg Ile Leu

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ischemia, mesenteric ischemia, peripheral vascular disease, renal artery stenosis, or stroke.

9. The method of claim 6 wherein the substrate is a microarray.

10. A method of using cDNAs to screen a sample to identify a ligand which specifically binds a cDNA, the method comprising:

- a) combining the combination of claim 1 with the sample under conditions to allow specific binding; and
- b) detecting specific binding between each cDNA and at least ligand, thereby identifying a ligand that specifically binds to each cDNA.

11. The method of claim 10 wherein the ligands are DNA molecules, proteins, RNA molecules, or transcription factors.

12. A vector containing the cDNA of claim 2.

13. A host cell containing the vector of claim 12.

14. A method for producing a protein, the method comprising the steps of:

- a) culturing the host cell of claim 13 under conditions for expression of protein; and
- b) recovering the protein from the host cell culture.

15. A protein comprising a polypeptide having the amino acid of SEQ ID NO:26.

16. A method for using a protein to screen a plurality of molecules or compounds to identify at least one ligand which specifically binds the protein, the method comprising:

- a) combining the protein encoded by a cDNA of claim 1 with the plurality of molecules or compounds under conditions to allow specific binding; and

- b) detecting specific binding between the protein and a molecule or compound, thereby identifying a ligand which specifically binds the protein.

17. The method of claim 16 wherein the plurality of molecules or compounds is selected from agonists, antagonists, antibodies, DNA molecules, small molecule drugs, immunoglobulins, inhibitors, mimetics, peptide nucleic acids, peptides, pharmaceutical agents, proteins, RNA molecules, and ribozymes.

18. A method of using a protein to produce an antibody, the method comprising:

- a) immunizing an animal with the protein encoded by a cDNA of claim 1 under conditions to elicit an antibody response;
- b) isolating animal antibodies; and
- c) screening the isolated antibodies with the protein, thereby identifying an antibody which specifically binds the protein.

19. An antibody produced by the method of claim 18.

20. A method for using an antibody to detect gene expression in a sample, the method comprising:

- a) combining the antibody of claim 19 with a sample under conditions which allow the formation of antibody:protein complexes; and
- b) detecting complex formation, wherein complex formation indicates expression of the protein in the sample.

* * * * *

专利名称(译)	动脉粥样硬化相关基因		
公开(公告)号	US20030129176A1	公开(公告)日	2003-07-10
申请号	US10/219664	申请日	2002-08-14
[标]申请(专利权)人(译)	Incyte公司基因组学		
申请(专利权)人(译)	Incyte公司基因组学, INC.		
当前申请(专利权)人(译)	Incyte公司基因组学, INC.		
[标]发明人	JONES KAREN ANNE VOLKMUTH WAYNE WALKER MICHAEL G MURRY LYNN E		
发明人	JONES, KAREN ANNE VOLKMUTH, WAYNE WALKER, MICHAEL G. MURRY, LYNN E.		
IPC分类号	G01N33/50 A61K38/00 A61P9/10 A61P43/00 C07K14/47 C12N1/15 C12N1/19 C12N1/21 C12N5/10 C12N15/09 C12P21/02 C12Q1/68 G01N33/15 G01N33/53 G01N33/566 G01N37/00 A61K38/43 C07H21/04 C12N9/00 C12N5/06		
CPC分类号	C07K14/47 A61K38/00		
外部链接	Espacenet USPTO		

摘要(译)

本发明涉及包含多种在心血管疾病中差异表达的cDNA的组合。所述组合和组合物可以完全或部分用于诊断,分期,治疗或监测与动脉粥样硬化相关的病症的进展或治疗。

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171   180   189   198   207   216
CAG CCA GCG CCG CCA GCC GTC TCG CCA CCA GTA CCG ACC CCT CCA GCA GCG GCT
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TGG CCC CCG CCG ACT TCA ACC ATT GCC CTC AAG GAT TGG GAC TAT AAT GGC CTT
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CCT GTC CTC ACC ACC AAC GCC ATC GGC CAG TGG GAT CTG GTC TGT GAC CTG GGC
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FIGURE 1A