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(54) **METHOD FOR EXPRESSION OF PROTEINS ON SPORE SURFACE**

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(57) **ABSTRACT**

The present invention relates to a method for display of protein on spore surface and a method for improving protein with rapidity using the same, which comprises the steps of (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein of interest, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein of interest, (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the protein of interest on a surface of a spore of the host cell; and (iv) recovering the spore displaying on its surface the protein of interest.

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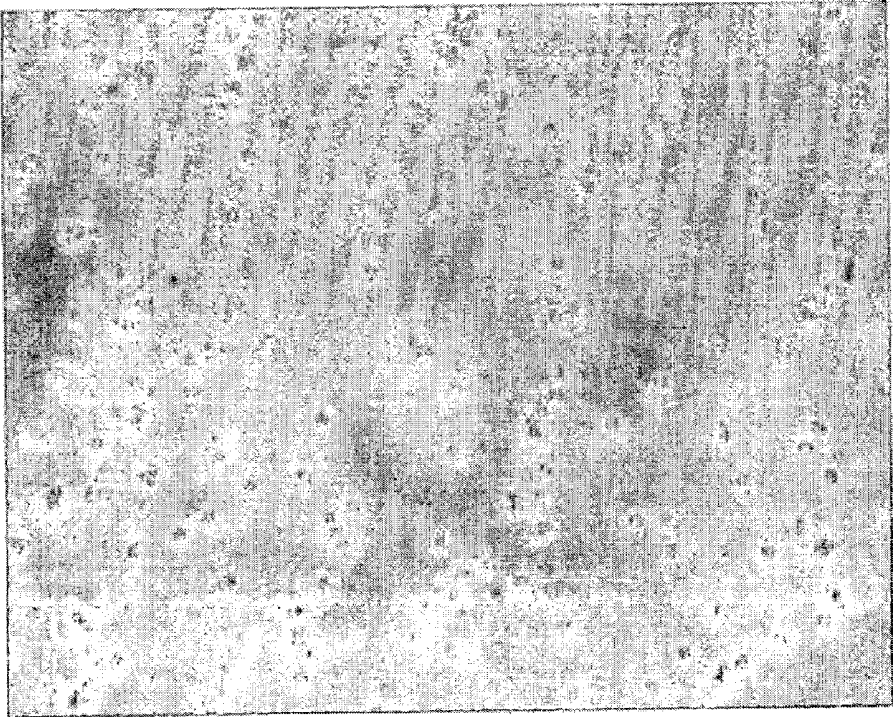
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FIG. 1



FIG. 2



# FIG. 3

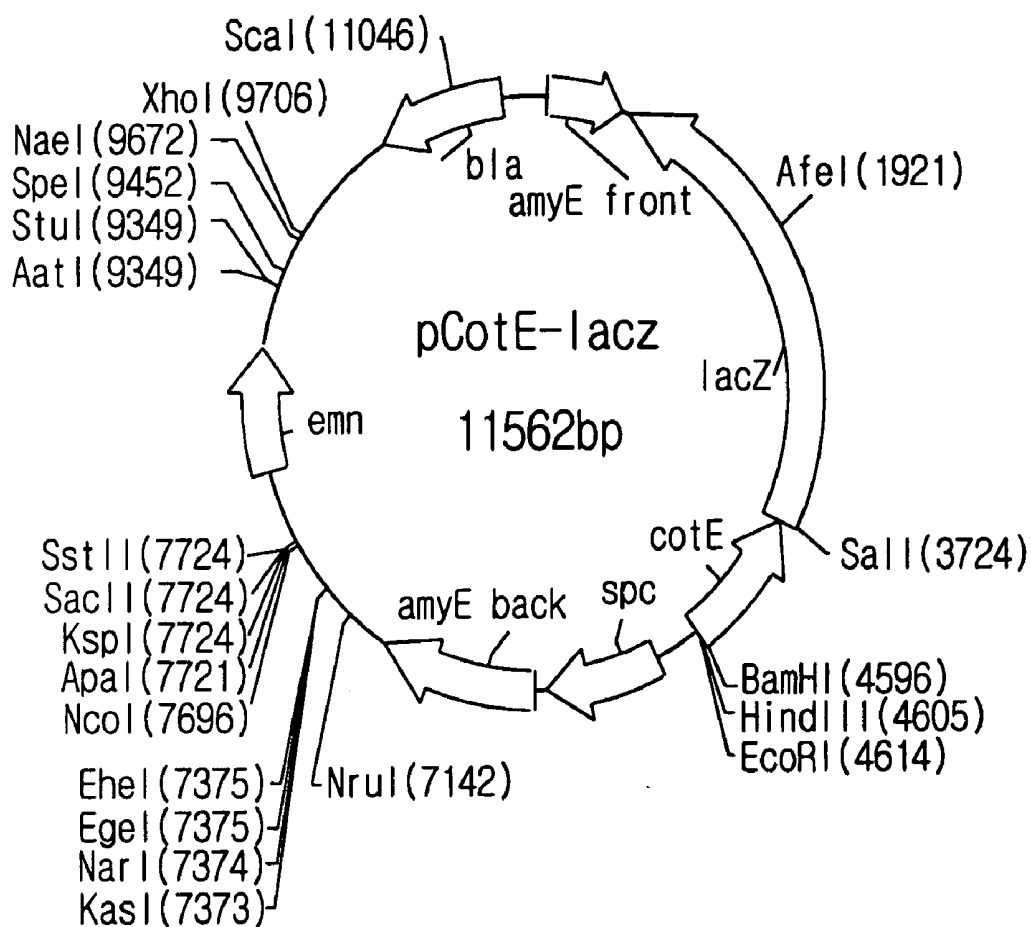
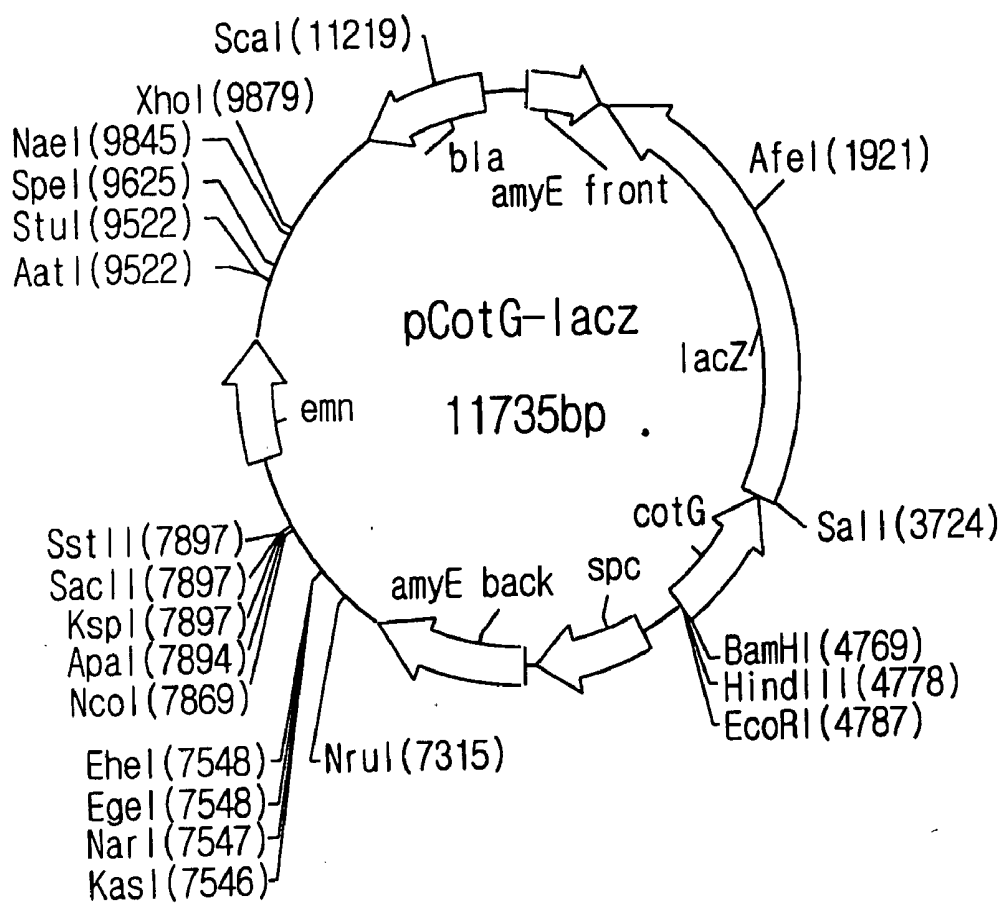
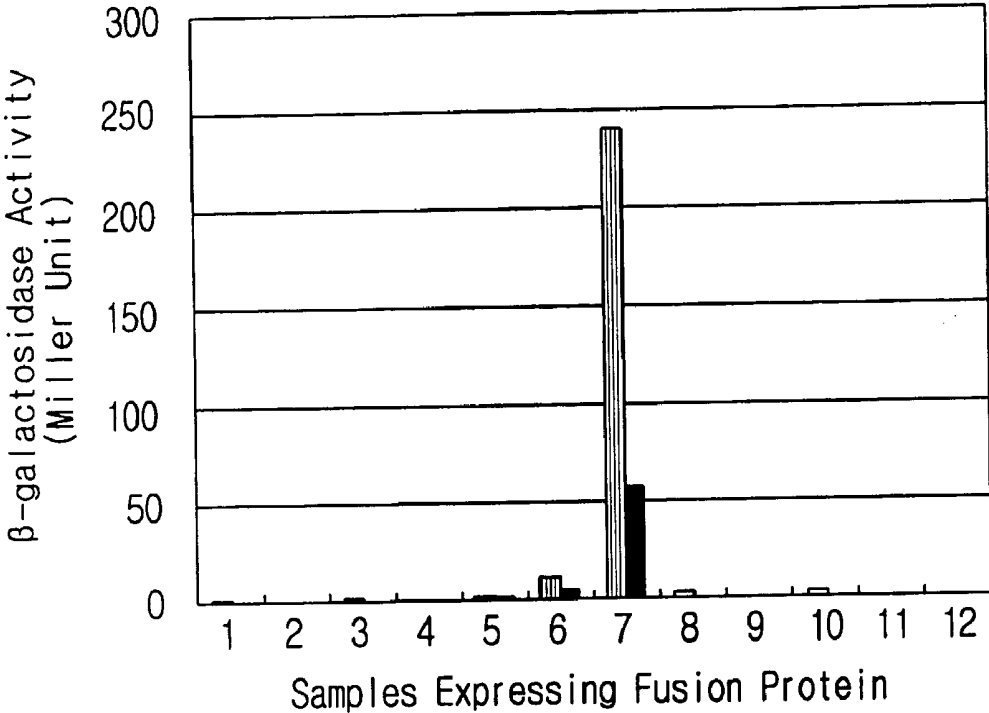


FIG. 4



# FIG.5



# FIG. 6

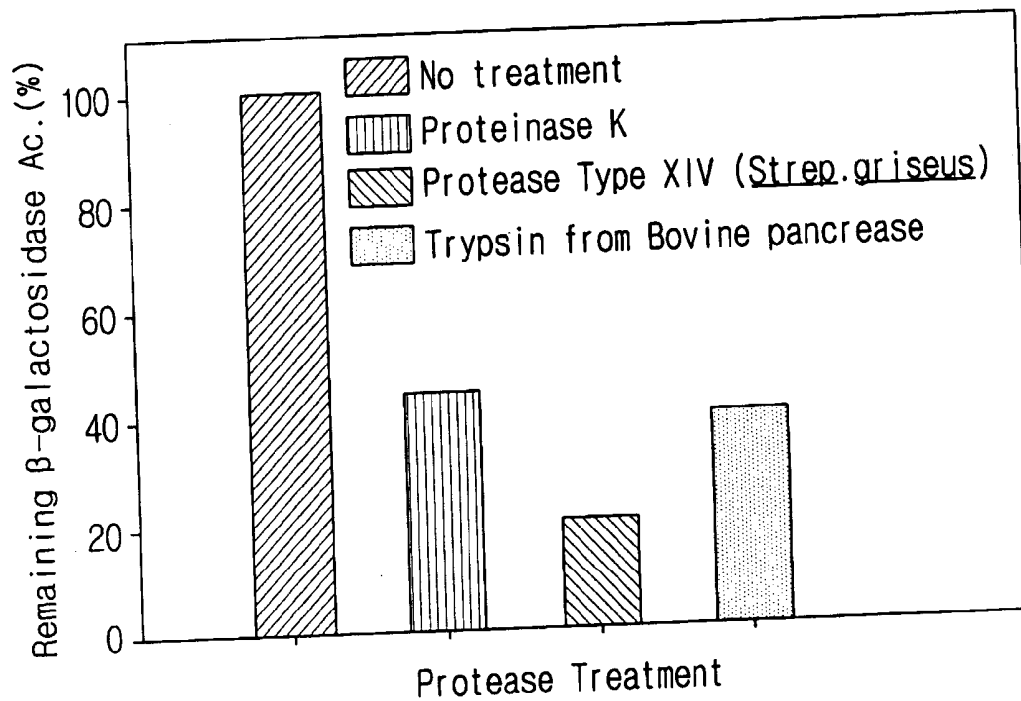
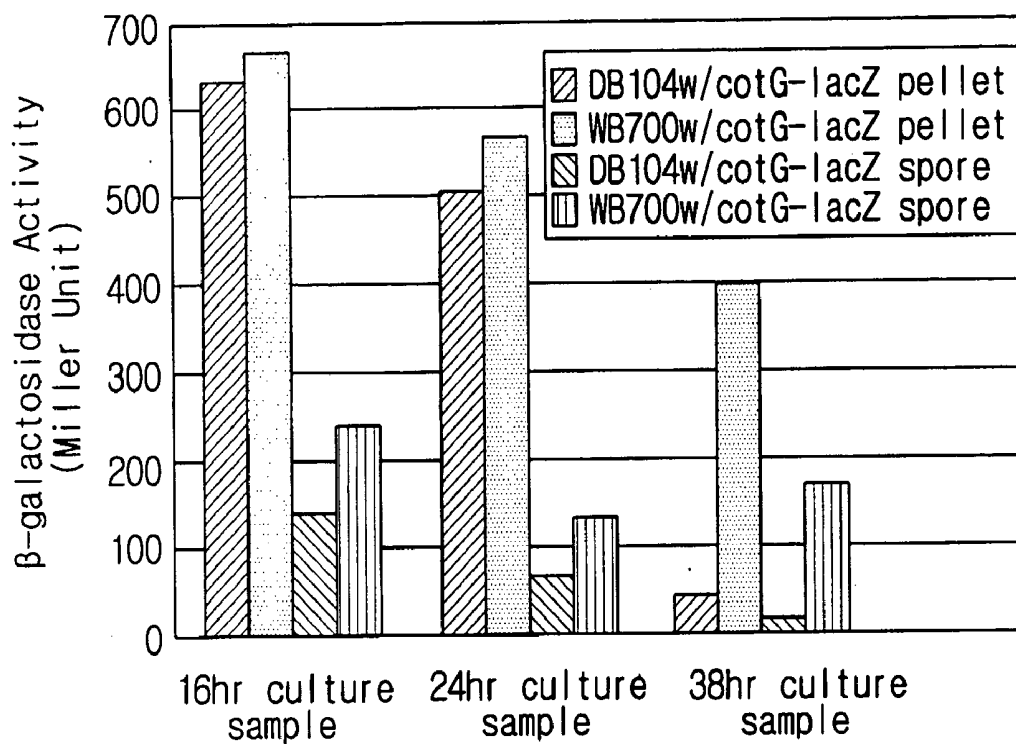
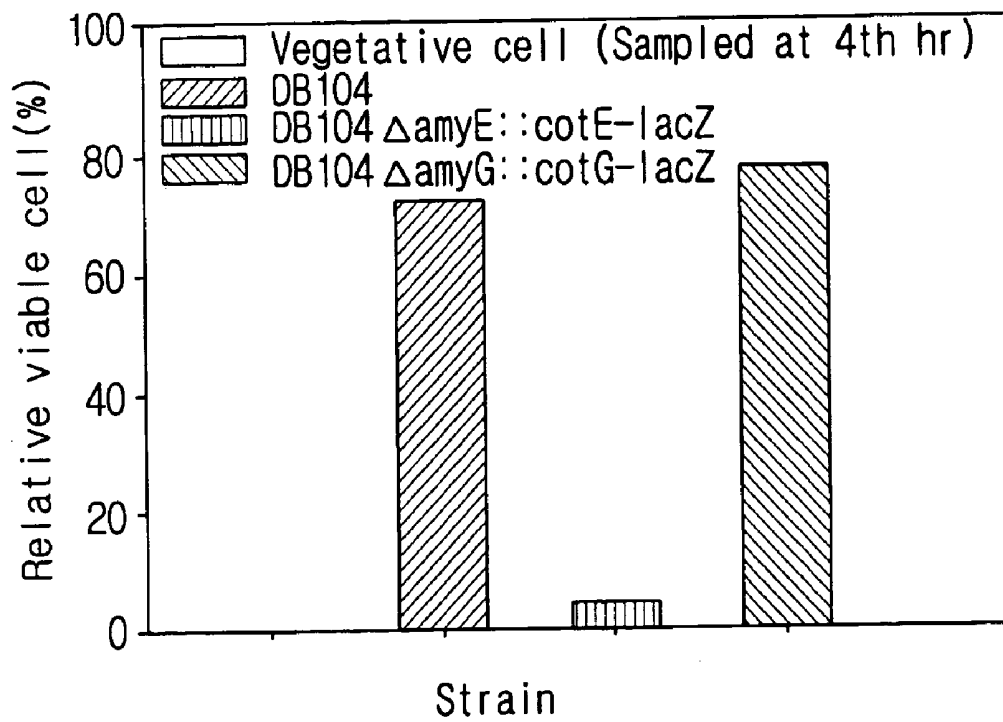


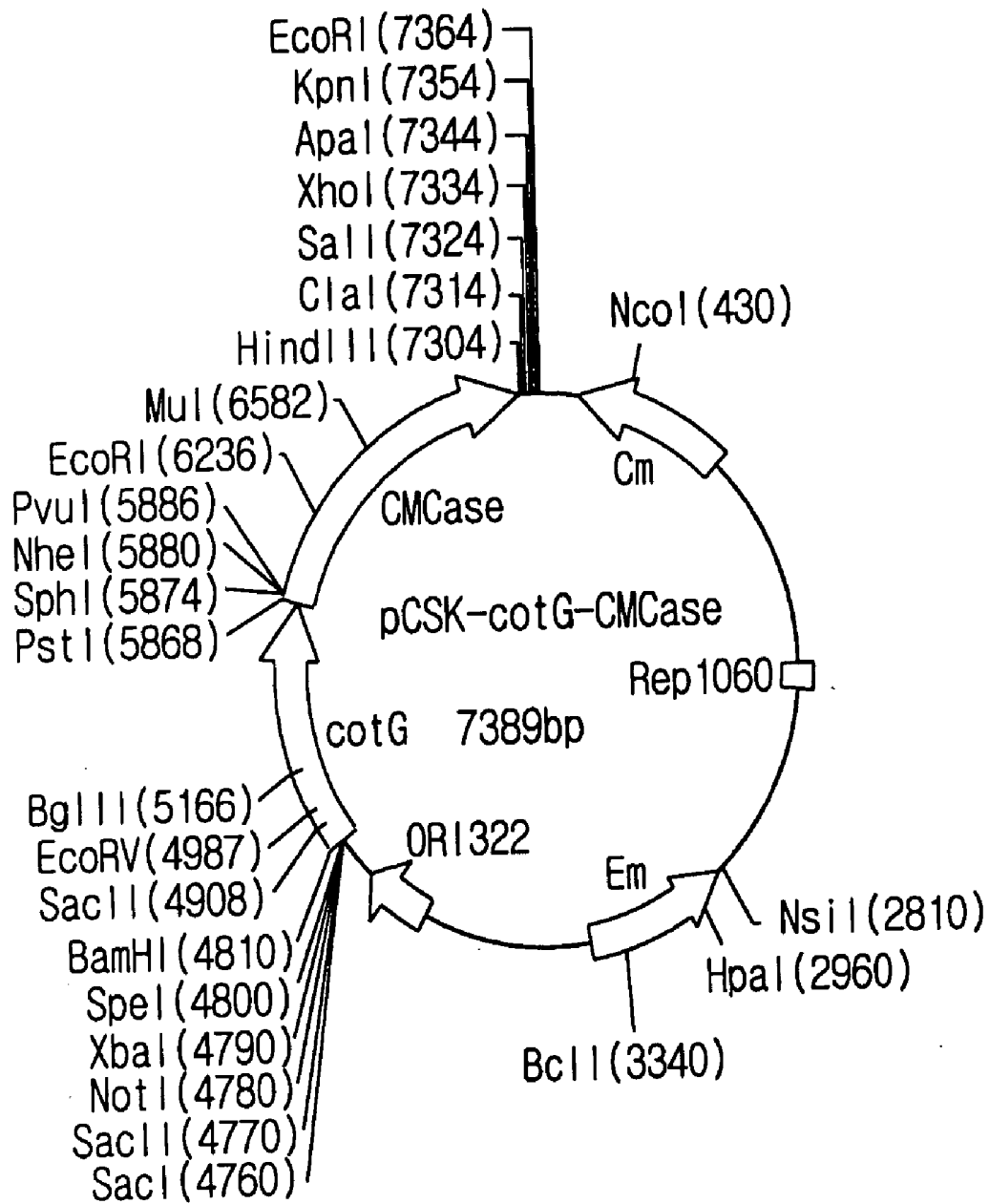
FIG. 7



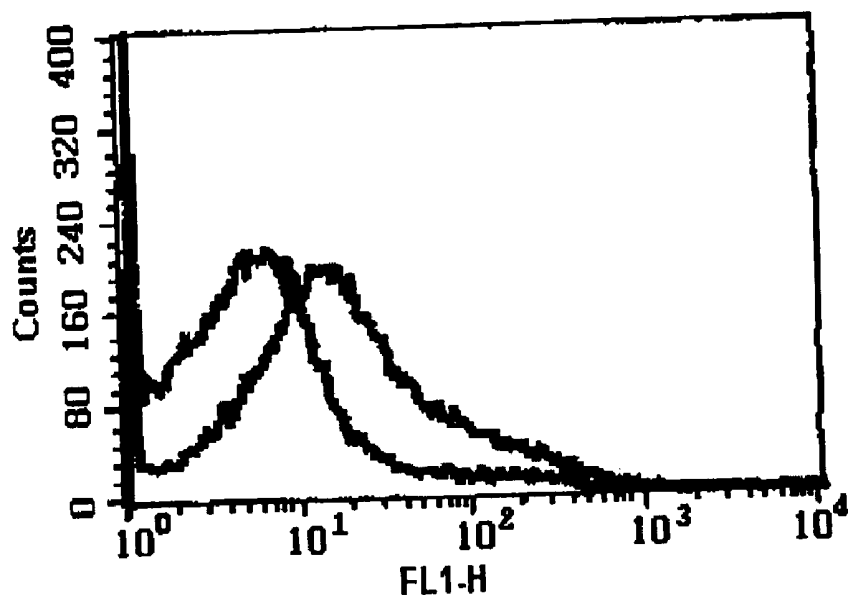
# FIG. 8



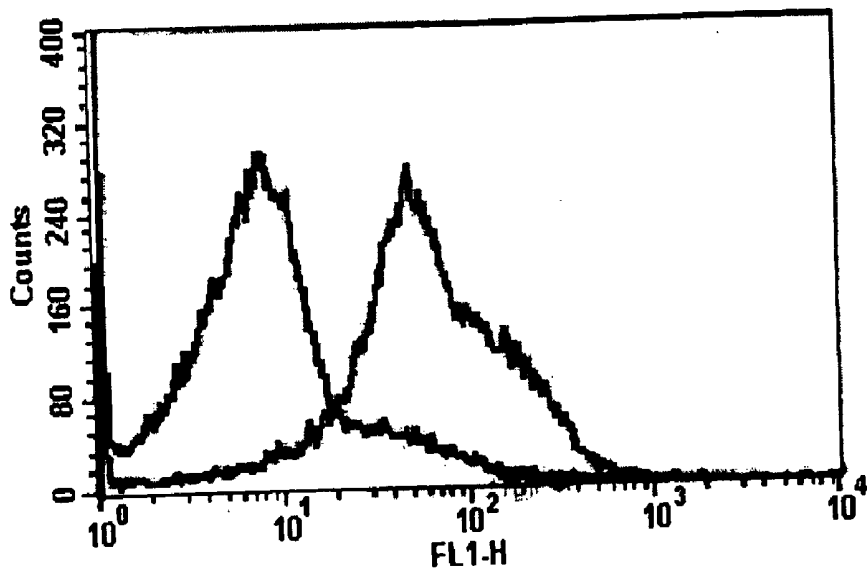
# FIG. 9



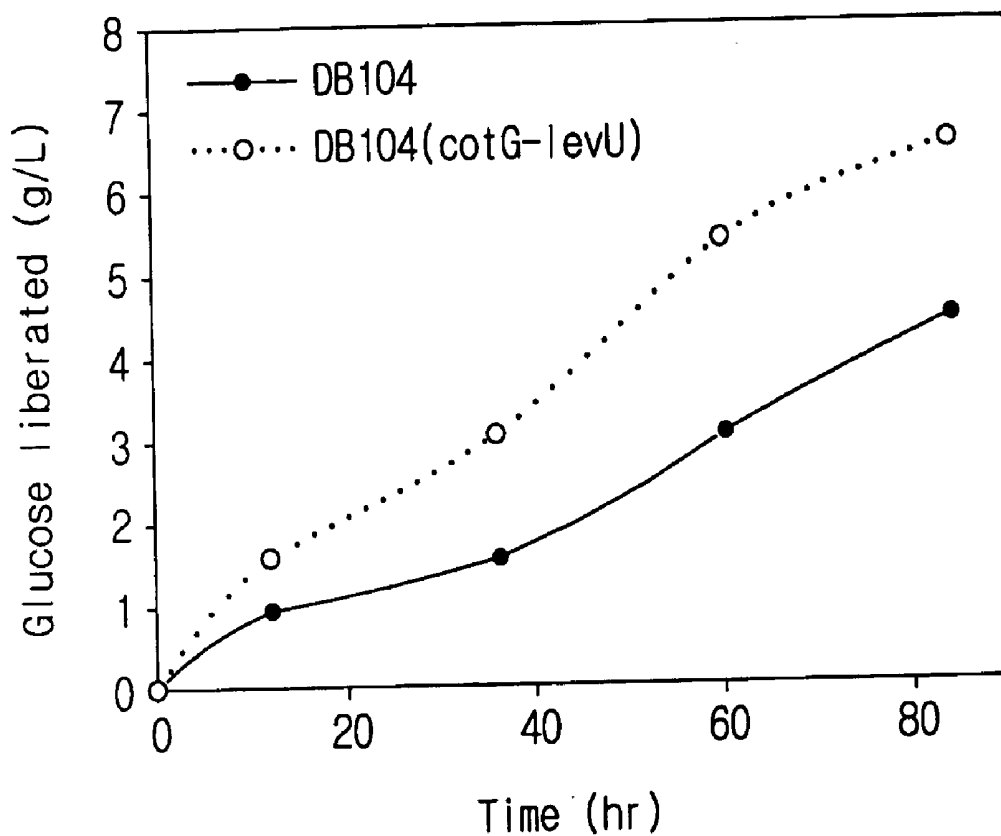
# FIG. 10



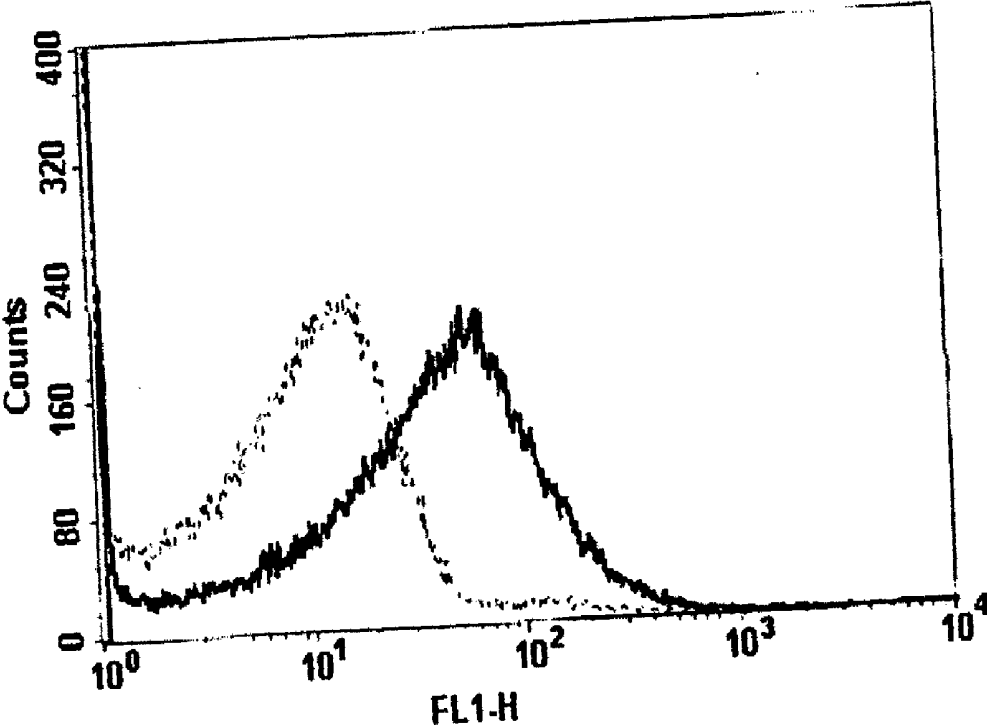
# FIG. 11



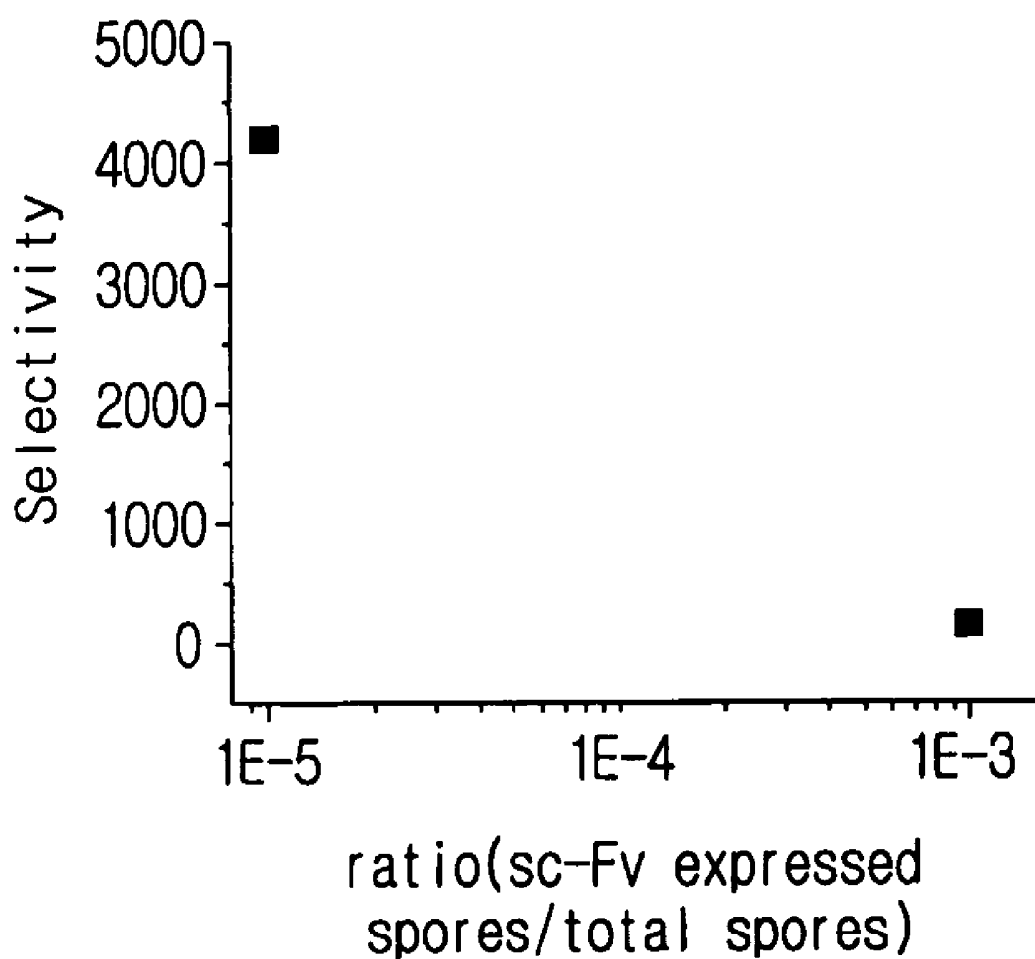
# FIG. 12



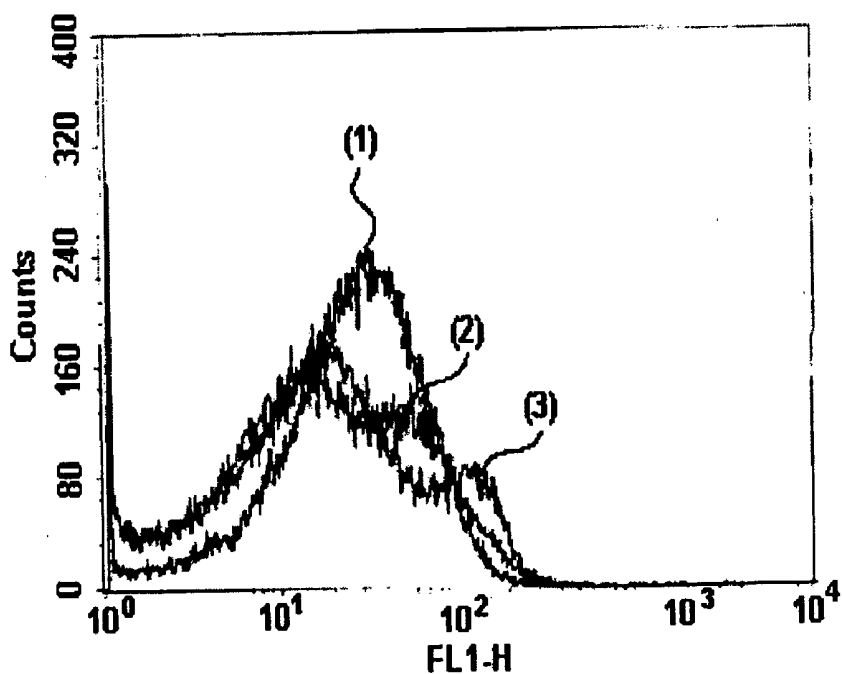
# FIG. 13



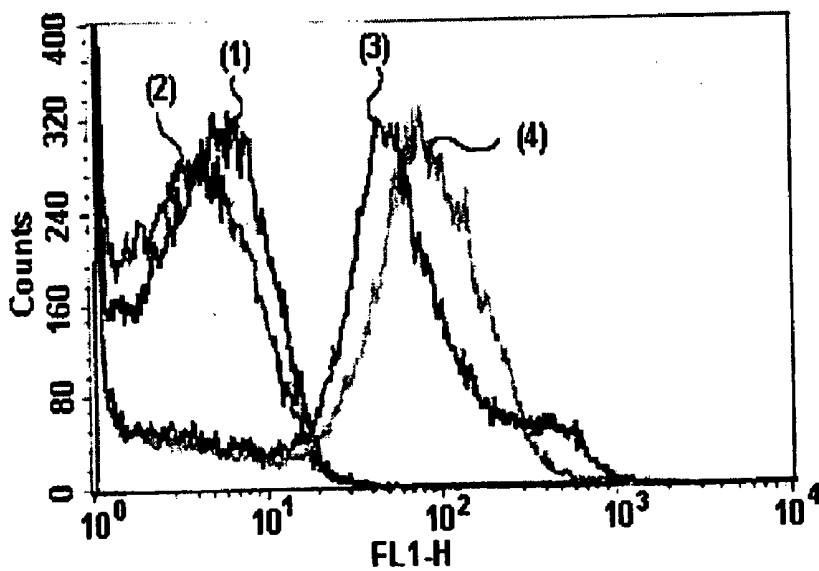
# FIG. 14



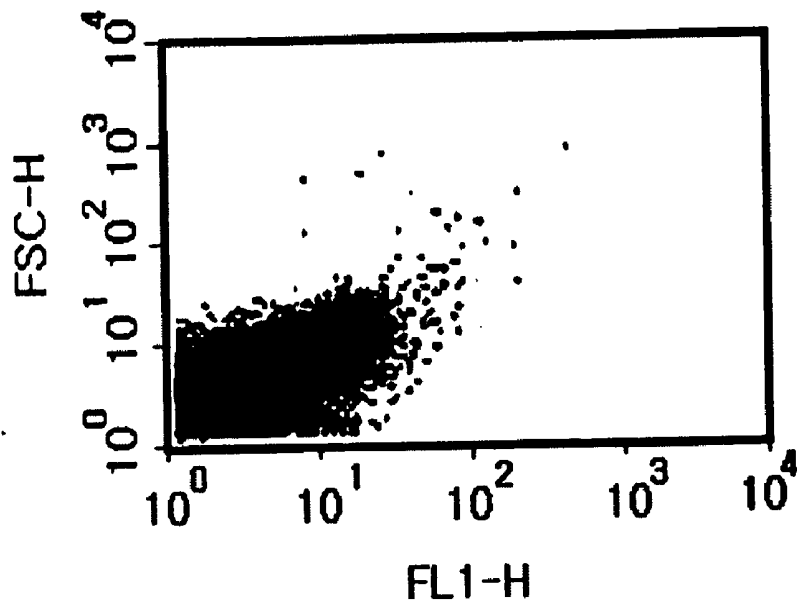
# FIG. 15



# FIG. 16



# FIG.17a



# FIG.17b

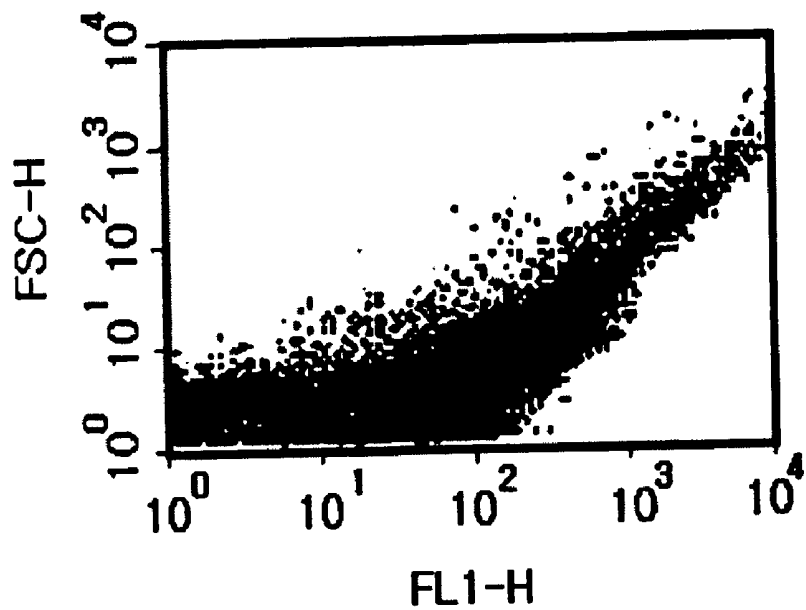


FIG. 17c

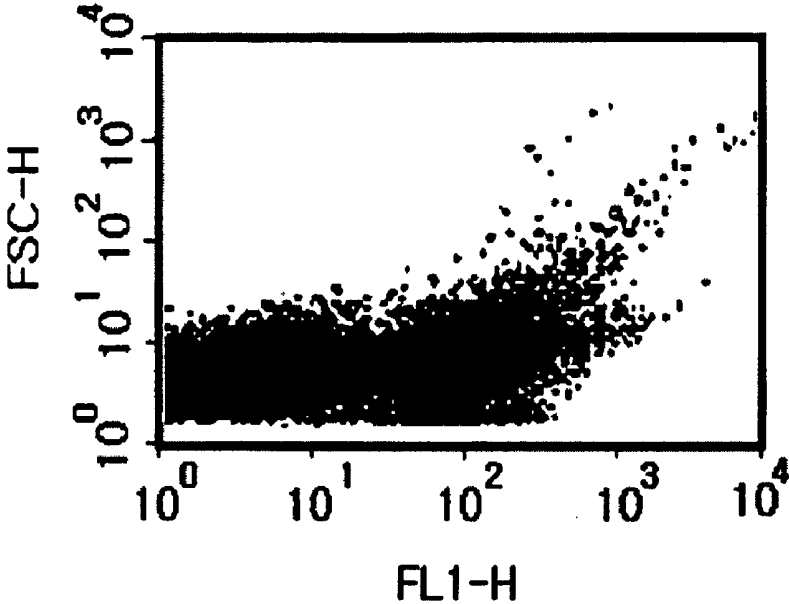
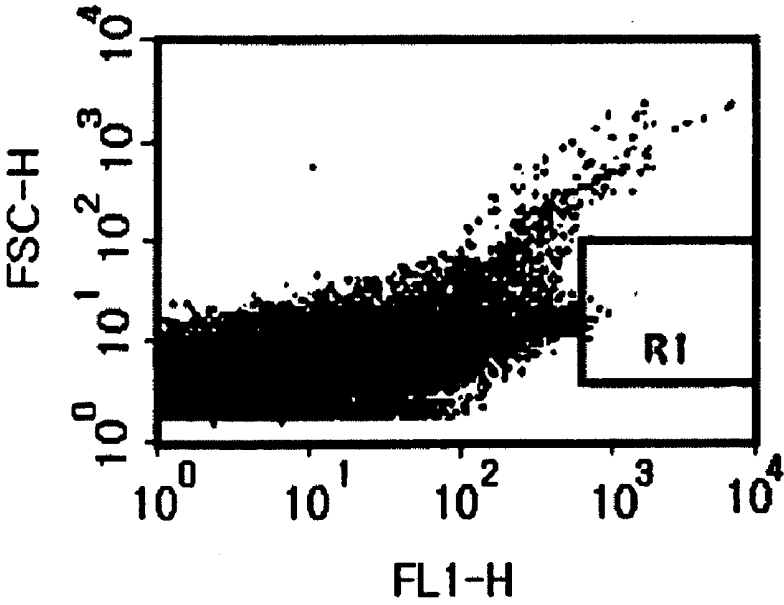


FIG. 17d



## METHOD FOR EXPRESSION OF PROTEINS ON SPORE SURFACE

### FIELD OF THE INVENTION

[0001] The present invention relates to a method for display of proteins on spore surface, in particular to a method for surface display using spore coat proteins as surface display motif and a high throughput method for improving protein.

### DESCRIPTION OF THE RELATED ART

[0002] The technology of surface display in which organism displays on its surface the desired proteinaceous substance such as peptide and polypeptide has wider application fields depending on the types of protein displayed or host organism (Georgiou et al., 1993, 1997; Fischetti et al., 1993; and Schreuder et al., 1996). The gene of protein to be displayed is contained in host organism and thus the host can be selectively screened using the characteristics of the protein displayed, thereby obtaining the desired gene from the selected host with easiness. Therefore, such surface display technology can guarantee a powerful tool on molecular evolution of protein (see WO 9849286; and U.S. Pat. No. 5,837,500).

[0003] High-Throughput Screening

[0004] For instance, phage displaying on its surface antibody having desired binding affinity is bound to immobilized antigen and then eluted, followed by propagating the eluted phage, thereby yielding the gene coding for target antibody from phage (U.S. Pat. No. 5,837,500). The bio panning method described above can provide a tool to select target antibody by surface displaying antibody library on phage surface in large amount and comprises the consecutive steps as follows: (1) constructing library; (2) surface displaying the library; (3) binding to immobilized antigen; (4) eluting the bound phage; finally (5) propagating selected clones.

[0005] The technology of phage surface display has been found to be useful in obtaining the desired monoclonal variant form enormous library (e.g.,  $10^6$ - $10^9$  variants) and thus applied to the field of high-throughput screening of antibody. Antibody has been used in various fields such as therapy, diagnosis, analysis, etc. and thus its demand has been largely increased. In this context, there has been a need for novel antibody to have binding affinity to new substance or catalyze biochemical reaction. The hybridoma technology to produce monoclonal antibody has been conventionally used so as to satisfy the need. However, the conventional method needs high expenditure and long time for performance whereas the yield of antibody is very low. In addition to this, to screen novel antibody, more than 1010 antibody libraries is generally used, as a result, the hybridoma technology has been thought to be inadequate in finding antibody exhibiting new binding property.

[0006] Many researches has focused on novel methods which is easier and more effective that the bio panning method described above and then developed novel technologies performed in such a manner that libraries are displayed on surface of bacteria or yeast and then cells displaying target protein is sorted with flow cytometry in a high-throughput manner. According to the technology, antigen

labeled with fluorescent dye is bound to surface-displaying cell and the antibody having the desired binding affinity is isolated with flow cytometry capable of analyzing more than  $10^8$  cells a hour. Francisco, et al., have demonstrated the usefulness of microbial display technology by revealing that surface-displayed monoclonal antibody could be concentrated with flow cytometry at rate of more than  $10^5$ , finally more than 79% have been proved to be the desired cells (Daugherty et al., 1998).

[0007] Live Vaccine

[0008] The surface display technology mentioned above can display antigen or fragment thereof and hence provide a delivery system for recombinant live vaccine. Up to now, attenuated pathogens or viruses have been predominantly employed as vaccine. Particularly, the bacteria have been found to express antigen intracellularly or extracellularly or on its cell membrane, thereby delivering antigen to host cell. The surface-displayed live vaccine induces a potential immune reaction and expresses continuously antigen during propagation in host cell; therefore, it has been highlighted as novel delivery system for vaccine. In particular, pathogen-derived antigenic epitope displayed on surface of nonpathogenic *E. coli* or Salmonella is administered orally in viable form and then exhibits to induce immune reaction in more continuous and powerful manner (Georgiou et al., 1997; and Lee et al., 2000).

[0009] Whole Cell Bioconversion

[0010] Whole cell as biocatalyst displaying on its surface enzyme capable of catalyzing chemical reaction can avoid necessities for direct expression, isolation and stabilization of enzyme. In case of expressing enzyme in cell for bioconversion, the cell is compelled to recovery and chemical (e.g., toluene) treatment to ensure impermeability of substrate. In addition, the lasting use renders the enzyme inactive or gives a problem on transference of substrate and product, thus dropping the productivity of overall process.

[0011] The above-mentioned shortcomings can be removed using enzyme displayed on cell surface (Jung et al, 1998a: 1998b) With whole cell displaying on its surface phosphodiesterase, organophosphorous-typed parathion and paraoxon with higher toxicity can be degraded, which is a typical example representing the applicability of cells displaying enzyme to environmental purification process (Richins et al., 1997).

[0012] Antipeptide Antibody

[0013] Martineau et al. have reported a highly simple method for production of antipeptide antibody using surface display technology of *E. coli* (Martineau et al., 1991). As described, the desired peptide is displayed on the protruding region of MalE and outer membrane protein, LamB and then whole cell or fragmented cell is administered to animal so as to generate antipeptide antibody. The method makes it possible to produce antibody with avoiding chemical synthesis of peptide and its linkage to carrier protein.

[0014] Whole Cell Absorber

[0015] To immobilize antibody or polypeptide on suitable carrier, which is useful in absorption chromatography, several subsequent steps must be performed, for example, protein production by fermentation, isolation of protein in

pure form, and immobilization on a carrier. Generally, it is difficult to prepare the bioabsorber.

[0016] As absorber, a whole cell displaying absorption protein has been developed. The whole cell absorber known mostly is *Staphylococcus aureus* displaying on its surface protein A naturally, which has a high binding affinity to Fc domain of mammalian antibody. Currently, novel method has been proposed to remove and recover heavy metals, which employs metallothionein or metal-absorption protein displayed on microbial cell surface in large amount (Sousa et al., 1996, 1998; and Samuelson et al., 2000). The method is more effective in removing and recovering heavy metals from contamination source in comparison with the conventional method using metal-absorption microbes.

[0017] As understood based on the matters described above, in order to display foreign protein on cell surface, a suitable surface protein and foreign protein must be linked each other in gene level to express fusion protein, and the fusion protein should pass stably across inner membrane of cell to be attached to cell surface. Preferably, the surface protein having the following characteristics is recommended as surface display motif: 1) existence of secretory signal enabling passage across inner membrane of cell, 2) existence of target signal enabling stable attachment to cell surface, 3) high expression level on cell surface, and 4) stable expression regardless of protein size (Georgiou et al., 1993).

[0018] Therefore, the surface display motif or novel recombinant protein, which meets the requirements described above, should be selected or prepared to develop novel surface display system overcoming disadvantages of the known systems. In addition, the selection of a suitable host cell to display is very pivotal.

[0019] Up to date, the developed surface display systems are as follows: phage surface display system (Chiswell and McCarferty, 1992), bacterial surface display system (Georgiou et al., 1993; Little et al., 1993; and Georgiou et al., 1997), surface display system of Gram negative bacteria (Francisco et al., 1992; Fuchs et al., 1991; Klausner et al., 1990, 1992; and Hedegaard et al., 1989), surface display system of Gram positive bacteria (Samuelson et al., 1995; Palva et al., 1994; and Sleytr and Sara, 1997), and surface display system of yeast (Ferguson, 1988; and Schreuder et al., 1996).

[0020] In the developed phage display system, the concentration of the desired clone from phage library has been found to be difficult and the antibody selected from phage library displaying has usually exhibited very low expression rate. According to a surface display system of Gram negative bacteria, the incorporation of foreign polypeptide into surface structure results in not only its steric limitation which makes it impossible to have stable membrane protein (Charbit et al., 1987; and Agterberg et al., 1990) but also drop of the stability of cell outer membrane and its viability. In addition, in surface display system of yeast, because the vector used has usually shown a low rate of transformation, which is unfavorable to surface display of library.

[0021] The surface display systems developed have been cooperatively used each other. For example, to screen antibody variant with enhanced binding affinity, a primary screening is performed using phage surface display system and additionally, the secondary screening is carried out using

cell surface display system (Georgiou, 2000). However, the phage display technology is encountered to difficulty in concentration of the desired clones from phage library. The reason is that the antibody displayed on phage surface does not show the elution pattern depending exactly on its binding affinity, which is ascribed to avidity of antibody displayed on phage surface. Therefore, there remains a need of novel methods ensuring screening the desired antibody from antibody library.

[0022] *E. coli* as display host, which has been intensively studied, uses generally cell outer membrane protein as surface display motif. However, the over-expression of cell outer membrane protein fused to foreign protein is likely to bring about structural instability of cell outer membrane, consequently, diving the viability of host cell (Georgiou et al., 1996). To be from the shortcomings, ice-nucleation protein with no effect on viability has been used as display motif, and has been applied to bioconversion process, surface display of enzyme library and screening enzyme variants (Jung et al., 1998a, 1998b; and Kim et al., 1998, 1999, 2000).

[0023] The size of library displayed on surface depends on the transformation efficiency of host cell with vector; thus *E. coli* as host has an advantage in view of the size of library to be displayed. Gram positive bacteria as host are relatively rigid and permit stable display of the desired protein; however, transformation efficiency is exhibited low, which results in smaller size of library than *E. coli*.

[0024] The host organisms having been developed are likely to be sensitive to a variety of physiochemical treatments, which makes it impossible to select proteins displayed on surface by virtue of direct physiochemical treatment. For example, in screening a variant of antibody with enhanced binding affinity, abrupt change of pH or adjustment of the concentration of base is generally performed to elute the variant, which are found to decrease the viability of phage or bacteria in medium.

[0025] In addition, the host organisms used conventionally have a complicated and weak structure of cell surface, which drops adaptability to extreme environment such as high temperature and high pressure. To employ *E. coli* displaying on its surface enzyme in bioconversion reaction, the cells must have represent stability in bioconversion system. In this context, the surface of *E. coli* displaying on, its surface enzyme is generally subject to immobilization, which does not lead to satisfying results (Freeman et al., 1998).

[0026] As described above, the known surface display technologies, based on applying fields, have used bacteriophage, Gram negative or positive bacterium, yeast, cilium or mammalian cell as host organism and surface proteins of each organism as surface display motif. However, in the surface display methods having been developed, the host organism does not have resistance to chemicals and physiochemical change such as pH change, and displaying protein on its surface in excess leads to disadvantages in cell surface, finally reducing the viability of host cell largely (Georgiou et al., 1996).

#### DETAILED DESCRIPTION OF THIS INVENTION

[0027] Under such situation, the present inventors have made intensive studies to be from the shortcoming of

conventional display methods, and as a result, we have developed novel display system using a spore as host and a coat protein as motif of surface display. Surprisingly, the developed display system has been found to have excellent stability to a variety of physiochemical stresses in surrounding environment and have much broader applicability.

[0028] Accordingly, it is an object of this invention to provide a method for displaying a protein of interest on spore surface using a system for spore surface display.

[0029] It is another object of this invention to provide a method for improving a protein of interest using a system for spore surface display.

[0030] It is still another object of this invention to provide a method for bioconversion using a system for spore surface display.

[0031] It is further object of this invention to provide a method for preparing protein microarray using a system for spore surface display.

[0032] It is still further object of this invention to provide a method producing an antibody to antigen in vertebrates using a system for spore surface display.

[0033] It is another object of this invention to provide a method for preparing a whole cell absorber using a system for spore surface display.

[0034] It is still another object of this invention to provide a microbial transformant for spore surface display of a protein of interest.

[0035] It is further object of this invention to provide a spore for spore surface display of a protein of interest.

[0036] It is still further object of this invention to provide a vector for spore surface display.

[0037] The principle of the present invention lies in the employment of microbial spore as host for surface display and spore coat protein as surface display motif. The present inventors have been compelled to select a system for spore surface display since the the spore has a following advantages (Driks, 1999): 1) a higher heat stability, 2) a significant stability to radioactivity, 3) a stability to toxins, 4) a higher stability to acid and base, 5) a significant stability to lysozyme, 6) a resistance to dryness, 7) a higher stability to organic solvents, 8) a fusion protein between a surface display motif and a protein of interest is displayed on spore surface immediately after expression without secretion in host cell, 9) no metabolic activity, and 10) shorter time for obtaining spore, e.g. within several hours.

[0038] In particular, the spore coat proteins used in this invention circumvent a necessity for passage across cell membrane, so that they do not need secretion signal and target signal which are prerequisites of surface display motif, thereby ensuring a surface display of protein such as  $\beta$ -galactosidase, in orderly fashion, which is difficult to pass across cell membrane.

[0039] U.S. Pat. No. 5,766,914 discloses a method of producing and purifying enzymes using fusion protein between cotC or cotD among spore coat proteins of *Bacillus subtilis* and lacZ as reporter. However, as disclosed, a purification method for demonstrating surface display of protein is not recognized to isolate spores specifically.

Furthermore, the activity of enzyme expressed has been very low and the display of enzyme on spore surface has never been demonstrated by means of reliable methods such as biochemical, physical and immunological methods. In addition to this, the inner coat protein, cotD is enclosed by outer coat protein of 70-200 nm thickness, which makes it difficult to be exposed to spore surface. In case of fusion protein expression using outer coat protein, cotC, the activity of enzyme is increased by four-fold in comparison with that of cotD; however, the activity, 0.02 U, is considered negligible, in particular, in consideration of industrial scale. Therefore, the matter disclosed in the document above cannot be considered to use and recognize a system for spore surface display. In other words, the patent document cannot be recognized to describe a system for spore surface display. U.S. Pat. Nos. 5,837,500 and 5,800,821 also indicate cotC and cotD as a preferable surface display motif, and therefore the patent documents cannot be recognized to describe a system for spore surface display because of the reasons mentioned above.

[0040] Furthermore, according to the purification method of spore proposed in U.S. Pat. No. 5,766,914, half of the purified resultant has been observed under microscope to have the complex forms between cells harboring spores and cell-lysis matters bound to spores (see FIG. 1; cells with blackish color and long side are those not forming spore and spores is observed to be white and circular), which has been demonstrated by the present inventors. The facts hereinabove reveals possibility to bring about the false results by measuring of the activity of reporter enzyme or analyzing of reporter enzyme with flow cytometry in vegetative cells rather than on spore surface. In contrast, the renografin gradient centrifugation as demonstrated in Examples below allows for the perfect purification of spores (see FIG. 2), thereby measuring the activity of enzyme displayed on spore surface solely.

[0041] Observations on lower enzyme activity in several documents including the patents above are likely to be resulted from the following reasons. First, it is suggested that the expression level of coat protein itself is low. The maximum expression levels of CotC and CotD are 40 and 147 Miller Units, respectively, which is considered to be largely low, in particular, in consideration of CotE of 6021 Miller Units (Zheng L and Losick R., *J. Mol. Biol.* 212:645-660(1990)). Furthermore, it is notable that the amount of enzyme displayed on spore surface has not been reported. Secondly, it is possible that the protein displayed on spore surface is cleaved by protease in host cell. Such suggestion is made based on the fact that at spore-forming stage of *Bacillus subtilis* a variety of proteases are expressed and reconstitution for spore formation is occurred. The suggestion can be demonstrated in Examples below in which a variant lack protease exhibits a much higher enzyme activity displayed on spore surface (see FIG. 7).

[0042] Using gene of GFP (green fluorescence protein) as reporter linked to cotE and spoIVA, the studies on gene expression and localization of the expressed protein in spore has been attempted (Webb et al., 1995; Lewis et al., 1996). The publications disclose that the fusion protein expressed is found in spore by means of observation under fluorescence microscope using fluorescence of GFP; however, they never describe if the fusion protein is displayed and linked on spore surface.

[0043] As another example of spore surface display using coat protein, U.S. Pat. No. 5,800,821 discloses a spore as delivery system of antigen. However, the publication does not disclose that the antigen expressed is displayed on spore surface and the spore containing antigen administered can induce immunization reaction in host.

[0044] The present inventors have recognized the shortcomings of the conventional arts described above and developed an efficient and optimized system for spore surface display, which have been confirmed by enzymological, immunological and physiochemical methods using various spore coat proteins.

[0045] In one aspect of this invention, there is provided a method for displaying a protein of interest on spore surface, which comprises the steps of: (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein of interest, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein of interest; (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the protein of interest on a surface of a spore of the host cell; and (iv) recovering the spore displaying on its surface the protein of interest.

[0046] In another aspect of this invention, there is provided a method for improving a protein of interest, which comprises the steps of: (i) constructing a gene library of the protein of interest; (ii) preparing a vector by linking the gene library to a gene encoding spore coat protein; (iii) transforming a spore-forming host cell with the vector; (iv) forming a spore in the transformed host cell and displaying the protein of interest on a surface of the spore; (v) recovering the spore displaying on its surface the protein of interest; and (vi) screening the spore displaying a variant of the protein of interest having a desired property.

[0047] In still another aspect of this invention, there is provided a method for improving a protein of interest using a resistance property of spore, which comprises the steps of: (i) constructing a gene library of the protein of interest; (ii) preparing a vector by linking the gene library to a gene encoding spore coat protein; (iii) transforming a spore-forming host cell with the vector; (iv) forming a spore in the transformed host cell and displaying the protein of interest on a surface of the spore; (v) treating the spore displaying on its surface the protein of interest with one or more selected from the group consisting of organic solvent, heat, acid, base, oxidant, dryness, surfactant and protease; (vi) recovering the spore displaying on its surface the protein of interest; and (vii) screening the spore displaying a variant of the protein of interest having a resistance to the treatment.

[0048] In further aspect of this invention, there is provided a method for bioconversion, which comprises the steps of: (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein of interest conducting a bioconversion reaction, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein of interest; (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the protein of interest on a surface of a spore of the host cell; (iv) recovering the spore displaying on its surface the protein

of interest; and (v) performing the bioconversion reaction using the spore displaying on its surface the protein of interest.

[0049] In still further aspect of this invention, there is provided a method for preparing protein microarray, which comprises the steps of: (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding antibody or antigen having binding affinity to a protein to be analyzed, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the antibody or antigen; (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the antibody or antigen on a surface of a spore of the host cell; (iv) recovering the spore displaying on its surface the antibody or antigen; and (v) immobilizing onto a solid surface the spore displaying on its surface the antibody or antigen.

[0050] In another aspect of this invention, there is provided a method producing an antibody to antigen in vertebrates, which comprises the steps of: (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding the antigen, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the antigen; (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the antigen on a surface of a spore of the host cell; (iv) recovering the spore displaying on its surface the antigen; and (v) administering to vertebrates a composition containing an immunologically effective amount of the spore displaying on its surface the antigen.

[0051] In still another aspect of this invention, there is provided a method for preparing a whole cell absorber, which comprises the steps of: (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein having a binding affinity to a certain substance, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein; (ii) transforming a host cell with the vector for spore surface display; (iii) displaying the protein on a surface of a spore of the host cell; (iv) recovering the spore displaying on its surface the protein; and (v) immobilizing onto a carrier the spore displaying on its surface the protein.

[0052] According to preferred embodiments of this invention, the gene encoding spore coat protein is derived from a spore-forming Gram negative bacterium including *Myxococcus*; a spore-forming Gram positive bacterium including *Bacillus*; a spore-forming Actinomycete; a spore-forming yeast including *Saccharomyces cerevisiae*, *Candida* and *Hansenulla* or a spore-forming fungus, but not limited to. More preferably, the gene encoding spore coat protein is derived from a spore-forming Gram positive bacterium, most preferably, *Bacillus* including *Bacillus subtilis* and *Bacillus polymyxa*, etc.

[0053] The gene of spore coat protein useful in this invention includes *cotA*, *cotB*, *cotC*, *cotD* (W. Donovan et al., *J. Mol. Biol.*, 196:1-10(1987)), *cotE* (L. Zheng et al., *Genes & Develop.*, 2:1047-1054(1988)), *cotF* (S. Cutting et al., *J. Bacteriol.*, 173:2915-2919(1991)), *cotG*, *cotH*, *cotJA*, *cotJC*, *cotK*, *cotL*, *cotM*, *cotS*, *cotT* (A. Aronson et al., *Mol.*

*Microbiol.*, 3:437-444(1989)), cotV, cotW, cotX, cotY, cotZ (J. Zhang et al., *J. Bacteriol.*, 175:3757-3766(1993)), spoIVA, spoVID and sodA, but not limited to.

[0054] In addition, the gene encoding spore coat protein useful in this invention is a modified form or a recombinant form of one selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, in which the modified form or the recombinant form has a more compatibility for spore surface display relative to wild type genes. The modified form of the gene is obtained by DNA shuffling method (Stemmer, *Nature*, 370: 389-391(1994)), StEP method (Zhao, H., et al., *Nat. Biotechnol.*, 16: 258-261 (1998)), RPR method (Shao, Z., et al., *Nucleic acids Res.*, 26: 681-683 (1998)), molecular breeding method (Ness, J. E., et al., *Nat. Biotechnol.*, 17: 893-896 (1999)), ITCHY method (Lutz S. and Benkovic S., *Current Opinion in Biotechnology*, 11: 319-324 (2000)), error prone PCR (Cadwell, R. C. and Joyce, G. F., *PCR Methods Appl.*, 2: 28-33 (1992)), point mutagenesis (Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor, N.Y., 1989), nucleotide mutagenesis (Smith M. *Annu. Rev. Genet.* 19: 423-462 (1985)), combinatorial cassette mutagenesis (Wells et al., *Gene* 34: 315-323 (1985)) and other suitable random mutagenesis.

[0055] Further to this, the gene encoding spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, in which the gene has a substituted promoter for its promoter to enhance spore surface display relative to wild type genes. The promoter for enhancing surface display, for example, includes the promoters of cotE or cotG genes, which show higher expression level.

[0056] In preferred embodiments of this invention, the gene encoding spore coat protein is selected from the group consisting of cotA, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, more preferably, cotE or cotG and most preferably, cotG.

[0057] According to the present methods, as linking a gene of coat protein and a gene of the protein of interest, the overall sequence, fragments, two or more repeated sequences of the gene of coat protein are useful. In two or more repeated sequences, the repeated sequences may be the same or different each other. The overall sequence, two or more repeated sequences of the gene of the protein of interest are also useful in the fusion sequence. In two or more repeated sequences, the repeated sequences may be the same or different each other. Other combinations also may be useful in the fusion sequence.

[0058] It is understood by one skilled in the art that the gene construct may exist as plasmid in host cell independently or as integrated form into chromosome of host cell. Additionally, in the gene construct, it is recognized by one skilled in the art that the gene of coat protein may be followed or preceded by the gene of the protein of interests Integrated form into the counterpart gene may be useful.

[0059] It is recognized by one skilled in the art that the expression of the fusion protein between coat protein and

protein of interest can be induced by virtue of promoters of coat protein gene and protein of interest or other suitable promoters inducible in host cell

[0060] The present methods is applicable to any protein, for example, including enzyme, enzyme inhibitor, hormone, hormone analogue, hormone receptor, signal transduction protein, antibody, monoclonal antibody, antigen, attachment protein, structural protein, regulatory protein, toxin protein, cytokine, transcription regulatory protein, blood clotting protein, plant defense-inducing protein and fragments thereof. The applicable proteins include multimer as well as monomer. The surface display of multimeric proteins has been rarely reported, for instance, the surface display of alkaline phosphatase in *E. coli*, has resulted the display toward inner part of cell outer membrane (Stathopoulos et al., 1996).  $\beta$ -galactosidase used as reporter enzyme in Examples of the present invention must form tetramer to exhibit its activity and has not been published to be successful in surface display.  $\beta$ -galactosidase generally cannot pass across cell membrane and comprises an amino acid sequence detrimental to cell membrane, as a result, the fusion protein between surface display motif and  $\beta$ -galactosidase has been recognized not to be displayed on cell surface. Therefore, the surface display of  $\beta$ -galactosidase described in Examples proves to be surprising.

[0061] The term used herein "protein" refers to molecule consisting of peptide bond, for example including oligopeptide and polypeptide.

[0062] The host cell suitable in this invention, includes spore-forming Gram negative bacterium including Myxococcus, a spore-forming Gram positive bacterium including Bacillus, a spore-forming Actinomycete, a spore-forming yeast and a spore-forming fungus, but not limited to. Preferably, the host cell is a spore-forming Gram positive bacterium, more preferably, Bacillus. In particular, *Bacillus subtilis* is advantageous in the senses that genetic knowledge and experimental methods on its spore forming as well as culturing method are well known.

[0063] According to the present methods, the spore may be reproductive or non-reproductive. In the method for improving a protein, the recovered coats are subject to reproduction but the methods using a spore as delivery means of protein of interest obviate the necessity for reproduction of spore. It is considerable that the organisms genetically engineered is likely to be regulated under laws and rules; hence non-reproductive spore is preferable. For example, *Bacillus subtilis* lack of cwID gene is preferably used due to being non-reproductive.

[0064] According preferred embodiments of this invention, the recovery of spore is performed in such a manner that the display of the protein of interest on the spore surface is maximized by controlling culture time, after which culturing is terminated and the spore is then recovered. Suitable culture time is varied depending upon the type of cell used, for example, in case of using *Bacillus subtilis* as host, the culture time of 16-25 hours is preferred.

[0065] In the present methods, the recovery of spore may be carried out according to the conventional methods known to one skilled in the art, more preferably, renografin gradients methods (C. R. Harwood, et al., "Molecular Biological Methods for Bacillus." John Wiley & Sons, New York, p.416(1990)).

[0066] As demonstrated in Examples, the stability of spore displaying the foreign protein of interest on its surface is very high in the present invention, indicating maintenance of the integrity of spore surface structure formed by cooperation of coat proteins while the foreign protein is displayed.

[0067] The protein of interest displayed on spore surface according to the present methods can be demonstrated with a wide variety of methods as follows: 1) A primary antibody is bound to the protein of interest displayed on spore surface and then reacted with a secondary antibody labeled with fluorescent chemical to stain the spore, followed by observation with fluorescence microscope or analysis with flow cytometry. 2) The protein of interest displayed on spore surface is treated with protease, followed by measurement of the activity of the protein or detecting lower signal with fluorescence microscope or flow cytometry. 3) In case that the protein of interest uses a substrate with higher molecular weight, the direct measurement of the activity of the protein can provide the level of display since the substrate cannot pass across outer coat of spore.

[0068] In the method for improving protein, the construction of gene library for the protein of interest is performed by a mutagenesis of the gene encoding the protein of interest of wild type, in which the mutagenesis includes DNA shuffling method (Stemmer, *Nature*, 370: 389-391(1994)), StEP method (Zhao, H., et al., *Nat. Biotechnol.*, 16: 258-261 (1998)), RPR method (Shao, Z., et al., *Nucleic acids Res.*, 26: 681-683 (1998)), molecular breeding method (Ness, J. E., et al., *Nat. Biotechnol.*, 17: 893-896 (1999)), ITCHY method (Lutz S. and Benkovic S., *Current Opinion in Biotechnology*, 11: 319-324 (2000)), error prone PCR (Cadwell, R. C. and Joyce, G. F., *PCR Methods Appl.*, 2: 28-33 (1992)), point mutagenesis (Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor, N.Y., 1989), nucleotide mutagenesis (Smith M. *Annu. Rev. Genet.* 19: 423-462 (1985)), combinatorial cassette mutagenesis (Wells et al., *Gene* 34: 315-323 (1985)) and other suitable random mutagenesis.

[0069] In the method for improving protein, the screening is performed in a rapid manner by means of measuring an activity of the protein or flow cytometry (Georgiou, 2000). In case of using an activity of the protein, the screening is carried out by measuring growth of host expressing the protein or colorimetric reaction catalyzed by the protein. In the method for improving protein using a resistance property of spore, the screening is carried out in a rapid manner by virtue of measuring an activity of the protein or using the structural stability of the protein.

[0070] The methods for improving protein provide in a high-throughput manner, from wild type, 1) enzymes catalyzing non-biological reaction (e.g., Diels-Alder condensation), 2) enzymes with non-natural stereoselectivity or regioselectivity, 3) enzymes with activity in organic solvent or organic solvent-aqueous solution two-phase system, and 4) enzymes with activity in extreme conditions such as high temperature or pressure.

[0071] In addition, to select a variant of antibody with enhanced binding affinity, it is general that pH is abruptly changed or the concentration of base is adjusted to elute the variant. In a method using phage or bacteria as carrier, such elution conditions are likely to decrease the viability of

phage or bacteria in medium. However, the methods for improving protein using system of spore surface display overcome the drawback.

[0072] In the meantime, the bioconversion process using surface-displayed enzymes requires a physiochemical stability of surface displaying host in extreme conditions because the process is usually executed in high temperature and/or organic solvent. In particular, a chemical synthesis valuable in current industry is mainly carried out in organic solvent and the synthesis of chiral compound or the resolution of racemic mixture is also performed in highly severe physiochemical conditions. Therefore, the surface-displayed enzyme as well as the organisms displaying enzyme is compelled to have stability in such extreme conditions. In this connection, it is demonstrated that the methods for bioconversion using system for spore surface display is largely advantageous.

[0073] The chemical processes using surface-displayed enzymes have been proposed (Georgiou et al., 1993). However, the proposed processes have generally required immobilization of cell surface with cross-linking agent since the host displaying enzyme is very unstable during process (Freeman et al., 1996). The present bioconversion process is free from the disadvantage mentioned above. Because the surface-displayed enzyme as well as the host displaying enzyme is largely stable, the present method avoids the immobilization. In Examples described hereinafter, the bioconversion reaction with  $\beta$ -galactosidase is exemplified and thus it is understood by one skilled in the art that the present method can be also applied to any type of enzyme such as lipase, protease, cellulase, glycosyltransferase, oxidoreductase and aldolase. In addition, the present method is useful in single step or multi-step reaction and in aqueous or non-aqueous solution. The present bioconversion method employs spore as free or immobilized form and can be performed with other microbes or enzymes.

[0074] Similar to DNA microarray, a protein microarray provides means for analyzing expression or expression level of target protein in certain cell. In order to fabricate protein array, the suitable proteins to be arrayed must be obtained and then immobilized on solid surface. During analysis using protein array, washing step is necessarily performed to remove unbound proteins and various treatments such as high temperature, higher salt concentration and pH adjustment are executed; therefore, it is pivotal to guarantee proteinaceous substance with higher stability in such detrimental environment.

[0075] In addition, the conventional process for preparing protein array needs tedious and repetitive works such as cloning genes of several thousands to tens of thousands of proteins and immobilizing of the proteins expressed. Therefore, there remains a need to improve simplicity and rapidity of the works.

[0076] According to the method for preparing protein microarray of this invention, it is ensured that the works described-above can be performed with much greater readiness. In the present method, a gene construct containing a gene encoding spore coat protein and a gene encoding the desired protein is introduced into host cell and the spore displaying on its surface the desired protein is isolated, followed by immobilization of the isolated spore onto a solid surface. In the method for preparing protein array, the

conventional steps may be used (see Wo 0061806, WO 0054046, U.S. Pat. No. 5,807,754, EP 0818467, WO 9742507, U.S. Pat. No. 5,114,674 and WO 9635953). The protein microarray manufactured by the present invention has a variety of applicable fields including diagnosis, analysis of gene expression, analysis of interaction between proteins, analysis of interaction between protein and ligand, study on metabolism, screening novel or improved enzymes, combinatorial biochemical synthesis and biosensor.

[0077] The solid substrate suitable in the present method includes, but not limited to, glasses (e.g., functionalized glasses), Si, Ge, GaAs, GaP, SiO<sub>2</sub>, SiN<sub>4</sub>, modified silicone nitrocellulose, polyvinylidene fluoride, polystyrene, polytetrafluoroethylene, polycarbonate, nylon, fiber and combinations thereof. The spore optionally may be attached to the array substrate through linker molecules. It is preferred that the regions of the array surface not being spotted are blocked. The amount of spores applied to each spot (or address) depends on the type of array. Interaction between the protein displayed on spore attached to solid substrate and the sample applied can be detected based on their inherent characteristics (e.g., immunogenicity) or can be rendered detectable by being labeled with an independently detectable tag (e.g., fluorescent, luminescent or radioactive molecules, and epitopes). The data generated with protein array of this invention can be analyzed using known computerized systems such as "reader" and "scanner".

[0078] According to the method producing an antibody of this invention, a composition containing an immunologically effective amount of the spore, preferably, further comprises adjuvant such as incomplete and complete Freund's adjuvants. In the present method, the mode of administration is, preferably, injection and more preferably, intravenous, intraperitoneal, subcutaneous and intramuscular injections. Boosting within suitable period after the first administration is preferable to yield a sufficient amount of antibody.

[0079] Meanwhile, in the process for preparing absorption chromatography, antibody or polypeptide is produced, purified and immobilized on a carrier. Generally, it is very difficult to prepare the bioabsorbers. The disadvantage may be overcome using whole cell displaying protein as described in Georgiou et al., 1997. Therefore, the system for spore surface display of this invention provides a whole cell absorber to solve the problems of the known absorbers.

[0080] In further aspect of this invention, there is provided a microbial transformant for spore surface display of a protein of interest, characterized in that the transformant is produced by transformation with a vector for spore surface display containing (i) a gene encoding a protein of interest and (ii) a gene encoding spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sods, in which when expressed, a fusion protein between the spore coat protein and the protein of interest is expressed.

[0081] According to preferred embodiment, the transformant is derived from a variant mutated to enhance spore surface display. For example, the mutation to enhance spore surface display eliminates a production of extracellular secretory protease in the transformant, so that the protein of interest displayed on spore surface is stably maintained. In

addition, the mutation to enhance spore surface display eliminates a production of intracellular protease in the transformant. It is also preferred that a gene or genes involved in spore forming is subject to mutation in order to the rate of spore forming (Perego, M., et al., *Mol. Microbiol.* 19: 1151-1157 (1996)).

[0082] In still further aspect of this invention, there is provide a spore for spore surface display of a protein of interest, characterized in that the spore displays the protein of interest on its surface.

[0083] According to the present invention, the spore may be reproductive or non-reproductive one which is selected based on its application field. Preferably, the non-reproductive spore can be obtained by virtue of one or more methods selected from the group consisting of genetic method (Popham D. L., et al., *J. Bacteriol.*, 181: 6205-6209 (1999)), chemical method (Setlow T. R., et al., *J. Appl. Microbiol.*, 89: 330-338 (2000)) and physical method (Munakata N, et al., *Photochem. Photobiol.*, 54: 761-768 (1991)). The genetic method to make the spore non-reproductive is accomplished by, for example, deleting a gene of host cell involved in reproduction of spore.

[0084] In the present invention, it is preferred that the spore is derived from a variant mutated to increase its agglutination property because in bioconversion performed in industrial scale, the separation between the resulting product and spores is rendered easier. The increase of the agglutination property in the spore is accomplished by one or more methods selected from the group consisting of genetic method, chemical method and physical method. As example of the physical method, the heat treatment can be proposed (Wiencek K. M., et. al., *Appl. Environ. Microbiol.*, 56: 2600-2605 (1990)).

[0085] In another aspect of this invention, there is provided a vector for spore surface display, characterized in that the vector comprises a replication origin, an antibiotic-resistance gene, a restriction site, a gene encoding a spore coat protein, a gene encoding a protein of interest and a promoter operatively linked to the gene encoding a spore coat protein, in which when expressed, a fusion protein between the spore coat protein and the protein of interest is expressed.

[0086] According to preferred embodiment, the gene encoding a spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cobZ, spoYA, spoVID and sodA, more preferably, cotE or cotG, and most preferably, cotG.

[0087] In the vector of this invention, the replication origin can include various origins known to one skilled in the art, for example, when the vector is introduced into a spore-forming yeast, 2 $\mu$ , ARS, ARS1 or ARS2 can be used as replication origin. In case of using Bacillus as host, ori 322, ColE1 origin, Rep1060, etc. can be used. The antibiotic-resistance gene used as selective marker, when prokaryote such as Bacillus is used as host, is a resistance gene to

antibiotics acting to prokaryotes, for example, including kanamycin, ampicillin, carbenicillin, chloramphenicol, streptomycin, geneticin, neomycin and tetracycline. The promoter used in the present vector includes a promoter of the gene of spore coat protein and a known promoter operable in host cell.

#### BRIEF DESCRIPTION OF THE DRAWINGS

[0088] FIG. 1 is a microscopic photograph showing spores of *Bacillus subtilis* purified according to method described in U.S. Pat. No. 5,766,914;

[0089] FIG. 2 is a microscopic photograph showing spores of *Bacillus subtilis* purified according to renografin gradients method;

[0090] FIG. 3 is a genetic map of the recombinant vector pCotE-lacZ of the present invention;

[0091] FIG. 4 is a genetic map of the recombinant vector pCotG-lacZ of the present invention;

[0092] FIG. 5 represents screening results demonstrating the preferred surface display motif in the present invention;

[0093] FIG. 6 is a graph showing the affect of protease to  $\beta$ -galactosidase displayed on spore surface;

[0094] FIG. 7 is a graph showing the activity of  $\beta$ -galactosidase displayed on spore surface in accordance with culture time;

[0095] FIG. 8 is a graph representing the heat stability of *Bacillus subtilis* DB104 strain displaying on its surface the protein;

[0096] FIG. 9 is a genetic map of recombinant vector pCSK-cotG-CMCase of this invention;

[0097] FIG. 10 is a graph showing analysis of spore surface-displayed carboxymethylcellulase using flow cytometry;

[0098] FIG. 11 is a graph showing analysis of spore surface-displayed levansucrase using flow cytometry;

[0099] FIG. 12 is a graph showing the activity of spore surface-displayed levansucrase;

[0100] FIG. 13 is a graph representing analysis of spore surface-displayed monoclonal antibody using flow cytometry;

[0101] FIG. 14 is a graph demonstrating selectivity to spore displaying single chain Fv;

[0102] FIG. 15 is a graph representing analysis with flow cytometry of monoclonal antibody library to have binding affinity to Pre-S region of hepatitis B virus;

[0103] FIG. 16 is a graph showing analysis of spore surface-displayed GFP using flow cytometry; and

[0104] FIGS. 17a to 17d are graphs representing isolation with flow cytometry of spores displaying improved GFP.

[0105] The following specific examples are intended to be illustrative of the invention and should not be construed as limiting the scope of the invention as defined by appended claims.

## EXAMPLES

### Example I

#### Isolation of the Gene Encoding Coat Proteins

[0106] I-1: Construction of the Vector for Spore Surface Display

[0107] To isolate the most appropriate coat protein for spore surface display among coat proteins consisting of spore, the recombinant vector having the gene encoding a fusion protein between coat protein and  $\beta$ -galactosidase was constructed as follow:

[0108] To begin with, the DNA was extracted from the *Bacillus subtilis* 168 strain provided from Dr. F. Kunst (Kunst F., et al., *Nature*, 390: 249-256(1997)) by Kalman's method (Kalman S., et al., *Appl. Environ. Microbiol.* 59, 1131-1137(1993)), and the purified DNA was served as template for PCR to spoIVA primers (SEQ ID NOs: 1 and 2), cotB primers (SEQ ID NOs: 3 and 4), cotC primers (SEQ ID NOs: 5 and 6), cotD primers (SEQ ID NOs: 7 and 8), cotE primers (SEQ ID NOs: 9 and 10), cotG primers (SEQ ID NOs: 11 and 12), cotH primers (SEQ ID NOs: 13 and 14), cotM primers (SEQ ID NOs: 15 and 16), cotV primers (SEQ ID NOs: 17 and 18), cotX primers (SEQ ID NOs: 19 and 20) and cotY primers (SEQ ID NOs: 21 and 22). Taq polymerase purchased from Boehringer Mannheim was used for total 35 cycles of PCR under condition of denaturation for 30 sec at 94° C., annealing for 30 sec at 55° C. and extension for 1 min at 72° C.

[0109] After then, each amplified PCR products were digested with BamHI and Sall and cloned between BamHI and Sall sites of plasmid pDG1728 which is a gratuitous gift by Dr. P. Stragier (Geurout-Fleury, A. M., et al., *Gene*, 180: 57-61(1996)), thus the constructed vectors express the fusion protein of coat protein and  $\beta$ -galactosidase. FIG. 3a shows the genetic map of pCotE-lacZ expressing fusion protein of CotE protein and  $\beta$ -galactosidase and FIG. 3b shows the genetic map of pCotG-lacZ expressing Fusion protein of CotG protein and  $\beta$ -galactosidase.

[0110] SEQ ID NO:23 shows the sequence of cotE-lacZ fused genes and SEQ ID NO:24 shows the amino acid sequence of CotE-LacZ fusion protein. In SEQ ID NO:23, promoter for cotE is 1-329, CotE structural gene is 330-872, restriction site is 873-878 and LacZ structural gene is 879-3902.

[0111] SEQ ID NO:25 shows the sequence of cotG-lacZ fused genes and SEQ ID NO:26 shows the amino acid sequence of CotG-LacZ fusion protein. In SEQ ID NO:25, promoter of cotG is 1-460, CotE structural gene is 461-1045, restriction site is 1046-1051 and LacZ structural gene is 1052-4075.

[0112] I-2: Pure Isolation of Spores

[0113] Constructed recombinant expression vectors were transformed into *Bacillus subtilis* DB104 (Kawamura F. and Doi R. H., *J. Bacteriol.* 160: 442-444(1984)) using natural transformation (C. R. Harwood, et al., *Molecular Biological Methods for Bacillus*, John Wiley & Sons, New York, p.416(1990)).

[0114] Other methods such as conjugation or transduction can be applied for introduction of the recombinant vectors into *Bacillus* strain.

[0115] Subsequently, each *Bacillus* strain comprising the fused gene between coat protein and  $\beta$ -galactosidase was cultured for 24 hr at a shaking incubator (37° C., 250 rpm) in GYS medium ((NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> 2 g/l, Yeast extract 2 g/l, K<sub>2</sub>HPO<sub>4</sub> 0.5 g/l, glucose 1 g/l, MgSO<sub>4</sub>·5H<sub>2</sub>O 0.07 g/l), and the only pure spores were isolated using renografin gradients method (C. R. Harwood, et al., "Molecular Biological Methods for *Bacillus*." John Wiley & Sons, New York, p.416(1990)).

#### [0116] I-3: Display of Proteins on Spore Surface

[0117] The spores isolated in the above-described Example and the cell pellet of *Bacillus subtilis* DB104 were subjected to evaluation of the activity of  $\beta$ -galactosidase using Miller's method (Miller, "Experiments in Molecular Genetics", Cold Spring Harbor, N.Y.: Cold Spring Harbor Laboratory, p.352-355(1972)) and the results are shown in FIG. 5. In FIG. 5, the gray bar indicates cell pellet, the black bar indicates the activity of  $\beta$ -galactosidase in purely isolated spores and '1' relates to result of control *Bacillus subtilis* DB104; '2' to result of SpoIVA-LacZ; '3' to result of CotB-LacZ; '4' to result of CotC-LacZ; '5' to result of CotD-LacZ; '6' to result of CotE-LacZ; '7' to result of CotG-LacZ; '8' to result of CotH-LacZ; '9' to result of CotM-LacZ; '10' to result of CotV-LacZ; '11' to result of CotX-LacZ; and '12' to result of CotY-LacZ fusion protein, respectively.

[0118] As shown in FIG. 5, it is known that Deits TL (U.S. Pat. No. 5,766,914) fails to induce the sufficient surface display of cotC and cotD since the expression levels of cotC and cotD are as low as the control. However, the expression level of cotE and cotG are comparatively high and especially, expression level of cotG is remarkably high comparing to other coat proteins. In addition, in the isolated spores, the surface display using cotG shows the highest enzyme activity, which demonstrates that CotG-LacZ fusion proteins are the highest level of display on spore surface.

[0119] Considering the expression level and the amount of fusion proteins displayed on spore surface, it is known that the cotG is the most preferable surface display motif. It is known to one skilled in the art that these results exclude other coat proteins other than cotG from applying to spore surface display.

#### [0120] I-4: Effect of Proteases on the Surface-Displayed Enzymes

[0121] To confirm whether the surface-displayed  $\beta$ -galactosidase is degraded or not, the purely isolated spore displaying CotG-LacZ was resuspended into 100  $\mu$ l of PBS solution, and each 10 mg/ml of protease K, protease type XIV or trypsin was treated. Thereafter, the activity of  $\beta$ -galactosidase was measured as described above and the results are shown in FIG. 6. As shown in FIG. 6, the activity of spore surface-displayed  $\beta$ -galactosidase is decreased with some variations in each result. These results give the evidence for the localization of  $\beta$ -galactosidase on spore surface.

[0122] DB104 strain lacking neutral and alkaline protease and WB700 strain (Ye, R., et al., *Biotechnology and Bioengineering*, 62:87-96(1999)) lacking 7 proteases among proteases secreted from *Bacillus subtilis* were transformed with the pCotG-lacZ expression vector using natural trans-

formation method as described in example I-1, and the activity of  $\beta$ -galactosidase in cell pellet and spores was measured as described in example I-3 (FIG. 7). As shown in FIG. 7, while the enzyme activity is abruptly decreased in DB104 strain as time goes, WB700 strain shows slight decrease in enzyme activity. These results indicate that the displayed  $\beta$ -galactosidases on spore surface are degraded in DB104 strain by the proteases secreted extracellularly; however, the displayed  $\beta$ -galactosidases in WB700 are stably maintained because of lack of the proteases secreted extracellularly. Therefore, the results also support the localization of  $\beta$ -galactosidase on spore surface.

#### Example II

##### Spore Production Depending on Culture Time

[0123] As shown in FIG. 7, it is required to stop incubation on a specific time point and isolate spores. In DB104, the enzyme activity of spores after 38 hr of incubation is significantly low comparing to that after 24 hr of incubation. Thus, it is demonstrated that the adjustment of incubation time makes it possible to yield spores displaying enzyme on its surface with the greatest enzyme activity.

#### Example III

##### Characterization of Spores Displaying $\beta$ -Galactosidase

[0124] Heat resistance was measured as follow in spores displaying  $\beta$ -galactosidase: 100  $\mu$ l of spores isolated by renografin gradients in Example I-2 were heated for 15 min and then spread on LB plates to evaluate viability of spores (FIG. 8). As shown in FIG. 8, spores displaying CotG-LacZ show similar heat resistance to spores without surface protein. In a result, the display of the foreign protein fused to coat protein on spore surface does not affect on its inherent characteristics such as heat resistance. Moreover, these results provide the promising usage of spore displaying on its surface enzyme in chemical reactions at high temperature. In addition, from these results, it is suggested that the spores transformed according to the present invention remain their inherent resistances to lysozyme, a bacterial cell wall-degrading enzyme and solvent.

#### Example IV

##### Displaying Various Enzymes on Spore Surface

##### [0125] IV-1: Construction of Recombinant Vectors

[0126] To use spores displaying various enzymes, it is prerequisite to confirm whether various enzymes in addition to  $\beta$ -galactosidase can be surface-displayed. Firstly, plasmid pHPS9 (Haima, et al., *Gene*, 86:63-69(1990)) was digested by EcoRI and HindIII and manipulated into blunt ends using Klenow enzyme. Then, DNA fragment containing multiple cloning sites, which was obtained from plasmid p123T (EMBL Z46733) with BssHII, was ligated to the blunt-ended pHPS9 plasmid to use as virgin vector named pCSK1 in the following experiments. The pCSK-cotG plasmid was prepared by restricting PCSK1 plasmid with BamHI and PstI and ligating PCR-amplified cotG gene. In the course of PCR for cotG gene amplification, a linker between cotG gene and target gene was incorporated using cotG-linker 5

primer (SEQ ID NO:27) and 3 primer (SEQ ID NO:12) with template of DNA in *Bacillus subtilis*.

[0127] In other experiments, genes encoding carboxymethyl cellulase, levansucrase and lipase was prepared as follows: Carboxymethyl cellulase cloned in pBS1 plasmid (S. H. Park et al., *Agric. Biol. Chem.*, 55: 441-448(1991)) was directly employed. The pBS1 plasmid contains the gene encoding carboxymethylcellulase cloned from *Bacillus subtilis* BSE616 strain. In the present Example, PCR was performed with the pBS1 as template using primer represented by SEQ ID NOs:28 and 29. In the case of PCR for levansucrase, pSST110 plasmid (Jung, H.-C., et al., *Nat. Biotech.*, 16: 576-580(1998)) was used as template and primers represented by SEQ ID NOs:30 and 31 were used. In PCR for lipase, pTOTAL (Ahn, J.-H., et al., *J. Bacteriol.*, 181: 1847-1852(1999)) was added as template and primers of SEQ ID NOs: 32 and 33 were used. All PCRs were performed in the same condition as described in Example I-1.

[0128] Recombinant vectors containing gene coding for fusion between CotG and the carboxymethylcellulase, levansucrase or lipase were prepared by cloning into pCSK-cotG using PstI and BamHI restriction enzymes both in vector and in the PCR-amplified inserts. As an example of the above construction, FIG. 9 shows pCSK-cotG-CMCase which is the recombinant vector encoding fusion protein between CotG and carboxymethylcellulase. Transformed *Bacillus subtilis* DB104 with pCSK-cotG-CMCase was named *Bacillus subtilis* GFSD18 and deposited at the Korean Collection for Type Cultures (KCTC, KR) with accession No. KCTC 0887BP (Nov. 16, 2000).

[0129] SEQ ID NO:34 shows nucleotide sequence of fused cotG-CMCase genes and SEQ ID NO:35 shows amino acid sequence of CotG-CMCase encoded by SEQ ID NO:34. In SEQ ID NO:34, promoter for cotG is 1-460, structural gene for CotG is 461-1045, linker is 1046-1084, and structural gene for CMCase is 1085-2491.

[0130] IV-2: Expression of Recombinant Vectors and Verification

[0131] The above-prepared recombinant vectors were employed for transformation of *Bacillus subtilis* DB104 with the same procedures as described in Example I-2. Subsequently, each transformed *Bacillus* strains was cultured for 24 hr at a shaking incubator (37° C., 250 rpm) in GYS medium, the only pure spores were isolated using renografin gradients method, and enzyme activity of carboxymethylcellulase (Kim, et al., *Appl. Environ. Microbiol.*, 66:788-793(2000)), levansucrase (Jung, et al., *Nat. Biotech.*, 16:576-580(1998)) or lipase was evaluated. The activity of lipase was evaluated as follow: The spores suspended in 10% PBS was mixed with 10% olive oil, reacted for 48 hr, treated with 0.2 ml cupric acid on supernatant solution and the observance of OD was performed at 715 nm.

[0132] In the case of carboxymethylcellulase, the activity of enzyme displayed on spore was 175 mU comparing to 0 mU in control. In other verifying method, carboxymethylcellulase-specific antibody (Kim, et al., *Appl. Environ. Microbiol.*, 66:788-793(2000)) was probed for flow cytometry (FACSort, Becton Dickinson, USA) and the carboxymethylcellulases were detected on the surface of spores transformed by pCSK-cotG-CMCase (FIG. 10).

[0133] The activity of levansucrase was also high in spores transformed by recombinant vector (FIG. 12) and the levansucrases were detected on the surface of transformed spores as verified with flow cytometry using levansucrase-specific antibody (Jung, et al., *Nat. Biotech.*, 16:576-580(1998)) in the same procedures as above-described % in carboxymethylcellulase (FIG. 11).

[0134] The activity of lipase was measured as  $A_{715}=0.14$  in spores transformed with recombinant vector.

[0135] On the basis of these results, it is demonstrated that various enzymes as well as  $\beta$ -galactosidase can be displayed on the surface of spore according to the present invention.

[0136] Based on the results in these examples and example I, it is known to one skilled in the art that the gene construct containing gene encoding fusion protein between coat protein and protein of interest may exist as plasmid in host cell independently or as integrated form into chromosome of host cell and both forms may lead to successful spore surface display. It is also recognizable that the gene of coat protein may be followed or preceded by the gene of the protein of interest. In addition, it is recognized that in the gene construct, the overall sequence, fragments, two or more repeated sequences of the gene of coat protein are useful. In two or more repeated sequences, the repeated sequences may be the same or different each other. The overall sequence, two or more repeated sequences of the gene of the protein of interest are also useful in the fusion sequence. In two or more repeated sequences, the repeated sequences may be the same or different each other.

[0137] It is recognized by one skilled in the art that the expression of the fusion protein between coat protein and protein of interest can be induced by virtue of promoters of coat protein gene and other suitable promoters operable in host cell. Any vector carrying the present gene construct may be used in this invention, which is recognized by one skilled in the art referring to these results.

[0138] It is known that both monomeric and multimeric enzyme can be applied for the present invention since the  $\beta$ -galactosidase used in example I is tetramer (U. Karlsson et al., *J. Ultrastruct. Res.*, 10:457-469(1964)) and the enzymes described in this Example are monomers.

#### Example V

##### Display of Antibody on Spore Surface and Screening for Directed Evolution

[0139] On the purpose of application of other proteins in addition to enzymes, the experiment to display antibody on spore surface was performed as follows:

[0140] V-1: Construction of Recombinant Vector for Surface Display of Single Chain Fv

[0141] Gene encoding single chain Fv, against Pre-S2 domain (SEQ ID NO:36) of hepatitis B virus (HBV) was linked to cotG gene encoding surface protein of *Bacillus subtilis* spore. Single chain Fv gene was amplified by PCR with pAScFv101 (WO 9737025) as template and with primers described in SEQ ID NOs:37 and 38. Taq polymerase purchased from Bioneer (Korea) was used for total 30 cycles of PCR under condition of denaturation for 30 sec at 94° C., annealing for 30 sec at 55° C. and extension for

1 min at 72° C. And then, each PCR product was restricted by *Apa*I and *Nhe*I, cloned into pCSK-CotG between the same restriction sites (pCSK-CotG-scFv) and transformed into JM109 using transformation method by Inoue, et al. (Inoue, H., et al., *Gene*, 96:23-28(1990)). The amplified vectors for displaying on spore surface were isolated by alkaline extraction method (Sambrook et al., *Molecular Cloning: A laboratory Manual*, Cold Spring Harbor, N.Y., 1989) and transformed into *Bacillus subtilis* DB104 by natural transformation as described in Example I-1.

[0142] V-2: Verification of Single Chain Fv Display on Spore Surface Using Flow Cytometry

[0143] Affinity of the displayed single chain Fv against the Pre-S2 of HBV was evaluated by FACSsort as the following procedures.

[0144] Firstly, Pre-S2 peptide was labeled with fluorescein (PanVera, USA) using fluorescein succinidimyl ester coupling method.

[0145] The transformed strains were inoculated into LB broth containing 5 µg/ml chloroamphenicol, pre-cultured for 8-10 hr at 37° C., 1% of seed culture was inoculated into GYS broth for sporulation, cultured for 24 hr at 37° C. and the cultured media was harvested. The pure spores were isolated using renografin gradients method, 100 µl pure spores were blocked with PBS containing 3% skim milk to inhibit non-specific binding and reacted with 10 µl of fluorescein labeled Pre-S2 peptide. Thereafter, the spores bound to fluorescein labeled Pre-S2 peptide were detected in the same procedures as described in example IV (FIG. 13). As shown in FIG. 13, it is demonstrated that the monoclonal antibody against Pre-S2 peptide is successfully displayed without reduction of the affinity to its antigen.

[0146] According to the above results, it is recognized that the present methods may be applicable to any protein, for example, enzyme, hormone, hormone analogue, enzyme inhibitor, signal transduction protein or its fragment, antibody or its fragment, antigen protein, attachment protein, structural protein, regulatory protein, toxin protein, plant defense-inducing protein.

[0147] V-3: Selection of Spores Displaying Single Chain Fv using Flow Cytometry

[0148] Whether the displayed single chain Fv has affinity to Pre-S2 of HBV was verified with FACSsort as follows:

[0149] The transformed strains were inoculated into LB broth containing 5 µg/ml chloroamphenicol, pre-cultured for 8-10 hr at 37° C., 1% of seed culture was inoculated into GYS broth for sporulation, cultured for 24 hr at 37° C. and the cultured media was harvested. And then, 50 ml of harvested culture medium was centrifuged at 10,000 g for 10 min supernatant was discarded, bacteria were resuspended in 500 µl of 20% renografin (Sigma, USA). 100 µl of resuspended cell was carefully flowed onto 500 µl of 50% renogrant in microtube to form layer, the microtube was centrifuged at 10,000 g for 30 min and pure spores were isolated from pellet.

[0150] To discard remained renografin, spores were rinsed 3 times with DW and resuspended in PBS buffer. And then, spores displaying single chain Fv were mixed with wild type spores at a ratio of 1:103 and 1:105 and the spores with

affinity to Pre-S2 of HBV were harvested using fluorescein-labeled Pre-S2 peptide and FACSsort.

[0151] The selectivity was evaluated by colony-forming assay on LB agar plates and LB agar plates containing 5 µg/ml of chloroamphenicol comparing to wild type. Spores displaying surface single chain Fv are resistant to chloroamphenicol owing to chloroamphenicol resistant gene contained in the recombinant vectors.

[0152] FIG. 14 shows the selectivity of spores displaying single chain Fv in each ratio (selectivity=ratio of spores displaying single chain Fv after flow cytometry/ratio of spores displaying single chain Fv before flow cytometry). In the case that the ratio of spores displaying single chain Fv before flow cytometry is 10<sup>-5</sup>, the selectivity was over 4,000, which indicates that spores with enhanced affinity can be selected by flow cytometry among spores displaying various antibody libraries.

[0153] V-4: Directed Evolution of Single Chain Fv Displayed on Spore Surface

[0154] To display single chain Fv library on spore surface, the gene encoding single chain Fv against Pre-S2 of HBV was amplified by error prone PCR. PCR was carried out using pAScFv101 plasmid described in the example V-1 as template and SEQ ID NOs:37 and 38 as primer. PCR mixture was prepared by mixing 0.3 µM of each primers, 5 ng of DNA template, PCR solution (10 mM Tris(pH 8.3), 50 mM KCl, 7 mM MgCl<sub>2</sub>, 0.01% (w/v) gelatin), 0.2 mM dGTP, 0.2 mM dATP, 1 mM dTTP, 1 mM dCTP, 5 U Taq polymerase from Bioneer (Korea) and DW up to 100 µl. Total 13 cycles of PCR was performed under condition of denaturation for 30 sec at 94° C., annealing for 30 sec at 50° C. and extension for 1 min at 72° C.

[0155] Subsequently, restricted PCR products with *Apa*I and *Nhe*I were cloned into pCSK-CotG, vector for displaying on spore surface, between the same restriction sites and library was prepared by transforming the cloned vectors into JM109 *E. coli* with the method of Inoue et al.

[0156] The vectors for displaying on spore surface were isolated by alkaline extraction method and transformed into *Bacillus subtilis* DB104 by natural transformation. And then, single chain Fv library against Pre-S2 of HBV was displayed on spore surface as described in example V-2 (FIG. 15).

[0157] As shown in FIG. 15, spores with increased fluorescence (i.e., increased affinity) were isolated. This result demonstrates the applicability of the present invention to prepare and select protein variants with improved characteristics.

## Example VI

### Bioconversion Using Spores Displaying Protein of Interest

[0158] Forte of transglycosylation by enzyme is the capability of formation of site-specific glycosidic linkage without protection/de-protection step. There have been studied for formation of glycosidic linkage by 1) induction of reverse hydrolysis in non-aqueous system using glycosidase which is conventionally available glycosidic hydrolyzing enzyme and 2) transglycosylation in which glycosidic linkage is

substituted with receptor alcohol instead of hydrolysis of glycosidic linkage by water (G. Ljunger et al., *Enzyme Microb. Technol.*, 16:1808-1814(1994); T. Usui et al., *Carbohydr. Res.*, 244:315-323(1993); and R. Lopez et al., *J. Org. Chem.*, 59:737-745(1994)). The above conventional methods usually use organic solvent to increase synthetic yield and inhibit hydrolysis. However, because the organic solvent inactivates enzyme, it is difficult to accomplish the high yield. Thus, it is necessary to inhibit the inactivation of glycosidase in organic solvent for higher glycosylation yield.

[0159] The purpose of the Example is to exemplify the higher glycosylation yield with improved enzyme stability even in organic solvent by virtue of displaying glycosidase on the surface of hydrophobic *Bacillus* spores.

[0160] VI-1: Stability of  $\beta$ -Galactosidase Displayed on Surface of Spores in Organic Solvent

[0161] Each of  $\beta$ -galactosidase in free form (Sigma, USA) and the  $\beta$ -galactosidase displayed on surface of *Bacillus* spore was dispersed into 500  $\mu$ l of Tris-HCl buffer (pH 7.5), added the same volume of the various solvents described in Table 1, mixed for 37° C. for 1 hr and the remained enzymatic activity was measured by Miller method described in Example I-3 (Table 1).

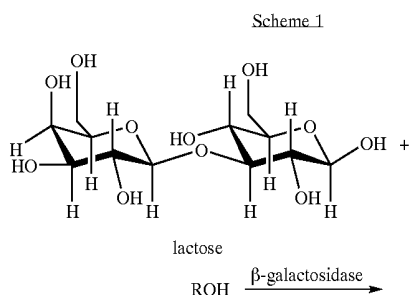
TABLE 1

	Residual activity (%)	
	Free form $\beta$ -galactosidase	Surface-Displayed $\beta$ -galactosidase
Control	100	100
Hexane	84.3	100
Ether	48.2	77.2
Toluene	4.2	51.9
Ethylacetate	0.1	9.6
Acetonitril	0.0	0.8
Ethanol	0.0	0.0

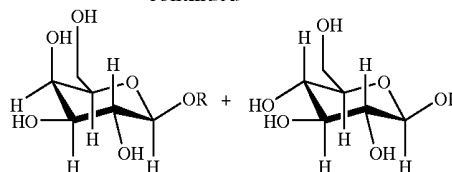
[0162] As shown in Table 1, the displayed  $\beta$ -galactosidase shows higher stability than that of free form  $\beta$ -galactosidase in various organic solvents.

[0163] VI-2: Transglycosylation Reaction in Water-Organic Solvent Two-phase System Using  $\beta$ -galactosidase Displayed on Spore Surface

[0164] To perform transglycosylation in two-phase system,  $\beta$ -galactosidase, which is one of conventional glycosidase, is used as a model for glycosylation reaction (Scheme 1).



-continued



[0165] At first, 1 ml of 1 M lactose in 10 mM phosphate buffer (pH 5.1) was mixed with 10 ml of 10 mM 5-phenyl-1-pentanol in hexane for reaction solution. And then,  $\beta$ -galactosidase displayed on spore surface (240 U; 1 U=the amount of enzyme capable of hydrolysis of 1  $\mu$ mol ONPG (o-nitrophenyl- $\beta$ -D-galactopyranoside) for 1 min at 37° C.) and free form  $\beta$ -galactosidase (240 U) was added into the above reaction solution, respectively, and reacted for 48 hr at 30° C. while stirring.

[0166] In results, the yield of 5-phenylpentyl- $\beta$ -D-galactopyranoside was 21% by  $\beta$ -galactosidase displayed on spore surface; however, in free form  $\beta$ -galactosidase, the hydrolysis of lactose only occurred with no transglycosylation. Such result is ascribed to the increased stability, in organic solvent, of  $\beta$ -galactosidase displayed on spore surface. Actually, after 72 hr reaction, about 5% of enzyme activity was detected in the displayed  $\beta$ -galactosidase while measured the complete inactivation in free form  $\beta$ -galactosidase. Another advantage of the displayed  $\beta$ -galactosidase owes to hydrophobicity of *Bacillus* spores. In other words, the distribution of displayed  $\beta$ -galactosidase at interface between water and organic solvent phase inhibits the hydrolysis comparing to free form  $\beta$ -galactosidase.

[0167] Based on the results of this Example, it is understood that variants and modifications thereof falling within the spirit of the invention may become apparent to those skilled in the art, for example, any enzymes in addition to  $\beta$ -galactosidase such as lipase and protease can be employed for bioconversion of the present invention. In addition, the present bioconversion is useful in single step or multi-step reaction and in aqueous or non-aqueous solution. The present bioconversion method can employ spore as free or immobilized form and can be performed with other microbes or enzymes.

#### Example VII

##### Display of Antigen on Spore Surface

[0168] By displaying antigen on spore surface, antigen capable of inducing immune response in vivo can be applied as live vaccine. *Bacillus subtilis* has been considered as safe strain to human body since it has been employed in food fermentation for a long time (Sonenshein A. L., et al., *Bacillus subtilis* and other gram-positive bacteria. *American society for Microbiology*, Washington, p871(1993)).

[0169] Gene for CotE-antigen fusion protein is constructed by cloning the gene for surface antigen of HBV into pCotG-lacZ vector constructed in Example I-1. Thereafter, the constructed recombinant vector is transformed into *Bacillus subtilis* and the transformants are cultured in GYS medium. And then, the antigen-displaying spores are purely isolated from culture medium by renografin gradients method.

## Example VIII

Protein Improvement Using Spore Displaying  
Protein of Interest

[0170] For example of application of the present invention to high-throughput screening of target protein and to protein improvement, GFP (Green Fluorescence Protein) was used as follows:

[0171] VIII-1: Construction of Vector for GFP Display on Spore Surface

[0172] gfp gene was cloned into pCSK-CotG vector constructed in Example IV-1 and the following sub-cloning procedures were performed for display on spore surface. Each primer was prepared for the purpose of fusing cotG gene to EGFP and GFPuv genes. The fluorescence intensity of EGFP (Excit./Emis. Maxima (nm): 488/509; Clontech, USA) has 35-fold stronger than that of wild type GFP and thus results in detection even in FITC filter and GFPuv (Excit./Emis. Maxima (nm): 395/509; Clontech, USA) is detectable with UV. For further manipulation, NheI and HindIII restriction sites were inserted into primers for egfp gene (SEQ ID NOs:39 and 40) and PstI and EcoRI restriction sites were inserted in primers for gfpuv gene (SEQ ID NOs:41 and 42).

[0173] Each of egfp (800 bp) and gfpuv (720 bp) genes was amplified by PCR (MJ Research PTC-100™ programmable Thermal Controller; 95° C. 30 sec, 55° C. 30 sec, 72° C. 2 min, 25 cycles) using Pfu Turbo polymerase (Stratagene, USA) and pEGFP-C1 (Clontech, USA) or pGFPuv (Clontech, USA) as template.

[0174] Thereafter, pCSK-CotG-EGFP or pCSK-CotG-GFPuv vectors were constructed by cloning the restricted PCR products into NheI/HindIII (egfp gene) or PstI/EcoRI (gfpuv gene) restriction sites of pCSK-CotG vector.

[0175] VIII-2: Display and Confirmation of GFP on Spore Surface

[0176] The constructed vectors were transformed into *Bacillus subtilis* DB104 by natural transformation. Transformants were selected on LB agar plate containing 5 µg/ml chloramphenicol. Through the selection, *Bacillus subtilis* DB104-SDG-EGFP strain for display of EGFP and *Bacillus subtilis* DB104-SDG-GFPuv strain for display of GFPuv on spore surface were obtained. As control strains, *Bacillus subtilis* DB104-SDC strain transformed with only pCSK vector and *Bacillus subtilis* DB104-SDG strain transformed for expressing only CotG protein were prepared.

[0177] For analysis of GFP display on spore surface, the above *Bacillus subtilis* DB104-SDC, -SDG, -SDG-EGFP and -SDG-GFPuv were inoculated into LB broth containing 5 µg/ml chloramphenicol and spores were then purified as described in Example V-4.

[0178] Subsequently, the display of GFP on spore surface was analyzed by measuring GFP fluorescence with flow cytometry in similar manner to Example IV (FIG. 16). In FIG. 16, curves (1)-(4) indicate the results of spores of DB104-SDC DB104-SDG, DB104-SDG-GFPuv and DB104-SDG-EGFP, respectively.

[0179] As shown in FIG. 16, the fluorescent intensity of spores derived from DB104-SDG-EGFP (recombinant

strain for EGFP-spore surface display) and DB104-SDG-GFPuv (recombinant strain for GFPuv-spore surface display) is significantly higher than that of DB104-SDC and DB104-SDG as control. In above results, the successful display of EGFP or GFPuv is validated by noticeable change of peaks indicating fluorescence in spore on its surface displaying EGFP or GFPuv comparing to controls.

[0180] VIII-3: Improvement of GFP

[0181] For the purpose of GFP improvement, error prone PCR was performed with template of pGFPuv vector (Clontech, USA) containing gfpuv gene using primers of SEQ ID NOs: 42 and 43. PCR mixture was prepared by mixing 0.3 µM of each primers, 5 ng of DNA template, PCR solution 10 mM Tris (pH 8.3), 50 mM KCl, 7 mM MgCl<sub>2</sub>, 0.01% (w/v) gelatin, 0.2 mM dGTP, 0.2 mM dATP, 1 mM dTTP, 1 mM dCTP, 0.15 mM MnCl<sub>2</sub>, 5 U Taq polymerase from Bioneer (Korea) and DW up to 100 µl. Total 13 cycles of PCR was performed under condition of denaturation for 30 sec at 94° C., annealing for 30 sec at 50° C. and extension for 1 min at 72° C.

[0182] Subsequently, the gfpuv genes were discarded from pCSK-CotG-GFPuv vectors by restriction with PstI/EcoRI, the above PCR-amplified inserts were cloned into the vectors with the same restriction sites and *Bacillus subtilis* DB104 was transformed with the cloned vectors by natural transformation to construct gfpuv library displayed on spore surface. Then, the prepared library was inoculated into GYS medium for sporulation and pure spores were isolated as described in Example V-4. Transformant spores displaying improved GFP variant were screened by measuring GFP fluorescence with flow cytometry (FIGS. 17a to 17d). FIGS. 17a to 17d indicates the analysis of flow cytometry from *Bacillus subtilis* DB104-SDC, DB104-SDG-GFPuv, DB104-SDG-EGFP and DB104-SDG-GFP with gfp library subject to error prone PCR, respectively.

[0183] To isolate spores with higher fluorescent intensity than spores derived from DB104-SDG-EGFP and DB104-SDG-GFP control strains, the isolation of spores with higher fluorescence (region R1) among spores displaying GFP library was repeated several times.

[0184] It is understood that using the above method, the improved GFP protein exhibiting higher fluorescence intensity or fluorescence with different wavelength can be screened in a high-throughput manner.

## Example VIII

Protein Array Using Spores Displaying on its  
Surface Protein of Interest

[0185] 106-109 spores displaying monoclonal antibodies against surface antigen of HBV are attached onto glass substrate for protein array (BMS, Germany) with aldehyde functional group on its surface using automated array apparatus. The attachment is made in a form of covalent linkage, which is Schiff base between amino group of protein on spore surface and aldehyde group on surface of slide glass. Although the displayed proteins attached on solid surface may be inactivated, they may have an orientation.

[0186] The protein array kit manufactured according to the present invention has a variety of applicable fields including diagnosis, analysis of gene expression, analysis of interac-

tion between proteins, analysis of interaction between protein and ligand, study on metabolism, screening novel or improved enzymes, combinatorial biochemical synthesis and biosensor.

#### Example IX

##### Production of Antibody Using Spores Displaying Antigen

[0187] The spores on its surface displaying surface antigen of HBV isolated in Example VII are suspended in PBS and the same volume of complete Freund's adjuvant is added. Thereafter, the mixture is well agitated to make emulsion formulation and the emulsion is injected i.v. into BALB/c mice with age of 6-8 week. After 4 weeks of the injection, the secondary administration is performed. Then, the additional boosting injection is performed about 2-3 times for induction of antibody.

[0188] As described above, the display method on spore surface of the present invention provides improvements in: a resistance against physiochemical change in environment of display host, a diversity of displayable proteins, a viability of display host and rapidity of screening.

[0189] Throughout this application, various patents and publications are referenced and citations are provided in parentheses. The disclosure of these patents and publications in their entities are hereby incorporated by references into this application in order to more fully describe this invention and the state of the art to which this invention pertains.

[0190] Having described a preferred embodiment of the present invention, it is to be understood that variants and modifications thereof falling within the spirit of the invention may become apparent to those skilled in this art, and the scope of this invention is to be determined by appended claims and their equivalents.

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 <211> LENGTH: 1191  
 <212> TYPE: PRT  
 <213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 24

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Met Ser Glu Tyr Arg Glu Ile Ile Thr Lys Ala Val Val Ala Lys Gly
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Arg Lys Phe Thr Gln Cys Thr Asn Thr Ile Ser Pro Glu Lys Lys Pro
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Ser Ser Ile Leu Gly Gly Trp Ile Ile Asn His Lys Tyr Asp Ala Glu

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35			40			45									
Lys	Ile	Gly	Lys	Thr	Val	Glu	Ile	Glu	Gly	Tyr	Tyr	Asp	Ile	Asn	Val
	50						55					60			
Trp	Tyr	Ser	Tyr	Ala	Asp	Asn	Thr	Lys	Thr	Glu	Val	Val	Thr	Glu	Arg
	65				70					75					80
Val	Lys	Tyr	Val	Asp	Val	Ile	Lys	Leu	Arg	Tyr	Arg	Asp	Asn	Asn	Tyr
				85						90				95	
Leu	Asp	Asp	Glu	His	Glu	Val	Ile	Ala	Lys	Val	Leu	Gln	Gln	Pro	Asn
			100					105					110		
Cys	Leu	Glu	Val	Thr	Ile	Ser	Pro	Asn	Gly	Asn	Lys	Ile	Val	Val	Gln
		115					120					125			
Ala	Glu	Arg	Glu	Phe	Leu	Ala	Glu	Val	Val	Gly	Glu	Thr	Lys	Val	Val
	130						135					140			
Val	Glu	Val	Asn	Pro	Asp	Trp	Glu	Glu	Asp	Asp	Glu	Glu	Asp	Trp	Glu
	145				150					155					160
Asp	Glu	Leu	Asp	Glu	Glu	Leu	Glu	Asp	Ile	Asn	Pro	Glu	Phe	Leu	Val
				165						170				175	
Gly	Asp	Pro	Glu	Glu	Val	Asp	Arg	Glu	Asn	Pro	Gly	Val	Thr	Gln	Leu
			180					185						190	
Asn	Arg	Leu	Ala	Ala	His	Pro	Pro	Phe	Ala	Ser	Trp	Arg	Asn	Ser	Glu
		195					200					205			
Glu	Ala	Arg	Thr	Asp	Arg	Pro	Ser	Gln	Gln	Leu	Arg	Ser	Leu	Asn	Gly
	210						215				220				
Glu	Trp	Arg	Phe	Ala	Trp	Phe	Pro	Ala	Pro	Glu	Ala	Val	Pro	Glu	Ser
	225				230					235					240
Trp	Leu	Glu	Cys	Asp	Leu	Pro	Glu	Ala	Asp	Thr	Val	Val	Val	Pro	Ser
				245					250					255	
Asn	Trp	Gln	Met	His	Gly	Tyr	Asp	Ala	Pro	Ile	Tyr	Thr	Asn	Val	Thr
		260						265					270		
Tyr	Pro	Ile	Thr	Val	Asn	Pro	Pro	Phe	Val	Pro	Thr	Glu	Asn	Pro	Thr
		275					280					285			
Gly	Cys	Tyr	Ser	Leu	Thr	Phe	Asn	Val	Asp	Glu	Ser	Trp	Leu	Gln	Glu
	290						295				300				
Gly	Gln	Thr	Arg	Ile	Ile	Phe	Asp	Gly	Val	Asn	Ser	Ala	Phe	His	Leu
	305				310					315					320
Trp	Cys	Asn	Gly	Arg	Trp	Val	Gly	Tyr	Gly	Gln	Asp	Ser	Arg	Leu	Pro
				325						330				335	
Ser	Glu	Phe	Asp	Leu	Ser	Ala	Phe	Leu	Arg	Ala	Gly	Glu	Asn	Arg	Leu
				340				345					350		
Ala	Val	Met	Val	Leu	Arg	Trp	Ser	Asp	Gly	Ser	Tyr	Leu	Glu	Asp	Gln
		355						360					365		
Asp	Met	Trp	Arg	Met	Ser	Gly	Ile	Phe	Arg	Asp	Val	Ser	Leu	Leu	His
	370						375				380				
Lys	Pro	Thr	Thr	Gln	Ile	Ser	Asp	Phe	His	Val	Ala	Thr	Arg	Phe	Asn
	385				390					395					400
Asp	Asp	Phe	Ser	Arg	Ala	Val	Leu	Glu	Ala	Glu	Val	Gln	Met	Cys	Gly
				405						410				415	
Glu	Leu	Arg	Asp	Tyr	Leu	Arg	Val	Thr	Val	Ser	Leu	Trp	Gln	Gly	Glu
			420					425					430		
Thr	Gln	Val	Ala	Ser	Gly	Thr	Ala	Pro	Phe	Gly	Gly	Glu	Ile	Ile	Asp
		435					440						445		

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Glu Arg Gly Gly Tyr Ala Asp Arg Val Thr Leu Arg Leu Asn Val Glu  
 450 455 460

Asn Pro Lys Leu Trp Ser Ala Glu Ile Pro Asn Leu Tyr Arg Ala Val  
 465 470 475 480

Val Glu Leu His Thr Ala Asp Gly Thr Leu Ile Glu Ala Glu Ala Cys  
 485 490 495

Asp Val Gly Phe Arg Glu Val Arg Ile Glu Asn Gly Leu Leu Leu Leu  
 500 505 510

Asn Gly Lys Pro Leu Leu Ile Arg Gly Val Asn Arg His Glu His His  
 515 520 525

Pro Leu His Gly Gln Val Met Asp Glu Gln Thr Met Val Gln Asp Ile  
 530 535 540

Leu Leu Met Lys Gln Asn Asn Phe Asn Ala Val Arg Cys Ser His Tyr  
 545 550 555 560

Pro Asn His Pro Leu Trp Tyr Thr Leu Cys Asp Arg Tyr Gly Leu Tyr  
 565 570 575

Val Val Asp Glu Ala Asn Ile Glu Thr His Gly Met Val Pro Met Asn  
 580 585 590

Arg Leu Thr Asp Asp Pro Arg Trp Leu Pro Ala Met Ser Glu Arg Val  
 595 600 605

Thr Arg Met Val Gln Arg Asp Arg Asn His Pro Ser Val Ile Ile Trp  
 610 615 620

Ser Leu Gly Asn Glu Ser Gly His Gly Ala Asn His Asp Ala Leu Tyr  
 625 630 635 640

Arg Trp Ile Lys Ser Val Asp Pro Ser Arg Pro Val Gln Tyr Glu Gly  
 645 650 655

Gly Gly Ala Asp Thr Thr Ala Thr Asp Ile Ile Cys Pro Met Tyr Ala  
 660 665 670

Arg Val Asp Glu Asp Gln Pro Phe Pro Ala Val Pro Lys Trp Ser Ile  
 675 680 685

Lys Lys Trp Leu Ser Leu Pro Gly Glu Thr Arg Pro Leu Ile Leu Cys  
 690 695 700

Glu Tyr Ala His Ala Met Gly Asn Ser Leu Gly Gly Phe Ala Lys Tyr  
 705 710 715 720

Trp Gln Ala Phe Arg Gln Tyr Pro Arg Leu Gln Gly Gly Phe Val Trp  
 725 730 735

Asp Trp Val Asp Gln Ser Leu Ile Lys Tyr Asp Glu Asn Gly Asn Pro  
 740 745 750

Trp Ser Ala Tyr Gly Gly Asp Phe Gly Asp Thr Pro Asn Asp Arg Gln  
 755 760 765

Phe Cys Met Asn Gly Leu Val Phe Ala Asp Arg Thr Pro His Pro Ala  
 770 775 780

Leu Thr Glu Ala Lys His Gln Gln Gln Phe Phe Gln Phe Arg Leu Ser  
 785 790 795 800

Gly Gln Thr Ile Glu Val Thr Ser Glu Tyr Leu Phe Arg His Ser Asp  
 805 810 815

Asn Glu Leu Leu His Trp Met Val Ala Leu Asp Gly Lys Pro Leu Ala  
 820 825 830

Ser Gly Glu Val Pro Leu Asp Val Ala Pro Gln Gly Lys Gln Leu Ile  
 835 840 845

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Glu Leu Pro Glu Leu Pro Gln Pro Glu Ser Ala Gly Gln Leu Trp Leu  
 850 855 860  
 Thr Val Arg Val Val Gln Pro Asn Ala Thr Ala Trp Ser Glu Ala Gly  
 865 870 875 880  
 His Ile Ser Ala Trp Gln Gln Trp Arg Leu Ala Glu Asn Leu Ser Val  
 885 890 895  
 Thr Leu Pro Ala Ala Ser His Ala Ile Pro His Leu Thr Thr Ser Glu  
 900 905 910  
 Met Asp Phe Cys Ile Glu Leu Gly Asn Lys Arg Trp Gln Phe Asn Arg  
 915 920 925  
 Gln Ser Gly Phe Leu Ser Gln Met Trp Ile Gly Asp Lys Lys Gln Leu  
 930 935 940  
 Leu Thr Pro Leu Arg Asp Gln Phe Thr Arg Ala Pro Leu Asp Asn Asp  
 945 950 955 960  
 Ile Gly Val Ser Glu Ala Thr Arg Ile Asp Pro Asn Ala Trp Val Glu  
 965 970 975  
 Arg Trp Lys Ala Ala Gly His Tyr Gln Ala Glu Ala Ala Leu Leu Gln  
 980 985 990  
 Cys Thr Ala Asp Thr Leu Ala Asp Ala Val Leu Ile Thr Thr Ala His  
 995 1000 1005  
 Ala Trp Gln His Gln Gly Lys Thr Leu Phe Ile Ser Arg Lys Thr Tyr  
 1010 1015 1020  
 Arg Ile Asp Gly Ser Gly Gln Met Ala Ile Thr Val Asp Val Glu Val  
 1025 1030 1035 1040  
 Ala Ser Asp Thr Pro His Pro Ala Arg Ile Gly Leu Asn Cys Gln Leu  
 1045 1050 1055  
 Ala Gln Val Ala Glu Arg Val Asn Trp Leu Gly Leu Gly Pro Gln Glu  
 1060 1065 1070  
 Asn Tyr Pro Asp Arg Leu Thr Ala Ala Cys Phe Asp Arg Trp Asp Leu  
 1075 1080 1085  
 Pro Leu Ser Asp Met Tyr Thr Pro Tyr Val Phe Pro Ser Glu Asn Gly  
 1090 1095 1100  
 Leu Arg Cys Gly Thr Arg Glu Leu Asn Tyr Gly Pro His Gln Trp Arg  
 1105 1110 1115 1120  
 Gly Asp Phe Gln Phe Asn Ile Ser Arg Tyr Ser Gln Gln Gln Leu Met  
 1125 1130 1135  
 Glu Thr Ser His Arg His Leu Leu His Ala Glu Glu Gly Thr Trp Leu  
 1140 1145 1150  
 Asn Ile Asp Gly Phe His Met Gly Ile Gly Gly Asp Asp Ser Trp Ser  
 1155 1160 1165  
 Pro Ser Val Ser Ala Glu Phe Gln Leu Ser Ala Gly Arg Tyr His Tyr  
 1170 1175 1180  
 Gln Leu Val Trp Cys Gln Lys  
 1185 1190

<210> SEQ ID NO 25  
 <211> LENGTH: 4173  
 <212> TYPE: DNA  
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 <220> FEATURE:  
 <221> NAME/KEY: CDS  
 <222> LOCATION: (461)..(4075)  
 <400> SEQUENCE: 25

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gggtctttat actccgcatt taagtgaatc tctcgcgcgc cgcggaatgt tttcggctga	120
taaaaggaaa tatggtatga cttctttttg aagtctctga tatgtgatcc ccgataagcg	180
atatcaatat ccagcctttt ttgatttacc ttcacacag ctggcaccgg atcatcgtcc	240
catatatacct tttttaattc acgcaagtct ttggatgaa caaacagctg ataaagcgg	300
aaattggatt gattcttcat ccataatcct cttacaaat tttaggcttt tatttttata	360
agatctcagc ggaacactta tacacttttt aaaaccgcgc gtactatgag ggtagtaagg	420
atcttcatcc ttaacatatt tttaaaagga ggatttcaaa ttg ggc cac tat tcc	475
	Leu Gly His Tyr Ser
	1 5
cat tct gac atc gaa gaa gcg gtg aaa tcc gca aaa aaa gaa ggt tta	523
His Ser Asp Ile Glu Glu Ala Val Lys Ser Ala Lys Lys Glu Gly Leu	
	10 15 20
aag gat tat tta tac caa gag cct cat gga aaa aaa cgc agt cat aaa	571
Lys Asp Tyr Leu Tyr Gln Glu Pro His Gly Lys Lys Arg Ser His Lys	
	25 30 35
aag tcg cac cgc act cac aaa aaa tct cgc agc cat aaa aaa tca tac	619
Lys Ser His Arg Thr His Lys Lys Ser Arg Ser His Lys Lys Ser Tyr	
	40 45 50
tgc tct cac aaa aaa tct cgc agt cac aaa aaa tca ttc tgt tct cac	667
Cys Ser His Lys Lys Ser Arg Ser His Lys Lys Ser Phe Cys Ser His	
	55 60 65
aaa aaa tct cgc agc cac aaa aaa tca tac tgc tct cac aag aaa tct	715
Lys Lys Ser Arg Ser His Lys Lys Ser Tyr Cys Ser His Lys Lys Ser	
	70 75 80 85
cgc agc cac aaa aaa tcg tac cgt tct cac aaa aaa tct cgc agc tat	763
Arg Ser His Lys Lys Ser Tyr Arg Ser His Lys Lys Ser Arg Ser Tyr	
	90 95 100
aaa aaa tct tac cgt tct tac aaa aaa tct cgt agc tat aaa aaa tct	811
Lys Lys Ser Tyr Arg Ser Tyr Lys Lys Ser Arg Ser Tyr Lys Lys Ser	
	105 110 115
tgc cgt tct tac aaa aaa tct cgc agc tac aaa aag tct tac tgt tct	859
Cys Arg Ser Tyr Lys Lys Ser Arg Ser Tyr Lys Lys Ser Tyr Cys Ser	
	120 125 130
cac aag aaa aaa tct cgc agc tat aag aag tca tgc cgc aca cac aaa	907
His Lys Lys Lys Ser Arg Ser Tyr Lys Lys Ser Cys Arg Thr His Lys	
	135 140 145
aaa tct tat cgt tcc cat aag aaa tac tac aaa aaa ccg cac cac cac	955
Lys Ser Tyr Arg Ser His Lys Lys Tyr Tyr Lys Lys Pro His His His	
	150 155 160 165
tgc gac gac tac aaa aga cac gat gat tat gac agc aaa aaa gaa tac	1003
Cys Asp Asp Tyr Lys Arg His Asp Asp Tyr Asp Ser Lys Lys Glu Tyr	
	170 175 180
tgg aaa gac ggc aat tgc tgg gta gtc aaa aag aaa tac aaa gtc gac	1051
Trp Lys Asp Gly Asn Cys Trp Val Val Lys Lys Lys Tyr Lys Val Asp	
	185 190 195
cgg gaa aac cct ggc gtt acc caa ctt aat cgc ctt gca gca cat ccc	1099
Arg Glu Asn Pro Gly Val Thr Gln Leu Asn Arg Leu Ala Ala His Pro	
	200 205 210
cct ttc gcc agc tgg cgt aat agc gaa gag gcc cgc acc gat cgc cct	1147
Pro Phe Ala Ser Trp Arg Asn Ser Glu Glu Ala Arg Thr Asp Arg Pro	
	215 220 225
tcc caa cag ttg cgc agc ctg aat ggc gaa tgg cgc ttt gcc tgg ttt	1195

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Ser 230	Gln	Gln	Leu	Arg	Ser 235	Leu	Asn	Gly	Glu	Trp 240	Arg	Phe	Ala	Trp	Phe 245	
ccg	gca	cca	gaa	gcg	gtg	ccg	gaa	agc	tgg	ctg	gag	tgc	gat	ctt	cct	1243
Pro	Ala	Pro	Glu	Ala	Val	Pro	Glu	Ser	Trp 255	Leu	Glu	Cys	Asp	Leu	Pro 260	
gag	gcc	gat	act	gtc	gtc	gtc	ccc	tca	aac	tgg	cag	atg	cac	ggg	tac	1291
Glu	Ala	Asp	Thr	Val	Val	Val	Pro	Ser	Asn 270	Trp	Gln	Met	His	Gly	Tyr 275	
gat	gcg	ccc	atc	tac	acc	aac	gta	acc	tat	ccc	att	acg	gtc	aat	ccg	1339
Asp	Ala	Pro	Ile	Tyr	Thr	Asn	Val	Thr	Tyr 285	Pro	Ile	Thr	Val	Asn	Pro 290	
ccg	ttt	ggt	ccc	acg	gag	aat	ccg	acg	ggt	tgt	tac	tcg	ctc	aca	ttt	1387
Pro	Phe	Val	Pro	Thr	Glu	Asn	Pro	Thr	Gly 300	Cys	Tyr	Ser	Leu	Thr	Phe 305	
aat	ggt	gat	gaa	agc	tgg	cta	cag	gaa	ggc	cag	acg	cga	att	att	ttt	1435
Asn	Val	Asp	Glu	Ser	Trp	Leu	Gln	Glu	Gly 310	Gln	Thr	Arg	Ile	Ile	Phe 325	
gat	ggc	ggt	aac	tcg	gcg	ttt	cat	ctg	tgg	tgc	aac	ggg	cgc	tgg	gtc	1483
Asp	Gly	Val	Asn	Ser	Ala	Phe	His	Leu	Trp 330	Cys	Asn	Gly	Arg	Trp	Val 340	
ggt	tac	ggc	cag	gac	agt	cgt	ttg	ccg	tct	gaa	ttt	gac	ctg	agc	gca	1531
Gly	Tyr	Gly	Gln	Asp	Ser	Arg	Leu	Pro	Ser 345	Glu	Phe	Asp	Leu	Ser	Ala 355	
ttt	tta	cgc	gcc	gga	gaa	aac	cgc	ctc	gcg	gtg	atg	gtg	ctg	cgt	tgg	1579
Phe	Leu	Arg	Ala	Gly	Glu	Asn	Arg	Leu	Ala 360	Val	Met	Val	Leu	Arg	Trp 370	
agt	gac	ggc	agt	tat	ctg	gaa	gat	cag	gat	atg	tgg	cgg	atg	agc	ggc	1627
Ser	Asp	Gly	Ser	Tyr	Leu	Glu	Asp	Gln	Asp 375	Met	Trp	Arg	Met	Ser	Gly 385	
att	ttc	cgt	gac	gtc	tcg	ttg	ctg	cat	aaa	ccg	act	aca	caa	atc	agc	1675
Ile	Phe	Arg	Asp	Val	Ser	Leu	Leu	His	Lys 390	Pro	Thr	Thr	Gln	Ile	Ser 405	
gat	ttc	cat	ggt	gcc	act	cgc	ttt	aat	gat	gat	ttc	agc	cgc	gct	gta	1723
Asp	Phe	His	Val	Ala	Thr	Arg	Phe	Asn	Asp 410	Asp	Phe	Ser	Arg	Ala	Val 420	
ctg	gag	gct	gaa	ggt	cag	atg	tgc	ggc	gag	ttg	cgt	gac	tac	cta	cgg	1771
Leu	Glu	Ala	Glu	Val	Gln	Met	Cys	Gly	Glu 425	Leu	Arg	Asp	Tyr	Leu	Arg 435	
gta	aca	ggt	tct	tta	tgg	cag	ggt	gaa	acg	cag	gtc	gcc	agc	ggc	acc	1819
Val	Thr	Val	Ser	Leu	Trp	Gln	Gly	Glu	Thr 440	Gln	Val	Ala	Ser	Gly	Thr 450	
gcg	cct	ttc	ggc	ggt	gaa	att	atc	gat	gag	cgt	ggt	ggt	tat	gcc	gat	1867
Ala	Pro	Phe	Gly	Gly	Glu	Ile	Ile	Asp	Glu 455	Arg	Gly	Gly	Tyr	Ala	Asp 465	
cgc	gtc	aca	cta	cgt	ctg	aac	gtc	gaa	aac	ccg	aaa	ctg	tgg	agc	gcc	1915
Arg	Val	Thr	Leu	Arg	Leu	Asn	Val	Glu	Asn 470	Pro	Lys	Leu	Trp	Ser	Ala 485	
gaa	atc	ccg	aat	ctc	tat	cgt	gcg	gtg	ggt	gaa	ctg	cac	acc	gcc	gac	1963
Glu	Ile	Pro	Asn	Leu	Tyr	Arg	Ala	Val	Val 490	Glu	Leu	His	Thr	Ala	Asp 500	
ggc	acg	ctg	att	gaa	gca	gaa	gcc	tgc	gat	gtc	ggt	ttc	cgc	gag	gtg	2011
Gly	Thr	Leu	Ile	Glu	Ala	Glu	Ala	Cys	Asp 505	Val	Gly	Phe	Arg	Glu	Val 515	
cgg	att	gaa	aat	ggt	ctg	ctg	ctg	ctg	aac	ggc	aag	ccg	ttg	ctg	att	2059
Arg	Ile	Glu	Asn	Gly	Leu	Leu	Leu	Leu	Asn 520	Gly	Lys	Pro	Leu	Leu	Ile 530	
cgg	ggc	ggt	aac	cgt	cac	gag	cat	cat	cct	ctg	cat	ggt	cag	gtc	atg	2107

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Arg 535	Gly	Val	Asn	Arg	His	Glu	His	His	Pro	Leu	His	Gly	Gln	Val	Met	
						540					545					
gat	gag	cag	acg	atg	gtg	cag	gat	atc	ctg	ctg	atg	aag	cag	aac	aac	2155
Asp	Glu	Gln	Thr	Met	Val	Gln	Asp	Ile	Leu	Leu	Met	Lys	Gln	Asn	Asn	
550					555					560					565	
ttt	aac	gcc	gtg	cgc	tgt	tcg	cat	tat	ccg	aac	cat	ccg	ctg	tggtac		2203
Phe	Asn	Ala	Val	Arg	Cys	Ser	His	Tyr	Pro	Asn	His	Pro	Leu	Trp	Tyr	
					570				575					580		
acg	ctg	tgc	gac	cgc	tac	ggc	ctg	tat	gtg	gtg	gat	gaa	gcc	aat	att	2251
Thr	Leu	Cys	Asp	Arg	Tyr	Gly	Leu	Tyr	Val	Val	Asp	Glu	Ala	Asn	Ile	
			585					590					595			
gaa	acc	cac	ggc	atg	gtg	cca	atg	aat	cgt	ctg	acc	gat	gat	ccg	cgc	2299
Glu	Thr	His	Gly	Met	Val	Pro	Met	Asn	Arg	Leu	Thr	Asp	Asp	Pro	Arg	
		600					605					610				
tgg	cta	ccg	gcg	atg	agc	gaa	cgc	gta	acg	cga	atg	gtg	cag	cgc	gat	2347
Trp	Leu	Pro	Ala	Met	Ser	Glu	Arg	Val	Thr	Arg	Met	Val	Gln	Arg	Asp	
	615					620					625					
cgt	aat	cac	ccg	agt	gtg	atc	atc	tgg	tcg	ctg	ggg	aat	gaa	tca	ggc	2395
Arg	Asn	His	Pro	Ser	Val	Ile	Ile	Trp	Ser	Leu	Gly	Asn	Glu	Ser	Gly	
630					635					640					645	
cac	ggc	gct	aat	cac	gac	gcg	ctg	tat	cgc	tgg	atc	aaa	tct	gtc	gat	2443
His	Gly	Ala	Asn	His	Asp	Ala	Leu	Tyr	Arg	Trp	Ile	Lys	Ser	Val	Asp	
			650						655					660		
cct	tcc	cgc	ccg	gtg	cag	tat	gaa	ggc	ggc	gga	gcc	gac	acc	acg	gcc	2491
Pro	Ser	Arg	Pro	Val	Gln	Tyr	Glu	Gly	Gly	Gly	Ala	Asp	Thr	Thr	Ala	
			665					670					675			
acc	gat	att	att	tgc	ccg	atg	tac	gcg	cgc	gtg	gat	gaa	gac	cag	ccc	2539
Thr	Asp	Ile	Ile	Cys	Pro	Met	Tyr	Ala	Arg	Val	Asp	Glu	Asp	Gln	Pro	
		680					685					690				
ttc	ccg	gct	gtg	ccg	aaa	tgg	tcc	atc	aaa	aaa	tgg	ctt	tcg	cta	cct	2587
Phe	Pro	Ala	Val	Pro	Lys	Trp	Ser	Ile	Lys	Lys	Trp	Leu	Ser	Leu	Pro	
		695				700					705					
gga	gag	acg	cgc	ccg	ctg	atc	ctt	tgc	gaa	tac	gcc	cac	gcg	atg	ggt	2635
Gly	Glu	Thr	Arg	Pro	Leu	Ile	Leu	Cys	Glu	Tyr	Ala	His	Ala	Met	Gly	
710					715					720				725		
aac	agt	ctt	ggc	ggt	ttc	gct	aaa	tac	tgg	cag	gcg	ttt	cgt	cag	tat	2683
Asn	Ser	Leu	Gly	Gly	Phe	Ala	Lys	Tyr	Trp	Gln	Ala	Phe	Arg	Gln	Tyr	
			730						735					740		
ccc	cgt	tta	cag	ggc	ggc	ttc	gtc	tgg	gac	tgg	gtg	gat	cag	tcg	ctg	2731
Pro	Arg	Leu	Gln	Gly	Gly	Phe	Val	Trp	Asp	Trp	Val	Asp	Gln	Ser	Leu	
			745					750					755			
att	aaa	tat	gat	gaa	aac	ggc	aac	ccg	tgg	tcg	gct	tac	ggc	ggt	gat	2779
Ile	Lys	Tyr	Asp	Glu	Asn	Gly	Asn	Pro	Trp	Ser	Ala	Tyr	Gly	Gly	Asp	
		760					765					770				
ttt	ggc	gat	acg	ccg	aac	gat	cgc	cag	ttc	tgt	atg	aac	ggt	ctg	gtc	2827
Phe	Gly	Asp	Thr	Pro	Asn	Asp	Arg	Gln	Phe	Cys	Met	Asn	Gly	Leu	Val	
		775				780					785					
ttt	gcc	gac	cgc	acg	ccg	cat	cca	gcg	ctg	acg	gaa	gca	aaa	cac	cag	2875
Phe	Ala	Asp	Arg	Thr	Pro	His	Pro	Ala	Leu	Thr	Glu	Ala	Lys	His	Gln	
		790				795				800				805		
cag	cag	ttt	ttc	cag	ttc	cgt	tta	tcc	ggg	caa	acc	atc	gaa	gtg	acc	2923
Gln	Gln	Phe	Phe	Gln	Phe	Arg	Leu	Ser	Gly	Gln	Thr	Ile	Glu	Val	Thr	
			810						815					820		
agc	gaa	tac	ctg	ttc	cgt	cat	agc	gat	aac	gag	ctc	ctg	cac	tggtat		2971
Ser	Glu	Tyr	Leu	Phe	Arg	His	Ser	Asp	Asn	Glu	Leu	Leu	His	Trp	Met	
			825					830					835			
gtg	gcg	ctg	gat	ggt	aag	ccg	ctg	gca	agc	ggt	gaa	gtg	cct	ctg	gat	3019

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Val Ala Leu Asp Gly Lys Pro Leu Ala Ser Gly Glu Val Pro Leu Asp	
840 845 850	
gtc gct cca caa ggt aaa cag ttg att gaa ctg cct gaa cta ccg cag	3067
Val Ala Pro Gln Gly Lys Gln Leu Ile Glu Leu Pro Glu Leu Pro Gln	
855 860 865	
ccg gag agc gcc ggg caa ctc tgg ctc aca gta cgc gta gtg caa ccg	3115
Pro Glu Ser Ala Gly Gln Leu Trp Leu Thr Val Arg Val Val Gln Pro	
870 875 880 885	
aac gcg acc gca tgg tca gaa gcc ggg cac atc agc gcc tgg cag cag	3163
Asn Ala Thr Ala Trp Ser Glu Ala Gly His Ile Ser Ala Trp Gln Gln	
890 895 900	
tgg cgt ctg gcg gaa aac ctc agt gtg acg ctc ccc gcc gcg tcc cac	3211
Trp Arg Leu Ala Glu Asn Leu Ser Val Thr Leu Pro Ala Ala Ser His	
905 910 915	
gcc atc ccg cat ctg acc acc agc gaa atg gat ttt tgc atc gag ctg	3259
Ala Ile Pro His Leu Thr Thr Ser Glu Met Asp Phe Cys Ile Glu Leu	
920 925 930	
ggt aat aag cgt tgg caa ttt aac cgc cag tca ggc ttt ctt tca cag	3307
Gly Asn Lys Arg Trp Gln Phe Asn Arg Gln Ser Gly Phe Leu Ser Gln	
935 940 945	
atg tgg att ggc gat aaa aaa caa ctg ctg acg ccg ctg cgc gat cag	3355
Met Trp Ile Gly Asp Lys Lys Gln Leu Leu Thr Pro Leu Arg Asp Gln	
950 955 960 965	
ttc acc cgt gca ccg ctg gat aac gac att ggc gta agt gaa gcg acc	3403
Phe Thr Arg Ala Pro Leu Asp Asn Asp Ile Gly Val Ser Glu Ala Thr	
970 975 980	
cgc att gac cct aac gcc tgg gtc gaa cgc tgg aag gcg gcg gcc cat	3451
Arg Ile Asp Pro Asn Ala Trp Val Glu Arg Trp Lys Ala Ala Gly His	
985 990 995	
tac cag gcc gaa gca gcg ttg ttg cag tgc acg gca gat aca ctt gct	3499
Tyr Gln Ala Glu Ala Ala Leu Leu Gln Cys Thr Ala Asp Thr Leu Ala	
1000 1005 1010	
gat gcg gtg ctg att acg acc gct cac gcg tgg cag cat cag ggg aaa	3547
Asp Ala Val Leu Ile Thr Thr Ala His Ala Trp Gln His Gln Gly Lys	
1015 1020 1025	
acc tta ttt atc agc cgg aaa acc tac cgg att gat ggt agt ggt caa	3595
Thr Leu Phe Ile Ser Arg Lys Thr Tyr Arg Ile Asp Gly Ser Gly Gln	
1030 1035 1040 1045	
atg gcg att acc gtt gat gtt gaa gtg gcg agc gat aca ccg cat ccg	3643
Met Ala Ile Thr Val Asp Val Glu Val Ala Ser Asp Thr Pro His Pro	
1050 1055 1060	
gcg gcg att ggc ctg aac tgc cag ctg gcg cag gta gca gag ccg gta	3691
Ala Arg Ile Gly Leu Asn Cys Gln Leu Ala Gln Val Ala Glu Arg Val	
1065 1070 1075	
aac tgg ctc gga tta ggg ccg caa gaa aac tat ccc gac cgc ctt act	3739
Asn Trp Leu Gly Leu Gly Pro Gln Glu Asn Tyr Pro Asp Arg Leu Thr	
1080 1085 1090	
gcc gcc tgt ttt gac cgc tgg gat ctg cca ttg tca gac atg tat acc	3787
Ala Ala Cys Phe Asp Arg Trp Asp Leu Pro Leu Ser Asp Met Tyr Thr	
1095 1100 1105	
ccg tac gtc ttc ccg agc gaa aac ggt ctg cgc tgc ggg acg cgc gaa	3835
Pro Tyr Val Phe Pro Ser Glu Asn Gly Leu Arg Cys Gly Thr Arg Glu	
1110 1115 1120 1125	
ttg aat tat ggc cca cac cag tgg cgc gcc gac ttc cag ttc aac atc	3883
Leu Asn Tyr Gly Pro His Gln Trp Arg Gly Asp Phe Gln Phe Asn Ile	
1130 1135 1140	
agc cgc tac agt caa cag caa ctg atg gaa acc agc cat cgc cat ctg	3931

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Ser Arg Tyr Ser Gln Gln Gln Leu Met Glu Thr Ser His Arg His Leu
      1145                1150                1155

ctg cac gcg gaa gaa ggc aca tgg ctg aat atc gac ggt ttc cat atg   3979
Leu His Ala Glu Glu Gly Thr Trp Leu Asn Ile Asp Gly Phe His Met
      1160                1165                1170

ggg att ggt ggc gac gac tcc tgg agc ccg tca gta tcg gcg gaa ttt   4027
Gly Ile Gly Gly Asp Asp Ser Trp Ser Pro Ser Val Ser Ala Glu Phe
      1175                1180                1185

cag ctg agc gcc ggt cgc tac cat tac cag ttg gtc tgg tgt caa aaa   4075
Gln Leu Ser Ala Gly Arg Tyr His Tyr Gln Leu Val Trp Cys Gln Lys
      1190                1195                1200                1205

      taata ataaccgggc aggccatgtc tgcccgtatt tcgctgaagg aaatccatta   4130

tgtactatcg atcagaccag tttttaattt gtgtgtttcc atg                   4173

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&lt;210&gt; SEQ ID NO 26

&lt;211&gt; LENGTH: 1205

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Bacillus subtilis

&lt;400&gt; SEQUENCE: 26

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Leu Gly His Tyr Ser His Ser Asp Ile Glu Glu Ala Val Lys Ser Ala
  1                5                10                15

Lys Lys Glu Gly Leu Lys Asp Tyr Leu Tyr Gln Glu Pro His Gly Lys
  20                25                30

Lys Arg Ser His Lys Lys Ser His Arg Thr His Lys Lys Ser Arg Ser
  35                40                45

His Lys Lys Ser Tyr Cys Ser His Lys Lys Ser Arg Ser His Lys Lys
  50                55                60

Ser Phe Cys Ser His Lys Lys Ser Arg Ser His Lys Lys Ser Tyr Cys
  65                70                75                80

Ser His Lys Lys Ser Arg Ser His Lys Lys Ser Tyr Arg Ser His Lys
  85                90                95

Lys Ser Arg Ser Tyr Lys Lys Ser Tyr Arg Ser Tyr Lys Lys Ser Arg
 100                105                110

Ser Tyr Lys Lys Ser Cys Arg Ser Tyr Lys Lys Ser Arg Ser Tyr Lys
 115                120                125

Lys Ser Tyr Cys Ser His Lys Lys Lys Ser Arg Ser Tyr Lys Lys Ser
 130                135                140

Cys Arg Thr His Lys Lys Ser Tyr Arg Ser His Lys Lys Tyr Tyr Lys
 145                150                155                160

Lys Pro His His His Cys Asp Asp Tyr Lys Arg His Asp Asp Tyr Asp
 165                170                175

Ser Lys Lys Glu Tyr Trp Lys Asp Gly Asn Cys Trp Val Val Lys Lys
 180                185                190

Lys Tyr Lys Val Asp Arg Glu Asn Pro Gly Val Thr Gln Leu Asn Arg
 195                200                205

Leu Ala Ala His Pro Pro Phe Ala Ser Trp Arg Asn Ser Glu Glu Ala
 210                215                220

Arg Thr Asp Arg Pro Ser Gln Gln Leu Arg Ser Leu Asn Gly Glu Trp
 225                230                235                240

Arg Phe Ala Trp Phe Pro Ala Pro Glu Ala Val Pro Glu Ser Trp Leu
 245                250                255

Glu Cys Asp Leu Pro Glu Ala Asp Thr Val Val Val Pro Ser Asn Trp

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Ala Asp Thr Thr Ala Thr Asp Ile Ile Cys Pro Met Tyr Ala Arg Val  
675 680 685

Asp Glu Asp Gln Pro Phe Pro Ala Val Pro Lys Trp Ser Ile Lys Lys  
690 695 700

Trp Leu Ser Leu Pro Gly Glu Thr Arg Pro Leu Ile Leu Cys Glu Tyr  
705 710 715 720

Ala His Ala Met Gly Asn Ser Leu Gly Gly Phe Ala Lys Tyr Trp Gln  
725 730 735

Ala Phe Arg Gln Tyr Pro Arg Leu Gln Gly Gly Phe Val Trp Asp Trp  
740 745 750

Val Asp Gln Ser Leu Ile Lys Tyr Asp Glu Asn Gly Asn Pro Trp Ser  
755 760 765

Ala Tyr Gly Gly Asp Phe Gly Asp Thr Pro Asn Asp Arg Gln Phe Cys  
770 775 780

Met Asn Gly Leu Val Phe Ala Asp Arg Thr Pro His Pro Ala Leu Thr  
785 790 795 800

Glu Ala Lys His Gln Gln Gln Phe Phe Gln Phe Arg Leu Ser Gly Gln  
805 810 815

Thr Ile Glu Val Thr Ser Glu Tyr Leu Phe Arg His Ser Asp Asn Glu  
820 825 830

Leu Leu His Trp Met Val Ala Leu Asp Gly Lys Pro Leu Ala Ser Gly  
835 840 845

Glu Val Pro Leu Asp Val Ala Pro Gln Gly Lys Gln Leu Ile Glu Leu  
850 855 860

Pro Glu Leu Pro Gln Pro Glu Ser Ala Gly Gln Leu Trp Leu Thr Val  
865 870 875 880

Arg Val Val Gln Pro Asn Ala Thr Ala Trp Ser Glu Ala Gly His Ile  
885 890 895

Ser Ala Trp Gln Gln Trp Arg Leu Ala Glu Asn Leu Ser Val Thr Leu  
900 905 910

Pro Ala Ala Ser His Ala Ile Pro His Leu Thr Thr Ser Glu Met Asp  
915 920 925

Phe Cys Ile Glu Leu Gly Asn Lys Arg Trp Gln Phe Asn Arg Gln Ser  
930 935 940

Gly Phe Leu Ser Gln Met Trp Ile Gly Asp Lys Lys Gln Leu Leu Thr  
945 950 955 960

Pro Leu Arg Asp Gln Phe Thr Arg Ala Pro Leu Asp Asn Asp Ile Gly  
965 970 975

Val Ser Glu Ala Thr Arg Ile Asp Pro Asn Ala Trp Val Glu Arg Trp  
980 985 990

Lys Ala Ala Gly His Tyr Gln Ala Glu Ala Ala Leu Leu Gln Cys Thr  
995 1000 1005

Ala Asp Thr Leu Ala Asp Ala Val Leu Ile Thr Thr Ala His Ala Trp  
1010 1015 1020

Gln His Gln Gly Lys Thr Leu Phe Ile Ser Arg Lys Thr Tyr Arg Ile  
1025 1030 1035 1040

Asp Gly Ser Gly Gln Met Ala Ile Thr Val Asp Val Glu Val Ala Ser  
1045 1050 1055

Asp Thr Pro His Pro Ala Arg Ile Gly Leu Asn Cys Gln Leu Ala Gln  
1060 1065 1070

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Val Ala Glu Arg Val Asn Trp Leu Gly Leu Gly Pro Gln Glu Asn Tyr  
 1075 1080 1085

Pro Asp Arg Leu Thr Ala Ala Cys Phe Asp Arg Trp Asp Leu Pro Leu  
 1090 1095 1100

Ser Asp Met Tyr Thr Pro Tyr Val Phe Pro Ser Glu Asn Gly Leu Arg  
 1105 1110 1115 1120

Cys Gly Thr Arg Glu Leu Asn Tyr Gly Pro His Gln Trp Arg Gly Asp  
 1125 1130 1135

Phe Gln Phe Asn Ile Ser Arg Tyr Ser Gln Gln Gln Leu Met Glu Thr  
 1140 1145 1150

Ser His Arg His Leu Leu His Ala Glu Glu Gly Thr Trp Leu Asn Ile  
 1155 1160 1165

Asp Gly Phe His Met Gly Ile Gly Gly Asp Asp Ser Trp Ser Pro Ser  
 1170 1175 1180

Val Ser Ala Glu Phe Gln Leu Ser Ala Gly Arg Tyr His Tyr Gln Leu  
 1185 1190 1195 1200

Val Trp Cys Gln Lys  
 1205

<210> SEQ ID NO 27  
 <211> LENGTH: 47  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: cotG-linker 5' primer

<400> SEQUENCE: 27

ctattgctgc agtgaacccc cacctccttt gtatttcttt ttgacta 47

<210> SEQ ID NO 28  
 <211> LENGTH: 49  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: CMCase 5' primer

<400> SEQUENCE: 28

ggcatgctgc aggcattgctc tagccgatcg gggacaaaaa cgccagtag 49

<210> SEQ ID NO 29  
 <211> LENGTH: 24  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: CMCase 3' primer

<400> SEQUENCE: 29

gccaaaaaaa agcttaacta attt 24

<210> SEQ ID NO 30  
 <211> LENGTH: 32  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: levU 5' primer

<400> SEQUENCE: 30

aagtgcctgc agatgtgaa taaagcaggc at 32

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<210> SEQ ID NO 31
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: levU 3' primer

<400> SEQUENCE: 31

aatgaaaagc ttttatttat tcaataaaga ca 32

<210> SEQ ID NO 32
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: tliA 5' primer

<400> SEQUENCE: 32

ctgcaggaat tcatgggtgt atttgactac aa 32

<210> SEQ ID NO 33
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: tliA 3' primer

<400> SEQUENCE: 33

gaagcttgcg caaggaagac tgagatg 27

<210> SEQ ID NO 34
<211> LENGTH: 2510
<212> TYPE: DNA
<213> ORGANISM: Bacillus subtilis
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (461)..(2491)

<400> SEQUENCE: 34

ggatccagtg tccctagctc cgagaaaaaa tccagagaca atttgtttct catcaaggaa 60
gggtctttat actccgcatt taagtgaatc tctcgcgcgc cgcggaatgt tttcggctga 120
taaaaggaaa tatggtatga cttctttttg aagtctctga tatgtgatcc cggataagcg 180
atatcaatat ccagcctttt ttgatttacc ttcatacag ctggcaccgg atcatcgtcc 240
catatatacct tttttaattc acgcaagtct tttggatgaa caaacagctg ataaagcggg 300
aaattggatt gattcttcat ccataatcct cttacaaaat tttaggcttt tatttttata 360
agatctcagc ggaacactta tacacttttt aaaaccgcgc gtactatgag ggtagtaagg 420
atcttcatcc ttaacatatt tttaaaagga ggatttcaaa ttg ggc cac tat tcc 475
Leu Gly His Tyr Ser
1 5

cat tct gac atc gaa gaa gcg gtg aaa tcc gca aaa aaa gaa ggt tta 523
His Ser Asp Ile Glu Glu Ala Val Lys Ser Ala Lys Lys Glu Gly Leu
10 15 20

aag gat tat tta tac caa gag cct cat gga aaa aaa cgc agt cat aaa 571
Lys Asp Tyr Leu Tyr Gln Glu Pro His Gly Lys Lys Arg Ser His Lys
25 30 35

aag tcg cac cgc act cac aaa aaa tct cgc agc cat aaa aaa tca tac 619
Lys Ser His Arg Thr His Lys Lys Ser Arg Ser His Lys Lys Ser Tyr
40 45 50

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tgc tct cac aaa aaa tct cgc agt cac aaa aaa tca ttc tgt tct cac	667
Cys Ser His Lys Lys Ser Arg Ser His Lys Lys Ser Phe Cys Ser His	
55 60 65	
aaa aaa tct cgc agc cac aaa aaa tca tac tgc tct cac aag aaa tct	715
Lys Lys Ser Arg Ser His Lys Lys Ser Tyr Cys Ser His Lys Lys Ser	
70 75 80 85	
cgc agc cac aaa aaa tcg tac cgt tct cac aaa aaa tct cgc agc tat	763
Arg Ser His Lys Lys Ser Tyr Arg Ser His Lys Lys Ser Arg Ser Tyr	
90 95 100	
aaa aaa tct tac cgt tct tac aaa aaa tct cgt agc tat aaa aaa tct	811
Lys Lys Ser Tyr Arg Ser Tyr Lys Lys Ser Arg Ser Tyr Lys Lys Ser	
105 110 115	
tgc cgt tct tac aaa aaa tct cgc agc tac aaa aag tct tac tgt tct	859
Cys Arg Ser Tyr Lys Lys Ser Arg Ser Tyr Lys Lys Ser Tyr Cys Ser	
120 125 130	
cac aag aaa aaa tct cgc agc tat aag aag tca tgc cgc aca cac aaa	907
His Lys Lys Lys Ser Arg Ser Tyr Lys Lys Ser Cys Arg Thr His Lys	
135 140 145	
aaa tct tat cgt tcc cat aag aaa tac tac aaa aaa ccg cac cac cac	955
Lys Ser Tyr Arg Ser His Lys Lys Tyr Tyr Lys Lys Pro His His His	
150 155 160 165	
tgc gac gac tac aaa aga cac gat gat tat gac agc aaa aaa gaa tac	1003
Cys Asp Asp Tyr Lys Arg His Asp Asp Tyr Asp Ser Lys Lys Glu Tyr	
170 175 180	
tgg aaa gac ggc aat tgc tgg gta gtc aaa aag aaa tac aaa gga ggt	1051
Trp Lys Asp Gly Asn Cys Trp Val Val Lys Lys Lys Tyr Lys Gly Gly	
185 190 195	
ggg ggt tca ctg cag gca tgc gct agc cga tcg ggg aca aaa acg cca	1099
Gly Gly Ser Leu Gln Ala Cys Ala Ser Arg Ser Gly Thr Lys Thr Pro	
200 205 210	
gta gcc aag aat ggc cag ctt agc ata aaa ggt aca cag ctc gtt aac	1147
Val Ala Lys Asn Gly Gln Leu Ser Ile Lys Gly Thr Gln Leu Val Asn	
215 220 225	
cga gac ggt aaa gcg gta cag ctg aag ggg atc agt tca cac gga ttg	1195
Arg Asp Gly Lys Ala Val Gln Leu Lys Gly Ile Ser Ser His Gly Leu	
230 235 240 245	
caa tgg tat gga gaa tat gtc aat aaa gac agc tta aaa tgg ctg agg	1243
Gln Trp Tyr Gly Glu Tyr Val Asn Lys Asp Ser Leu Lys Trp Leu Arg	
250 255 260	
gac gat tgg ggt atc acc gtt ttc cgt gca gcg atg tat acg gca gat	1291
Asp Asp Trp Gly Ile Thr Val Phe Arg Ala Ala Met Tyr Thr Ala Asp	
265 270 275	
ggc ggt ata att gac aac ccg tcc gtg aaa aat aaa atg aaa gaa gcg	1339
Gly Gly Ile Ile Asp Asn Pro Ser Val Lys Asn Lys Met Lys Glu Ala	
280 285 290	
gtt gaa gcg gca aaa gag ctt ggg ata tat gtc atc att gac tgg cat	1387
Val Glu Ala Ala Lys Glu Leu Gly Ile Tyr Val Ile Ile Asp Trp His	
295 300 305	
atc tta aat gac ggt aat cca aac caa aat aaa gag aag gca aaa gaa	1435
Ile Leu Asn Asp Gly Asn Pro Asn Gln Asn Lys Glu Lys Ala Lys Glu	
310 315 320 325	
ttc ttc aag gaa atg tca agc ctt tac gga aac acg cca aac gtc att	1483
Phe Phe Lys Glu Met Ser Ser Leu Tyr Gly Asn Thr Pro Asn Val Ile	
330 335 340	
tat gaa att gca aac gaa cca aac ggt gat gtg aac tgg aag cgt gat	1531
Tyr Glu Ile Ala Asn Glu Pro Asn Gly Asp Val Asn Trp Lys Arg Asp	
345 350 355	

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att aaa ccg tat gcg gaa gaa gtg att tcc gtt atc cgc aaa aat gat Ile Lys Pro Tyr Ala Glu Glu Val Ile Ser Val Ile Arg Lys Asn Asp 360 365 370	1579
cca gac aac att atc att gtc gga acc ggt aca tgg agc cag gat gtg Pro Asp Asn Ile Ile Ile Val Gly Thr Gly Thr Trp Ser Gln Asp Val 375 380 385	1627
aat gat gct gcc gat gac cag cta aaa gat gca aac gtt atg gac gca Asn Asp Ala Ala Asp Asp Gln Leu Lys Asp Ala Asn Val Met Asp Ala 390 395 400 405	1675
ctt cat ttt tat gcc gcc aca cac gcc caa ttt tta cgg gat aaa gca Leu His Phe Tyr Ala Gly Thr His Gly Gln Phe Leu Arg Asp Lys Ala 410 415 420	1723
aac tat gca ctc agc aaa gga gca cct att ttt gtg aca gag tgg gga Asn Tyr Ala Leu Ser Lys Gly Ala Pro Ile Phe Val Thr Glu Trp Gly 425 430 435	1771
aca agc gac gcg tct gcc aat gcc ggt gta ttc ctt gat caa tcg agg Thr Ser Asp Ala Ser Gly Asn Gly Gly Val Phe Leu Asp Gln Ser Arg 440 445 450	1819
gaa tgg ctg aaa tat ctc gac agc aag acc atc agc tgg gtg aac tgg Glu Trp Leu Lys Tyr Leu Asp Ser Lys Thr Ile Ser Trp Val Asn Trp 455 460 465	1867
aat ctt tct gat aag cag gaa tca tcc tca gct tta aag ccg ggg gca Asn Leu Ser Asp Lys Gln Glu Ser Ser Ser Ala Leu Lys Pro Gly Ala 470 475 480 485	1915
tct aaa aca gcc gcc tgg cgg ttg tca gat tta tct gct tca gga aca Ser Lys Thr Gly Gly Trp Arg Leu Ser Asp Leu Ser Ala Ser Gly Thr 490 495 500	1963
ttc gtt aga gaa aac att ctc gcc acc aaa gat tcg acg aag gac att Phe Val Arg Glu Asn Ile Leu Gly Thr Lys Asp Ser Thr Lys Asp Ile 505 510 515	2011
cct gaa acg cca gca aaa gat aaa ccc aca cag gaa aac ggt att tct Pro Glu Thr Pro Ala Lys Asp Lys Pro Thr Gln Glu Asn Gly Ile Ser 520 525 530	2059
gta caa tac aga gca ggg gat ggg agt atg aac agc aac caa atc cgt Val Gln Tyr Arg Ala Gly Asp Gly Ser Met Asn Ser Asn Gln Ile Arg 535 540 545	2107
ccg cag ctt caa ata aaa aat aac gcc aat acc acg gtt gat tta aaa Pro Gln Leu Gln Ile Lys Asn Asn Gly Asn Thr Thr Val Asp Leu Lys 550 555 560 565	2155
gat gtc act gcc cgt tac tgg tat aac gcg aaa aac aaa gcc caa aac Asp Val Thr Ala Arg Tyr Trp Tyr Asn Ala Lys Asn Lys Gly Gln Asn 570 575 580	2203
gtt gac tgt gac tac gcg cag ctt gga tgc gcc aat gtg aca tac aag Val Asp Cys Asp Tyr Ala Gln Leu Gly Cys Gly Asn Val Thr Tyr Lys 585 590 595	2251
ttt gtg acg ttg cat aaa cca aag caa ggt gca gat acc tat ctg gaa Phe Val Thr Leu His Lys Pro Lys Gln Gly Ala Asp Thr Tyr Leu Glu 600 605 610	2299
ctt gga ttt aaa aac gga acg ctg gca ccg gga gca agc aca ggg aat Leu Gly Phe Lys Asn Gly Thr Leu Ala Pro Gly Ala Ser Thr Gly Asn 615 620 625	2347
att cag ctt cgt ctt cac aat gat gac tgg agc aat tat gca caa agc Ile Gln Leu Arg Leu His Asn Asp Asp Trp Ser Asn Tyr Ala Gln Ser 630 635 640 645	2395
ggc gat tat tcc ttt ttc aaa tca aat acg ttt aaa aca acg aaa aaa Gly Asp Tyr Ser Phe Phe Lys Ser Asn Thr Phe Lys Thr Thr Lys Lys 650 655 660	2443



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Thr Pro Asn Val Ile Tyr Glu Ile Ala Asn Glu Pro Asn Gly Asp Val  
 340 345 350  
 Asn Trp Lys Arg Asp Ile Lys Pro Tyr Ala Glu Glu Val Ile Ser Val  
 355 360 365  
 Ile Arg Lys Asn Asp Pro Asp Asn Ile Ile Ile Val Gly Thr Gly Thr  
 370 375 380  
 Trp Ser Gln Asp Val Asn Asp Ala Ala Asp Asp Gln Leu Lys Asp Ala  
 385 390 395 400  
 Asn Val Met Asp Ala Leu His Phe Tyr Ala Gly Thr His Gly Gln Phe  
 405 410 415  
 Leu Arg Asp Lys Ala Asn Tyr Ala Leu Ser Lys Gly Ala Pro Ile Phe  
 420 425 430  
 Val Thr Glu Trp Gly Thr Ser Asp Ala Ser Gly Asn Gly Gly Val Phe  
 435 440 445  
 Leu Asp Gln Ser Arg Glu Trp Leu Lys Tyr Leu Asp Ser Lys Thr Ile  
 450 455 460  
 Ser Trp Val Asn Trp Asn Leu Ser Asp Lys Gln Glu Ser Ser Ser Ala  
 465 470 475 480  
 Leu Lys Pro Gly Ala Ser Lys Thr Gly Gly Trp Arg Leu Ser Asp Leu  
 485 490 495  
 Ser Ala Ser Gly Thr Phe Val Arg Glu Asn Ile Leu Gly Thr Lys Asp  
 500 505 510  
 Ser Thr Lys Asp Ile Pro Glu Thr Pro Ala Lys Asp Lys Pro Thr Gln  
 515 520 525  
 Glu Asn Gly Ile Ser Val Gln Tyr Arg Ala Gly Asp Gly Ser Met Asn  
 530 535 540  
 Ser Asn Gln Ile Arg Pro Gln Leu Gln Ile Lys Asn Asn Gly Asn Thr  
 545 550 555 560  
 Thr Val Asp Leu Lys Asp Val Thr Ala Arg Tyr Trp Tyr Asn Ala Lys  
 565 570 575  
 Asn Lys Gly Gln Asn Val Asp Cys Asp Tyr Ala Gln Leu Gly Cys Gly  
 580 585 590  
 Asn Val Thr Tyr Lys Phe Val Thr Leu His Lys Pro Lys Gln Gly Ala  
 595 600 605  
 Asp Thr Tyr Leu Glu Leu Gly Phe Lys Asn Gly Thr Leu Ala Pro Gly  
 610 615 620  
 Ala Ser Thr Gly Asn Ile Gln Leu Arg Leu His Asn Asp Asp Trp Ser  
 625 630 635 640  
 Asn Tyr Ala Gln Ser Gly Asp Tyr Ser Phe Phe Lys Ser Asn Thr Phe  
 645 650 655  
 Lys Thr Thr Lys Lys Ile Thr Leu Tyr Asp Gln Gly Lys Leu Ile Trp  
 660 665 670  
 Gly Thr Glu Pro Asn  
 675

&lt;210&gt; SEQ ID NO 36

&lt;211&gt; LENGTH: 24

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Hepatitis B virus

&lt;400&gt; SEQUENCE: 36

Met Gln Trp Asn Ser Thr Thr Phe His Leu Gln Asp Pro Arg Val Arg

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1	5	10	15
Gly Leu Tyr Phe Pro Ala Gly Gly			
20			
<210> SEQ ID NO 37 <211> LENGTH: 30 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Fv 5' primer  <400> SEQUENCE: 37  gaggctagct cgactgagga gtctggagga <span style="float: right;">30</span>			
<210> SEQ ID NO 38 <211> LENGTH: 30 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Fv 3' primer  <400> SEQUENCE: 38  ggagggccct taacgtttta tttccaggta <span style="float: right;">30</span>			
<210> SEQ ID NO 39 <211> LENGTH: 29 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: egfp 5' primer  <400> SEQUENCE: 39  cggctagcgc tatggtgagc aagggcgag <span style="float: right;">29</span>			
<210> SEQ ID NO 40 <211> LENGTH: 32 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: egfp 3' primer  <400> SEQUENCE: 40  gcgggcccaa gcttttactt gtacagctcg tc <span style="float: right;">32</span>			
<210> SEQ ID NO 41 <211> LENGTH: 32 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: gfpuv 5' primer  <400> SEQUENCE: 41  gcggatccct gcagatgagt aaaggagaag aa <span style="float: right;">32</span>			
<210> SEQ ID NO 42 <211> LENGTH: 32 <212> TYPE: DNA			

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<213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: gfpuv 3' primer

<400> SEQUENCE: 42

cgaagcttga attcttattt gtagagctca tc

32

What is claimed is:

1. A method for displaying a protein of interest on spore surface, which comprises the steps of:

- (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein of interest, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein of interest;
- (ii) transforming a host cell with the vector for spore surface display;
- (iii) displaying the protein of interest on a surface of a spore of the host cell; and
- (iv) recovering the spore displaying on its surface the protein of interest.

2. A method for improving a protein of interest, which comprises the steps of:

- (i) constructing a gene library of the protein of interest;
- (ii) preparing a vector by linking the gene library to a gene encoding spore coat protein;
- (iii) transforming a spore-forming host cell with the vector;
- (iv) forming a spore in the transformed host cell and displaying the protein of interest on a surface of the spore;
- (v) recovering the spore displaying on its surface the protein of interest; and
- (vi) screening the spore displaying a variant of the protein of interest having a desired property.

3. The method according to claim 2, wherein the screening is performed by means of measuring an activity of the protein or flow cytometry.

4. A method for improving a protein of interest using a resistance property of spore, which comprises the steps of:

- (i) constructing a gene library of the protein of interest;
- (ii) preparing a vector by linking the gene library to a gene encoding spore coat protein;
- (iii) transforming a spore-forming host cell with the vector;
- (iv) forming a spore in the transformed host cell and displaying the protein of interest on a surface of the spore;

- (v) treating the spore displaying on its surface the protein of interest with one or more selected from the group consisting of organic solvent, heat, acid, base, oxidant, dryness, surfactant and protease;

- (vi) recovering the spore displaying on its surface the protein of interest; and

- (vii) screening the spore displaying a variant of the protein of interest having a resistance to the treatment.

5. The method according to claim 4, wherein the screening is performed using an activity of the protein or a structural stability of the protein.

6. A method for bioconversion, which comprises the steps of:

- (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein of interest conducting a bioconversion reaction, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein of interest;

- (ii) transforming a host cell with the vector for spore surface display;

- (iii) displaying the protein of interest on a surface of a spore of the host cell;

- (iv) recovering the spore displaying on its surface the protein of interest; and

- (v) performing the bioconversion reaction using the spore displaying on its surface the protein of interest.

7. A method for preparing protein microarray, which comprises the steps of:

- (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding antibody or antigen having binding affinity to a protein to be analyzed, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the antibody or antigen;

- (ii) transforming a host cell with the vector for spore surface display;

- (iii) displaying the antibody or antigen on a surface of a spore of the host cell;

- (iv) recovering the spore displaying on its surface the antibody or antigen; and

- (v) immobilizing onto a solid surface the spore displaying on its surface the antibody or antigen.

8. A method producing an antibody to antigen in vertebrates, which comprises the steps of:

- (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding the antigen, wherein, when

expressed, the gene construct expresses a fusion protein between the spore coat protein and the antigen;

- (ii) transforming a host cell with the vector for spore surface display;
- (iii) displaying the antigen on a surface of a spore of the host cell;
- (iv) recovering the spore displaying on its surface the antigen; and
- (v) administering to vertebrates a composition containing an immunologically effective amount of the spore displaying on its surface the antigen.

9. A method for preparing a whole cell absorber, which comprises the steps of:

- (i) preparing a vector for spore surface display comprising a gene construct containing a gene encoding spore coat protein and a gene encoding a protein having a binding affinity to a certain substance, wherein, when expressed, the gene construct expresses a fusion protein between the spore coat protein and the protein;
- (ii) transforming a host cell with the vector for spore surface display;
- (iii) displaying the protein on a surface of a spore of the host cell;
- (iv) recovering the spore displaying on its surface the protein; and
- (v) immobilizing onto a carrier the spore displaying on its surface the protein.

10. The method according to any one of claims 1-9, wherein the gene encoding spore coat protein is derived from a spore-forming Gram negative bacterium including Myxococcus, a spore-forming Gram positive bacterium including Bacillus, a spore-forming Actinomycete, a spore-forming yeast or a spore-forming fungus.

11. The method according to claim 10, wherein the gene encoding spore coat protein is derived from a spore-forming Gram positive bacterium.

12. The method according to claim 11, wherein the gene encoding spore coat protein is derived from Bacillus.

13. The method according to any one of claims 1-9, wherein the gene encoding spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA.

14. The method according to any one of claims 1-9, wherein the gene encoding spore coat protein is selected from the group consisting of cotA, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA.

15. The method according to any one of claims 1-9, wherein the gene encoding spore coat protein is a modified form or a recombinant form of one selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, in which the modified form or the recombinant form has a more compatibility for spore surface display relative to wild type genes.

16. The method according to claim 15, wherein the modified form of the gene encoding spore coat protein is obtained by a method selected from the group consisting of DNA shuffling method, StEP method, RPR method, molecu-

lar breeding method, ITCHY method, error-prone PCR, point mutagenesis, nucleotide mutagenesis, combinatorial cassette mutagenesis and other suitable random mutagenesis.

17. The method according to any one of claims 1-9, wherein the gene encoding spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, in which the gene has a substituted promoter for its promoter to enhance spore surface display relative to wild type genes.

18. The method according to claim 13, wherein the gene encoding spore coat protein is cotE or cotG.

19. The method according to claim 14, wherein the gene encoding spore coat protein is cotE or cotG.

20. The method according to claim 15, wherein the gene encoding spore coat protein is cotE or cotG.

21. The method according to claim 16, wherein the gene encoding spore coat protein is cotE or cotG.

22. The method according to claim 16, wherein the gene encoding spore coat protein is cotE or cotG.

23. The method according to any one of claims 1-5, wherein the protein of interest is selected from the group consisting of enzyme, enzyme inhibitor, hormone, hormone analogue, hormone receptor, signal transduction protein, antibody, monoclonal antibody, antigen, adhesive protein, structural protein, regulatory protein, toxin protein, cytokine, transcription regulatory protein, blood clotting protein, plant protection-inducing protein and fragments thereof.

24. The method according to any one of claims 1-9, wherein the host cell is selected from the group consisting of a spore-forming Gram negative bacterium including Myxococcus, a spore-forming Gram positive bacterium including Bacillus, a spore-forming Actinomycete, a spore-forming yeast or a spore-forming fungus.

25. The method according to claim 24, wherein the host cell is a spore-forming Gram positive bacterium.

26. The method according to claim 25, wherein the host cell is Bacillus.

27. The method according to any one of claims 1-9, wherein the spore is reproductive or non-reproductive one.

28. The method according to any one of claims 1-9, wherein the recovering is performed in such a manner that the display of the protein of interest on the spore surface is maximized by regulating culture time, after which culturing is terminated and the spore is then recovered.

29. The method according to any one of claims 2-5, wherein the constructing a gene library is performed by a mutagenesis of the gene encoding the protein of interest of wild type, in which the mutagenesis is selected from the group consisting of DNA shuffling method, StEP method, RPR method, molecular breeding method, ITCHY method, error-prone PCR, point mutagenesis, nucleotide mutagenesis, combinatorial cassette mutagenesis and other suitable random mutagenesis.

30. A microbial transformant for spore surface display of a protein of interest, characterized in that the transformant is produced by transformation with a vector for spore surface display containing (i) a gene encoding a protein of interest and (ii) a gene encoding spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA, in

which when expressed, a fusion protein between the spore coat protein and the protein of interest is expressed.

**31.** The transformant according to claim 30, wherein the transformant is derived from a variant mutated to enhance spore surface display.

**32.** The transformant according to claim 31, wherein the mutation to enhance spore surface display eliminates a production of extracellular secretory protease in the transformant, so that the protein of interest displayed on spore surface is stably maintained.

**33.** The transformant according to claim 31, wherein the mutation to enhance spore surface display eliminates a production of intracellular protease in the transformant.

**34.** A spore for spore surface display of a protein of interest, characterized in that the spore displays the protein of interest on its surface.

**35.** The spore according to claim 34, wherein the spore is reproductive or non-reproductive one.

**36.** The spore according to claim 35, wherein the spore is non-reproductive one by virtue of one or more methods selected from the group consisting of genetic method, chemical method and physical method.

**37.** The spore according to claim 36, wherein the genetic method to make the -spore non-reproductive is accomplished by deleting a gene involved in reproduction of spore.

**38.** The spore according to claim 34, wherein the spore is derived from a variant mutated to increase its agglutination property.

**39.** The spore according to claim 38, wherein the increase of the agglutination property in the spore is accomplished by one or more methods selected from the group consisting of genetic method, chemical method and physical method.

**40.** A vector for spore surface display, characterized in that the vector comprises a replication origin, an antibiotic-resistance gene, a restriction site, a gene encoding a spore coat protein, a gene encoding a protein of interest and a promoter operatively linked to the gene encoding a spore coat protein, in which when expressed, a fusion protein between the spore coat protein and the protein of interest is expressed.

**41.** The vector according to claim 40, wherein the gene encoding a spore coat protein is selected from the group consisting of cotA, cotB, cotC, cotD, cotE, cotF, cotG, cotH, cotJA, cotJC, cotK, cotL, cotM, cotS, cotT, cotV, cotW, cotX, cotY, cotZ, spoIVA, spoVID and sodA.

**42.** The vector according to claim 41, wherein the gene encoding a spore coat protein is cotE or cotG.

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专利名称(译)	在孢子表面上表达蛋白质的方法		
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摘要(译)

本发明涉及一种在孢子表面上展示蛋白质的方法和使用该方法的快速改善蛋白质的方法，该方法包括以下步骤：(i) 制备用于孢子表面展示的载体，其包含含有编码孢子外壳的基因的基因构建体蛋白质和编码目的蛋白质的基因，其中，当表达时，基因构建体在孢子外壳蛋白和目标蛋白质之间表达融合蛋白，(ii) 用载体转化宿主细胞用于孢子表面展示；(iii) 在宿主细胞的孢子表面上展示目的蛋白质；(iv) 回收在其表面上显示目的蛋白质的孢子。

