

US006825008B2

(12) United States Patent

Davis et al.

(10) Patent No.: US 6,825,008 B2

(45) **Date of Patent:** Nov. 30, 2004

(54) EXPRESSED LIGAND—VASCULAR INTERCELLULAR SIGNALLING MOLECULE

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(*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 136 days.

(21) Appl. No.: 10/225,060

(22) Filed: Aug. 21, 2002

(65) **Prior Publication Data**

US 2003/0092891 A1 May 15, 2003

Related U.S. Application Data

(62) Division of application No. 09/709,188, filed on Nov. 9, 2000, now Pat. No. 6,441,137, which is a continuation of application No. 08/740,223, filed on Oct. 25, 1996, now Pat. No. 6,265,564.

(60) Provisional application No. 60/022,999, filed on Aug. 2,

(51) **Int. Cl.**⁷ **G12P 21/06**; C12N 15/00; C12N 5/02; C12N 1/20; C07H 21/04

(52) **U.S. Cl.** **435/69.1**; 536/23.4; 435/320.1; 435/252.3; 435/325; 435/455

(56) References Cited

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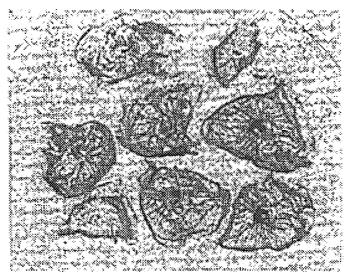
Primary Examiner—Christina Chan Assistant Examiner—Maher Haddad (74) Attorney, Agent, or Firm—Valeta Gregg, Esq.

(57) ABSTRACT

The present invention provides for a modified TIE-2 ligand which has been altered by addition, deletion or substitution of one or more amino acids, or by way of tagging, with for example, the Fc portion of human IgG-1, but which retains its ability to bind the TIE-2 receptor. The invention further provides for a modified TIE-2 ligand which is a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a portion of a second TIE-2 ligand which is different from the first. In a specific embodiment, the invention further provides for a chimeric TIE ligand comprising at least a portion of TIE-2 Ligand-1 and a portion of TIE-2 Ligand-2. In addition the present invention provides for isolated nucleic acid molecule encoding the modified TIE-2 ligands described. The invention also provides for therapeutic compositions as well as a method of blocking blood vessel growth, a method of promoting neovascularization, a method of promoting the growth or differentiation of a cell expressing the TIE receptor, a method of blocking the growth or differentiation of a cell expressing the TIE receptor and a method of attenuating or preventing tumor growth in a human.

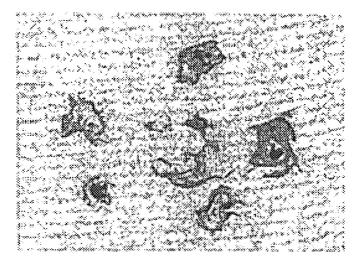
7 Claims, 47 Drawing Sheets

Fig. 1 A



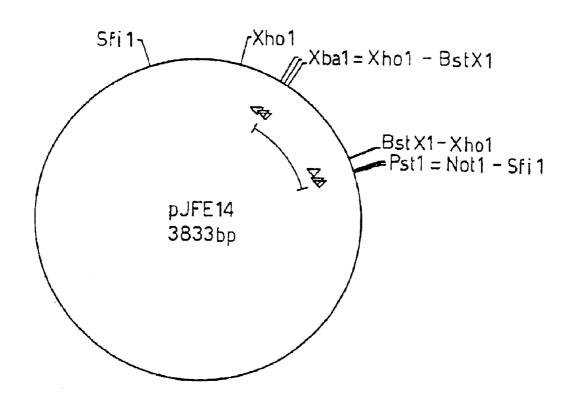
r EHK-1 lecto/h lgG1 Fc Gelloam (6ug)

Fig. 1 B



r TiE-2 ecto/h lgG1 Fc Gelloam (6ug)

Fig. 2



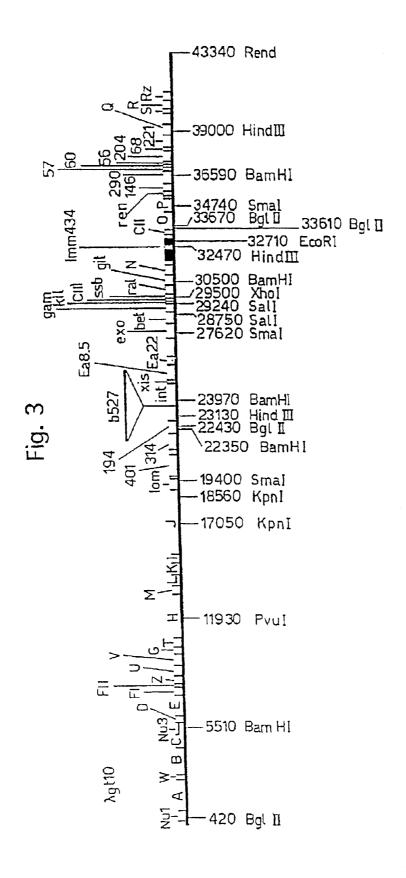


Fig. 4 A

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Nov. 30, 2004

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. 6 D 1600 1610 1620 1630 1640 NANA ATA AGC AGC ATC AGG CAA CGA AAT GAT TIT AGC ACA ANG GAT GAT GAC AAC GAC K I S S I S O P G N D F S T K D G D N D>	1660 1670 1680 1690 1700 ATT TGC AAA TGT TCA CAA ATG CTA ACA GGA GGC TGG TTT GAT GCA TGT GGT I C K C S Q H L T G G W W F D A C G>	1720 1730 1740 1750 1760 AAC TTG AAC GGA ATG TAC TAT CCA CAG AGG CAG AAC ACA AAT AAG TTC AAC GGC N L N G M Y Y P Q R Q N T N K F N G>	10 1780 1890 1800 1810 1820 ATT ANA TGG TAC TGG ANA GGC TCA GGC TAT TCG CTC ANG GCC ACA ACC ATG ATG ATC ATG	1840 1850 1860 1870 1880 1890 1900 GCA GAT TIC TANACATCCCAGTCCACTGAGGAACTGTCTCGAACTATTTTCAAAGACTTAAAGCCCAGT A D F>	1910 1920 1930 1940 1950 1960 1970 1980 GCACTGAAAGTCACGACTGTCCTCTCTCCACAGAGGCGTGTGCTCGGTGTTGAGGGGGACCCACATGCT	1990 2000 2010 2020 2030 2040 2050 2060 CCAGATTAGAGCCTAAACTTTATCATCATCATCATCATCATAATT	2010 2080 2090 2100 2110 2120 2130 2140 creatyragacacaccetaticcaaacatcaacccaacctgacaatcagactgacaatcagactgacaa	2150 2160 2170 2180 2190 2200 2210 2220 ccaagaatgttatgtgaataataactaaaatgaactttatgaactaaataactaaaatgaactttatgaactaaaatgaactaaaaataacaaaatgaactttaaaaatgaacaaaaataaaaatgaacaaaaatgaacaaaaatgaacaaaaatgaacaaaaatgaacaaaaaaaa	2230 2240 2250 2260 2270 2280 ACTIGGATTICTICTICAGGACTIGTTATAGACTIGTGTÄAATAGCCATATGTCCTGAATTIC
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Fig. 6 L	1650 AAA K	1710 CC7	1770 • ATT	1830 CGA R	٦ <u>ئ</u>	1 5	2 GTG	8	., 5
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Fig. 7

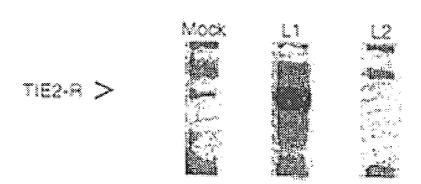


Fig. 8

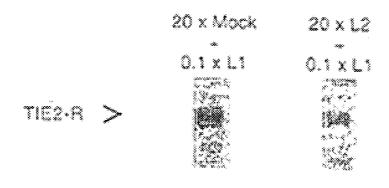
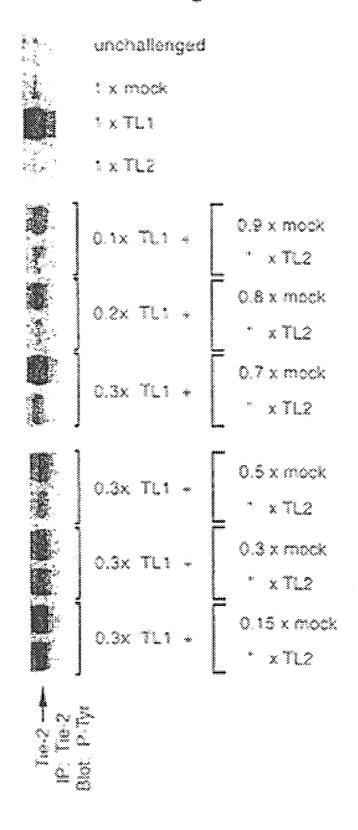
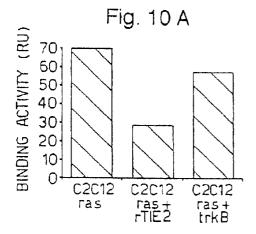
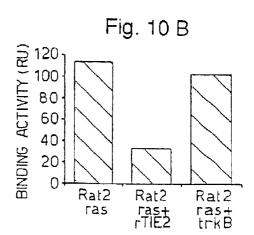
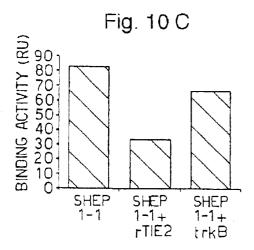


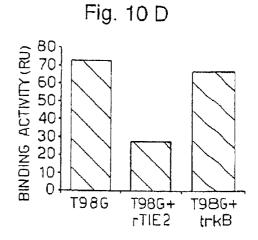
Fig. 9

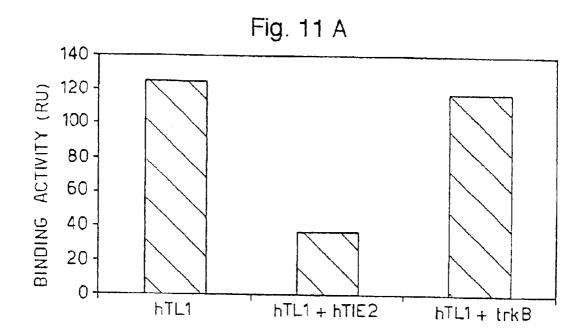












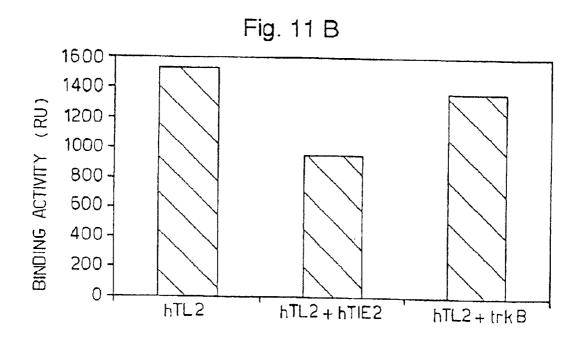
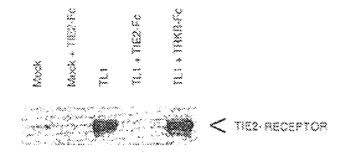
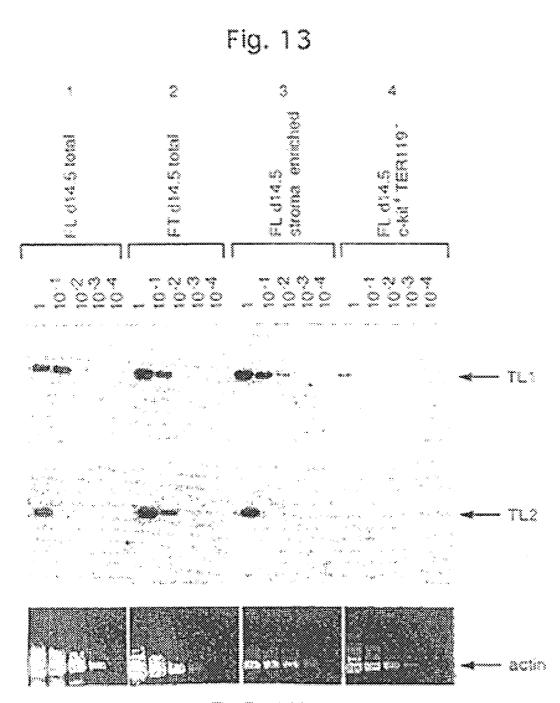


Fig. 12





FL: Fetal Liver

Fig. 14

Fetal Thymus E17.5

COR!*: Cortical stromal cells

A285 *: Medulia stromal delis

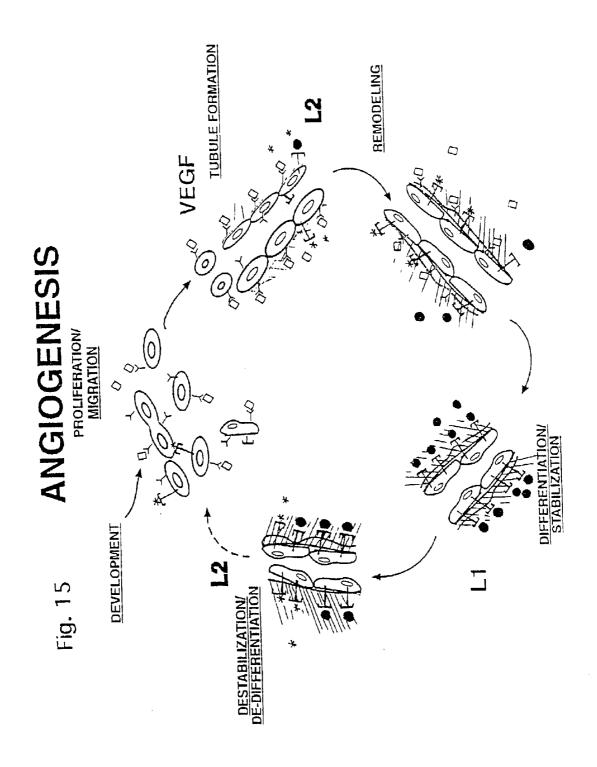


Fig. 16 C 0 244 244 500

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30 * * * CAYTFILPEH	30 . CaYTF]LPE	110 * * PATMIEI GTS	110 Pavadei Gyn	190 * * HKEELD TLK	190 Hilqîq siko	270	PEDCADVY ONG	270 RDCAeVf ksG	350	TSQRQ YMLR	350 InQqr Yulk	430 GWPD ACCE	430 GWMFD ACGPS
20 * * RRYNRIQHGQ	20 kkgygvQHGs	100 ICONAV ONE:	100 IQONAV QNQI	180 * HKILE MEGK	180 Krvla medik	260	GKREEE KPF	260 vakeeq isf	340	ALGNE FIFAI	340 WLGNE Fysgl	420 * * ICKCA LMLTGG	420 1CKCs GALAG
10 PRRSPERISG	10 FRkSmds1G	90 • 20aks emaq	90 NYKK Ewel	170 * LHEK NSLLED	170 lqdk nsfle	250	KEGV LLKG	n Smaa kdpto	330	PGN PSGEY	D FGN PSGEYW	410 TOA DADACM	og probect
Ħ	BaymM	* ENYIVE	BNYIqd	TWEILK	I TSEInK	*	LVML-CT	250 LltmsTs	*	WKGYKUAGI	330 WREYRVGF	* HGADFSTE	410 pgnDFSTRI
17.1	71.2	TE.1	TL2	111	TL2		11.1	TL2		1777	41.2	TLI	17.
7													

Fig. 18

COVALENT MULTIMERIC STRUCTURE OF TL1 AND TL2 AND THEIR INTERCONVERSION BY THE MUTATION OF ONE CYSTEINE

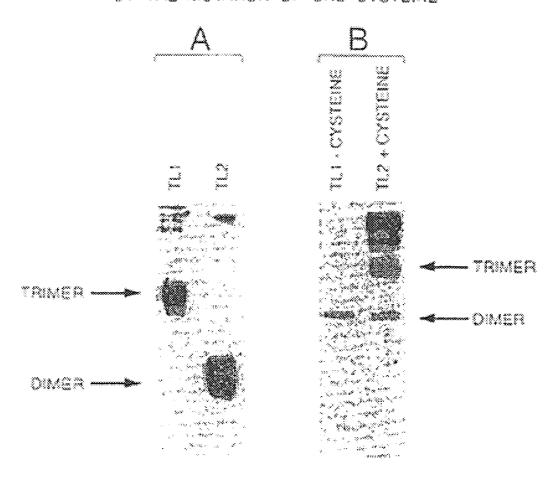


Fig. 19

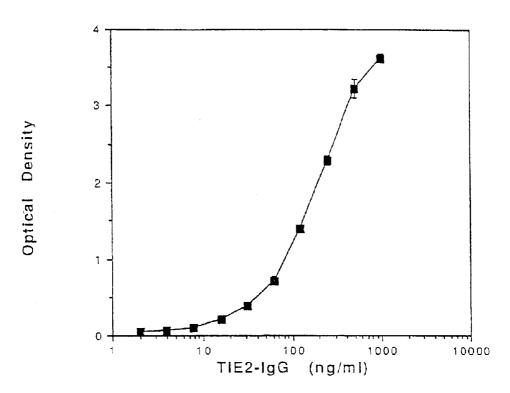
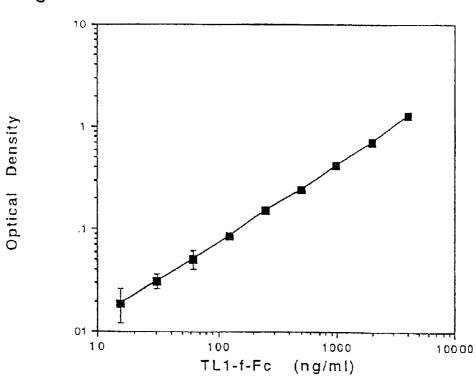


Fig. 20



5 * CTG	* AGC	CTC J.	CTG E	cAG \$	510 * 3 ACC T>	* 55 t	e g	chG Q
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ACC1	<i>\bar{U}</i> ≈	* GTO >	270 * : TTG	, gr	GRC 5	ACT	* 85 m '	690 4 G GAA
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* GAG GCT GTG CAC CGC C * TTA L ATC TAT (1070 * CGG GAG 6 820 * AGC 5 1060 * * * 3 ACC CTC ATC CAG CAC C * * CCG CAT CCG 810 * AGC S * Grc > AGT * * * * CTG ART GAC AGC AGC 1050 * * * . GGT GGC TGG P * CGG ATT R I 1040 * GAT GGA D G 950 * * * 3 ATC RAG CGC 1 1190 1200 * * * * 3 GCC TAC TIG CTA CGC G'

GGC CTC TCC AAC CTC AAT GGC ATC TAC TAT TCA GTT CAT CAG CAC TTG CAC AAG ATC AAT GGC ATC CGC TGG CAC TAC TTC CGA G L S N L N G I Y Y S V N Q H L K K I N G I R W H Y F R> CCGTAGGAGG ATTCTCAACC CAGGIGACTC TGTGCACGCT GGGCCCTGCC CAGANATCAG TGCCCAGGGC TCATCTTGAC ATTCTGGAAC ATCGGAACCA OCITACCITG CCCCIGARIT ACAAGAAITC ACCIGCCICC CIGITGCCCI CIARITGIGA AAITGCIGGG IGCIIGAAGG CACCIGCCIC IGIIGGAACC 1670 1680 1690 1580 1590 * * * * 1760 1770 1660 * 1610 1620 1630 1640 1650 * * * * * * * * * * 1710 1720 · 1730 1740 * * * * * * 1840 1830 1820

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Fig. 21 C

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Fig. 22 B

400 ENFQLG Jr.hi.> Jr.hi.> Jr.hi.> Jh.y.a>	480 TINGIR 1k> 1k> fk> fk>	
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DMETUGGGWT LIQHREDGSV NFQRTWEEVK EGFGNVAREH WLGNEAVHRL TSRTAYLLRV ELHDWEGAQT SIQYENFQLG n.dvnvn. vl dg.k mpsg.yfifaiqrq.m.i .mnra ysdr.hi.>n.vn vl dg.k mpsg.yfifaiqrq.m.i .mnra ysdr.hi.>n.dvn vl dg.k mplg.yfifai .qrq.m.i .mnra ysdr.hi.>dvg vqrg.x i.g.knea hsl.dh.y.a>agi.rnea yslh.y.s>	SERORYSLSV NDSSSSAGRK NSLAPQCTKF STKDMDNDNC MCKCAQMLSG GWWFDACGLS NLNGITYSVH QHLHFKINGIR n.k.n.r.yl kghtgt.kq s.ilh.adal.tpmf.tag .nhg.lk> n.k.n.r.yl kghtgt.kq s.ilh.adal.tpmf.tag .nhg.lk> n.k.n.r.yl kghtgt.kq s.ilh.adal.tpmf.tag .nhg.lk> g.esn.rihl tgltgt.aki s.isqp.sdsk. istpqpqk .ntn.fk>eln.rihl kgltgt.ki s.isqp.ndgk. istpmpqr .ntn.fk>	490 500 WHYFRGPSYS INGTRMHLRP MGA*k.r. rs.t.i. ldfk.r. rs.t.i. ldf> .y.wk.sg. ka.t.i. adf>
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730 740 750 760 760 770 780 790 GGT GTC AGG CAC CAG CAG CAG CAG CAG CAG CAG CTG CTG GTG TTG GTG TTG GT TTG 930 940 950 960 970 980 990 GTG TCC ART GCA AGG AGG TGG TGG ACC CTG CAG AGC AGT GGA GGC AGG TGG ACC V S N A T K P R K V F C D L Q S S G G R W T GAG AAG CGG TTG CAG GCC CTG GAG ACC AAG CAG CAG GAG GTG GCC AGC ATC CTC AGC AAG AAG E K R L Q A L E T K Q Q E E L A S I L S K K GCG AAG CTG TA S R Q S A L T N I E R G L R 1000 1010 1020 1030 1040 1050 CCC ATC CAG CGC CGT GAG AAT GGC ACC GTG AAT TTT CAG CGG AAC TGG AAG GAT TAC AAA CAG GGC L I Q R R E N G T V N F Q R N W K D Y K Q G 1060 1070 1080 1090 1100 1110 1120 TTC GGA GAC CCA GCT GGG CTG GGC AAT GAA GTG GTG CAC CAG CTC ACC AGA AGG GCA F G D P A G E H W L G N E V V H Q L T R R A 1130 1140 1150 1160 1170 1180 GCC TAC TCT CTG CGT GTG GAG GAG GCC GAG GCC TAT GCC CAG TAC GAA CAT A Y S L R V E L Q D W E G H E A Y A Q Y E H \Box

Nov. 30, 2004

Fig. 23 (

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EXPRESSED LIGAND— VASCULAR INTERCELLULAR SIGNALLING MOLECULE

This application is a divisional application of U.S. Ser. 5 No. 09/709,188 filed 9 Nov. 2000, now U.S. Pat. No. 6,441,137, which is a continuation application of U.S. Ser. No. 08/740,223, filed on Oct. 25, 1996, now U.S. Pat. No. 6,265,564, which claims the priority of U.S. Provisional application No. 60/022,999 filed Aug. 2, 1996, now abandoned.

INTRODUCTION

The present invention relates generally to the field of genetic engineering and more particularly to genes for receptor tyrosine kinases and their cognate ligands, their insertion into recombinant DNA vectors, and the production of the encoded proteins in recipient strains of microorganisms and recipient eukaryotic cells. More specifically, the present invention is directed to a novel modified TIE-2 ligand that binds the TIE-2 receptor, as well as to methods of making and using the modified ligand. The invention further provides a nucleic acid sequence encoding the modified ligand, and methods for the generation of nucleic acid encoding the modified ligand and the gene product. The modified TIE-2 ligand, as well as nucleic acid encoding it, may be useful in the diagnosis and treatment of certain diseases involving endothelial cells and associated TIE receptors, such as neoplastic diseases involving tumor angiogenesis, wound healing, thromboembolic diseases, atherosclerosis and inflammatory diseases. In addition, the modified ligand may be used to promote the proliferation and/or differentiation of hematopoietic stem cells.

More generally, the receptor activating modified TIE-2 ligands described herein may be used to promote the growth, survival, migration, and/or differentiation and/or stabilization or destabilization of cells expressing TIE receptor. Biologically active modified TIE-2 ligand may be used for the in vitro maintenance of TIE receptor expressing cells in culture. Cells and tissues expressing TIE receptor include, for example, cardiac and vascular endothelial cells, lens epithelium and heart epicardium and early hematopoietic cells. Alternatively, such human ligand may be used to support cells which are engineered to express TIE receptor. Further, modified TIE-2 ligand and its cognate receptor may be used in assay systems to identify further agonists or antagonists of the receptor.

BACKGROUND OF THE INVENTION

The cellular behavior responsible for the development, maintenance, and repair of differentiated cells and tissues is regulated, in large part, by intercellular signals conveyed via growth factors and similar ligands and their receptors. The receptors are located on the cell surface of responding cells and they bind peptides or polypeptides known as growth factors as well as other hormone-like ligands. The results of this interaction are rapid biochemical changes in the responding cells, as well as a rapid and a long-term readjustment of cellular gene expression. Several receptors associated with various cell surfaces may bind specific growth factors.

The phosphorylation of tyrosine residues in proteins by tyrosine kinases is one of the key modes by which signals are transduced across the plasma membrane. Several currently known protein tyrosine kinase genes encode transmembrane receptors for polypeptide growth factors and

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hormones such as epidermal growth factor (EGF), insulin, insulin-like growth factor-I (IGF-I), platelet derived growth factors (PDGF-A and -B), and fibroblast growth factors (FGFs). (Heldin et al., Cell Regulation, 1: 555-566 (1990); Ullrich, et al., Cell, 61: 243-54 (1990)). In each instance, these growth factors exert their action by binding to the extracellular portion of their cognate receptors, which leads to activation of the intrinsic tyrosine kinase present on the cytoplasmic portion of the receptor. Growth factor receptors of endothelial cells are of particular interest due to the possible involvement of growth factors in several important physiological and pathological processes, such as vasculogenesis, angiogenesis, atherosclerosis, and inflammatory diseases. (Folkman, et al. Science, 235: 442-447 (1987)). Also, the receptors of several hematopoietic growth factors are tyrosine kinases; these include c-fms, which is the colony stimulating factor 1 receptor, Sherr, et al., Cell, 41: 665-676 (1985), and c-kit, a primitive hematopoietic growth factor receptor reported in Huang, et al., Cell, 63: 225-33 (1990).

The receptor tyrosine kinases have been divided into evolutionary subfamilies based on the characteristic structure of their ectodomains. (Ullrich, et al. Cell, 61: 243-54 (1990)). Such subfamilies include, EGF receptor-like kinase (subclass I) and insulin receptor-like kinase (subclass II), each of which contains repeated homologous cysteine-rich sequences in their extracellular domains. A single cysteinerich region is also found in the extracellular domains of the eph-like kinases. Hirai, et al., Science, 238: 1717-1720 (1987); Lindberg, et al. Mol. Cell. Biol., 10: 6316-24 (1990); Lhotak, et al., Mol. Cell. Biol. 11: 2496-2502 (1991). PDGF receptors as well as c-fms and c-kit receptor tyrosine kinases may be grouped into subclass III; while the FGF receptors form subclass IV. Typical for the members of both of these subclasses are extracellular folding units stabilized by intrachain disulfide bonds. These so-called immunoglobulin (Ig)-like folds are found in the proteins of the immunoglobulin superfamily which contains a wide variety of other cell surface receptors having either cellbound or soluble ligands. Williams, et al., Ann. Rev. Immunol., 6: 381-405 (1988).

Receptor tyrosine kinases differ in their specificity and affinity. In general, receptor tyrosine kinases are glycoproteins which consist of (1) an extracellular domain capable of binding the specific growth factor(s); (2) a transmembrane domain which usually is an alpha-helical portion of the protein; (3) a juxtamembrane domain where the receptor may be regulated by, e.g., protein phosphorylation; (4) a tyrosine kinase domain which is the enzymatic component of the receptor; and (5) a carboxyterminal tail which in many receptors is involved in recognition and binding of the substrates for the tyrosine kinase.

Processes such as alternative exon splicing and alternative choice of gene promoter or polyadenylation sites have been reported to be capable of producing several distinct polypeptides from the same gene. These polypeptides may or may not contain the various domains listed above. As a consequence, some extracellular domains may be expressed as separate, secreted proteins and some forms of the receptors may lack the tyrosine kinase domain and contain only the extracellular domain inserted in the plasma membrane via the transmembrane domain plus a short carboxyl terminal tail.

A gene encoding an endothelial cell transmembrane tyrosine kinase, originally identified by RT-PCR as an unknown tyrosine kinase-homologous cDNA fragment from human leukemia cells, was described by Partanen, et al.,

Proc. Natl. Acad. Sci. USA, 87: 8913–8917 (1990). This gene and its encoded protein are called "TIE" which is an abbreviation for "tyrosine kinase with Ig and EGF homology domains." Partanen, et al. Mol. Cell. Biol. 12: 1698–1707 (1992).

It has been reported that tie mRNA is present in all human fetal and mouse embryonic tissues. Upon inspection, tie message has been localized to the cardiac and vascular endothelial cells. Specifically, tie mRNA has been localized to the endothelia of blood vessels and endocardium of 9.5 to 18.5 day old mouse embryos. Enhanced tie expression was shown during neovascularization associated with developing ovarian follicles and granulation tissue in skin wounds. Korhonen, et al. Blood 80: 2548–2555 (1992). Thus the TIEs have been suggested to play a role in angiogenesis, which is important for developing treatments for solid tumors and several other angiogenesis-dependent diseases such as diabetic retinopathy, psoriasis, atherosclerosis and arthritis

Two structurally related rat TIE receptor proteins have been reported to be encoded by distinct genes with related profiles of expression. One gene, termed tie-1, is the rat homolog of human tie. Maisonpierre, et al., Oncogene 8: 1631–1637 (1993). The other gene, tie-2, may be the rat homolog of the murine tek gene, which, like tie, has been reported to be expressed in the mouse exclusively in endothelial cells and their presumptive progenitors. Dumont, et al. Oncogene 8: 1293–1301 (1993). The human homolog of tie-2 is described in Ziegler, U.S. Pat. No. 5,447,860 which issued on Sep. 5, 1995 (wherein it is referred to as "ork"), which is incorporated in its entirety herein.

Both genes were found to be widely expressed in endothelial cells of embryonic and postnatal tissues. Significant levels of tie-2 transcripts were also present in other embryonic cell populations, including lens epithelium, heart epicardium and regions of mesenchyme. Maisonpierre, et al., Oncogene 8: 1631–1637 (1993).

The predominant expression of the TIE receptor in vascular endothelia suggests that TIE plays a role in the development and maintenance of the vascular system. This could include roles in endothelial cell determination, proliferation, differentiation and cell migration and patterning into vascular elements. Analyses of mouse embryos deficient in TIE-2 illustrate its importance in angiogenesis, particularly for vascular network formation in endothelial cells. Sato, T. N., et al., Nature 376:70–74 (1995). In the mature vascular system, the TIEs could function in endothelial cell survival, maintenance and response to pathogenic influences.

The TIE receptors are also expressed in primitive hematopoietic stem cells, B cells and a subset of megakaryocytic cells, thus suggesting the role of ligands which bind these receptors in early hematopoiesis, in the differentiation and/or proliferation of B cells, and in the megakaryocytic 55 differentiation pathway. Iwama, et al. Biochem. Biophys. Research Communications 195:301–309 (1993); Hashiyama, et al. Blood 87:93–101 (1996), Batard, et al. Blood 87:2212–2220 (1996).

SUMMARY OF THE INVENTION

The present invention provides for a composition comprising a modified TIE-2 ligand substantially free of other proteins. As used herein, modified TIE-2 ligand refers to a ligand of the TIE family of ligands, whose representatives 65 comprise ligands TL1, TL2, TL3 and TL4 as described herein, which has been altered by addition, deletion or

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substitution of one or more amino acids, or by way of tagging, with for example, the Fc portion of human IgG-1, but which retains its ability to bind the TIE-2 receptor. Modified TIE-2 ligand also includes a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a portion of a second TIE-2 ligand which is different from the first. By way of non-limiting example, the first TIE-2 ligand is TL1 and the second TIE-2 ligand is TL2. The invention envisions other combinations using additional TIE-2 ligand family members. For example, other combinations for creating a chimeric TIE-2 ligand are possible, including but not limited to those combinations wherein the first ligand is selected from the group consisting of TL1, TL2, TL3 and TL4, and the second ligand, different from the first ligand, is selected from the group consisting of TL1, TL2, TL3 and TL4.

The invention also provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand. In one embodiment, the isolated nucleic acid molecule encodes a TIE-2 ligand of the TIE family of ligands, whose representatives comprise ligands TL1, TL2, TL3 and TL4 as described herein, which has been altered by addition, deletion or substitution of one or more amino acids, or by way of tagging, with for example, the Fc portion of human IgG-1, but which retains its ability to bind the TIE-2 receptor. In another embodiment, the isolated nucleic acid molecule encodes a modified TIE-2 ligand which is a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a portion of a second TIE-2 ligand which is different from the first. By way of non-limiting example, the first TIE-2 ligand is TL1 and the second TIE-2 ligand is TL2. The invention envisions other combinations using additional TIE-2 ligand family members. For example, other combinations are possible, including but not limited to those combinations wherein the isolated nucleic acid molecule encodes a modified TIE-2 ligand which is a chimeric TIE-2 ligand comprising a portion of a first ligand selected from the group consisting of TL1, TL2, TL3 and TL4, and a portion of a second ligand, different from the first ligand, selected from the group consisting of TL1, TL2, TL3 and TL4.

The isolated nucleic acid may be DNA, cDNA or RNA. The invention also provides for a vector comprising an isolated nucleic acid molecule encoding a modified TIE-2 ligand. The invention further provides for a host-vector system for the production in a suitable host cell of a polypeptide having the biological activity of a modified TIE-2 ligand. The suitable host cell may be bacterial, yeast, insect or mammalian. The invention also provides for a method of producing a polypeptide having the biological activity of a modified TIE-2 ligand which comprises growing cells of the host-vector system under conditions permitting production of the polypeptide and recovering the polypeptide so produced.

The invention herein described of an isolated nucleic acid molecule encoding a modified TIE-2 ligand further provides for the development of the ligand as a therapeutic for the treatment of patients suffering from disorders involving cells, tissues or organs which express the TIE-2 receptor. The present invention also provides for an antibody which specifically binds such a therapeutic molecule. The antibody may be monoclonal or polyclonal. The invention also provides for a method of using such a monoclonal or polyclonal antibody to measure the amount of the therapeutic molecule in a sample taken from a patient for purposes of monitoring the course of therapy.

The present invention also provides for an antibody which specifically binds a modified TIE-2 ligand as described

herein. The antibody may be monoclonal or polyclonal. Thus the invention further provides for therapeutic compositions comprising an antibody which specifically binds a modified TIE-2 ligand, in a pharmaceutically acceptable vehicle. The invention also provides for a method of blocking blood vessel growth in a mammal by administering an effective amount of a therapeutic composition comprising an antibody which specifically binds a receptor activating modified TIE-2 ligand as described herein, in a pharmaceutically acceptable vehicle.

The invention further provides for therapeutic compositions comprising a modified TIE-2 ligand as described herein, in a pharmaceutically acceptable vehicle. The invention also provides for a method of promoting neovascularization in a patient by administering an effective amount of a therapeutic composition comprising a receptor activating modified TIE-2 ligand as described herein, in a pharmaceutically acceptable vehicle. In one embodiment, the method may be used to promote wound healing. In another embodiment, the method may be used to treat ischemia. In yet another embodiment, a receptor activating modified TIE-2 ligand as described herein is used, alone or in combination with other hematopoietic factors, to promote the proliferation or differentiation of hematopoietic stem cells, B cells or megakaryocytic cells.

Alternatively, the invention provides that a modified TIE-2 ligand may be conjugated to a cytotoxic agent and a therapeutic composition prepared therefrom. The invention further provides for a receptorbody which specifically binds a modified TIE-2 ligand. The invention further provides for therapeutic compositions comprising a receptorbody which specifically binds a modified TIE-2 ligand in a pharmaceutically acceptable vehicle. The invention also provides for a method of blocking blood vessel growth in a mammal by administering an effective amount of a therapeutic composition comprising a receptorbody which specifically binds a modified TIE-2 ligand in a pharmaceutically acceptable vehicle.

The invention also provides for a TIE-2 receptor antagonist as well as a method of inhibiting TIE-2 biological activity in a mammal comprising administering to the mammal an effective amount of a TIE-2 antagonist. According to the invention, the antagonist may be a modified TIE-2 ligand as described herein which binds to, but does not activate, the TIE-2 receptor.

BRIEF DESCRIPTION OF THE FIGURES

FIGS. 1A and 1B—TIE-2 receptorbody (TIE-2 RB) inhibits the development of blood vessels in the embryonic chicken chorioallantoic membrane (CAM). A single piece of resorbable gelatin foam (Gelfoam) soaked with 6 µg of RB was inserted immediately under the CAM of 1-day chick embryos. After 3 further days of incubation, 4 day old embryos and surrounding CAM were removed and examined. FIG. 1A: embryos treated with EHK-1 RB (rEHK-1 ecto/hlgG1 Fc) were viable and possessed normally developed blood vessels in their surrounding CAM. FIG. 1B: all embryos treated with TIE-2 RB (r TIE-2 ecto/hlgG1 Fc) were dead, diminished in size and were almost completely devoid of surrounding blood vessels.

FIG. 2-Vector pJFE14.

FIG. 3—Restriction map of λgt10.

FIGS. 4A–4D—Nucleic acid (SEQ ID NO: 1) and deduced amino acid (SEQ ID NO: 2) (single letter code) 65 sequences of human TIE-2 ligand 1 from clone $\lambda gt10$ encoding htie-2 ligand 1.

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FIGS. **5A–5D**—Nucleic acid (SEQ ID NO: 3) and deduced amino acid (SEQ ID NO: 4) (single letter code) sequences of human TIE-2 ligand 1 from T98G clone.

FIGS. 6A–6D—Nucleic acid and deduced amino acid (single letter code) sequences of human TIE-2 ligand 2 from clone pBluescript KS encoding human TIE 2 ligand 2.

FIG. 7—Western blot showing activation of TIE-2 receptor by TIE-2 ligand 1 (Lane L1) but not by TIE-2 ligand 2 (Lane L2) or control (Mock).

FIG. 8—Western blot showing that prior treatment of HAEC cells with excess TIE-2 ligand 2 (Lane 2) antagonizes the subsequent ability of dilute TIE-2 ligand 1 to activate the TIE-2 receptor (TIE2-R) as compared with prior treatment of HAEC cells with MOCK medium (Lane 1).

FIG. 9—Western blot demonstrating the ability of TL2 to competitively inhibit TL1 activation of the TIE-2 receptor using the human cell hybrid line, EA.hy926.

FIGS. 10A–10D—Histogram representation of binding to rat TIE-2 IgG immobilized surface by TIE-2 ligand in C2C12 ras (FIG. 10A), Rat2 ras (FIG. 10B), SHEP (FIG. 10C), and T98G (FIG. 10D) concentrated (10x) conditioned medium. Rat TIE-2 (rTIE2) specific binding is demonstrated by the significant reduction in the binding activity in the presence of 25 µg/ml soluble rat TIE-2 RB as compared to a minor reduction in the presence of soluble trkB RB.

FIGS. 11A–11B—Binding of recombinant human TIE-2 ligand 1 (hTL1) (FIG. 11A) and human TIE-2 ligand 2 (hTL2) (FIG. 11B), in COS cell supernatants, to a human TIE-2 receptorbody (RB) immobilized surface. Human TIE-2-specific binding was determined by incubating the samples with 25 μ g/ml of either soluble human TIE-2 RB or trkB RB; significant reduction in the binding activity is observed only for the samples incubated with human TIE-2 RB

FIG. 12—Western blot showing that TIE-2 receptorbody (denoted TIE-2 RB or, as here, TIE2-Fc) blocks the activation of TIE-2 receptors by TIE-2 ligand 1 (TL1) in HUVEC cells, whereas an unrelated receptorbody (TRKB-Fc) does not block this activation.

FIG. 13—Agarose gels showing serial dilutions [undiluted (1) to 10^{-4}] of the TL1 and TL2 RT-PCR products obtained from E14.5 mouse fetal liver (Lanes 1-total, Lanes 3-stromal enriched, and Lanes 4—c-kit+TER119 hematopoietic precursor cells) and E14.5 mouse fetal thymus (Lanes 2-total).

FIG. 14—Agarose gels showing serial dilutions [undiluted (1) to 10^{-3}] of the TL1 and TL2 RT-PCR products obtained from E17.5 mouse fetal thymus cortical stromal cells (Lanes 1-CDR1+/A2B5-) and medullary stromal cells (Lane CDR1-/A2B5+).

FIG. 15—A schematic representation of the hypothesized role of the TIE-2/TIE ligands in angiogenesis. TL1 is represented by (•), TL2 is represented by (*), TIE-2 is represented by (T), VEGF is represented by ([]), and flk-1 (a VEGF receptor) is represented by (Y).

FIG. 16—In situ hybridization slides showing the temporal expression pattern of TIE-2, TL1, TL2, and VEGF during angiogenesis associated with follicular development and corpus luteum formation in the ovary of a rat that was treated with pregnant mare serum. Column 1: Early preovulatory follicle; Column 2: pre-ovulatory follicle; Column 3: early corpus luteum; and Column 4: atretic follicle; Row A: bright field; Row B: VEGF; Row C: TL2; Row D: TL1 and Row E: TIE-2 receptor.

FIG. 17—Comparison of amino acid sequences of mature TL1 protein (SEQ ID NO: 7) and mature TL2 protein (SEQ

ID NO: 8). The TL1 sequence is the same as that set forth in FIGS. 4A–4D (SEQ ID NO: 1 AND SEQ ID NO: 2), except that the putative leader sequence has been removed. Similarly, the TL2 sequence is the same as that set forth in FIGS. 6A–6D (SEQ ID NO: 5 and SEQ ID NO: 6), except that the putative leader sequence has been removed. Arrows indicate residues Arg49, Cys245 and Arg264 of TL1, which correspond to the residues at amino acid positions 69, 265 and 284, respectively, of TL1 as set forth in FIGS. 4A–4D (SEQ ID NO: 1 and SEQ ID NO: 2).

FIG. 18—Western blot of the covalent multimeric structure of TL1 and TL2 (Panel A) and the interconversion of TL1 and TL2 by the mutation of one cysteine (Panel B).

FIG. 19—A typical curve of TIE-2-IgG binding to immobilized TL1 in a quantitative cell-free binding assay.

FIG. **20**—A typical curve showing TIE-2 ligand 1 ligandbody comprising the fibrinogen-like domain of the ligandbound to the Fc domain of IgG (TL1-fFc) binding to immobilized TIE-2 ectodomain in a quantitative cell-free binding assay.

FIGS. 21A–21C—Nucleotide (SEQ ID NO: 9) and deduced amino acid (SEQ ID NO: 10) (single letter code) sequences of TIE ligand-3. The coding sequence starts at position 47. The fibrinogen-like domain starts at position 25 929.

FIGS. 22A–22B—Comparison of Amino Acid Sequences of TIE Ligand Family Members. mTL3=mouse TIE ligand-3 (SEQ ID NO: 11); hTL1=human TIE-2 ligand1 (SEQ ID NO: 12); chTL1=chicken TIE-2 ligand1 (SEQ ID NO: 13); 30 mTL1=mouse TIE-2 ligand 1 (SEQ ID NO: 14); mTL2=mouse TIE-2 ligand 2 (SEQ ID NO: 15); hTL2=human TIE-2 ligand 2 (SEQ ID NO: 16). The boxed regions indicate conserved regions of homology among the family members.

FIGS. 23A–23C—Nucleotide (SEQ ID NO: 17) and deduced amino acid (SEQ ID NO: 18) (single letter code) sequences of TIE ligand-4. Arrow indicates nucleotide position 569.

FIGS. 24A–24C—Nucleotide (SEQ ID NO: 19) and ⁴⁰ deduced amino acid (SEQ ID NO: 20) (single letter code) sequences of chimeric TIE ligand designated 1N1C2F (chimera 1). The putative leader sequence is encoded by nucleotides 1–60.

FIGS. 25A-25C—Nucleotide (SEQ ID NO: 21) and deduced amino acid (SEQ ID NO: 22) (single letter code) sequences of chimeric TIE ligand designated 2N2C1F (chimera 2). The putative leader sequence is encoded by nucleotides 1-48.

FIGS. 26A–26C—Nucleotide (SEQ ID NO: 23) and deduced amino acid (SEQ ID NO: 24) (single letter code) sequences of chimeric TIE ligand designated 1N2C2F (chimera 3). The putative leader sequence is encoded by nucleotides 1–60.

FIGS. 27A–27C—Nucleotide (SEQ ID NO: 25) and deduced amino acid (SEQ ID NO: 26) (single letter code) sequences of chimeric TIE ligand designated 2N1C1F (chimera 4). The putative leader sequence is encoded by nucleotides 1–48.

DETAILED DESCRIPTION OF THE INVENTION

As described in greater detail below, applicants have created novel modified TIE-2 ligands that bind the TIE-2 65 receptor. The present invention provides for a composition comprising a modified TIE-2 ligand substantially free of

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other proteins. As used herein, modified TIE-2 ligand refers to a ligand of the TIE family of ligands, whose representatives comprise ligands TL1, TL2, TL3 and TL4 as described herein, which has been altered by addition, deletion or substitution of one or more amino acids, or by way of tagging, with for example, the Fc portion of human IgG-1, but which retains its ability to bind the TIE-2 receptor. Modified TIE-2 ligand also includes a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a 10 portion of a second TIE-2 ligand which is different from the first. By way of non-limiting example, the first TIE-2 ligand is TL1 and the second TIE-2 ligand is TL2. The invention envisions other combinations using additional TIE-2 ligand family members. For example, other combinations for creating a chimeric TIE-2 ligand are possible,)including but not limited to those combinations wherein the first ligand is selected from the group consisting of TL1, TL2, TL3 and TL4, and the second ligand, different from the first ligand, is selected from the group consisting of TL1, TL2, TL3 and TL4.

The invention also provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand. In one embodiment, the isolated nucleic acid molecule encodes a TIE-2 ligand of the TIE family of ligands, whose representatives comprise ligands TL1, TL2, TL3 and TL4 as described herein, which has been altered by addition, deletion or substitution of one or more amino acids, or by way of tagging, with for example, the Fc portion of human IgG-1, but which retains its ability to bind the TIE-2 receptor. In another embodiment, the isolated nucleic acid molecule encodes a modified TIE-2 ligand which is a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a portion of a second TIE-2 ligand which is different from the first. By way of non-limiting example, the first TIE-2 ligand is TL1 and the second TIE-2 ligand is TL2. The invention envisions other combinations using additional TIE-2 ligand family members. For example, other combinations are possible, including but not limited to those combinations wherein the isolated nucleic acid molecule encodes a modified TIE-2 ligand which is a chimeric TIE-2 ligand comprising a portion of a first ligand selected from the group consisting of TL1, TL2, TL3 and TL4, and a portion of a second ligand, different from the first ligand, selected from the group consisting of TL1, TL2, TL3 and

The present invention comprises the modified TIE-2 ligands and their amino acid sequences, as well as functionally equivalent variants thereof, as well as proteins or peptides comprising substitutions, deletions or insertional mutants of the described sequences, which bind TIE-2 receptor and act as agonists or antagonists thereof. Such variants include those in which amino acid residues are substituted for residues within the sequence resulting in a silent change. For example, one or more amino acid residues within the sequence can be substituted by another amino acid(s) of a similar polarity which acts as a functional equivalent, resulting in a silent alteration. Substitutes for an amino acid within the sequence may be selected from other members of the class to which the amino acid belongs. For 60 example, the class of nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid.

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Also included within the scope of the invention are proteins or fragments or derivatives thereof which exhibit the same or similar biological activity as the modified TIE-2 ligands described herein, and derivatives which are differentially modified during or after translation, e.g., by glycosylation, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand, etc. Functionally equivalent molecules also include molecules that contain modifications, including N-terminal modifications, which result from expression in a particular recombinant host, such as, for example, N-terminal methylation which occurs in certain bacterial (e.g. *E. coli*) expression systems.

The present invention also encompasses the nucleotide sequences that encode the proteins described herein as modified TIE-2 ligands, as well as host cells, including yeast, bacteria, viruses, and mammalian cells, which are genetically engineered to produce the proteins, by e.g. transfection, transduction, infection, electroporation, or microinjection of nucleic acid encoding the modified TIE-2 ligands described herein in a suitable expression vector. The present invention also encompasses introduction of the nucleic acid encoding modified TIE-2 ligands through gene therapy techniques such as is described, for example, in Finkel and Epstein FASEB J. 9:843–851 (1995); Guzman, et al. PNAS (USA) 91:10732–10736 (1994).

One skilled in the art will also recognize that the present invention encompasses DNA and RNA sequences that hybridize to a modified TIE-2 ligand encoding nucleotide sequence, under conditions of moderate stringency, as defined in, for example, Sambrook, et al. Molecular Clon- 30 ing: A Laboratory Manual, 2 ed. Vol. 1, pp. 101-104, Cold Spring Harbor Laboratory Press (1989). Thus, a nucleic acid molecule contemplated by the invention includes one having a nucleotide sequence deduced from an amino acid sequence of a modified TIE-2 ligand prepared as described herein, as 35 well as a molecule having a sequence of nucleotides that hybridizes to such a nucleotide sequence, and also a nucleotide sequence which is degenerate of the above sequences as a result of the genetic code, but which encodes a ligand that binds TIE-2 receptor and which has an amino acid 40 sequence and other primary, secondary and tertiary characteristics that are sufficiently duplicative of a modified TIE-2 ligand described herein so as to confer on the molecule the same biological activity as the modified TIE-2 ligand described herein.

The present invention provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds and activates TIE-2 receptor comprising a nucleotide sequence encoding TIE-2 ligand 1 wherein the portion of the nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 1 is replaced by a nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 2. The invention also provides for such a nucleic acid molecule, with a further modification such that the portion of the nucleotide sequence that encodes the coiled-coil domain of 55 TIE-2 ligand 1 is replaced by a nucleotide sequence that encodes the coiled-coil domain of TIE-2 ligand 2.

The present invention also provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds and activates TIE-2 receptor comprising a nucleotide 60 sequence encoding TIE-2 ligand 1 wherein the portion of the nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 1 is replaced by a nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 2 and which is further modified to encode a different amino acid instead 65 of the cysteine residue encoded by nucleotides 784–786 as set forth in FIGS. 27A–27C (SEQ ID NO: 25 and SEQ ID

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NO: 26). A serine residue is preferably substituted for the cysteine residue. In another embodiment, the nucleic acid molecule is further modified to encode a different amino acid instead of the arginine residue encoded by nucleotides 199–201 as set forth in FIGS. 27A–27C (SEQ ID NO: 25 and SEQ ID NO: 26). A serine residue is preferably substituted for the arginine residue.

The present invention also provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds and activates TIE-2 receptor comprising a nucleotide sequence encoding TIE-2 ligand 1 which is modified to encode a different amino acid instead of the cysteine residue at amino acid position 245. A serine residue is preferably substituted for the cysteine residue.

The invention further provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds but does not activate TIE-2 receptor comprising a nucleotide sequence encoding TIE-2 ligand 1 wherein the portion of the nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 1 is deleted. The invention also provides for such a nucleic acid molecule further modified so that the portion of the nucleotide sequence that encodes the coiled-coil domain of TIE-2 ligand 1 is deleted and the portion encoding the fibrinogen-like domain is fused in-frame to a nucleotide sequence encoding a human immunoglobulin gamma-1 constant region (IgG1 Fc).

The invention further provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds but does not activate TIE-2 receptor comprising a nucleotide sequence encoding TIE-2 ligand 2 wherein the portion of the nucleotide sequence that encodes the N-terminal domain of TIE-2 ligand 2 is deleted. The invention also provides for such a nucleic acid molecule further modified so that the portion of the nucleotide sequence that encodes the coiled-coil domain of TIE-2 ligand 2 is deleted and the portion encoding the fibrinogen-like domain is fused in-frame to a nucleotide sequence encoding a human immunoglobulin gamma-1 constant region (IgG1 Fc).

The invention further provides for an isolated nucleic acid molecule encoding a modified TIE-2 ligand that binds but does not activate TIE-2 receptor comprising a nucleotide sequence encoding TIE-2 ligand 1 wherein the portion of the nucleotide sequence that encodes the fibrinogen-like domain of TIE-2 ligand 1 is replaced by a nucleotide sequence that encodes the fibrinogen-like domain of TIE-2 ligand 2. The invention also provides for such a nucleic acid molecule further modified so that the portion of the nucleotide sequence that encodes the coiled-coil domain of TIE-2 ligand 1 is replaced by a nucleotide sequence that encodes the coiled-coil domain of TIE-2 ligand 2.

The invention further provides for a modified TIE-2 ligand encoded by any of nucleic acid molecules of the invention

The present invention also provides for a chimeric TIE-2 ligand comprising at least a portion of a first TIE-2 ligand and a portion of a second TIE-2 ligand which is different from the first, wherein the first and second TIE-2 ligands are selected from the group consisting of TIE-2 Ligand-1, TIE-2 Ligand-2, TIE Ligand-3 and TIE Ligand-4. Preferably, the chimeric TIE ligand comprises at least a portion of TIE-2 Ligand-1 and a portion of TIE-2 Ligand-2.

The invention also provides a nucleic acid molecule that encodes a chimeric TIE ligand as set forth in FIGS. **24A–24**C (SEQ ID NO: 19 and SEQ ID NO: 20), **25A–25**C (SEQ ID NO: 21 and SEQ ID NO: 22), **26A–26**C (SEQ ID NO: 23 and SEQ ID NO: 24), or **27A–27**C (SEQ ID NO: 25

and (SEQ ID NO: 26). The invention also provides a chimeric TIE ligand as set forth in FIGS. **24**A–**24**C (SEQ ID NO: 19 and SEQ ID NO: 20), **25**A–**25**C (SEQ ID NO: 21 and SEQ ID NO: 22), **26**A–**26**C (SEQ ID NO: 23 and SEQ ID NO: 24), or **27**A–**27**C (SEQ ID NO: 25 and SEQ ID NO: 26). The invention further provides a chimeric TIE ligand as set forth in FIGS. **27**A–**27**C (SEQ ID NO: 25 and SEQ ID NO: 26), modified to have a different amino acid instead of the cysteine residue encoded by nucleotides 784–786.

Any of the methods known to one skilled in the art for the 10 insertion of DNA fragments into a vector may be used to construct expression vectors encoding a modified TIE-2 ligand using appropriate transcriptional/translational control signals and the protein coding sequences. These methods may include in vitro recombinant DNA and synthetic tech- 15 niques and in vivo recombinations (genetic recombination). Expression of a nucleic acid sequence encoding a modified TIE-2 ligand or peptide fragments thereof may be regulated by a second nucleic acid sequence which is operably linked to the a modified TIE-2 ligand encoding sequence such that 20 the modified TIE-2 ligand protein or peptide is expressed in a host transformed with the recombinant DNA molecule. For example, expression of a modified TIE-2 ligand described herein may be controlled by any promoter/enhancer element known in the art. Promoters which may be used to control 25 expression of the ligand include, but are not limited to the long terminal repeat as described in Squinto et al., (Cell 65:1-20 (1991)); the SV40 early promoter region (Bernoist and Chambon, Nature 290:304-310), the CMV promoter, the M-MuLV 5' terminal repeat, the promoter contained in 30 the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, et al., Cell 22:787-797 (1980)), the herpes thymidine kinase promoter (Wagner et al., Proc. Natl. Acad. Sci. U.S.A. 78:144–1445 (1981)), the adenovirus promoter, the regulatory sequences of the metallothionein gene 35 (Brinster et al., Nature 296:39-42 (1982)); prokaryotic expression vectors such as the β-lactamase promoter (Villa-Kamaroff, et al., Proc. Natl. Acad. Sci. U.S.A. 75:3727-3731 (1978)), or the tac promoter (DeBoer, et al., Proc. Natl. Acad. Sci. U.S.A. 80:21-25 (1983)), see also 40 "Useful proteins from recombinant bacteria" in Scientific American, 242:74–94 (1980); promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADH (alcohol dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following 45 animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals; elastase I gene control region which is active in pancreatic acinar cells (Swift et al., Cell 38:639-646 (1984); Ornitz et al., Cold Spring Harbor Symp. Quant. Biol. 50:399-409 50 (1986); MacDonald, Hepatology 7:425-515 (1987); insulin gene control region which is active in pancreatic beta cells [Hanahan, Nature 315:115-122 (1985)]; immunoglobulin gene control region which is active in lymphoid cells (Grosschedl et al., 1984, Cell 38:647-658; Adames et al., 55 1985, Nature 318:533-538; Alexander et al., 1987, Mol. Cell. Biol. 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et al., 1986, Cell 45:485-495), albumin gene control region which is active in liver (Pinkert et 60 al., 1987, Genes and Devel. 1:268-276), alpha-fetoprotein gene control region which is active in liver (Krumlauf et al., 1985, Mol. Cell. Biol. 5:1639-1648; Hammer et al., 1987, Science 235:53-58); alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al, 1987, Genes and 65 Devel. 1:161–171), beta-globin gene control region which is active in myeloid cells (Mogram et al., 1985, Nature

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315:338–340; Kollias et al., 1986, Cell 46:89–94); myelin basic protein gene control region which is active in oligodendrocytes in the brain (Readhead et al., 1987, Cell 48:703–712); myosin light chain-2 gene control region which is active in skeletal muscle (Shani, 1985, Nature 314:283–286), and gonadotropic releasing hormone gene control region which is active in the hypothalamus (Mason et al., 1986, Science 234:1372–1378). The invention further encompasses the production of antisense compounds which are capable of specifically hybridizing with a sequence of RNA encoding a modified TIE-2 ligand to modulate its expression. Ecker, U.S. Pat. No. 5,166,195, issued Nov. 24, 1992.

Thus, according to the invention, expression vectors capable of being replicated in a bacterial or eukaryotic host comprising a nucleic acid encoding a modified TIE-2 ligand as described herein, are used to transfect a host and thereby direct expression of such nucleic acid to produce a modified TIE-2 ligand, which may then be recovered in a biologically active form. As used herein, a biologically active form includes a form capable of binding to TIE receptor and causing a biological response such as a differentiated function or influencing the phenotype of the cell expressing the receptor. Such biologically active forms could, for example, induce phosphorylation of the tyrosine kinase domain of TIE receptor. Alternatively, the biological activity may be an effect as an antagonist to the TIE receptor. In alternative embodiments, the active form of a modified TIE-2 ligand is one that can recognize TIE receptor and thereby act as a targeting agent for the receptor for use in both diagnostics and therapeutics. In accordance with such embodiments, the active form need not confer upon any TIE expressing cell any change in phenotype.

Expression vectors containing the gene inserts can be identified by four general approaches: (a) DNA-DNA hybridization, (b) presence or absence of "marker" gene functions, (c) expression of inserted sequences and (d) PCR detection. In the first approach, the presence of a foreign gene inserted in an expression vector can be detected by DNA-DNA hybridization using probes comprising sequences that are homologous to an inserted modified TIE-2 ligand encoding gene. In the second approach, the recombinant vector/host system can be identified and selected based upon the presence or absence of certain "marker" gene functions (e.g., thymidine kinase activity, resistance to antibiotics, transformation phenotype, occlusion body formation in baculovirus, etc.) caused by the insertion of foreign genes in the vector. For example, if a nucleic acid encoding a modified TIE-2 ligand is inserted within the marker gene sequence of the vector, recombinants containing the insert can be identified by the absence of the marker gene function. In the third approach, recombinant expression vectors can be identified by assaying the foreign gene product expressed by the recombinant. Such assays can be based, for example, on the physical or functional properties of a modified TIE-2 ligand gene product, for example, by binding of the ligand to TIE receptor or a portion thereof which may be tagged with, for example, a detectable antibody or portion thereof or by binding to antibodies produced against the modified TIE-2 ligand protein or a portion thereof. Cells of the present invention may transiently or, preferably, constitutively and permanently express a modified TIE-2 ligand as described herein. In the fourth approach, DNA nucleotide primers can be prepared corresponding to a tie specific DNA sequence. These primers could then be used to PCR a tie gene fragment. (PCR Protocols: A Guide To Methods and Applications, Edited by Michael A. Innis et al., Academic Press (1990)).

The recombinant ligand may be purified by any technique which allows for the subsequent formation of a stable, biologically active protein. Preferably, the ligand is secreted into the culture medium from which it is recovered. Alternatively, the ligand may be recovered from cells either as soluble proteins or as inclusion bodies, from which it may be extracted quantitatively by 8M guanidinium hydrochloride and dialysis in accordance with well known methodology. In order to further purify the ligand, affinity chromatography, conventional ion exchange chromatography, hydrophobic interaction chromatography, reverse phase chromatography or gel filtration may be used.

In additional embodiments of the invention, as described in greater detail in the Examples, a modified TIE-2 ligand encoding gene may be used to inactivate or "knock out" an endogenous gene by homologous recombination, and thereby create a TIE ligand deficient cell, tissue, or animal. For example, and not by way of limitation, the recombinant TIE ligand-4 encoding gene may be engineered to contain an insertional mutation, for example the neo gene, which would inactivate the native TIE ligand-4 encoding gene. Such a construct, under the control of a suitable promoter, may be introduced into a cell, such as an embryonic stem cell, by a technique such as transfection, transduction, or injection. Cells containing the construct may then be selected by G418 resistance. Cells which lack an intact TIE ligand-4 encoding gene may then be identified, e.g. by Southern blotting, PCR detection, Northern blotting or assay of expression. Cells lacking an intact TIE ligand-4 encoding gene may then be fused to early embryo cells to generate transgenic animals 30 deficient in such ligand. Such an animal may be used to define specific in vivo processes, normally dependent upon the ligand.

The present invention also provides for antibodies to a modified TIE-2 ligand described herein which are useful for 35 detection of the ligand in, for example, diagnostic applications. For preparation of monoclonal antibodies directed toward a modified TIE-2 ligand, any technique which provides for the production of antibody molecules by continuous cell lines in culture may be used. For example, the 40 hybridoma technique originally developed by Kohler and Milstein (1975, Nature 256:495–497), as well as the trioma technique, the human B-cell hybridoma technique (Kozbor et al., 1983, Immunology Today 4:72), and the EBVhybridoma technique to produce human monoclonal anti- 45 bodies (Cole et al., 1985, in "Monoclonal Antibodies and Cancer Therapy," Alan R. Liss, Inc. pp. 77-96) and the like are within the scope of the present invention.

The monoclonal antibodies may be human monoclonal antibodies or chimeric human-mouse (or other species) 50 a modified TIE-2 ligand which activates the TIE-2 receptor monoclonal antibodies. Human monoclonal antibodies may be made by any of numerous techniques known in the art (e.g., Teng et al., 1983, Proc. Natl. Acad. Sci. U.S.A. 80:7308-7312; Kozbor et al., 1983, Immunology Today 4:72-79; Olsson et al., 1982, Meth. Enzymol. 92:3-16). 55 Chimeric antibody molecules may be prepared containing a mouse antigen-binding domain with human constant regions (Morrison et al., 1984, Proc. Natl. Acad. Sci. U.S.A. 81:6851, Takeda et al., 1985, Nature 314:452).

Various procedures known in the art may be used for the 60 production of polyclonal antibodies to epitopes of a modified TIE-2 ligand described herein. For the production of antibody, various host animals, including but not limited to rabbits, mice and rats can be immunized by injection with a modified TIE-2 ligand, or a fragment or derivative thereof. 65 Various adjuvants may be used to increase the immunological response, depending on the host species, and including

but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (Bacille Calmette-Guerin) and Corynebacterium parvum.

A molecular clone of an antibody to a selected a modified TIE-2 ligand epitope can be prepared by known techniques. Recombinant DNA methodology (see e.g., Maniatis et al., 1982, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.) may be used to construct nucleic acid sequences which encode a monoclonal antibody molecule, or antigen binding region

The present invention provides for antibody molecules as well as fragments of such antibody molecules. Antibody fragments which contain the idiotype of the molecule can be generated by known techniques. For example, such fragments include but are not limited to: the F(ab'), fragment which can be produced by pepsin digestion of the antibody molecule; the Fab' fragments which can be generated by reducing the disulfide bridges of the F(ab')2 fragment, and the Fab fragments which can be generated by treating the antibody molecule with papain and a reducing agent. Antibody molecules may be purified by known techniques, es, immunoabsorption or immunoaffinity chromatography, chromatographic methods such as HPLC (high performance liquid chromatography), or a combination thereof.

The present invention further encompasses an immunoassay for measuring the amount of a modified TIE-2 ligand in a biological sample by

- a) contacting the biological sample with at least one antibody which specifically binds a modified TIE-2 ligand so that the antibody forms a complex with any modified TIE-2 ligand present in the sample; and
- b) measuring the amount of the complex and thereby measuring the amount of the modified TIE-2 ligand in the biological sample.

The invention further encompasses an assay for measuring the amount of TIE receptor in a biological sample by

- a) contacting the biological sample with at least one ligand of the invention so that the ligand forms a complex with the TIE receptor; and
- b) measuring the amount of the complex and thereby measuring the amount of the TIE receptor in the biological sample.

The present invention also provides for the utilization of as described herein, to support the survival and/or growth and/or migration and/or differentiation of TIE-2 receptor expressing cells. Thus, the ligand may be used as a supplement to support, for example, endothelial cells in culture.

Further, the creation by applicants of a modified TIE-2 ligand for the TIE-2 receptor enables the utilization of assay systems useful for the identification of agonists or antagonists of the TIE-2 receptor. Such assay systems would be useful in identifying molecules capable of promoting or inhibiting angiogenesis. For example, in one embodiment, antagonists of the TIE-2 receptor may be identified as test molecules that are capable of interfering with the interaction of the TIE-2 receptor with a modified TIE-2 ligand that binds the TIE-2 receptor. Such antagonists are identified by their ability to 1) block the binding of a biologically active modified TIE-2 ligand to the receptor as measured, for example, using BIAcore biosensor technology (BIAcore; =

Pharmacia Biosensor, Piscataway, N.J.); or 2) block the ability of a biologically active modified TIE-2 ligand to cause a biological response. Such biological responses include, but are not limited to, phosphorylation of the TIE receptor or downstream components of the TIE signal transduction pathway, or survival, growth or differentiation of TIE receptor bearing cells.

In one embodiment, cells engineered to express the TIE receptor may be dependent for growth on the addition of a modified TIE-2 ligand. Such cells provide useful assay systems for identifying additional agonists of the TIE receptor, or antagonists capable of interfering with the activity of the modified TIE-2 ligand on such cells. Alternatively, autocrine cells, engineered to be capable of co-expressing both a modified TIE-2 ligand and receptor, may provide useful systems for assaying potential agonists or antagonists.

Therefore, the present invention provides for introduction of a TIE-2 receptor into cells that do not normally express this receptor, thus allowing these cells to exhibit profound and easily distinguishable responses to a ligand which binds 20 this receptor. The type of response elicited depends on the cell utilized, and not the specific receptor introduced into the cell. Appropriate cell lines can be chosen to yield a response of the greatest utility for assaying, as well as discovering, molecules that can act on tyrosine kinase receptors. The 25 molecules may be any type of molecule, including but not limited to peptide and non-peptide molecules, that will act in systems to be described in a receptor specific manner.

One of the more useful systems to be exploited involves the introduction of a TIE receptor (or a chimeric receptor 30 comprising the extracellular domain of another receptor tyrosine kinase such as, for example, trkC and the intracellular domain of a TIE receptor) into a fibroblast cell line (e.g., NIH3T3 cells) thus such a receptor which does not normally mediate proliferative or other responses can, fol- 35 lowing introduction into fibroblasts, nonetheless be assayed by a variety of well established methods to quantitate effects of fibroblast growth factors (e.g. thymidine incorporation or other types of proliferation assays; see van Zoelen, 1990, "The Use of Biological Assays For Detection Of Polypep- 40 tide Growth Factors" in Progress Factor Research, Vol. 2, pp. 131-152; Zhan and M. Goldfarb, 1986, Mol. Cell. Biol.. Vol. 6, pp. 3541-3544). These assays have the added advantage that any preparation can be assayed both on the cell line having the introduced receptor as well as the parental cell 45 line lacking the receptor; only specific effects on the cell line with the receptor would be judged as being mediated through the introduced receptor. Such cells may be further engineered to express a modified TIE-2 ligand, thus creating an autocrine system useful for assaying for molecules that 50 act as antagonists/agonists of this interaction. Thus, the present invention provides for host cells comprising nucleic acid encoding a modified TIE-2 ligand and nucleic acid encoding TIE receptor.

The TIE receptor/modified TIE-2 ligand interaction also 55 provides a useful system for identifying small molecule agonists or antagonists of the TIE receptor. For example, fragments, mutants or derivatives of a modified TIE-2 ligand may be identified that bind TIE receptor but do not induce any other biological activity. Alternatively, the characterization of a modified TIE-2 ligand enables the further characterization of active portions of the molecule. Further, the identification of a ligand enables the determination of the X-ray crystal structure of the receptor/ligand complex, thus enabling identification of the binding site on the receptor. 65 Knowledge of the binding site will provide useful insight into the rational design of novel agonists and antagonists.

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The specific binding of a test molecule to TIE receptor may be measured in a number of ways. For example, the actual binding of test molecule to cells expressing TIE may be detected or measured, by detecting or measuring (i) test molecule bound to the surface of intact cells; (ii) test molecule cross-linked to TIE protein in cell lysates; or (iii) test molecule bound to TIE in vitro. The specific interaction between test molecule and TIE may be evaluated by using reagents that demonstrate the unique properties of that interaction.

As a specific, nonlimiting example, the methods of the invention may be used as follows. Consider a case in which a modified TIE-2 ligand in a sample is to be measured. Varying dilutions of the sample (the test molecule), in parallel with a negative control (NC) containing no modified TIE-2 ligand activity, and a positive control (PC) containing a known amount of a modified TIE-2 ligand, may be exposed to cells that express TIE in the presence of a detectably labeled modified TIE-2 ligand (in this example, radioiodinated ligand). The amount of modified TIE-2 ligand in the test sample may be evaluated by determining the amount of ¹²⁵I-labeled modified TIE-2 ligand that binds to the controls and in each of the dilutions, and then comparing the sample values to a standard curve. The more modified TIE-2 ligand in the sample, the less ¹²⁵I-ligand that will bind to TIE.

The amount of ¹²⁵I-ligand bound may be determined by measuring the amount of radioactivity per cell, or by crosslinking a modified TIE-2 ligand to cell surface proteins using DSS, as described in Meakin and Shooter, 1991, Neuron 6:153-163, and detecting the amount of labeled protein in cell extracts using, for example, SDS polyacrylamide gel electrophoresis, which may reveal a labeled protein having a size corresponding to TIE receptor/modified TIE-2 ligand. The specific test molecule/TIE interaction may further be tested by adding to the assays various dilutions of an unlabeled control ligand that does not bind the TIE receptor and therefore should have no substantial effect on the competition between labeled modified TIE-2 ligand and test molecule for TIE binding. Alternatively, a molecule known to be able to disrupt TIE receptor/modified TIE-2 ligand binding, such as, but not limited to, anti-TIE antibody, or TIE receptorbody as described herein, may be expected to interfere with the competition between ¹²⁵I-modified TIE-2 ligand and test molecule for TIE receptor binding.

Detectably labeled modified TIE-2 ligand includes, but is not limited to, a modified TIE-2 ligand linked covalently or noncovalently to a radioactive substance, a fluorescent substance, a substance that has enzymatic activity, a substance that may serve as a substrate for an enzyme (enzymes and substrates associated with colorimetrically detectable reactions are preferred) or to a substance that can be recognized by an antibody molecule that is preferably a detectably labeled antibody molecule.

Alternatively, the specific binding of test molecule to TIE may be measured by evaluating the secondary biological effects of a modified TIE-2 ligand/TIE receptor binding, including, but not limited to, cell growth and/or differentiation or immediate early gene expression or phosphorylation of TIE. For example, the ability of the test molecule to induce differentiation can be tested in cells that lack tie and in comparable cells that express tie; differentiation in tie-expressing cells but not in comparable cells that lack tie would be indicative of a specific test molecule/TIE interaction. A similar analysis could be performed by detecting immediate early gene (e.g. fos and jun) induction in tieminus and tie-plus cells, or by detecting phosphorylation of

TIE using standard phosphorylation assays known in the art. Such analysis might be useful in identifying agonists or antagonists that do not competitively bind to TIE.

Similarly, the present invention provides for a method of identifying a molecule that has the biological activity of a 5 modified TIE-2 ligand comprising (i) exposing a cell that expresses tie to a test molecule and (ii) detecting the specific binding of the test molecule to TIE receptor, in which specific binding to TIE positively correlates with TIE-like activity. Specific binding may be detected by either assaying 10 for direct binding or the secondary biological effects of binding, as discussed supra. Such a method may be particularly useful in identifying new members of the TIE ligand family or, in the pharmaceutical industry, in screening a large array of peptide and non-peptide molecules (e.g., 15 peptidomimetics) for TIE associated biological activity. In a preferred, specific, nonlimiting embodiment of the invention, a large grid of culture wells may be prepared that contain, in alternate rows, PC12 (or fibroblasts, see infra) cells that are either tie-minus or engineered to be tie-plus. A 20 variety of test molecules may then be added such that each column of the grid, or a portion thereof, contains a different test molecule. Each well could then be scored for the presence or absence of growth and/or differentiation. An extremely large number of test molecules could be screened 25 for such activity in this manner.

In additional embodiments, the invention provides for methods of detecting or measuring TIE ligand-like activity or identifying a molecule as having such activity comprising (i) exposing a test molecule to a TIE receptor protein in vitro 30 under conditions that permit binding to occur and (ii) detecting binding of the test molecule to the TIE receptor protein, in which binding of test molecule to TIE receptor correlates with TIE ligand-like activity. According to such methods, the TIE receptor may or may not be substantially 35 purified, may be affixed to a solid support (e.g. as an affinity column or as an ELISA assay), or may be incorporated into an artificial membrane. Binding of test molecule to TIE receptor may be evaluated by any method known in the art. In preferred embodiments, the binding of test molecule may 40 be detected or measured by evaluating its ability to compete with detectably labeled known TIE ligands for TIE receptor

The present invention also provides for a method of detecting the ability of a test molecule to function as an 45 antagonist of TIE ligand-like activity comprising detecting the ability of the molecule to inhibit an effect of TIE ligand binding to TIE receptor on a cell that expresses the receptor. Such an antagonist may or may not interfere with TIE receptor/modified TIE-2 ligand binding. Effects of a modified TIE-2 ligand binding to TIE receptor are preferably biological or biochemical effects, including, but not limited to, cell survival or proliferation, cell transformation, immediate early gene induction, or TIE phosphorylation.

The invention further provides for both a method of 55 identifying antibodies or other molecules capable of neutralizing the ligand or blocking binding to the receptor, as well as the molecules identified by the method. By way of nonlimiting example, the method may be performed via an assay which is conceptually similar to an ELISA assay. For example, TIE receptorbody may be bound to a solid support, such as a plastic multiwell plate. As a control, a known amount of a modified TIE-2 ligand which has been Myctagged may then be introduced to the well and any tagged modified TIE-2 ligand which binds the receptorbody may 65 then be identified by means of a reporter antibody directed against the Myctag. This assay system may then be used to

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screen test samples for molecules which are capable of i) binding to the tagged ligand or ii) binding to the receptor-body and thereby blocking binding to the receptor-body by the tagged ligand. For example, a test sample containing a putative molecule of interest together with a known amount of tagged ligand may be introduced to the well and the amount of tagged ligand which binds to the receptor-body may be measured. By comparing the amount of bound tagged ligand in the test sample to the amount in the control, samples containing molecules which are capable of blocking ligand binding to the receptor may be identified. The molecules of interest thus identified may be isolated using methods well known to one of skill in the art.

Once a blocker of ligand binding is found, one of skill in the art would know to perform secondary assays to determine whether the blocker is binding to the receptor or to the ligand, as well as assays to determine if the blocker molecule can neutralize the biological activity of the ligand. For example, by using a binding assay which employs BIAcore biosensor technology (or the equivalent), in which either TIE receptorbody or a modified TIE-2 ligand or ligandbody is covalently attached to a solid support (e.g. carboxymethyl dextran on a gold surface), one of skill in the art would be able to determine if the blocker molecule is binding specifically to the ligand, ligandbody or to the receptorbody. To determine if the blocker molecule can neutralize the biological activity of the ligand, one of skill in the art could perform a phosphorylation assay (see Example 5) or alternatively, a functional bioassay, such as a survival assay, by using primary cultures of, for example, endothelial cells. Alternatively, a blocker molecule which binds to the receptorbody could be an agonist and one of skill in the art would know to how to determine this by performing an appropriate assay for identifying additional agonists of the TIE receptor.

In addition, the invention further contemplates compositions wherein the TIE ligand is the receptor binding domain of a TIE-2 ligand described herein. For example, TIE-2 ligand 1 contains a "coiled coil" domain (beginning at the 5' end and extending to the nucleotide at about position 1160 of FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2] and about position 1157 of FIGS. 5A-5D [SEQ ID NO: 3 and SEQ ÎD NO: 4]) and a fibrinogen-like domain (which is encoded by the nucleotide sequence of FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2] beginning at about position 1161 and about position 1158 of FIGS. 5A-5D [SEQ ID NO: 3 and SEQ ID NO: 4]). The fibrinogen-like domain of TIE-2 ligand 2 is believed to begin on or around the same amino acid sequence as in ligand 1 (FRDCA) which is encoded by nucleotides beginning around 1197 of FIGS. 6A-6D (SEQ ID NO: 5 and SEQ ID NO: 6). The fibrinogen-like domain of TIE ligand-3 is believed to begin on or around the amino acid sequence which is encoded by nucleotides beginning around position 929 as set forth in FIGS. 21A-21C (SEO ID NO: 9 and SEQ ID NO: 10). Multimerization of the coiled coil domains during production of the ligand hampers purification. As described in Example 19, Applicants have discovered, however, that the fibrinogen-like domain comprises the TIE-2 receptor binding domain. The monomeric forms of the fibrinogen-like domain do not, however, appear to bind the receptor. Studies utilizing myc-tagged fibrinogen-like domain, which has been "clustered" using anti-myc antibodies, do bind the TIE-2 receptor. [Methods of production of "clustered ligands and ligandbodies are described in Davis, et al. Science 266:816-819 (1994)]. Based on these finding, applicants produced "ligandbodies' which comprise the fibrinogen-like domain of the TIE-2 ligands coupled to the Fc domain of IgG ("fFc's"). These

ligandbodies, which form dimers, efficiently bind the TIE-2 receptor. Accordingly, the present invention contemplates the production of modified TIE ligandbodies which may be used as targeting agents, in diagnostics or in therapeutic applications, such as targeting agents for tumors and/or associated vasculature wherein a TIE antagonist is indicated.

The invention herein further provides for the development of the ligand, a fragment or derivative thereof, or another molecule which is a receptor agonist or antagonist, as a therapeutic for the treatment of patients suffering from 10 disorders involving cells, tissues or organs which express the TIE receptor. Such molecules may be used in a method of treatment of the human or animal body, or in a method of diagnosis.

Because TIE receptor has been identified in association 15 with endothelial cells and, as demonstrated herein, blocking of TIE-2 ligand 1 appears to prevent vascularization, applicants expect that a modified TIE-2 ligand described herein may be useful for the induction of vascularization in diseases or disorders where such vascularization is indicated. 20 Such diseases or disorders would include wound healing, ischaemia and diabetes. The ligands may be tested in animal models and used therapeutically as described for other agents, such as vascular endothelial growth factor (VEGF), another endothelial cell-specific factor that is angiogenic. 25 Ferrara, et al. U.S. Pat. No. 5,332,671 issued Jul. 26, 1994. The Ferrara reference, as well as other studies, describe in vitro and in vivo studies that may be used to demonstrate the effect of an angiogenic factor in enhancing blood flow to ischemic myocardium, enhancing wound healing, and in 30 other therapeutic settings wherein neoangiogenesis is desired. [see Sudo, et al. European Patent Application 0 550 296 A2 published Jul. 7, 1993; Banai, et al. Circulation 89:2183-2189 (1994); Unger, et al. Am. J. Physiol. 266:H1588-H1595 (1994); Lazarous, et al. Circulation 35 91:145-153 (1995)]. According to the invention, a modified TIE-2 ligand may be used alone or in combination with one or more additional pharmaceutically active compounds such as, for example, VEGF or basic fibroblast growth factor (bFGF), as well as cytokines, neurotrophins, etc.

Conversely, antagonists of the TIE receptor, such as modified TIE-2 ligands which bind but do not activate the receptor as described herein, receptorbodies as described herein in Examples 2 and 3, and TIE-2 ligand 2 as described in Example 9, would be useful to prevent or attenuate 45 vascularization, thus preventing or attenuating, for example, tumor growth. These agents may be used alone or in combination with other compositions, such as anti-VEGF antibodies, that have been shown to be useful in treating conditions in which the therapeutic intent is to block angiogenesis. Applicants expect that a modified TIE-2 ligand described herein may also be used in combination with agents, such as cytokine antagonists such as IL-6 antagonists, that are known to block inflammation.

For example, applicants have determined that TIE ligands 55 are expressed in cells within, or closely associated with, tumors. For example, TIE-2 ligand 2 appears to be tightly associated with tumor endothelial cells. Accordingly, it and other TIE antagonists may also be useful in preventing or attenuating, for example, tumor growth. In addition, TIE 60 ligands or ligandbodies may be useful for the delivery of toxins to a receptor bearing cell. Alternatively, other molecules, such as growth factors, cytokines or nutrients, may be delivered to a TIE receptor bearing cell via TIE ligands or ligandbodies. TIE ligands or ligandbodies such as 65 modified TIE-2 ligand described herein may also be used as diagnostic reagents for TIE receptor, to detect the receptor in

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vivo or in vitro. Where the TIE receptor is associated with a disease state, TIE ligands or ligandbodies such as a modified TIE-2 ligand may be useful as diagnostic reagents for detecting the disease by, for example, tissue staining or whole body imaging. Such reagents include radioisotopes, flurochromes, dyes, enzymes and biotin. Such diagnostics or targeting agents may be prepared as described in Alitalo, et al. WO 95/26364 published Oct. 5, 1995 and Burrows, F. and P. Thorpe, PNAS (USA) 90:8996–9000 (1993) which is incorporated herein in its entirety.

In other embodiments, the TIE ligands, a receptor activating modified TIE-2 ligand described herein are used as hematopoietic factors. A variety of hematopoietic factors and their receptors are involved in the proliferation and/or differentiation and/or migration of the various cells types contained within blood. Because the TIE receptors are expressed in early hematopoietic cells, the TIE ligands are expected to play a comparable role in the proliferation or differentiation or migration of these cells. Thus, for example, TIE containing compositions may be prepared, assayed, examined in in vitro and in vivo biological systems and used therapeutically as described in any of the following: Sousa, U.S. Pat. No. 4,810,643, Lee, et al., Proc. Natl. Acad. Sci. USA 82:4360-4364 (1985) Wong, et al. Science, 228:810-814 (1985); Yokota, et al. Proc. Natl. Acad. Sci (USA) 81:1070 (1984); Bosselman, et al. WO 9105795 published May 2, 1991 entitled "Stem Cell Factor" and Kirkness, et al. WO 95/19985 published Jul. 27, 1995 entitled "Haemopoietic Maturation Factor". Accordingly, receptor activating modified TIE-2 ligand may be used to diagnose or treat conditions in which normal hematopoiesis is suppressed, including, but not limited to anemia, thrombocytopenia, leukopenia and granulocytopenia. In a preferred embodiment, receptor activating modified TIE-2 ligand may be used to stimulate differentiation of blood cell precursors in situations where a patient has a disease, such as acquired immune deficiency syndrome (AIDS) which has caused a reduction in normal blood cell levels, or in clinical settings in which enhancement of hematopoietic populations 40 is desired, such as in conjunction with bone marrow transplant, or in the treatment of aplasia or myelosuppression caused by radiation, chemical treatment or chemo-

The receptor activating modified TIE-2 ligands of the present invention may be used alone, or in combination with another pharmaceutically active agent such as, for example, ctyokines, neurotrophins, interleukins, etc. In a preferred embodiment, the ligands may be used in conjunction with any of a number of the above referenced factors which are known to induce stem cell or other hematopoietic precursor proliferation, or factors acting on later cells in the hematopoietic pathway, including, but not limited to, hemopoietic maturation factor, thrombopoietin, stem cell factor, erythropoietin, G-CSF, GM-CSF, etc.

In an alternative embodiment, TIE receptor antagonists are used to diagnose or treat patients in which the desired result is inhibition of a hematopoietic pathway, such as for the treatment of myeloproliferative or other proliferative disorders of blood forming organs such as thrombocythemias, polycythemias and leukemias. In such embodiments, treatment may comprise use of a therapeutically effective amount of the a modified TIE-2 ligand, TIE antibody, TIE receptorbody, a conjugate of a modified TIE-2 ligand, or a ligandbody or fFC as described herein.

The present invention also provides for pharmaceutical compositions comprising a modified TIE-2 ligand or ligandbodies described herein, peptide fragments thereof, or

derivatives in a pharmacologically acceptable vehicle. The modified TIE-2 ligand proteins, peptide fragments, or derivatives may be administered systemically or locally. Any appropriate mode of administration known in the art may be used, including, but not limited to, intravenous, intrathecal, intraarterial, intranasal, oral, subcutaneous, intraperitoneal, or by local injection or surgical implant. Sustained release formulations are also provided for.

The present invention also provides for an antibody which specifically binds such a therapeutic molecule. The antibody may be monoclonal or polyclonal. The invention also provides for a method of using such a monoclonal or polyclonal antibody to measure the amount of the therapeutic molecule in a sample taken from a patient for purposes of monitoring the course of therapy.

The invention further provides for a therapeutic composition comprising a modified TIE-2 ligand or ligandbody and a cytotoxic agent conjugated thereto. In one embodiment, the cytotoxic agent may be a radioisotope or toxin.

The invention also provides for an antibody which specifically binds a modified TIE-2 ligand. The antibody may 20 be monoclonal or polyclonal. The invention further provides for a method of purifying a modified TIE-2 ligand comprising:

- a) coupling at least one TIE binding substrate to a solid matrix;
- b) incubating the substrate of a) with a cell lysate so that the substrate forms a complex with any modified TIE-2 ligand in the cell lysate;
- c) washing the solid matrix; and
- d) eluting the modified TIE-2 ligand from the coupled 30 substrate.

The substrate may be any substance that specifically binds the modified TIE-2 ligand. In one embodiment, the substrate is selected from the group consisting of anti-modified TIE-2 ligand antibody, TIE receptor and TIE receptorbody. The 35 invention further provides for a receptorbody which specifically binds a modified TIE-2 ligand, as well as a therapeutic composition comprising the receptorbody in a pharmaceutically acceptable vehicle, and a method of blocking blood vessel growth in a human comprising administering an 40 effective amount of the therapeutic composition.

The invention also provides for a therapeutic composition comprising a receptor activating modified TIE-2 ligand or ligandbody in a pharmaceutically acceptable vehicle, as well as a method of promoting neovascularization in a patient 45 comprising administering to the patient an effective amount of the therapeutic composition.

In addition, the present invention provides for a method for identifying a cell which expresses TIE receptor which comprises contacting a cell with a detectably labeled modified TIE-2 ligand or ligandbody, under conditions permitting binding of the detectably labeled ligand to the TIE receptor and determining whether the detectably labeled ligand is bound to the TIE receptor, thereby identifying the cell as one which expresses TIE receptor. The present invention also 55 provides for a therapeutic composition comprising a modified TIE-2 ligand or ligandbody and a cytotoxic agent conjugated thereto. The cytotoxic agent may be a radioisotope or toxin.

The invention also provides a method of detecting expression of a modified TIE-2 ligand by a cell which comprises obtaining mRNA from the cell, contacting the mRNA so obtained with a labeled nucleic acid molecule encoding a modified TIE-2 ligand, under hybridizing conditions, determining the presence of mRNA hybridized to the labeled 65 molecule, and thereby detecting the expression of a modified TIE-2 ligand in the cell.

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The invention further provides a method of detecting expression of a modified TIE-2 ligand in tissue sections which comprises contacting the tissue sections with a labeled nucleic acid molecule encoding a modified TIE-2 ligand, under hybridizing conditions, determining the presence of mRNA hybridized to the labelled molecule, and thereby detecting the expression of a modified TIE-2 ligand in tissue sections.

EXAMPLE 1

Identification of the ABAE Cell Line as Reporter Cells for the TIE-2 Receptor

Adult BAE cells are registered in the European Cell Culture Repository, under ECACC#92010601. (See PNAS 75:2621 (1978)). Northern (RNA) analyses revealed moderate levels of tie-2 transcripts in the ABAE (Adult Bovine Arterial Endothelial) cell line, consistent with in situ hybridization results that demonstrated almost exclusive localization of tie-2 RNAs to vascular endothelial cells. We therefore examined ABAE cell lysates for the presence of TIE-2 protein, as well as the extent to which this TIE-2 protein is tyrosine-phosphorylated under normal versus serumdeprived growth conditions. ABAE cell lysates were harvested and subjected to immunoprecipitation, followed by Western blot analyses of immunoprecipitated proteins with TIE-2 specific and phosphotyrosine-specific antisera. Omission or inclusion of TIE-2 peptides as specific blocking molecules during TIE-2 immunoprecipitation allowed unambiguous identification of TIE-2 as a moderately detectable protein of ~150 kD whose steady-state phosphotyrosine levels diminish to near undetectable levels by prior serumstarvation of the cells.

Culture of ABAE cells and harvest of cell lysates was done as follows. Low-passage-number ABAE cells were plated as a monolayer at a density of 2×10^6 cells/150 mm plastic petri plate (Falcon) and cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% bovine calf serum (10% BCS), 2 mM L-glutamine (Q) and 1% each of penicillin and streptomycin (P-S) in an atmosphere of 5% CO₂. Prior to harvest of cell lysates, cells were serumstarved for 24 hours in DMEM/Q/P-S, followed by aspiration of the medium and rinsing of the plates with ice-cold phosphate buffered saline (PBS) supplemented with sodium orthovanadate, sodium fluoride and sodium benzamidine. Cells were lysed in a small volume of this rinse buffer that had been supplemented with 1% NP40 detergent and the protease inhibitors PMSF and aprotinin. Insoluble debris was removed from the cell Iysates by centrifugation at 14,000×G for 10 minutes, at 4° C. and the supernatants were subjected to immunoprecipitation with antisera specific for TIE-2 receptor, with or without the presence of blocking peptides added to ~20 µg/ml lysate. Immunoprecipitated proteins were resolved by PAGE (7.5% Laemmli gel), and then electro-transferred to PVDF membrane and incubated either with various TIE-2- or phosphotyrosine-specific antisera. TIE-2 protein was visualized by incubation of the membrane with HRP-linked secondary antisera followed by treatment with ECL reagent (Amersham).

EXAMPLE 2

Cloning and Expression of TIE-2 Receptorbody for Affinity-based Study of TIE-2 Ligand Interactions

An expression construct was created that would yield a secreted protein consisting of the entire extracellular portion

of the rat TIE-2 receptor fused to the human immunoglobulin gamma-1 constant region (IgG1 Fc). This fusion protein is called a TIE-2 "receptorbody" (RB), and would be normally expected to exist as a dimer in solution based on formation of disulfide linkages between individual IgG1 Fc tails. The Fc portion of the TIE-2 RB was prepared as follows. A DNA fragment encoding the Fc portion of human IgG1 that spans from the hinge region to the carboxyterminus of the protein, was amplified from human placental cDNA by PCR with oligonucleotides corresponding to the 10 published sequence of human IgG1; the resulting DNA fragment was cloned in a plasmid vector. Appropriate DNA restriction fragments from a plasmid encoding the fulllength TIE-2 receptor and from the human IgG1 Fc plasmid were ligated on either side of a short PCR-derived fragment that was designed so as to fuse, in-frame, the TIE-2 and human IgG1 Fc protein-coding sequences. Thus, the resulting TIE-2 ectodomain-Fc fusion protein precisely substituted the IgG1 Fc in place of the region spanning the TIE-2 transmembrane and cytoplasmic domains. An alternative 20 method of preparing RBs is described in Goodwin, et. al. Cell 73:447-456 (1993).

Milligram quantities of TIE-2 RB were obtained by cloning the TIE-2 RB DNA fragment into the pVL1393 baculovirus vector and subsequently infecting the 25 Spodoptera frugiperda SF-21AE insect cell line. Alternatively, the cell line SF-9 (ATCC Accession No. CRL-1711) or the cell line BTI-TN-5b1-4 may be used. DNA encoding the TIE-2 RB was cloned as an Eco RI-NotI fragment into the baculovirus transfer plasmid pVL1393. 30 Plasmid DNA purified by cesium chloride density gradient centrifugation was recombined into viral DNA by mixing 3 μg of plasmid DNA with 0.5 μg of Baculo-Gold DNA (Pharminigen), followed by introduction into liposomes using 30 µg Lipofectin (GIBCO-BRL). DNA-liposome mix- 35 tures were added to SF-21AE cells (2×10^6 cells/60 mm dish) in TMN-FH medium (Modified Grace's Insect Cell Medium (GIBCO-BRL) for 5 hours at 27° C., followed by incubation at 27° C. for 5 days in TMN-FH medium supplemented with 5% fetal calf serum. Tissue culture medium was harvested 40 for plaque purification of recombinant viruses, which was carried out using methods previously described (O'Reilly, D. R., L. K. Miller, and V. A. Luckow, Baculovirus Expression Vectors—A Laboratory Manual. 1992, New York: W. H. Freeman) except that the agarose overlay contained 125 45 μg/mL X-gal (5-bromo-4-chloro-3-indolyl-β-Dgalactopyranoside; GIBCO-BRL). After 5 days of incubation at 27° C., non-recombinant plaques were scored by positive chromogenic reaction to the X-gal substrate, and their positions marked. Recombinant plaques were then 50 visualized by addition of a second overlay containing 100 $\mu g/mL$ MTT (3-[4,5-dimethylthiazol-2-yl]2,5, diphenyltetrazolium bromide; Sigma). Putative recombinant virus plaques were picked by plug aspiration, and purified by multiple rounds of plaque isolation to assure homogeneity. 55 Virus stocks were generated by serial, low-multiplicity passage of plaque-purified virus. Low passage stocks of one virus clone (vTIE-2 receptorbody) were produced.

SF-21AE cells were cultured in serum free medium (SF-900 II, Gibco BRL) containing 1× antibiotic/ 60 antimycotic solution (Gibco BRL) and 25 mg/L Gentamycin (Gibco BRL). Pluronic F-68 was added as a surfactant to a final concentration of 1 g/L. Cultures (4L) were raised in a bioreactor (Artisan Cell Station System) for at least three days prior to infection. Cells were grown at 27° C., with 65 gassing to 50% dissolved oxygen, at a gas flow rate of 80 mL/min (aeration at a sparge ring). Agitation was by means

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of a marine impeller at a rate of 100 rpm. Cells were harvested in mid-logarithmic growth phase ($\sim 2\times 10^6$ cells/mL), concentrated by centrifugation, and infected with 5 plaque forming units of vTIE-2 receptorbody per cell. Cells and inoculum were brought to 400 mL with fresh medium, and virus was adsorbed for 2 hours at 27° C. in a spinner flask. The culture was then resuspended in a final volume of 8L with fresh serum-free medium, and the cells incubated in the bioreactor using the previously described conditions.

Culture medium from vTIE-2 receptorbody-infected SF21AE cells were collected by centrifugation (500×g, 10 minutes) at 72 hours post-infection. Cell supernatants were brought to pH 8 with NaOH. EDTA was added to a final concentration of 10 mM and the supernatant pH was readjusted to 8. Supernatants were filtered (0.45 µm, Millipore) and loaded on a protein A column (protein A sepharose 4 fast flow or HiTrap protein A, both from Pharmacia). The column was washed with PBS containing 0.5 M NaCl until the absorbance at 280 nm decreased to baseline. The column was washed in PBS and eluted with 0.5 M acetic acid. Column fractions were immediately neutralized by eluting into tubes containing 1 M Tris pH 9. The peak fractions containing the TIE-2 receptorbody were pooled and dialyzed versus PBS.

EXAMPLE 3

Demonstration that TIE-2 has a Critical Role in Development of the Vasculature

Insight into the function of TIE-2 was gained by introduction of "excess" soluble TIE-2 receptorbody (TIE-2 RB) into a developing system. The potential ability of TIE-2 RB to bind, and thereby neutralize, available TIE-2 ligand could result in an observable disruption of normal vascular development and characterization of the ligand. To examine whether TIE-2 RB could be used to disrupt vascular development in early chick embryos, small pieces of a biologically resorbable foam were soaked with TIE-2 RB and inserted immediately beneath the chorioallantoic membrane at positions just lateral to the primitive embryo.

Early chicken embryos develop atop the yolk from a small disk of cells that is covered by the chorioallantoic membrane (CAM). The endothelial cells that will come to line the vasculature in the embryo arise from both extra- and intraembryonic cell sources. Extra-embryonically-derived endothelial cells, which provide the major source of endothelial cells in the embryo, originate from accretions of mesenchyme that are situated laterally around the embryo-proper, just underneath the CAM. As these mesenchyme cells mature, they give rise to a common progenitor of both the endothelial and hematopoietic cell lineages, termed the hemangioblast. In turn, the hemangioblast gives rise to a mixed population of angioblasts (the endothelial cell progenitor) and hematoblasts (the pluripotential hematopoietic precursor). Formation of rudiments of the circulatory system begins when endothelial cell progeny segregate to form a one-cell-thick vesicle that surrounds the primitive blood cells. Proliferation and migration of these cellular components eventually produces a vast network of bloodfilled microvessels under the CAM that will ultimately invade the embryo to join with limited, intra-embryonicallyderived vascular elements.

Newly fertilized chicken eggs obtained from Spafas, Inc. (Boston, Mass.) were incubated at 99.5° F., 55% relative humidity. At about 24 hrs. of development, the egg shell was wiped down with 70% ethanol and a dentist's drill was used

to make a 1.5 cm. hole in the blunt apex of each egg. The shell membrane was removed to reveal an air space directly above the embryo. Small rectangular pieces of sterile Gelfoam (Upjohn) were cut with a scalpel and soaked in equal concentrations of either TIE-2- or EHK-1 receptorbody. EHK-1 receptorbody was made as set forth in Example 2 using the EHK-1 extracellular domain instead of the TIE-2 extracellular domain (Maisonpierre et al., Oncogene 8:3277-3288 (1993). Each Gelfoam piece absorbed approximately 6 ug of protein in 30 ul. Sterile watchmakers forceps 10 were used to make a small tear in the CAM at a position several millimeters lateral to the primitive embryo. The majority of the piece of RB-soaked Gelfoam was inserted under the CAM and the egg shell was sealed over with a piece of adhesive tape. Other similarly-staged eggs were 15 treated in parallel with RB of the unrelated, neuronally expressed receptor tyrosine kinase, EHK-1 (Maisonpierre et al., Oncogene 8:3277-3288 (1993). Development was allowed to proceed for 4 days and then the embryos were examined by visual inspection. Embryos were removed by 20 carefully breaking the shells in dishes of warmed PBS and carefully cutting away the embryo with surrounding CAM. Of 12 eggs treated with each RB, 6 TIE-2 RB and 5 EHK-1 RB treated embryos had developed beyond the stage observed at the start of the experiment. A dramatic difference 25 was seen between these developed embryos, as shown in FIGS. 1A and 1B. Those treated with EHK-1 RB appeared to have developed relatively normally. Four out of five EHK-1 embryos were viable as judged by the presence of a beating heart. Furthermore, the extra-embryonic 30 vasculature, which is visually obvious due to the presence of red blood cells, was profuse and extended several centimeters laterally under the CAM. By contrast, those treated with TIE-2 RB were severely stunted, ranging from 2-5 mm. in diameter, as compared with more than 10 mm in diameter 35 for the EHK-1 RB embryos. All of the TIE-2 RB treated embryos were dead and their CAMs were devoid of blood vessels. The ability of TIE-2 RB to block vascular development in the chicken demonstrates that TIE-2 ligand is necessary for development of the vasculature.

EXAMPLE 4

Identification of a TIE-2-Specific Binding Activity in Conditioned Medium from the ras Oncogene-Transformed C2C12 Mouse Myoblast Cell Line

Screening of ten-fold-concentrated cell-conditioned media (10× CCM) from various cell lines for the presence of soluble, TIE-2-specific binding activity (BIAcore; Pharmacia Biosensor, Piscataway, N.J.) revealed binding activity in serum-free medium from oncogenic-ras-transformed C2C12 cells (C2C12-ras), RAT 2-ras (which is a ras transformed fibroblast cell line), human glioblastoma T98G and the human neuroblastoma cell line known as SHEP-1.

The C2C12-ras 10× CCM originated from a stably transfected line of C2C12 myoblasts that was oncogenically transformed by transfection with the T-24 mutant of H-ras by standard calcium phosphate-based methods. An SV40 based neomycin-resistance expression plasmid was physically linked with the ras expression plasmid in order to 60 permit selection of transfected clones. Resulting G418-resistant ras-C2C12 cells were routinely maintained as a monolayer on plastic dishes in DMEM/glutamine/penicillinstreptomycin supplemented with 10% fetal calf serum (FCS). Serum-free C2C12-ras 10× CCM was made by 65 plating the cells at 60% confluence in a serum free defined media for 12 hours. [Zhan and Goldfarb, Mol. Cell. Biol. 6:

3541–3544 (1986)); Zhan, et al. Oncogene 1: 369–376 (1987)]. The medium was discarded and replaced with fresh DMEM/Q/P-S for 24 hours. This medium was harvested and cells were re-fed fresh DMEM/Q/P-S, which was also harvested after a further 24 hours. These CCM were supplemented with the protease inhibitors PMSF (1 mM) and aprotinin (10 μg/ml), and ten-fold concentrated on sterile size-exclusion membranes (Amicon). TIE-2-binding activity could be neutralized by incubation of the medium with an excess of TIE-2 RB, but not by incubation with EHK-1 RB, prior to BIAcore analysis.

Binding activity of the 10× CCM was measured using biosensor technology (BIAcore; Pharmacia Biosensor, Piscataway, N.J.) which monitors biomolecular interactions in real-time via surface plasmon resonance. Purified TIE-2 RB was covalently coupled through primary amines to the carboxymethyl dextran layer of a CM5 research grade sensor chip (Pharmacia Biosensor; Piscataway, N.J.). The sensor chip surface was activated using a mixture of N-hydroxysuccinimide (NHS) and N-ethyl-N'-(3-dimethylaminopropyl)carbodiimide (EDC), followed by immobilization of TIE-2 RB (25 µg/mL, pH 4.5) and deactivation of unreacted sites with 1.0 M ethanolamine (pH 8.5). A negative control surface of the EHK-1 receptorbody was prepared in a similar manner.

The running buffer used in the system was HBS (10 mM Hepes, 3.4 mM EDTA, 150 mM NaCl, 0.005% P20 surfactant, pH 7.4). The $10\times$ CCM samples were centrifuged for 15 min at 4° C. and further clarified using a sterile, low protein-binding 0.45 μ m filter (Millipore; Bedford, Mass.). Dextran (2 mg/ml) and P20 surfactant (0.005%) were added to each CCM sample. Aliquots of 40 μ L were injected across the immobilized surface (either TIE-2 or EHK-1) at a flow rate of 5 μ L/min and the receptor binding was monitored for 8 min. The binding activity (resonance units, RU) was measured as the difference between a baseline value determined 30 s prior to the sample injection and a measurement taken at 30 s post-injection. Regeneration of the surface was accomplished with one 12- μ L pulse of 3 M MgCl₂.

The instrument noise level is 20 RU; therefore, any binding activity with a signal above 20 RU may be interpreted as a real interaction with the receptor. For C2C12-ras conditioned media, the binding activities were in the range 60-90 RU for the TIE-2 RB immobilized surface. For the same samples assayed on a EHK-1 RB immobilized surface, the measured activities were less than 35 RU. Specific binding to the TIE-2 receptorbody was evaluated by incubating the samples with an excess of either soluble TIE-2 or EHK-1 RB prior to assaying the binding activity. The addition of soluble EHK-1 RB had no effect on the TIE-2 binding activity of any of the samples, while in the presence of soluble TIE-2 binding to the surface is two-thirds less than that measured in the absence of TIE-2. A repeat assay using >50x concentrated C2C12-ras CCM resulted in a four-fold enhancement over background of the TIE-2 specific binding signal.

EXAMPLE 5

C2C12-ras CCM Contains an Activity that Induces Tyrosine Phosphorylation of TIE-2 Receptor

C2C12-ras 10× CCM was examined for its ability to induce tyrosine phosphorylation of TIE-2 in ABAE cells. Serum-starved ABAE cells were briefly incubated with C2C12-ras CCM, lysed and subjected to immunoprecipitation and Western analyses as described above. Stimulation

of serum-starved ABAE cells with serum-free C2C12-ras 10× CCM was done as follows. The medium of ABAE cells starved as described above was removed and replaced with either defined medium or 10× CCM that had been prewarmed to 37° C. After 10 minutes, the media were removed and the cells were twice rinsed on ice with an excess of chilled PBS supplemented with orthovanadate/NaF/benzamidine. Cell lysis and TIE-2-specific immunoprecipitation was done as described above.

ABAE cells incubated for 10 minutes with defined 10 medium showed no induction of TIE-2 tyrosine phosphorylation, whereas incubation with C2C12-ras CCM stimulated at least a $100\times$ increase in TIE-2 phosphorylation. This activity was almost totally depleted by pre-incubation of the C2C12-ras $10\times$ CCM for 90 minutes at room temperature with 13 μ g of TIE-2 RB coupled to protein G-Sepharose beads. Medium incubated with protein G Sepharose alone was not depleted of this phosphorylating activity.

EXAMPLE 6

Expression Cloning of TIE-2 Ligand

COS-7 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS), 1% each of penicillin and streptomycin (P/S) and 2 mM glutamine in an atmosphere of 5% CO2. The mouse myoblast C2C12 ras cell line was cultured in Eagle's minimal essential medium (EMEM) with 10% FBS, (P/S) and 2 mM glutamine. Full length mouse TIE-2 ligand cDNA clones were obtained by screening a C2C12 ras cDNA library in the pJFE14 vector expressed in COS cells. This vector, as shown in FIG. 2, is a modified version of the vector pSR $_{\alpha}$ (Takebe, et al. 1988, Mol. Cell. Biol. 8:466–472). The library was created using the two BSTX1 restriction sites in the pJFE14 vector.

COS-7 cells were transiently transfected with either the pJFE14 library or control vector by the DEAE-dextran transfection protocol. Briefly, COS-7 cells were plated at a density of 1.0×10^6 cells/100 mm plate 24 hours prior to transfection. For transfection, the cells were cultured in serum-free DMEM containing 400 μ g/ml of DEAE-dextran, 1 μ M chloroquine, and 2 mM glutamine, and 1 μ g of the appropriate DNA for 3–4 hours at 37° C. in an atmosphere of 5% CO₂. The transfection media was aspirated and replaced with PBS with 10% DMSO for 2–3 min. Following this DMSO "shock", the COS-7 cells were placed into DMEM with 10% FBS, 1% each of penicillin and streptomycin, and 2 mM glutamine for 48 hours.

Because the TIE-2 ligand is secreted it was necessary to permeabilize the cells to detect binding of the receptorbody probe to the ligand. Two days after transfection the cells were rinsed with PBS and then incubated with PBS containing 1.8% formaldehyde for 15–30 min. at room temperature. Cells were then washed with PBS and incubated for 15 min. with PBS containing 0.1% Triton X-100 and 10% Bovine Calf Serum to permeabilize the cells and block non-specific binding sites.

The screening was conducted by direct localization of 60 staining using a TIE-2 receptorbody (RB), which consisted of the extracellular domain of TIE-2 fused to the IgG1 constant region. This receptorbody was prepared as set forth in Example 2. A 100 mm dish of transfected, fixed and permeabilized COS cells was probed by incubating them for 65 30 min with TIE-2 RB. The cells were then washed twice with PBS and incubated for an additional 30 min with

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PBS/10% Bovine Calf Serum/anti-human IgG-alkaline phosphatase conjugate. After three PBS washes, cells were incubated in alkaline-phosphatase substrate for 30-60 min. The dish was then inspected microscopically for the presence of stained cells. For each stained cell, a small area of cells including the stained cell was scraped from the dish using a plastic pipette tip and plasmid DNA was then rescued and used to electroporate bacterial cells. Single bacterial colonies resulting from the electroporation were picked and plasmid DNA prepared from these colonies was used to transfect COS-7 cells which were probed for TIE-2 ligand expression as evidenced by binding to TIE-2 receptorbodies. This allowed identification of single clones coding for TIE-2 ligand. Confirmation of TIE-2 ligand expression was obtained by phosphorylation of the TIE-2 receptor using the method set forth in Example 5. A plasmid clone encoding the TIE-2 ligand was deposited with the ATCC on Oct. 7, 1994 and designated as "pJFE14 encoding TIE-2 ligand" under ATCC Accession No. 75910.

EXAMPLE 7

Isolation and Sequencing of Full Length cDNA Clone Encoding Human TIE-2 Ligand

A human fetal lung cDNA library in lambda gt-10 (see FIG. 3) was obtained from Clontech Laboratories, Inc. (Palo Alto, Calif.). Plaques were plated at a density of 1.25×10⁶/20×20 cm plate, and replica filters taken following standard procedures (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., page 8.46, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.).

Isolation of human tie-2 ligand clones was carried out as follows. A 2.2 kb Xhol fragment from the deposited tie-2 ligand clone (ATCC NO. 75910—see Example 6 above) was labeled by random priming to a specific activity of approximately 5×10⁸ cpm/ng. Hybridization was carried out at 65° C. in hybridization solution containing 0.5 mg/ml salmon sperm DNA. The filters were washed at 65° C. in 2×SSC, 0.1% SDS and exposed to Kodak XAR-5 film overnight at -70° C. Positive phage were plaque purified. High titre phage lysates of pure phage were used for isolation of DNA via a Qiagen column using standard techniques (Qiagen, Inc., Chatsworth, Calif., 1995 catalog, page 36). Phage DNA was digested with EcoRI to release the cloned cDNA fragment for subsequent subcloning. A lambda phage vector containing human tie-2 ligand DNA was deposited with the ATCC on Oct. 26, 1994 under the designation λgt10 encoding htie-2 ligand 1 (ATCC Accession No. 75928). Phage DNA may be subjected directly to DNA sequence analysis by the dideoxy chain termination method (Sanger, et al., 1977, Proc. Natl. Acad. Sci. U.S.A. 74: 5463-5467).

Subcloning of the human tie-2 ligand DNA into a mammalian expression vector may be accomplished as follows. The clone λgt10 encoding htie-2 ligand 1 contains an EcoRI site located 490 base pairs downstream from the start of the coding sequence for the human TIE-2 ligand. The coding region may be excised using unique restriction sites upstream and downstream of the initiator and stop codons respectively. For example, an Spel site, located 70 bp 5' to the initiator codon, and a Bpu1102i (also known as Blpl) site, located 265 bp 3' to the stop codon, may be used to excise the complete coding region. This may then be subcloned into the pJFE14 cloning vector, using the Xbal (compatible to the Spel overhang) and the Pstl sites (the Pstl and Bpu1102i sites are both made blunt ended).

The coding region from the clone λgt10 encoding htie-2 ligand 1 was sequenced using the ABI 373A DNA sequencer

and Taq Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, Inc., Foster City, Calif.). The nucleotide and deduced amino acid sequence of human TIE-2 ligand from the clone $\lambda gt10$ encoding htie-2 ligand 1 is shown in FIGS. 4A–4D (SEQ ID NO: 1 and SEQ ID NO: 2).

In addition, full length human tie-2 ligand cDNA clones were obtained by screening a human glioblastoma T98G cDNA library in the pJFE14 vector. Clones encoding human TIE-2 ligand were identified by DNA hybridization using a 2.2 kb XhoI fragment from the deposited tie-2 ligand clone 10 (ATCC NO. 75910) as a probe (see Example 6 above). The coding region was sequenced using the ABI 373A DNA sequencer and Taq Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, Inc., Foster City, Calif.). This sequence was nearly identical to that of clone λgt10 encod- 15 ing htie-2 ligand 1. As shown in FIGS. 4A-4D (SEQ ID NO: 1 and SEQ ID NO: 2), the clone λgt10 encoding htie-2 ligand 1 contains an additional glycine residue which is encoded by nucleotides 1114-1116. The coding sequence of the T98G clone does not contain this glycine residue but 20 otherwise is identical to the coding sequence of the clone λgt10 encoding htie-2 ligand 1. FIGS. 5A-5D (SEQ ID NO: 3 and SEQ ID NO: 4) sets forth the nucleotide and deduced amino acid sequence of human TIE-2 ligand from the T98G clone.

EXAMPLE 8

Isolation and Sequencing of Second Full Length cDNA Clone a Encoding Human TIE-2 Ligand

A human fetal lung cDNA library in lambda gt-10 (see FIG. 3) was obtained from Clontech Laboratories, Inc. (Palo Alto, Calif.). Plaques were plated at a density of 1.25×10⁶ 20×20 cm plate, and replica filters taken following standard procedures (Sambrook, et al., Molecular Cloning: A Labo- 35 ratory Manual, 2nd Ed., page 8.46, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.). Duplicate filters were screened at low stringency (2xSSC, 55° C.) with probes made to the human TIE-2 ligand 1 sequence. One of the duplicate filters was probed with a 5' probe, encoding 40 amino acids 25-265 of human TIE-2 ligand 1 as set forth in FIGS. 4A-4D (SEQ ID NO: 1 and SEQ ID NO: 2). The second duplicate filter was probed with a 3' probe, encoding amino acids 282-498 of human TIE-2 ligand 1 sequence (see FIGS. 4A-4D (SEQ ID NO: 1 and SEQ ID NO: 2). Both 45 probes were hybridized at 55° C. in hybridization solution containing 0.5 mg/ml salmon sperm DNA. Filters were washed in 2×SSC at 55° C. and exposed overnight to X-ray film. In addition, duplicate filters were also hybridized at normal stringency (2×SSC, 65° C.) to the full length coding 50 probe of mouse TIE-2 ligand 1 (F3-15, XhoI insert). Three positive clones were picked that fulfilled the following criteria: i. hybridization had not been seen to the full length (mouse) probe at normal stringency, and ii. hybridization was seen at low stringency to both 5' and 3' probes. EcoRI 55 digestion of phage DNA obtained from these clones indicated two independent clones with insert sizes of approximately 2.2 kb and approximately 1.8 kb. The 2.2 kb EcoRI insert was subcloned into the EcoRI sites of both pBluescript KS (Stratagene) and a mammalian expression vector suit- 60 able for use in COS cells. Two orientations were identified for the mammalian expression vector. The 2.2 kb insert in pBluescript KS was deposited with the ATCC on Dec. 9, 1994 and designated as pBluescript KS encoding human TIE 2 ligand 2. The start site of the TIE-2 ligand 2 coding 65 sequence is approximately 355 base pairs downstream of the pBluescript EcoRI site.

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COS-7 cells were transiently transfected with either the expression vector or control vector by the DEAE-dextran transfection protocol. Briefly, COS-7 cells were plated at a density of 1.0×10^6 cells/100 mm plate 24 hours prior to transfection. For transfection, the cells were cultured in serum-free DMEM containing 400 μ g/ml of DEAE-dextran, 1 μ M chloroquine, and 2 mM glutamine, and 1 μ g of the appropriate DNA for 3–4 hours at 37° C. in an atmosphere of 5% CO₂. The transfection media was aspirated and replaced with phosphate-buffered saline with 10% DMSO for 2–3 min. Following this DMSO "shock", the COS-7 cells were placed into DMEM with 10% FBS, 1% each of penicillin and streptomycin, and 2 mM glutamine for 48 hours

Because the TIE-2 ligand is secreted it was necessary to permeabilize the cells to detect binding of the receptorbody probe to the ligand. Transfected COS-7 cells were plated at a density of 1.0×10^6 cells/100 mm plate. The cells were rinsed with PBS and then incubated with PBS containing 1.8% formaldehyde for 15-30 min. at room temperature. Cells were then washed with PBS and incubated for 15 min. with PBS containing 0.1% Triton X-100 and 10% Bovine Calf Serum to permeabilize the cells and block non-specific binding sites. The screening was conducted by direct localization of staining using a TIE-2 receptorbody, which consisted of the extracellular domain of TIE-2 fused to the IgG1 constant region. This receptorbody was prepared as set forth in Example 2. Transfected COS cells were probed by incubating them for 30 min with TIE-2 receptorbody. The cells were then washed twice with PBS, fixed with methanol, and then incubated for an additional 30 min with PBS/10% Bovine Calf Serum/anti-human IgG-alkaline phosphatase conjugate. After three PBS washes, cells were incubated in alkaline-phosphatase substrate for 30-60 min. The dish was then inspected microscopically for the presence of stained cells. Cells expressing one orientation of the clone, but not the other orientation, were seen to bind the TIE-2 receptorbody.

One of skill in the art will readily see that the described methods may be used to further identify other related members of the TIE ligand family.

The coding region from the clone pBluescript KS encoding human TIE-2 ligand 2 was sequenced using the ABI 373A DNA sequencer and Taq Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, Inc., Foster City. Calif.). The nucleotide and deduced amino acid sequence of human TIE-2 ligand from the clone pBluescript KS encoding human TIE-2 ligand 2 is shown in FIGS. 6A–6D (SEQ ID NO: 5 and SEQ ID NO: 6).

EXAMPLE 9

TIE-2 Ligand 2 is a Receptor Antagonist

Conditioned media from COS cells expressing either TIE-2 ligand 2 (TL2) or TIE-2 ligand 1 (TL1) were compared for their ability to activate TIE-2 receptors naturally present in human endothelial cell lines.

Lipofectamine reagent (GIBCO-BRL, Inc.) and recommended protocols were used to transfect COS-7 cells with either the pJFE14 expression vector alone, pJFE14 vector containing the human TIE-2 ligand 1 cDNA, or with a pMT21 expression vector (Kaufman, R. J., 1985, Proc. Natl. Acad. Sci. USA 82: 689–693) containing the human TIE-2 ligand 2 cDNA. COS media containing secreted ligands were harvested after three days and concentrated 20-fold by diafiltration (DIAFLO ultrafiltration membranes, Amicon,

Inc.). The quantity of active TIE-2 ligand 1 and TIE-2 ligand 2 present in these media was determined and expressed as the amount (in resonance units, R.U.) of TIE-2 receptor specific binding activity measured by a BIAcore binding assay.

Northern (RNA) analyses revealed significant levels of TIE-2 transcripts in HAEC (Human Aortic Endothelial Cell) human primary endothelial cells (Clonetics, Inc.). Therefore, these cells were used to examine whether TIE-2 receptor is tyrosine-phosphorylated when exposed to COS media con- $_{10}$ taining the TIE-2 ligands. HAEC cells were maintained in a complete endothelial cell growth medium (Clonetics, Inc.) that contained 5% fetal bovine serum, soluble bovine brain extract, 10 ng/ml human EGF, 1 mg/ml hydrocortisone, 50 mg/ml gentamicin and 50 ng/ml amphotericin-B. Assessment of whether TL1 and TL2 could activate TIE-2 receptor in the HAEC cells was done as follows. Semi-confluent HAEC cells were serum-starved for two hours in highglucose Dulbecco's MEM with added L-glutamine and penicillin-streptomycin at 37° C. followed by replacement 20 of the starvation medium with ligand-containing conditioned COS media for 7 minutes at 37° C. in a 5% CO2 incubator. The cells were subsequently lysed and TIE-2 receptor protein was recovered by immunoprecipitation of the lysates with TIE-2 peptide antiserum, followed by Western blotting with antiphosphotyrosine antiserum, exactly as described in example 1. The results are shown in FIG. 7. Phosphotyrosine levels on the TIE-2 receptor (TIE-2-R) were induced by treatment of HEAC cells with TIE-2 ligand 1 (Lane L1) but not by TIE-2 ligand 2 (Lane L2) conditioned 30 COS media. MOCK is conditioned media from COS transfected with JFE14 empty vector.

Evidence that both TL1 and TL2 specifically bind to the TIE-2 receptor was demonstrated by using a BIAcore to assay the TIE-2 receptor specific binding activities in trans-35 fected COS media and by immunostaining of TL1- and TL2-expressing COS cells with TIE-2 receptorbodies.

Because TL2 did not activate the TIE-2 receptor, applicants set out to determine whether TL2 might be capable of serving as an antagonist of TL1 activity. HAEC phospho- 40 rylation assays were performed in which cells were first incubated with an "excess" of TL2, followed by addition of dilute TL1. It was reasoned that prior occupancy of TIE-2 receptor due to high levels of TL2 might prevent subsequent stimulation of the receptor following exposure to TL1 45 present at a limiting concentration.

Semi-confluent HAEC cells were serum-starved as described above and then incubated for 3 min., at 37° C. with 1-2 ml. of 20× COS/JFE14-TL2 conditioned medium. Control plates were treated with 20x COS/JFE14-only 50 medium (MOCK). The plates were removed from the incubator and various dilutions of COS/JFE14-TL1 medium were then added, followed by further incubation of the plates for 5-7 min. at 37° C. Cells were subsequently rinsed, lysed was examined by receptor immunoprecipitation and Western blotting, as described above. TL1 dilutions were made using 20× COS/JFE14-TL1 medium diluted to 2×, 0.5×, 0.1×, or 0.02× by addition of 20× COS/JFE14-alone medium. An assay of the initial 20x TL1 and 20x TL2 COS media using 60 BIAcore biosensor technology indicated that they contained similar amounts of TIE-2-specific binding activities, i.e., 445 R.U. and 511 R.U. for TL1 and TL2, respectively. The results of the antiphosphotyrosine Western blot, shown in FIG. 8, indicate that when compared to prior treatment of 65 HAEC cells with MOCK medium (lane 1), prior treatment of HAEC cells with excess TIE-2 ligand 2 (lane 2) antago-

nizes the subsequent ability of dilute TIE-2 ligand 1 to activate the TIE-2 receptor (TIE-2-R).

The ability of TL2 to competitively inhibit TL1 activation of the TIE-2-R was further demonstrated using the human cell hybrid line, EA.hy926 (see Example 21 for detailed description of this cell line and its maintenance). Experiments were performed in which unconcentrated COS cell media containing TL1 were mixed at varying dilutions with either MOCK- or TL2-conditioned media and placed on serum-starved EA.hy926 cell monolayers for 5 minutes at 37° C. The media were then removed, the cells were harvested by lysis and TIE-2-specific tyrosine phosphorylation was examined by Western blots, as described above. FIG. 9 shows an experiment which contains three groups of treatments, as viewed from left to right. As shown in the four lanes at the left, treatment of the EA.hy926 cells with 1× COS-TL1 alone robustly activated the endogenous TIE-2-R in these cells, whereas 1x TL2 COS medium was inactive. However, mixture of TL1 with either MOCK or TL2 demonstrated that TL2 can block the activity of TL1 in a dose-dependent fashion. In the central three pairs of lanes the ratio of TL2 (or MOCK) was decreased while the amount of TL1 in the mixture was correspondingly increased from 0.1x to 0.3x. At any of these mixture ratios the TL1:TL2 lanes showed a reduced level of TIE-2-R phosphorylation compared to that of the corresponding TL1:MOCK lanes. When the amount TL1 was held steady and the amount of TL2 (or MOCK) was decreased, however (shown in the three pairs of lanes at the right), a point was reached at which the TL2 in the sample was too dilute to effectively inhibit TL1 activity. The relative amount of each ligand present in these conditioned COS media could be estimated from their binding units as measured by the BIAcore assay and from Western blots of the COS media with ligand-specific antibodies. Consequently, we can infer that only a few-fold molar excess of TL2 is required to effectively block the activity of TL1 in vitro. This is significant because we have observed distinct examples in vivo (see Example 17 and FIG. 16) where TL2 mRNAs achieve considerable abundance relative to those of TL1. Thus, TL2 may be serving an important physiological role in effectively blocking signaling by the TIE-2-R at these sites.

Taken together these data confirm that, unlike TL1, TL2 is unable to stimulate endogenously expressed TIE-2-R on endothelial cells. Furthermore, at a few fold molar excess TL2 can block TL1 stimulation of the TIE-2 receptor, indicating that TL2 is a naturally occurring TIE-2 receptor antagonist.

EXAMPLE 10

Identification of TIE-2-Specific Binding Activity in Conditioned Medium and COS Cell Supernatants

Binding activity of 10x CCM from the cell lines C2C12and TIE-2-specific tyrosine phosphorylation in the lysates 55 ras, Rat2 ras, SHEP, and T98G, or COS cell supernatants after transfection with either human TIE-2 ligand 1 (hTL1) or human TIE-2 ligand 2 (hTL2) was measured using biosensor technology (BIAcore; Pharmacia Biosensor, Piscataway, N.J.) which monitors biomolecular interactions in real-time via surface plasmon resonance (SPR). Purified rat or human TIE-2 RB was covalently coupled through primary amines to the carboxymethyl dextran layer of a CM5 research grade sensor chip (Pharmacia Biosensor; Piscataway, N.J.). The sensor chip surface was activated using a mixture of N-hydroxysuccinimide (NHS) and N-ethyl-N'-(3-dimethylaminopropyl)carbodiimide (EDC), followed by immobilization of TIE-2 RB (25 μ g/mL, pH

4.5) and deactivation of unreacted sites with 1.0 M ethanolamine (pH 8.5). In general, 9000–10000 RU of each receptorbody was coupled to the sensor chip.

The running buffer used in the system was HBS (10 mM Hepes, 150 mM NaCl, 0.005% P20 surfactant, pH 7.4). The samples were centrifuged for 15 min at 4° C. and further clarified using a sterile, low protein-binding 0.45 μ m filter (Millipore; Bedford, Mass.). Dextran (2 mg/ml) and P20 surfactant (0.005%) were added to each sample. Aliquots of 40 μ L were injected across the immobilized surface (either rat or human TIE-2) at a flow rate of 5 μ L/min and the receptor binding was monitored for 8 min. The binding activity (resonance units, RU) was measured as the difference between a baseline value determined 30 s prior to the sample injection and a measurement taken at 30 s postinjection. Regeneration of the surface was accomplished with one 15- μ L pulse of 3 M MgCl₂.

The CCM samples (C2C12-ras, Rat2-ras, SHEP, T98G) were tested on the rat TIE-2 RB immobilized surface, while the recombinant hTL1 and hTL2 were tested on the human TIE-2 RB immobilized surface. In each case, specific binding to the TIE-2 receptorbody was evaluated by incubating the samples with 25 μ g/ml of either soluble TIE-2 (rat or human) RB or trkB RB prior to assaying the binding activity. As shown in FIGS. **10A–10D** and FIGS. **11A–11B**, the addition of soluble trkB RB causes a slight decrease in the TIE-2 binding activity, while the addition of soluble TIE-2 RB significantly reduces the binding activity as compared to that measured in the absence of TIE-2 RB.

EXAMPLE 11

TIE-2 RB Specifically Blocks Activation of the TIE-2 Receptor by TIE-2 Ligand 1

The applicants sought to determine whether soluble TIE-2 RB can serve as a competitive inhibitor to block activation of TIE-2 receptor by TIE-2 ligand 1 (TL1). To do this, TL1-containing COS media were preincubated with either TIE-2- or TrkB-RB and then compared for their ability to activate TIE-2 receptors naturally present in a human endothelial cell line.

Conditioned COS media were generated from COS-7 cells transfected with either the pJFE14 expression vector alone (MOCK), or pJFE14 vector containing the human 45 TIE-2 ligand 1 cDNA (TL1) and harvested as described in Example 9 hereinabove, with the exception that the media were sterile filtered but not concentrated. The quantity of TL1 was determined and expressed as the amount (in resonance units, R.U.) of TIE-2 receptor-specific binding 50 activity measured by BIAcore binding assay.

Northern (RNA) analyses revealed significant levels of tie-2 transcripts in HUVEC (Human Umbilical Vein Endothelial Cell) human primary endothelial cells (Clonetics, Inc.). Therefore, these cells were used to examine whether 55 TIE-2 receptor can be tyrosine-phosphorylated when exposed in the presence of TIE-2- or TrkB-RBs to COS media containing TL1. HUVEC cells were maintained at 37° C., 5% CO₂ in a complete endothelial cell growth medium (Clonetics, Inc.) that contained 5% fetal bovine serum, 60 soluble bovine brain extract with $10 \,\mu\text{g/ml}$ heparin, $10 \,\text{ng/ml}$ human EGF, 1 ug/ml hydrocortisone, 50 µg/ml gentamicin and 50 ng/ml amphotericin-B. Assessment of whether TL1 could activate TIE-2 receptor in the HUVEC cells was done as follows. Confluent dishes of HUVEC cells were serum- 65 starved for two-to-four hours in low-glucose Dulbecco's MEM at 37° C., 5% $\rm CO_2$, followed by 10 minute incubation

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in starvation medium that included 0.1 mM sodium orthovanadate, a potent inhibitor of phosphotyrosine phosphatases. Meanwhile, conditioned COS media were preincubated 30 min. at room temperature with either TIE-2- or TrkB-RB added to 50 μ g/ml. The starvation medium was then removed from the HUVEC dishes and incubated with the RB-containing COS media for 7 minutes at 37° C. HUVEC cells were subsequently lysed and TIE-2 receptor protein was recovered by immunoprecipitation with TIE-2 peptide antiserum, followed by Western blotting with an anti-phosphotyrosine antibody, as described in Example 1.

The results are shown in FIG. 12. Phosphotyrosine levels on the TIE-2 receptor were induced by treatment of HUVEC cells with TIE-2 ligand 1 (TL1) relative to that seen with control medium (MOCK) and this induction is specifically blocked by prior incubation with TIE-2-RB (TIE-2-Fc) but not by incubation with TrkB-RB (TrkB-Fc). These data indicate that soluble TIE-2 RB can serve as a selective inhibitor to block activation of TIE-2 receptor by TIE-2 ligand 1.

EXAMPLE 12

Construction of TIE-2 Ligandbodies

An expression construct was created that would yield a secreted protein consisting of the entire coding sequence of human TIE-2 ligand 1 (TL1) or TIE-2 ligand 2 (TL2) fused to the human immunoglobulin gamma-1 constant region (IgG1 Fc). These fusion proteins are called TIE-2 "ligandbodies" (TL1-Fc or TL2-Fc). The Fc portion of TL1-Fc and TL2-Fc was prepared as follows. ADNA fragment encoding the Fc portion of human IgG1 that spans from the hinge region to the carboxy-terminus of the protein, was amplified from human placental cDNA by PCR with oligonucleotides corresponding to the published sequence of human IgG1; the resulting DNA fragment was cloned in a plasmid vector. Appropriate DNA restriction fragments from a plasmid encoding full-length TL1 or TL2 and from the human IgG1 Fc plasmid were ligated on either side of a short PCRderived fragment that was designed so as to fuse, in-frame, TL1 or TL2 with human IgG1 Fc protein-coding sequences.

Milligram quantities of TL2-Fc were obtained by cloning the TL2-Fc DNA fragment into the pVL1393 baculovirus vector and subsequently infecting the Spodoptera frugiperda SF-21AE insect cell line. Alternatively, the cell line SF-9 (ATCC Accession No. CRL-1711) or the cell line BTI-TN-5b1-4 may be used. DNA encoding the TL2-Fc was cloned as an Eco RI-NotI fragment into the baculovirus transfer plasmid pVL1393. Plasmid DNA was recombined into viral DNA by mixing 3 μ g of plasmid DNA with 0.5 μ g of Baculo-Gold DNA (Pharminigen), followed by introduction into liposomes using 30 µg Lipofectin (GIBCO-BRL). DNA-liposome mixtures were added to SF-21AE cells (2×106 cells/60 mm dish) in TMN-FH medium (Modified Grace's Insect Cell Medium (GIBCO-BRL) for 5 hours at 27° C., followed by incubation at 27° C. for 5 days in TMN-FH medium supplemented with 5% fetal calf serum. Tissue culture medium was harvested for plaque purification of recombinant viruses, which was carried out using methods previously described (O'Reilly, D. R., L. K. Miller, and V. A. Luckow, Baculovirus Expression Vectors—A Laboratory Manual. 1992, New York: W. H. Freeman) except that the agarose overlay contained 125 mg/mL X-gal (5-bromo-4-chloro-3-indolyl-b-D-galactopyranoside; GIBCO-BRL). After 5 days of incubation at 27° C., non-recombinant plaques were scored by positive chromogenic reaction to the X-gal substrate, and their positions marked. Recombinant

plaques were then visualized by addition of a second overlay containing 100 mg/mL MTT (3-[4,5-dimethylthiazol-2-yl] 2,5,diphenyltetrazolium bromide; Sigma). Putative recombinant virus plaques were picked by plug aspiration, and purified by multiple rounds of plaque isolation to assure 5 homogeneity. Virus stocks were generated by serial, low-multiplicity passage of plaque-purified virus. Low passage stocks of one virus clone (vTL2-Fc Clone #7) were produced

SF-21AE cells were cultured in serum-free medium (SF- 10 900 II, Gibco BRL) containing 1× antibiotic/antimycotic solution (Gibco BRL) and 25 mg/L Gentamycin (Gibco BRL). Pluronic F-68 was added as a surfactant to a final concentration of 1 g/L. Cultures (4 L) were raised in a bioreactor (Artisan Cell Station System) for at least three days prior to infection. Cells were grown at 27° C., with gassing to 50% dissolved oxygen, at a gas flow rate of 80 mL/min (aeration at a sparge ring). Agitation was by means of a marine impeller at a rate of 100 rpm. Cells were harvested in mid-logarithmic growth phase (~2×10 6 cells/ 20 mL), concentrated by centrifugation, and infected with 5 plaque forming units of vTL2-Fc per cell. Cells and inoculum were brought to 400 mL with fresh medium, and virus was adsorbed for 2 hours at 27° C. in a spinner flask. The culture was then resuspended in a final volume of 8 L with $\,^{25}$ fresh serum-free medium, and the cells incubated in the bioreactor using the previously described conditions.

Culture medium from vTL2-Fc-infected SF21AE cells were collected by centrifugation ($500\times g$, 10 minutes) at 72 hours post-infection. Cell supernatants were brought to pH 8 with NaOH. EDTA was added to a final concentration of 10 mM and the supernatant pH was readjusted to 8. Supernatants were filtered ($0.45~\mu m$, Millipore) and loaded on a protein A column (protein A sepharose 4 fast flow or HiTrap protein A, both from Pharmacia). The column was washed with PBS containing 0.5~M NaCl until the absorbance at 280 nm decreased to baseline. The column was washed in PBS and eluted with 0.5~M acetic acid. Column fractions were immediately neutralized by eluting into tubes containing 1 M Tris pH 9. The peak fractions containing the TL2-Fc were pooled and dialyzed versus PBS.

EXAMPLE 13

Expression of TIE-1, TIE-2, TL1, and TL2 in Renal Cell Carcinoma

In situ hybridization experiments were performed on human renal cell carcinoma tumor tissue using TIE-1, TIE-2, TL1, and TL2 cDNA probes. TIE-2, TIE-1, TL1, and TL2 50 expression were all up-regulated in the tumor vasculature. Ligand expression appeared to be localized to either the vascular endothelial cells (TL2) or very near the vascular endothelial cells in the mesenchyme (TL1). VEGF has been shown to be dramatically up-regulated in this tumor tissue. 55 Brown, et al. Am. J. Pathol. 143:1255–1262 (1993).

EXAMPLE 14

Expression of TIE-1, TIE-2, TL1, and TL2 in Wound Healing

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In situ hybridization experiments were performed on cross-sectional tissue slices obtained from a rat cutaneous wound model using TIE-1, TIE-2, TL1, and TL2 cDNA probes. The wound healing model involves pressing a small 65 cork bore against the skin of a rat and removing a small, cylindrical plug of skin. As healing begins at the base of the

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wound, a vertical slice of tissue is taken and used for in situ hybridization. In the tested tissue sample, TL1 and TL2 appeared to be slightly up-regulated by four days postinjury. In contrast to the slightly up-regulated expression of TL1 and TL2 in this tissue, VEGF expression, which may precede TL1 and TL2 expression, is dramatically up-regulated.

EXAMPLE 15

Expression of TIE Ligands in Fetal Liver and Thymus

Reverse transcription-PCR (RT-PCR) was performed on mouse E14.5 fetal liver and mouse E17.5 fetal thymus. Agarose gel electrophoresis of the RT-PCR products revealed that in the mouse fetal liver, TIE-2 ligand 1 (TL1) RNA is enriched in the stromal region, but is absent in c-kit+TER119 hematopoietic precursor cells. In this same tissue, TIE-2 ligand 2 (TL2) RNA is enriched in the stromal cells, but absent in the hematopoietic precursor cells (FIG. 13). In the mouse fetal thymus, TL2 is enriched in the stromal cells (FIG. 14).

EXAMPLE 16

The TIE Receptor/Ligand System in Angiogenesis

Although the TIE-2/TIE ligand system appears to play an important role in endothelial cell biology, it has not been shown to play a significant, active role in the early to intermediate stages of vascularization (f angioblast or endothelial cell proliferation and migration, tubule formation, and other early stage events in vascular modeling). In contrast to the receptors and factors known to mediate these aspects of vascular development, the temporally late pattern of expression of TIE-2 and TL1 in the course of vascularization suggests that this system plays a distinct role in the latter stages vascular development, including the structural and functional differentiation and stabilization of new blood vessels. The pattern of expression of TIE-2/TL1 also is consistent with a continuing role in the maintenance of the structural integrity and/or physiological characteristics of an established vasculature.

TIE Ligand 2 (TL2) appears to be a competitive inhibitor of TL1. The spatiotemporal characteristics of TL2 expression suggest that this single inhibitory molecule may play multiple, context-dependent roles essential to appropriate vascular development or remodeling (e.g. de-stabilization/ de-differentiation of mature endothelial cells allowing the formation of new vessels from existing vasculature, inhibition of inappropriate blood vessels formation, and regression/ involution of mature blood vessels). FIG. 15 is a schematic representation of the hypothesized role of the TIE-2/TIE ligands in angiogenesis. In this figure TL1 is represented by (*), TL2 is represented by (*), TIE-2 is represented by (T), VEGF is represented by ([]), and flk-1 (a VEGF receptor) is represented by (Y).

EXAMPLE 17

Expression of TIE Ligands in the Female Reproductive System: Expression in the Ovary

Preliminary observations made in experiments examining the expression of the TIE receptors and ligands in the female reproductive system are consistent with the hypothesis the TL1 plays a role in neovascularization which temporally follows that of VEGF. The pattern of TL2 expression is also

consistent with an antagonism of the action of TL1, and a specific role in vascular regression. To verify this, expression of relevant mRNAs can be examined following experimental induction of follicular and luteal development so that their temporal relation to various aspects of 5 neovascularization/vascular regression can be more clearly defined (e g in conjunction with endothelial cell staining, vascular fills). Angiogenesis associated with follicular development and corpus luteum formation in staged ovaries of mature, female rats or following induced ovulation in 10 pre-pubertal animals was followed using in situ hybridization. FIG. 16 contains photographs of in situ hybridization slides showing the temporal expression pattern of TIE-2, TL1, TL2, and VEGF during the ovarian cycle [Column 1: Early pre-ovulatory follicle; Column 2: pre-ovulatory fol- 15 licle; Column 3: early corpus luteum; and Column 4: atretic follicle; Row A:bright field; Row B:VEGF; Row C: TL2;

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Row D: TL1 and Row E: TIE-2 receptor]. These studies revealed that VEGF, TL1 and TL2 are expressed in a temporally and spatially coordinate fashion with respect to ²⁰ the development and regression of vasculature in the ovary, specifically with respect to the establishment of the vascular system which is generated in the course of the conversion of an ovarian follicle to a corpus luteum (CL).

Briefly, VEGF expression increases in the follicular granule layer prior to its vascularization during the process of luteinization. During the process of CL formation, highest levels of VEGF expression are apparent in the center of the developing CL in the vicinity of luteinizing cells which are not yet vascularized. VEGF levels remain moderately high and are diffusely distributed in the developed CL. In contrast, noticeably enhanced expression of TIE-2 ligand 1 occurs only late in process of CL formation, after a primary vascular plexus has been established. Later, TL1 expression is apparent throughout the CL at which time the definitive capillary network of the CL has been established.

TL2 exhibits a more complex pattern of expression than either VEGF or TL1. In the developing CL, TL2 is expressed at highest levels at the front of the developing capillary plexus between the central avascular region of the CL where VEGF expression is highest, and the most peripheral portion of the CL where TL1 expression is dominant and where the luteinization process is complete and the vascular system is most mature. TL2 also appears to be expressed at high levels in the follicular layer of large follicles which are undergoing atresia. While TL1 is also apparent in atretic follicles, VEGF is not expressed.

The pattern of expression described above is most consistent with a role for VEGF in the initiation of angiogenesis, with TL1 acting late in this process-for example in modeling and/or stabilization of the definitive vascular network. In contrast, TL2 is present both in areas of active expansion of a newly forming vascular network (during CL formation), and in regions which fail to establish a new vasculature and vascular regression is in progress (atretic follicles). This suggests a more dynamic and complex role for TL2, possibly involving destabilization of existing vasculature (necessary for regression) or developing vasculature (necessary for the dynamic modeling of newly forming vessels).

EXAMPLE 18

A Receptorbody Binding Assay and a Ligand Binding and Competition Assay

A quantitative cell-free binding assay with two alternate formats has been developed for detecting either TIE-2

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receptorbody binding or ligand binding and competition. In the receptorbody binding version of the assay, TIE-2 ligands (purified or partially purified; either TL1 or TL2) are coated onto an ELISA plate. Receptorbody at varying concentrations is then added, which binds to the immobilized ligand in a dose-dependent manner. At the end of 2 hours, excess receptorbody is washed away, then the amount bound to the plate is reported using a specific anti-human Fc antibody which is alkaline phosphatase tagged. Excess reporter antibody is washed away, then the AP reaction is developed using a colored substrate. The assay is quantitated using a spectrophotometer. FIG. 19 shows a typical TIE-2-IgG binding curve. This assay has been used to evaluate the integrity of TIE-2-IgG after injection into rats and mice. The assay can also be used in this format as a ligand competition assay, in which purified or partially-purified TIE ligands compete with immobilized ligand for receptorbody. In the ligand binding and competition version of the binding assay, TIE-2 ectodomain is coated onto the ELISA plate. The Fc-tagged fibrinogen-like domain fragments of the TIE ligands (TL1-fFc and TL2-fFc) then bind to the ectodomain, and can be detected using the same anti-human Fc antibody as described above. FIG. 20 shows an example of TL1-fFc binding to TIE-2 ectodomain. This version of the assay can also be used to quantitate levels of TL1-fFc in serum or other samples. If untagged ligand (again, either purified or unpurified) is added at the same time as the TL1-fFc, then a competition is set up between tagged ligand fragment and full-length ligand. The full-length ligand can displace the Fc-tagged fragment, and a competition curve is generated.

EXAMPLE 19

EA.hy926 Cell Line can be Used as a Reporter Cell Line for TIE Ligand Activity

EA.hy926 is a cell hybrid line that was established by fusion of HUVEC with the human lung carcinoma-derived line, A549 [Edgell, et al. Proc. Natl. Acad. Sci. (USA) 80, 3734–3737 (1983). EA.hy926 cells have been found to express significant levels of TIE-2 receptor protein with low basal phosphotyrosine levels. The density at which EA.hy926 cells are passaged prior to their use for receptor assays, as well as their degree of confluency at the time of assay, can affect TIE-2 receptor abundance and relative inducibility in response to treatment with ligand. By adopting the following regimen for growing these cells the EA.hy926 cell line can be used as a dependable system for assay of TIE-2 ligand activities.

EA.hy926 cells are seeded at 1.5×10⁶ cells in T-75 flasks (Falconware) and re-fed every other day with high-glucose Dulbecco's MEM, 10% fetal bovine serum, L-glutamine, penicillin-streptomycin, and 1× hypoxanthine-aminopterinthymidine (HAT, Gibco/BRL). After three to four days of growth, the cells are passaged once again at 1.5×10^6 cells per T-75 flask and cultured an additional three to four days. For phosphorylation assays, cells prepared as described above were serum-starved by replacement of the culture medium with high-glucose DMEM and incubation for 2-3 hours at 37° C. This medium was aspirated from the flask and samples of conditioned media or purified ligand were added to the flask in a total volume of 1.5 ml followed by incubation at 37° C. for 5 minutes. Flasks were removed from the incubator and placed on a bed of ice. The medium was removed and replaced with 1.25 ml Lysis Buffer containing 1% nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS in 20 mM Tris, pH 7.6, 150 mM NaCl, 50 mM NaF, 1 mM sodium orthovanadate, 5 mM benzamidine, and 1 mM

EDTA containing the protease inhibitors PMSF, aprotinin, and leupeptin. After 10 minutes on ice to allow membrane solubilization, plates were scraped and cell lysates were clarified by microcentrifugation at top speed for 10 minutes at 4° C. TIE-2 receptor was immunoprecipitated from the clarified supernatant by incubation in the cold with an anti-TIE-2 polyclonal antiserum and Protein G-conjugated Sepharose beads. The beads were washed three times with cold cell lysis buffer and boiled 5 minutes in Laemmli sample buffer, which was then loaded on 7.5% SDS- 10 polyacrylamide gels. Resolved proteins were electrotransferred to PVDF (Lamblia-P) membrane and then subjected to Western blot analysis using anti-phosphotyrosine antibody and the ECL reagent. Subsequent comparison of total TIE-2 protein levels on the same blots was done by stripping 15 the anti-phosphotyrosine antibody and reincubating with a polyclonal antiserum specific to the ectodomain of TIE-2.

EXAMPLE 20

Isolation and Sequencing of Full Length cDNA Clone Encoding Mammalian TIE Ligand-3

TIE ligand-3 (TL3) was cloned from a mouse BAC genomic library (Research Genetics) by hybridizing library duplicates, with either mouse TL1 or mouse TL2 probes 25 corresponding to the entire coding sequence of those genes. Each copy of the library was hybridized using phosphate buffer at 55° C. overnight. After hybridization, the filters were washed using 2×SSC, 0.1% SDS at 60° C., followed by exposure of X ray film to the filters. Strong hybridization signals were identified corresponding to mouse TL1 and mouse TL2. In addition, signals were identified which weakly hybridized to both mouse TL1 and mouse TL2. DNA corresponding to these clones was purified, then digested with restriction enzymes, and two fragments which hybridized to the original probes were subcloned into a bacterial plasmid and sequenced. The sequence of the fragments contained two exons with homology to both mouse TL1 and mouse TL2. Primers specific for these sequences were used as PCR primers to identify tissues containing transcripts corresponding to TL3. A PCR band corresponding to TL3 was identified in a mouse uterus cDNA library in lambda gt-11. (Clontech Laboratories, Inc., Palo Alto, Calif.).

Plaques were plated at a density of 1.25×10⁶/20×20 cm plate and replica filters taken following standard procedures 45 (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., page 8.46, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.). Duplicate filters were screened at "normal" stringency (2×SSC, 65° C.) with a 200 bp PCR radioactive probe made to the mouse TL3 sequence. Hybridization was at 65° C. in a solution containing 0.5 mg/ml salmon sperm DNA. Filters were washed in 2×SSC at 65° C. and exposed for 6 hours to X-ray film. Two positive clones that hybridized in duplicate were picked. EcoRI digestion of phage DNA obtained from these clones indicated two independent clones with insert sizes of approximately 1.2 kb and approximately 2.2 kb. The 2.2 kb EcoRI insert was subcloned into the EcoRI site of pBluescript KS (Stratagene). Sequence analysis showed that the longer clone was lacking an initiator methionine and signal peptide but otherwise 60 encoded a probe homologous to both mouse TL1 and mouse TL2.

Two TL3-specific PCR primers were then synthesised as follows: US2: cctctgggctcgccagtttgttagg (SEQ ID NO: 29) US1: ccagctggcagatatcagg (SEQ ID NO: 30)

The following PCR reactions were performed using expression libraries derived from the mouse cell lines

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C2C12ras and MG87. In the primary PCR reaction, the specific primer US2 was used in conjunction with vectorspecific oligos to allow amplification in either orientation. PCR was in a total volume of 100 ml using 35 cycles of 94° C., 1 mm; 42° C. or 48° C. for 1 mm; 72° C, 1 mm. The secondary PCR reaction included the second specific primer, US1, which is contained within the primary PCR product, in conjunction with the same vector oligos. The secondary reactions were for 30 cycles, using the same temperatures and times as previous. PCR products were gel isolated and submitted for sequence analysis. On the basis of sequences obtained from a total of four independent PCR reactions using two different cDNA libraries, the 5' end of the TL3 sequence was deduced. Northern analysis revealed moderate to low levels of mouse TL3 transcript in mouse placenta. The expression of mouse TL3 consisted of a transcript of approximately 3 kb. The full length TL3 coding sequence is set forth in FIGS. 21A-21C (SEQ ID NO: 9 and SEQ ID NO: 10).

The mouse TL3 sequence may then be used to obtain a human clone containing the coding sequence of human TL3 by hybridizing either a human genomic or cDNA library with a probe corresponding to mouse TL3 as has been described previously, for example, in Example 8 supra.

EXAMPLE 21

Isolation of Full Length Genomic Clone Encoding Human TIE Ligand-4

TIE ligand-4 (TL4) was cloned from a mouse BAC genomic library (BAC HUMAN (II), Genome Systems Inc.) by hybridizing library duplicates, with either a human TL1 radioactive probe corresponding to the entire fibrinogen coding sequence of TL1 (nucleotides 1153 to 1806 of FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2]) or a mouse TL3 radioactive probe corresponding to a segment of 186 nucleotides from the fibrinogen region of mouse TL3 (nucleotides 1307 to 1492 of FIGS. 21A-21C [SEQ ID NO: 9 and SEQ ID NO: 10]). Each probe was labeled by PCR using exact oligonucleotides and standard PCR conditions, except that dCTP was replaced by P³²dCTP. The PCR mixture was then passed through a gel filtration column to separate the probe from free P³² dCTP. Each copy of the library was hybridized using phosphate buffer, and radioactive probe at 55° C. overnight using standard hybridization conditions. After hybridization, the filters were washed using 2×SSC, 0.1% SDS at 55° C., followed by exposure of X ray film. Strong hybridization signals were observed corresponding to human TL1. In addition, signals were identified which weakly hybridized to both human TL1 and mouse TL3. DNA corresponding to these clones was purified using standard procedures, then digested with restriction enzymes, and one fragment which hybridized to the original probes was subcloned into a bacterial plasmid and sequenced. The sequence of the fragments contained one exon with homology to both human TL1 and mouse TL3 and other members of the TIE ligand family. Primers specific for these sequences may be used as PCR primers to identify tissues containing transcripts corresponding to TL4.

The complete sequence of human TL4 may be obtained by sequencing the full BAC clone contained in the deposited bacterial cells. Exons may be identified by homology to known members of the TIE-ligand family such as TL1, TL2 and TL3. The full coding sequence of TL4 may then be determined by splicing together the exons from the TL4 genomic clone which, in turn, may be used to produce the TL4 protein. Alternatively, the exons may be used as probes

to obtain a full length cDNA clone, which may then be used to produce the TL4 protein. Exons may also be identified from the BAC clone sequence by homology to protein domains such as fibrinogen domains, coiled coil domains, or protein signals such as signal peptide sequences. Missing 5 exons from the BAC clone m,ay be obtained by identification of contiguous BAC clones, for example, by using the ends of the deposited BAC clone as probes to screen a human genomic library such as the one used herein, by using the exon sequence contained in the BAC clone to screen a 10 cDNA library, or by performing either 5' or 3' RACE procedure using oligonucleotide primers based on the TL4 exon sequences.

Identification of Additional TIE Ligand Family Members

The novel TIE ligand-4 sequence may be used in a 15 rational search for additional members of the TIE ligand family using an approach that takes advantage of the existence of conserved segments of strong homology between the known family members. For example, an alignment of the amino acid sequences of the TIE ligands shows several 20 regions of conserved sequence (see boxed regions of FIGS.

22A-22B (SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, and SEQ ID NO: 16). Degenerate oligonucleotides essentially based on these boxes in combination with either previously known or novel 25 TIE ligand homology segments may be used to identify new TIE ligands.

The highly conserved regions among TL1, TL2 and TL3 may be used in designing degenerate oligonucleotide primers with which to prime PCR reactions using cDNAs. cDNA 30 templates may be generated by reverse transcription of tissue RNAs using oligo d(T) or other appropriate primers. Aliquots of the PCR reactions may then be subjected to electrophoresis on an agarose gel. Resulting amplified DNA fragments may be cloned by insertion into plasmids, 35 sequenced and the DNA sequences compared with those of all known TIE ligands.

Size-selected amplified DNA fragments from these PCR reactions may be cloned into plasmids, introduced into *E. coli* by electroporation, and transformants plated on selective agar. Bacterial colonies from PCR transformation may be analyzed by sequencing of plasmid DNAs that are purified by standard plasmid procedures.

Cloned fragments containing a segment of a novel TIE ligand may be used as hybridization probes to obtain full length cDNA clones from a cDNA library. For example, the human TL4 genomic sequence may be used to obtain a human cDNA clone containing the complete coding sequence of human TL4 by hybridizing a human cDNA library with a probe corresponding to human TL4 as has been described previously.

To study these differences to obtain full to sufficiently the constructed as follows. 23.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 24.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 24.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 25.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 25.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 25.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows. 25.1. Cysteine substitutions might be contributed biological properties of presence in TL1 of a constructed as follows.

EXAMPLE 22

Cloning of the Full Coding Sequence of hTL4

Both 5' and 3' coding sequence from the genomic human TL-4 clone encoding human TIE ligand-4 (hTL-4 ATCC Accession No. 98095) was obtained by restriction enzyme digestion, Southern blotting and hybridization of the hTL-4 clone to coding sequences from mouse TL3, followed by 60 subcloning and sequencing the hybridizing fragments. Coding sequences corresponding to the N-terminal and C-terminal amino acids of hTL4 were used to design PCR primers (shown below), which in turn were used for PCR amplification of TL4 from human ovary cDNA. A PCR band 65 was identified as corresponding to human TL4 by DNA sequencing using the ABI 373A DNA sequencer and Taq

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Dideoxy Terminator Cycle Sequencing Kit (Applied Biosystems, Inc., Foster City, Calif.). The PCR band was then subcloned into vector pCR-script and several plasmid clones were analyzed by sequencing. The complete human TL4 coding sequence was then compiled and is shown in FIGS. 23A–23C (SEQ ID NO: 17 and SEQ ID NO: 18). In another embodiment of the invention, the nucleotide at position 569 is changed from A to G, resulting in an amino acid change from Q to R.

The PCR primers used as described above were designed as follows: hTL4atg 5'-geatgetatetegagecaccATGCTCTCCCAGCTAGCCA TGCTGCAG-3'(SEQ ID NO: 27)

hTL4not

5'-gtgtcgacgcgccctctagatcagacTTAGATGTCCAAA GGCCGTATCATCAT-3'(SEQ ID NO: 28)

Lowercase letters indicate "tail" sequences added to the PCR primers to facilitate cloning of the amplified PCR fragments.

EXAMPLE 23

Construction and Characterization of Modified TIE Ligands

A genetic analysis of TIE-2 ligand-1 and TIE-2 ligand-2 (TL1 and TL2) was undertaken to gain insight into a number of their observed properties. Although TL1 and TL2 share similar structural homology, they exhibit different physical and biological properties. The most prominent feature that distinguishes the two ligands is that although they both bind to the TIE-2 receptor, TL1 is an agonist while TL2 is an antagonist. Under non-reducing electrophoretic conditions both proteins exhibit covalent, multimeric structures. TL1 is produced as a mixture of disulfide cross-linked multimers, primarily trimers and higher order species, without any dimeric species. But TL2 is produced almost exclusively as a dimeric species. Also, while TL2 is produced well in most expression systems, TL1 is expressed poorly and is difficult to produce in large quantities. Finally, production and purification conditions also appear to predispose TL1 to inactivation by proteolytic cleavage at a site near the amino terminus.

To study these differences, several modified ligands were

23.1. Cysteine substitution—Investigations into what factors might be contributing to the different physical and biological properties of the two molecules revealed the presence in TL1 of a cysteine residue (CYS 265 in FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2]; CYS 245 in FIG. 17 [SEQ ID NO: 7 and SEQ ID NO: 8]) preceding the fibrinogen-like domain in TL1 but absent in TL2—i.e., there was no corresponding cysteine residue in TL2. The CYS265 residue in TL1 is encoded by TGC and is located at about nucleotides 1102-1104 (see FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2]) at the approximate junction between the coiled-coil and fibrinogen-like domains. Because cysteine residues are generally involved in disulfide bond formation, the presence of which can contribute to both the tertiary structure and biological properties of a molecule, it was thought that perhaps the presence of the CYS265 residue in TL1 might be at least partially responsible for the different properties of the two molecules.

To test this hypothesis, an expression plasmid was constructed which contained a mutation in TL1 in which the CYS (residue 265 in FIGS. 4A–4D [SEQ ID NO: 1 and SEQ ID NO: 2]; residue 245 in FIG. 17) was replaced with an

amino acid (serine) which does not form disulfide bonds. In addition to this TL1/CYS mutant, a second expression plasmid was constructed which mutated the approximately corresponding position in TL2 (Met247 in FIG. 17 [SEQ ID NO: 7 and SEQ ID NO: 8]) so that this residue was now a cysteine. Both non-mutated and mutated expression plasmids of TL1 and TL2 were transiently transfected into COS7 cells, cell supernatants containing the recombinant proteins were harvested, and samples were subjected to both reducing and non-reducing SDS/PAGE electrophoresis and subsequent Western blotting.

FIG. 18 shows the Western blots under non-reducing conditions of both non-mutated and mutated TL1 and TL2 proteins, revealing that the TL1/CYS⁻ mutant runs as a dimer much like TL2 and that the TL2/CYS+mutant is able to form a trimer, as well as higher-order multimers, more like TL1. When the two mutant proteins were tested for their ability to induce phosphorylation in TIE-2 expressing cells, the TL1/CYS⁻ mutant was able to activate the TIE-2 receptor, whereas the TL2/CYS⁺ mutant was not.

Thus, when the cysteine residue (residue 265 in FIGS. 4A–4D [SEQ ID NO: 1 and SEQ ID NO: 2]; residue 245 in [SEQ ID NO: 7 and SEQ ID NO: 8]) of TL1 was genetically altered to a serine, it was found that the covalent structure of TL1 became similar to that of TL2, i.e., primarily dimeric. The modified TL1 molecule still behaved as an agonist, thus the trimeric and/or higher order multimeric structure was not the determining factor giving TL1 the ability to activate. Although the removal of the cysteine did make a molecule with more desirable properties, it did not improve the 30 production level of TH1.

23.2. Domain deletions—The nucleotide sequences encoding TL1 and TL2 share a genetic structure that can be divided into three domains, based on the amino acid sequences of the mature proteins. The last approximately 35 215 amino acid residues of each mature protein contains six cysteines and bears strong resemblance to a domain of fibrinogen. This region was thus denoted the "fibrinogenlike" domain or "F-domain." A central region of the mature protein containing approximately 205 residues had a high 40 probability of assuming a "coiled-coil" structure and was denoted the "coiled-coil" domain or "C-domain." The amino-terminal approximately 55 residues of the mature protein contained two cysteines and had a low probability of having a coiled-coil structure. This region was designated 45 the "N-terminal" domain or "N-domain." The modified ligands described herein are designated using a terminology wherein N=N-terminal domain, C=coiled-coil domain, F=fibrinogen-like domain and the numbers 1 and 2 refer to TL1 and TL2 respectively. Thus 1N indicates the N-terminal 50 domain from TL1, 2F indicates the fibrinogen-like domain of TL2, and so forth.

In order to test whether the fibrinogen-like domain (F-domain) of the TIE2 ligands contained TIE-2 activating activity, expression plasmids were constructed which 55 deleted the coiled-coil and N-terminal domains, leaving only that portion of the DNA sequence encoding the F-domain (for TL1, beginning in FIGS. 4A-4D [SEQ ID NO: 1 and SEQ ID NO: 2] at about nucleotide 1159, amino acid residue ARG284; for TL2, corresponding to about nucleotide 1200 in FIGS. 6A-6D [SEQ ID NO: 5 and SEQ ID NO: 6], amino acid residue 282). This mutant construct was then transiently transfected into COS cells. The supernatant containing the recombinant protein was harvested. The TL1/F-domain mutant was tested for its ability to bind the TIE-2 receptor. 65 The results showed that, as a monomer, the TL1/F-domain mutant was not able to bind TIE-2 at a detectable level.

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But when the TL1/F-domain monomer was myc-tagged and subsequently clustered with an antibody directed against the myc tag, it exhibited detectable binding to TIE-2. However, the antibody-clustered TL1/F-domain mutant was not able to induce phosphorylation in a TIE-2 expressing cell line

Thus it was determined that the F-domain of the TIE-2 ligands is involved in binding the receptor but that a truncation consisting of just the F-domain alone is not sufficient for receptor binding. This raised the possibility that the coiled-coil domain was responsible for holding together several fibrinogen-like domains, which might be essential for receptor binding. In an attempt to confirm this hypothesis, the F-domain was fused with the Fc section of human antibody IgG1. Because Fc sections dimerize upon expression by mammalian cells, these recombinant proteins mimicked the theoretical configuration of the F-domains were the native ligands to dimerize. This F-domain-Fc construct bound but failed to activate the receptor. Apparently, multimerization caused by other regions of the ligands is necessary to enable the ligands to bind the TIE receptor. In addition, some other factor outside of the F-domain must contribute to phosphorylation of the recep-

Mutants were then constructed which were missing the fibrinogen-like domain, and therefore contained only the N-terminal and coiled-coil domains. They were not capable of binding to the receptor. To assess the role of the N-terminal domain in receptor binding and activation, the ligands were truncated to just their C- and F-domains and tagged with a FLAG tag at the N-terminus, creating constructs termed FLAG-1C1F and FLAG-2C2F. Although these molecules stained robustly in COS7 cells transfected transiently to express the TIE receptor, they failed to respond in a phosphorylation assay. Thus the N-domain does contain an essential factor for receptor activation although, as disclosed infra, the ability of chimeric molecule 2N2C1F to activate the receptor shows that even the N-domain of an inactive ligand can fill that role.

The differences in behavior between the myc-tagged F-domain truncation and the Fc-tagged F-domain truncation described previously suggested that the TIE ligands can only bind in dimeric or higher multimeric forms. Indeed, non-reducing SDS-PAGE showed that the TIE ligands exist naturally in dimeric, trimeric, and multimeric forms. That the FLAG-1C1F and FLAG-2C2F truncations can bind to the TIE-2 receptor without dimerization by a synthetic tag (such as Fc), whereas the F truncations cannot, suggests that the C-region is at least partly responsible for the aggregation of the F-domains.

23.3. Swapping Constructs (chimeras)

Applicants had noted that the level of production of TL1 in COS7 cells was approximately tenfold lower than production of TL2. Therefore, chimeras of TL1 and TL2 were constructed in an attempt to explain this difference and also to further characterize the agonist activity of TL1 as compared to the antagonist activity of TL2.

Four chimeras were constructed in which either the N-terminal domain or the fibrinogen domain was exchanged between TL1 and TL2 and were designated using the terminology described previously such that, for example, 1N1C2F refers to a chimera having the N-terminal and coiled-coil domains of TL1, together with the fibrinogen-like domain from TL2.

The four chimeras were constructed as follows: chimera 1—1N1C2F

chimera 2-2N2C1F

chimera 3—1N2C2F chimera 4—2N1C1F

The nucleotide and amino acid sequences of chimeras 1–4 are shown in FIGS. **24**A–**24**C (SEQ ID NO: 19 and SEQ ID NO: 20), FIGS. **25**A–**25**C (SEQ ID NO: 21 and SEQ ID NO: 5 22), FIGS. **26**A–**26**C (SEQ ID NO: 23 and SEQ ID NO: 24), and FIGS. **27**A–**27**C (SEQ ID NO: 25 and SEQ ID NO: 26) respectively.

Each chimera was inserted into a separate expression vector pJFE14. The chimeras were then transfected into 10 COS7 cells, along with the empty pJFE14 vector, native TL1, and native TL2 as controls, and the culture supernatants were collected.

In order to determine how the swapping affected the level of expression of the ligands, a 1:5 dilution and a 1:50 15 dilution of the COS7 supernatants were dot-blotted onto nitrocellulose. Three ligands that contained the TL1 N-domain (i.e. native TL1, 1N2C2F and 1N1C2F) were then probed with a rabbit antibody specific to the N-terminus of TL1. Three ligands containing the TL2 N-domain, (i.e. 20 native TL2, 2N1C1F and 2N2C1F) were probed with a rabbit antibody specific for the N-terminus of TL2. The results demonstrated that the COS7 cells were expressing any molecule containing the N-domain of TL2 at roughly ten times the level of any molecule containing the TL1 25 N-domain, regardless of the makeup of the rest of the protein. The conclusion was that the N-domain must principally control the level of expression of the ligand.

The next question addressed was the chimeras' ability or inability to activate the TIE-2 receptor. EAhy926 cells were 30 challenged with the four chimeras, as well as TL1 as a positive control for phosphorylation and TL2 or an empty pJFE14-transfected COS7 cell supernatant as negative controls for phosphorylation. The cells were lysed, and the TIE-2 receptor was immunoprecipitated out of the cell lysate 35 and run on an SDS-PAGE. The samples were Western blotted and probed with an anti-phosphotyrosine antibody to detect any receptors that had been phosphorylated. Surprisingly, only the constructs containing the TL1 fibrinogen-like domain (2N1C1F and 2N2C1F) could phos- 40 phorylate the TIE-2 receptor. Thus, although the N-terminal region of TL1 is essential for activation, it can be replaced by the N-terminal region of TL2, i.e., the information that determines whether the ligand is an agonist or an antagonist is actually contained in the fibrinogen-like domain. Thus it 45 was determined that the F-domain, in addition to binding the TIE-2 receptor, is responsible for the phosphorylation activity of TL1. Further, when TL2, an otherwise inactive molecule, was altered by replacing its F-domain with the TL1 F-domain, the altered TL2 acted as an agonist.

The 2N1C1F construct was somewhat more potent, however. The signal caused by chimera 2N1C1F appeared slightly stronger than that of chimera 2N2C1F, leading to speculation that the C-domain of TL1, though not crucial for phosphorylation, might enhance the potency of TL1. 55 However, since the samples used for the phosphorylation assay were not normalized in terms of the concentration of ligand, it was possible that a stronger phosphorylation signal only indicated the presence of more ligand. The phosphorylation assay was therefore repeated with varying amounts 60 of ligand to determine whether the active chimeras displayed different potencies. The concentration of ligand in the COS7 supernatants of ligand transfections was determined through BIAcore biosenser technology according to methods previously described (Stitt, T. N., et al. (1995) Cell 80: 661-670). 65 BIAcore measured the binding activity of a supernatant to the TIE-2 receptor in arbitrary units called resonance units

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(RU). Fairly good correlation between RU's and ligand concentration has been generally observed, with 400 RU of activity corresponding to about 1 μ g of protein per mL of supernatant. Samples were diluted to concentrations of 100 RU, 20 RU, and 5 RU each and the phosphorylation assay was repeated. The results demonstrated that chimera 2N2C1F was clearly more potent than either the native TL1 or chimera 1N1C2F at the same concentrations.

Another interesting aspect of these exchange constructs is in their levels of expression. Each of the four chimeras was tested for its level of production in COS cells, its ability to bind to TIE2, and its ability to phosphorylate TIE2. The results of these experiments showed that chimeras 1 and 3 were produced at levels comparable to TL1, whereas chimeras 2 and 4 were produced at levels comparable to TL2. Thus a high level of protein production was correlated with the TL2 N-terminal domain. Additionally, when tested on endothelial EAhy926 cells, chimeras 2 and 4 were active, whereas 1 and 3 were not. Thus activity (phosphorylation of the receptor) correlates with the TL1 fibrinogen-like domain. Chimeras 2 and 4 therefore each had the desirable properties of high production levels as well as agonist activity.

23.4. Proteolytic resistant constructs—Based on the observation that a large fraction of TL1 preparations was often proteolytically cleaved near the N-terminus, it was proposed that an arginine residue located at position 49 of the mature protein (see FIG. 17 [SEQ ID NO: 7 and SEQ ID NO: 8]) was a candidate cleavage site that might be involved in the regulation of the protein's activity in vixo, and that replacing the arginine with a serine (R49—>S) might increase the stability of the protein without necessarily affecting its activity. Such a mutant of TL1 was constructed and was found to be about as active as the native TL1 but did not exhibit resistance to proteolytic cleavage.

23.5. Combination mutants—The most potent of the chimeric constructs, 2N1C1F, was additionally altered so that the cysteine encoded by nucleotides 784–786 as shown in FIGS. 27A–27C (SEQ ID NO: 25 and SEQ ID NO: 26) was converted to a serine. This molecule (denoted 2N1C1F (C246S)) was expressed well, potently activated the receptor, was resistant to proteolytic cleavage and was primarily dimeric, rather than higher-order multimeric. Thus the 2N domain appeared to confer protease resistance on the molecule. Finally, this molecule was further altered to eliminate the potentially protease sensitive site encoded by nucleotides 199–201 as shown in FIGS. 27A–27C (SEQ ID NO: 25 and SEQ ID NO: 26), to give a molecule (denoted 2N1C1F (R51->S,C246->S)) which was expected to be activating, well expressed, dimeric, and protease resistant.

Table 1 summarizes the modified TIE-2 ligand constructs that were made and characterizes each of them in terms of ability to bind the TIE-2 receptor, ability to activate the TIE-2 receptor, the type of structure formed (monomer, dimer, etc.) and their relative production levels. Unmodified TL1 (plain) and TL2 (striped) are shown with the three domains as boxes. Thus striped boxes indicate domains from TL2. The cysteine located at position 245 of the mature TL1 protein is indicated by a "C." An "X" through the "C" indicates that that cysteine residue was substituted for by another amino acid as in, for example, the TL1 CYS-mutant. Similarly, an "X" through the "R" in the last construct indicates the substitution for an Arg residue at position 49 of the mature TL1 protein. The "C." is present in one modified TL2 construct showing the TL2 CYS+ mutant. Constructs having Fc tails or flag tagging are also indicated.

Based upon the teachings herein, one of skill in the art can readily see that further constructs may be made in order to

create additional modified and chimeric TIE-2 ligands which have altered properties. For example, one may create a construct comprised of the N-terminal domain of TL2 and the F-domain of TL1 fused with the Fc section of human antibody IgG1. This construct would be expected to bind 5 and activate the TIE-2 receptor. Similarly, other constructs may be created using the teachings herein and are therefore considered to be within the scope of this invention.

23.6. Materials and Methods

Construction of Chimeras

Swapping constructs were inserted into a pJFE14 vector in which the Xbal site was changed to an AscI site. This vector was then digested with AscI and NotI yielding an AscI-NotI backbone. DNA fragments for the chimeras were generated by PCR using appropriate oligonucleotides.

The FLAG-1C1F and FLAG-2C2F inserts were subcloned into a pMT21 vector backbone that had been digested with EcoRI and NotI. The "CF" truncations were obtained through PCR, and the FLAG tag and a preceding trypsin signalling sequence were constructed by annealing synthetic 20 oligonucleotides.

Transfections

All constructs were transfected transiently into COS7 cells using either DEAE-Dextran or LipofectAMINE according to standard protocols. Cell cultures were har- 25 vested 3 days after the transfection and spun down at 1000 rpm for 1 minute, and the supernatants were transferred to fresh tubes and stored at -20° C.

Staining of FLAG-1C1F-Transfected and FLAG-2C2F-Transfected Cells 6-well dishes of COS7 cells were transfected transiently with the TIE-2 receptor. The COS7 super-

natant from various ligand tansfections was incubated on the cells for 30 minutes, followed by two washes with Phosphate Buffered Saline (PBS) without magnesium or calcium. The cells were fixed in -20° C. methanol for 3 minutes, washed once with PBS, and incubated with anti-FLAG M2 antibody (IBI;1:3000 dilution) in PBS/10% Bovine Calf Serum (BCS) for 30 minutes. The cells were washed once with PBS and incubated with goat anti-mouse IgG Alkaline Phosphatase (AP) conjugated antibody (Promega;1:1000) in PBS/10% BCS. The cells were washed twice with PBS and incubated with the phosphate substrate, BCIP/NBT, with 1 mM levamisole.

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Phosphorylation Assays

Dilution of COS7 supernatants for the dose response study was done in the supernatants of COS7 cells transfected with the empty vector pJFE14. EA cells that naturally express the TIE-2 receptor were starved for >2 hours in serum-free medium, followed by challenge with the appropriate COS7 supernatant for 10 minutes at 37° C. in an atmosphere of 5% CO2. The cells were then rinsed in ice-cold PBS and lysed with 1% NP40 lysis buffer containing protease inhibitors (10 μ g/ml leupeptin, 10 μ g/ml aprotinin, 1 mM PMSF) followed by immunoprecipitation with an antibody specific for the TIE-2 receptor. Samples were then subjected to immunoblot analysis, using anti pTyr antibodies.

Dot Blots

Samples were applied to a nitrocellulose membrane, which was blocked and probed with the appropriate antibodies.

TABLE 1

	TI BEE 1								
	MUTATION ANALYSIS OF TIE LIGANDS								
	COILED- N COIL	FIBRINOGEN- LIKE	TIE2 Binding	TIE2 Activation	Multimeric Structure	Production Levels			
TL1	С		+	+	HIGHER ORDER	LOW			
TL2			+	-	DIMER	HIGH			
			+	+	DIMER	LOW			
			+	-	HIGHER ORDER	HIGH			
	С		-	N.D.	N.D.	LOW			
			-	N.D.	N.D.	HIGH			
			-	-	MONOMER	HIGH			
			-	-	MONOMER	HIGH			
		Fc	+	-	DIMER	HIGH			
		Fc	+	-	DIMER	HIGH			
	С	Fc	+	+	HIGHER ORDER	LOW			

TABLE 1-continued

	MUTATION ANALYS	SIS OF TI	E LIGANDS		
	COILED- FIBRINOGEN- N COIL LIKE	TIE2 Binding	TIE2 Activation	Multimeric Structure	Production Levels
	Fc Fc	+	-	HIGHER ORDER	LOW
flag-	С	+	+	N.D.	LOW
flag-		+	-	N.D.	HIGH
	С	+	-	N.D.	HIGH
		+	=	N.D.	HIGH
	c///////	+	-	N.D.	LOW
		+	+	N.D.	HIGH*
		+	-	N.D.	LOW
	c	+	+**	N.D.	HIGH
		+	+**	DIMER	HIGH
	X c	+	+	N.D.	LOW

*HIGHEST PRODUCTION OF RU **MOST POTENTLY ACTIVATING

N.D. = NOT DETERMINED

Deposits

The following have been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Md. 20852 in accordance with the Budapest Treaty. A plasmid clone encoding a TIE-2 ligand was deposited with encoding TIE-2 ligand" under ATCC Accession No. 75910. Recombinant Autographa californica baculovirus encoding TIE-2 receptorbody was deposited with the ATCC on Oct. 7, 1994 and designated as "vTIE-2 receptorbody" under ATCC Accession No. VR2484. A lambda phage vector containing 50 human tie-2 ligand DNA was deposited with the ATCC on Oct. 26, 1994 and designated as "\(\lambda \text{gt10} \) encoding htie-2 ligand 1" under ATCC Accession No. 75928. A plasmid clone encoding a second TIE-2 ligand was deposited with

the ATCC on Dec. 9, 1994 and designated as "pBluescript KS encoding human TIE 2 ligand 2" under ATCC Accession No. 75963. E. coli strain DH10B containing plasmid pBe-LoBac11 with a human TL-4 gene insert encoding human TIE ligand-4 was deposited with the ATCC on Jul. 2, 1996 the ATCC on Oct. 7, 1994 and designated as "pJFE14 45 and designated as "hTL-4" under ATCC Accession No. 98095.

> The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

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	ctg Leu															735

							cag Gln							783	
							tca Ser							831	
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							tta Leu							927	
							gag Glu 215							975	
_		-					cag Gln	 _	_	_				1023	
							ctt Leu							1071	
_	-		-			_	aat Asn	-			-	-		1119	
	_				_	 _	gag Glu			_	-	_	-	1167	
							aaa Lys 295							1215	
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							tat Tyr							1359	
		-			_		gag Glu			_			-	1407	
							gag Glu 375							1455	
							ttc Phe							1503	
			_				cac His			-			_	1551	
							gat Asp							1599	
							gcc Ala							1647	
		_	-	_			aat Asn 455			_				1695	

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Glu His Asp Gly .	Asn Cys Arg 55	Glu Ser Thr	Thr Asp Gln 60	Tyr Asn Thr	
Asn Ala Leu Gln . 65	Arg Asp Ala 70	Pro His Val	Glu Pro Asp : 75	Phe Ser Ser 80	
Gln Lys Leu Gln	His Leu Glu 85	His Val Met 90	Glu Asn Tyr	Thr Gln Trp 95	
Leu Gln Lys Leu 100	Glu Asn Tyr	Ile Val Glu 105		Ser Glu Met 110	
Ala Gln Ile Gln (Gln Asn Ala	Val Gln Asr 120	His Thr Ala	Thr Met Leu	
Glu Ile Gly Thr	Ser Leu Leu 135		Ala Glu Gln	Thr Arg Lys	
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Ile Gln Leu Leu	Glu Asn Ser 165	Leu Ser Thr		Glu L y s Gln 175	
Leu Leu Gln Gln '	Thr Asn Glu	Ile Leu Lys 185		L y s Asn Ser 190	
Leu Leu Glu His : 195	Lys Ile Leu	Glu Met Glu 200	Gly Lys His 1 205	L y s Glu Glu	
Leu Asp Thr Leu 2	L y s Glu Glu 215		Leu Gln Gly	Leu Val Thr	
Arg Gln Thr Tyr 225	Ile Ile Gln 230	Glu Leu Glu	Lys Gln Leu . 235	Asn Arg Ala 240	
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Gln	Ty r 370	Met	Leu	Arg	Ile	Glu 375	Leu	Met	Asp	Trp	Glu 380	Gly	Asn	Arg	Ala	
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Arg	Leu	Tyr	Leu	L y s 405	Gly	His	Thr	Gly	Thr 410	Ala	Gly	Lys	Gln	Ser 415	Ser	
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Asn	Cys	Met 435	Сув	Lys	Cys	Ala	Leu 440	Met	Leu	Thr	Gly	Gly 445	Trp	Trp	Phe	
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tct	gggg	aga 🤉	gagga	aacaa	aa g	gacc	gtgaa	a ago	etget	ctg	taaa	aagc	ga o	cacaç	gccctc	120
ccaa	agtga	agc a	aggad	ctgti	tc tt	ccca	actgo	c aat	ctga	acag	ttta	actgo	cat o	gcct	ggagag	180
aaca	acago	cag t	aaaa	aacca	ag gt	ttg	ctact	gga	aaaa	agag	gaaa	agaga	aag a	actti	cattg	240
acg	gacco	cag o	ccato	ggca	gc gt	agca	agcco	t tg	gttt	cag	acg	gcag	cag o	ctcg	ggactc	300
tgga	acgto	gtg t	ttg	ccct	ca aq	gttt	gctaa	a gct	get	ggtt	tati	tact	gaa q	gaaaq	ga atg Met 1	359
														gcc Ala		407

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	tat Tyr												455
	cag Gln 35												503
	gac Asp												551
	agg Arg												599
	ctg Leu												647
	aat Asn		_	-		_	_	-	_	-		_	695
	aat Asn 115												743
	ctg Leu												791
	gcc Ala												839
	cac His												887
	agt Ser												935
_	gtg Val 195	-	_	-	_	_					_		983
	gaa Glu												1031
	att Ile												1079
	gtt Val		_				-	_			-		1127
	ctg Leu												1175
	aaa Lys 275												1223
	gga Gly												1271
	gaa Glu												1319
	aca Thr												1367

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cac ctt aaa gga ctt aca ggg aca gcc ggc aaa ata agc agc atc agc His Leu Lys Gly Leu Thr Gly Thr Ala Gly Lys Ile Ser Ser Ile Ser 405 410 415	1607
caa cca gga aat gat ttt agc aca aag gat gga gac aac gac aaa tgt Gln Pro Gly Asn Asp Phe Ser Thr Lys Asp Gly Asp Asn Asp Lys Cys 420 425 430	1655
att tgc aaa tgt tca caa atg cta aca gga ggc tgg tgg ttt gat gca Ile Cys Lys Cys Ser Gln Met Leu Thr Gly Gly Trp Trp Phe Asp Ala 435 440 445	1703
tgt ggt cct tcc aac ttg aac gga atg tac tat cca cag agg cag aac Cys Gly Pro Ser Asn Leu Asn Gly Met Tyr Tyr Pro Gln Arg Gln Asn 450 460 465	1751
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tat tcg ctc aag gcc aca acc atg atg atc cga cca gca gat ttc Tyr Ser Leu Lys Ala Thr Thr Met Met Ile Arg Pro Ala Asp Phe 485 490 495	1844
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Gln Tyr Gln Val Gln His Gly Ser Cys Ser Tyr Thr Phe Leu Leu Pro $35 \hspace{1cm} 40 \hspace{1cm} 45$	
Glu Met Asp Asn Cys Arg Ser Ser Ser Pro Tyr Val Ser Asn Ala 50 55 60	
Val Gln Arg Asp Ala Pro Leu Glu Tyr Asp Asp Ser Val Gln Arg Leu 65 70 75 80	

Gln	Val	Leu	Glu	Asn 85	Ile	Met	Glu	Asn	Asn 90	Thr	Gln	Trp	Leu	Met 95	Lys
Leu	Glu	Asn	Ty r 100	Ile	Gln	Asp	Asn	Met 105	Lys	Lys	Glu	Met	Val	Glu	Ile
Gln	Gln	Asn 115	Ala	Val	Gln	Asn	Gln 120	Thr	Ala	Val	Met	Ile 125	Glu	Ile	Gly
Thr	Asn 130	Leu	Leu	Asn	Gln	Thr 135	Ala	Glu	Gln	Thr	Arg 140	Lys	Leu	Thr	Asp
Val 145	Glu	Ala	Gln	Val	Leu 150	Asn	Gln	Thr	Thr	Arg 155	Leu	Glu	Leu	Gln	Leu 160
Leu	Glu	His	Ser	Leu 165	Ser	Thr	Asn	Lys	Leu 170	Glu	Lys	Gln	Ile	Leu 175	Asp
Gln	Thr	Ser	Glu 180	Ile	Asn	Lys	Leu	Gln 185	Asp	Lys	Asn	Ser	Phe 190	Leu	Glu
Lys	Lys	Val 195	Leu	Ala	Met	Glu	Asp 200	Lys	His	Ile	Ile	Gln 205	Leu	Gln	Ser
Ile	L y s 210	Glu	Glu	Lys	Asp	Gln 215	Leu	Gln	Val	Leu	Val 220	Ser	Lys	Gln	Asn
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Ser 305	Thr	Glu	Glu	Ile	Lys 310	Ala	Tyr	Суѕ	Asp	Met 315	Glu	Ala	Gly	Gly	Gly 320
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Tyr	Trp	Leu 355	Gly	Asn	Glu	Phe	Val 360	Ser	Gln	Leu	Thr	Asn 365	Gln	Gln	Arg
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His 65	Leu	Glu	His	Val	Met 70	Glu	Asn	Tyr	Thr	Gln 75	Trp	Leu	Gln	Lys	Leu 80
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Glu 145	Asn	Ser	Leu	Ser	Thr 150	Tyr	Lys	Leu	Glu	L y s 155	Gln	Leu	Leu	Gln	Gln 160
Thr	Asn	Glu	Ile	Leu 165	Lys	Ile	His	Glu	Lys 170	Asn	Ser	Leu	Leu	Glu 175	His
Lys	Ile	Leu	Glu 180	Met	Glu	Gly	Lys	His 185	Lys	Glu	Glu	Leu	Asp 190	Thr	Leu
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Ile	Ile 210	Gln	Glu	Leu	Glu	Lys 215	Gln	Leu	Asn	Arg	Ala 220	Thr	Thr	Asn	Asn
Ser 225	Val	Leu	Gln	Lys	Gln 230	Gln	Leu	Glu	Leu	Met 235	Asp	Thr	Val	His	Asn 240
Leu	Val	Asn	Leu	Cys 245	Thr	Lys	Glu	Gly	Val 250	Leu	Leu	Lys	Gly	Gly 255	Lys
Arg	Glu	Glu	Glu 260	Lys	Pro	Phe	Arg	Asp 265	Cys	Ala	Asp	Val	Ty r 270	Gln	Ala
Gly	Phe	Asn 275	Lys	Ser	Gly	Ile	Ty r 280	Thr	Ile	Tyr	Ile	Asn 285	Asn	Met	Pro
Glu	Pro 290	Lys	Lys	Val	Phe	C y s 295	Asn	Met	Asp	Val	Asn 300	Gly	Gly	Gly	Trp
Thr 305	Val	Ile	Gln	His	Arg 310	Glu	Asp	Gly	Ser	Leu 315	Asp	Phe	Gln	Arg	Gly 320
Trp	Lys	Glu	Tyr	L y s 325	Met	Gly	Phe	Gly	Asn 330	Pro	Ser	Gly	Glu	Ty r 335	Trp
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Leu	Arg	Ile 355	Glu	Leu	Met	Asp	Trp 360	Glu	Gly	Asn	Arg	Ala 365	Tyr	Ser	Gln
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		275					280					285						
Ser	Thr 290	Glu	Glu	Ile	Lys	Ala 295	Tyr	Cys	Asp	Met	Glu 300	Ala	Gly	Gly	Gly			
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er	Gln	Pro	Gly	Asn 405	Asp	Phe	Ser	Thr	Lys 410	Asp	Gly	Asp	Asn	Asp 415	Lys			
ys	Ile	Cys	L y s 420	Cys	Ser	Gln	Met	Leu 425	Thr	Gly	Gly	Trp	Trp 430	Phe	Asp			
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21 21 22 22 22 22 22 22	l> LE 2> TY 3> OF 0> FE 1> NA 2> LO 92 0> SE	ENGTH PE: RGANI EATUR AME/K DCATI THER 29.	DNA SM: RE: RE: CON: INFO	Homo CDS (47) DRMAT)(1 FION:	1573) : The	i e fibr					cag a		ctc t	- -	ition	55	
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Phe Ile Phe Ala Ile Thr Ser Gln Arg Gln Tyr Met Leu Arg Ile Glu His Ile Gly Asn Glu Lys Gln Asn Tyr Arg Leu Tyr Leu Lys Gly His 385 390395400 Leu Asn Gly Met Phe Tyr Thr Ala Gly Gln Asn His Gly Lys Leu Asn 450 $\,$ 460 $\,$ Gly Ile Lys Trp His Tyr Phe Lys Gly Pro Ser Tyr Ser Ile Arg Ser 465 470470475 Thr Thr Met Met Ile Arg Pro Leu Asp Phe 485 490 <210> SEQ ID NO 13 <211> LENGTH: 491 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEQUENCE: 13 Ala Phe Leu Ala Ala Ile Leu Ala His Ile Gly Cys Thr Thr Gln Arg 1 $$ 5 $$ 10 $$ 15 Arg Ser Pro Glu Asn Ser Gly Arg Arg Phe Asn Arg Ile Gln His Gly $20 \hspace{1.5cm} 25 \hspace{1.5cm} 30$ Gln Cys Thr Tyr Thr Phe Ile Leu Pro Glu Gln Asp Gly Asn Cys Arg 35 40 45Glu Ser Thr Thr Asp Gln Tyr Asn Thr Asn Ala Leu Gln Arg Asp Ala 50 55 Pro His Val Glu Gln Asp Phe Ser Phe Gln Lys Leu Gln His Leu Glu 65 70 75 80 His Val Met Glu Asn Tyr Thr Gln Trp Leu Gln Lys Leu Glu Ser Tyr 85 90 95 Ile Val Glu Asn Met Lys Ser Glu Met Ala Gln Leu Gln Gln Asn Ala 100 $$105\$ Val Gln Asn His Thr Ala Thr Met Leu Glu Ile Gly Thr Ser Leu Leu 115 120 125 Ser Gln Thr Ala Glu Gln Thr Arg Lys Leu Thr Asp Val Glu Thr Gln $130\,$ Val Leu Asn Gln Thr Ser Arg Leu Glu Ile Gln Leu Leu Glu Asn Ser 145 150 160 Leu Ser Thr Tyr Lys Leu Glu Lys Gln Leu Leu Gln Gln Thr Asn Glu 165 170 175Ile Leu Lys Ile His Glu Lys Asn Ser Leu Leu Glu His Lys Ile Leu 180 \$185\$Glu Met Glu Glu Arg His Lys Glu Glu Met Asp Thr Leu Lys Glu Glu 195 200 205 Lys Glu Asn Leu Gln Gly Leu Val Thr Arg Gln Ser Tyr Ile Ile Gln 210 215 220Glu Leu Glu Lys Gln Leu Asn Lys Ala Thr Thr Asn Asn Ser Val Leu

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Glu	Lys	Pro 275	Phe	Arg	Asp	Суѕ	Ala 280	Asp	Val	Tyr	Gln	Ala 285	Gly	Phe	Asn
Lys	Ser 290	Gly	Ile	Tyr	Thr	Ile 295	Tyr	Ile	Asn	Asn	Val 300	Ser	Asp	Pro	Lys
Lys 305	Val	Phe	Суѕ	Asn	Met 310	Asp	Val	Asn	Gly	Gly 315	Gly	Trp	Thr	Val	Ile 320
Gln	His	Arg	Glu	Asp 325	Gly	Ser	Leu	Asp	Phe 330	Gln	Lys	Gly	Trp	Lys 335	Glu
Tyr	Lys	Met	Gly 340	Phe	Gly	Ser	Pro	Ser 345	Gly	Glu	Tyr	Trp	Leu 350	Gly	Asn
Glu	Phe	Ile 355	Phe	Ala	Ile	Thr	Ser 360	Gln	Arg	Gln	Tyr	Ser 365	Leu	Arg	Ile
Glu	Leu 370	Met	Asp	Trp	Glu	Gly 375	Asn	Arg	Ala	Tyr	Ser 380	Gln	Tyr	Asp	Arg
Phe 385	His	Ile	Gly	Asn	Glu 390	Lys	Gln	Asn	Tyr	Arg 395	Leu	Tyr	Leu	Lys	Gly 400
His	Ser	Gly	Thr	Ala 405	Gly	Lys	Gln	Ser	Ser 410	Leu	Ile	Leu	His	Gly 415	Ala
Glu	Phe	Ser	Thr 420	Lys	Asp	Ala	Asp	Asn 425	Asp	Asn	Сув	Met	Cys 430	Lys	Cys
Ala	Leu	Met 435	Leu	Thr	Gly	Gly	Trp 440	Trp	Phe	Asp	Ala	Cys 445	Gly	Pro	Ser
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Asn 465	Gly	Ile	Lys	Trp	His 470	Tyr	Phe	Lys	Gly	Pro 475	Arg	Tyr	Ser	Ile	Arg 480
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Tyr	Asn	Arg 35	Ile	Gln	His	Gly	Gln 40	Сув	Ala	Tyr	Thr	Phe 45	Ile	Leu	Pro
Glu	His 50	Asp	Gly	Asn	Сув	Arg 55	Glu	Ser	Thr	Thr	Asp 60	Gln	Tyr	Asn	Thr
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Gln	Lys	Leu	Gln	His 85	Leu	Glu	His	Val	Met 90	Glu	Asn	Tyr	Thr	Gln 95	Trp
Leu	Gln	Lys	Leu 100	Glu	Asn	Tyr	Ile	Val 105	Glu	Asn	Met	Lys	Ser 110	Glu	Met

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		115					120					125			
Glu	11e 130	Gly	Thr	Ser	Leu	Leu 135	Ser	Gln	Thr	Ala	Glu 140	Gln	Thr	Arg	Lys
Leu 145	Thr	Asp	Val	Glu	Thr 150	Gln	Val	Leu	Asn	Gln 155	Thr	Ser	Arg	Leu	Glu 160
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Leu	Glu	His 195	Lys	Ile	Leu	Glu	Met 200	Glu	Gly	Lys	His	Lys 205	Glu	Glu	Met
Asp	Thr 210	Leu	Lys	Glu	Glu	Lys 215	Glu	Asn	Leu	Gln	Gl y 220	Leu	Val	Ser	Arg
Gln 225	Ser	Phe	Ile	Ile	Gln 230	Glu	Leu	Glu	Lys	Gln 235	Leu	Ser	Arg	Ala	Thr 240
Asn	Asn	Asn	Ser	Ile 245	Leu	Gln	Lys	Gln	Gln 250	Leu	Glu	Leu	Met	A sp 255	Thr
Val	His	Asn	Leu 260	Ile	Ser	Leu	Cys	Thr 265	Lys	Glu	Gly	Val	Leu 270	Leu	Lys
Gly	Gly	Lys 275	Arg	Glu	Glu	Glu	L y s 280	Pro	Phe	Arg	Asp	Cys 285	Ala	Asp	Val
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Asn 305	Val	Pro	Glu	Pro	Lys 310	Lys	Val	Phe	Cys	Asn 315	Met	Asp	Val	Asn	Gl y 320
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Glu	Tyr	Trp 355	Leu	Gly	Asn	Glu	Phe 360	Ile	Phe	Ala	Ile	Thr 365	Ser	Gln	Arg
Gln	Ty r 370	Met	Leu	Arg	Ile	Glu 375	Leu	Met	Asp	Trp	Glu 380	Gly	Asn	Arg	Ala
Ty r 385	Ser	Gln	Tyr	Asp	Arg 390	Phe	His	Ile	Gly	Asn 395	Glu	Lys	Gln	Asn	Ty r 400
Arg	Leu	Tyr	Leu	Lys 405	Gly	His	Thr	Gly	Thr 410	Ala	Gly	Lys	Gln	Ser 415	Ser
Leu	Ile	Leu	His 420	Gly	Ala	Asp	Phe	Ser 425	Thr	Lys	Asp	Ala	Asp 430	Asn	Asp
Asn	Cys	Met 435	Суѕ	Lys	Cys	Ala	Leu 440	Met	Leu	Thr	Gly	Gly 445	Trp	Trp	Phe
Asp	Ala 450	Cys	Gly	Pro	Ser	Asn 455	Leu	Asn	Gly	Met	Phe 460	Tyr	Thr	Ala	Gly
Gln 465	Asn	His	Gly	Lys	Leu 470	Asn	Gly	Ile	Lys	Trp 475	His	Tyr	Phe	Lys	Gly 480
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Arg	Tyr	Arg 35	Ile	Gln	Asn	Gly	Pro 40	Cys	Ala	Tyr	Thr	Phe 45	Leu	Leu	Pro
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Val 65	Gln	Arg	Asp	Ala	Pro 70	Pro	Asp	Tyr	Glu	Asp 75	Ser	Val	Gln	Ser	Leu 80
Gln	Leu	Leu	Glu	Asn 85	Val	Met	Glu	Asn	Ty r 90	Thr	Gln	Trp	Leu	Met 95	Lys
Leu	Glu	Asn	Tyr 100	Ile	Gln	Asp	Asn	Met 105	Lys	Lys	Glu	Met	Ala 110	Glu	Ile
Gln	Gln	Asn 115	Val	Val	Gln	Asn	His 120	Thr	Ala	Val	Met	Ile 125	Glu	Ile	Gly
Thr	Ser 130	Leu	Leu	Ser	Gln	Thr 135	Ala	Glu	Gln	Thr	Arg 140	Lys	Leu	Thr	Asp
Val 145	Glu	Thr	Gln	Val	Leu 150	Asn	Gln	Thr	Thr	A rg 155	Leu	Glu	Leu	Gln	Leu 160
Leu	Gln	His	Ser	Ile 165	Ser	Thr	Tyr	Lys	Leu 170	Glu	Lys	Gln	Ile	Leu 175	Asp
Gln	Thr	Ser	Glu 180	Ile	Asn	Lys	Ile	His 185	Asn	Lys	Asn	Ser	Phe 190	Leu	Glu
Gln	Lys	Val 195	Leu	Asp	Met	Glu	Gly 200	Lys	His	Ser	Glu	Glu 205	Met	Gln	Thr
Met	L y s 210	Glu	Gln	Lys	Asp	Glu 215	Leu	Gln	Val	Leu	Val 220	Ser	Lys	Gln	Ser
Ser 225	Val	Ile	Asp	Glu	Leu 230	Glu	Lys	Lys	Leu	Val 235	Thr	Ala	Thr	Val	Asn 240
Asn	Ser	Leu	Leu	Gln 245	Lys	Gln	Gln	His	Asp 250	Leu	Met	Asp	Thr	Val 255	Asn
Ser	Leu	Leu	Thr 260	Met	Met	Ser	Ser	Pro 265	Asn	Ser	Lys	Ser	Ser 270	Leu	Ala
Ile	Arg	A rg 275	Glu	Glu	Gln	Thr	Thr 280	Phe	Arg	Asp	Cys	Ala 285	Asp	Val	Phe
Lys	Ala 290	Gly	Leu	Thr	Lys	Ser 295	Gly	Ile	Tyr	Thr	Leu 300	Thr	Phe	Pro	Asn
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Lys	Gly	Trp	Lys 340	Glu	Tyr	Lys	Met	Gly 345	Phe	Gly	Asn	Pro	Leu 350	Gly	Glu
Tyr	Trp	Leu 355	Gly	Asn	Glu	Phe	Ile 360	Ser	Gln	Ile	Thr	Gly 365	Gln	His	Arg
Tyr	Val 370	Leu	Lys	Ile	Gln	Leu 375	Lys	Asp	Trp	Glu	Gly 380	Asn	Glu	Ala	His
Ser 385	Leu	Tyr	Asp	His	Phe 390	Tyr	Ile	Ala	Gly	Glu 395	Glu	Ser	Asn	Tyr	Arg 400
Ile	His	Leu	Thr	Gly	Leu	Thr	Gly	Thr	Ala	Ala	Lys	Ile	Ser	Ser	Ile

				405					410					415	
Ser	Gln	Pro	Gly 420	Ser	Asp	Phe	Ser	Thr 425	Lys	Asp	Ser	Asp	Asn 430	Asp	Lys
Cys	Ile	Cys 435	Lys	Сув	Ser	Leu	Met 440	Leu	Thr	Gly	Gly	Trp 445	Trp	Phe	Asp
Ala	Cys 450	Gly	Pro	Ser	Asn	Leu 455	Asn	Gly	Gln	Phe	Ty r 460	Pro	Gln	Lys	Gln
Asn 465	Thr	Asn	Lys	Phe	Asn 470	Gly	Ile	Lys	Trp	Ty r 475	Tyr	Trp	Lys	Gly	Ser 480
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Arg	Tyr	Arg 35	Ile	Gln	His	Gly	Ser 40	Сув	Ala	Tyr	Thr	Phe 45	Leu	Leu	Pro
Glu	Met 50	Asp	Asn	Gly	Arg	Ser 55	Ser	Ser	Ser	Thr	Tyr 60	Val	Thr	Asn	Ala
Val 65	Gln	Arg	Asp	Ala	Pro 70	Pro	Glu	Tyr	Glu	Asp 75	Ser	Val	Gln	Ser	Leu 80
Gln	Leu	Leu	Glu	Asn 85	Val	Met	Glu	Asn	Ty r 90	Thr	Gln	Trp	Leu	Met 95	Lys
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Thr	Ser 130	Leu	Leu	Ser	Gln	Thr 135	Ala	Glu	Gln	Thr	Arg 140	Lys	Leu	Thr	Asp
Val 145	Glu	Thr	Gln	Val	Leu 150	Asn	Gln	Thr	Thr	Arg 155	Leu	Glu	Leu	Gln	Leu 160
Leu	Gln	His	Ser	Ile 165	Ser	Thr	Tyr	Lys	Leu 170	Glu	Lys	Gln	Ile	Leu 175	Asp
Gln	Thr	Ser	Glu 180	Ile	Asn	Lys	Ile	His 185	Asp	Lys	Asn	Ser	Phe 190	Leu	Glu
Lys	Lys	Val 195	Leu	Asp	Met	Glu	Asp 200	Lys	His	Ile	Ile	Glu 205	Met	Gln	Thr
Ile	L y s 210	Glu	Glu	Lys	Asp	Glu 215	Leu	Gln	Val	Leu	Val 220	Ser	Lys	Gln	Asn
Ser 225	Ile	Ile	Glu	Glu	Leu 230	Glu	Lys	Lys	Ile	Val 235	Thr	Ala	Thr	Val	Asn 240
Asn	Ser	Val	Leu	Gln 245	Lys	Gln	Gln	His	Asp 250	Leu	Met	Asp	Thr	Val 255	Asn
Asn	Leu	Leu	Thr 260	Met	Met	Ser	Thr	Ser 265	Asn	Ser	Ala	Lys	Asp 270	Ser	Thr
Val	Ala	Arg 275	Glu	Glu	Gln	Ile	Ser 280	Phe	Arg	Asp	Cys	Ala 285	Asp	Val	Phe

290 295 300	
Ser Pro Glu Glu Ile Lys Ala Tyr Cys Asn Met Asp Ala Gly Gly Gly 305 310 315 320	
Gly Trp Thr Ile Ile Gln Arg Arg Glu Asp Gly Ser Leu Asp Phe Gln 325 330 335	
Lys Gly Trp Lys Glu Tyr Lys Val Gly Phe Gly Ser Pro Ser Gly Glu 340 345 350	
Tyr Trp Leu Gly Asn Glu Phe Ile Ser Gln Ile Thr Asn Gln Gln Arg 355 360 365	
Tyr Val Leu Lys Ile His Leu Lys Asp Trp Glu Gly Asn Glu Ala Tyr 370 375 380	
Ser Leu Tyr Asp His Phe Tyr Ile Ser Gly Glu Glu Leu Asn Tyr Arg 385 390 395 400	
Ile His Leu Lys Gly Leu Thr Gly Thr Ala Ala Lys Ile Ser Ser Ile 405 410 415	
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Cys Ile Cys Lys Cys Ser Leu Met Leu Thr Gly Gly Trp Trp Phe Asp 435 440 445	
Ala Cys Gly Pro Ser Asn Leu Asn Gly Met Phe Tyr Pro Gln Arg Gln 450 455 460	
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	ccc Pro 130														432
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	gag Glu														576
	caa Gln														624
	cag Gln 210														672
	acg Thr														720
	cgc Arg														768
_	ctg Leu	-	_	_	_		_	_			_			-	 816
	aac Asn														864
	gac Asp 290														912
	acc Thr														960
_	ctg Leu	_	-	_								_	_	-	 1008
	ggc Gly														1056
	gga Gly														1104
	ctc Leu 370														1152
	gaa Glu														1200
	gag Glu														1248
	ggg Gly														1296

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Сув	Glu	Thr 35	Leu	Val	Val	Gln	His 40	Gly	His	Суѕ	Ser	Ty r 45	Thr	Phe	Leu	
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Lys	Leu	Pro	Thr	Gln 85	Gln	Val	Lys	Gln	Leu 90	Glu	Gln	Ala	Leu	Gln 95	Asn	
Asn	Thr	Gln	Trp 100	Leu	Lys	Lys	Leu	Glu 105	Arg	Ala	Ile	Lys	Thr 110	Ile	Leu	
Arg	Ser	Lys 115	Leu	Glu	Gln	Val	Gln 120	Gln	Gln	Met	Ala	Gln 125	Asn	Gln	Thr	
Ala	Pro 130	Met	Leu	Glu	Leu	Gly 135	Thr	Ser	Leu	Leu	Asn 140	Gln	Thr	Thr	Ala	
Gln 145	Ile	Arg	Lys	Leu	Thr 150	Asp	Met	Glu	Ala	Gln 155	Leu	Leu	Asn	Gln	Thr 160	
Ser	Arg	Met	Asp	Ala 165	Gln	Met	Pro	Glu	Thr 170	Phe	Leu	Ser	Thr	Asn 175	Lys	
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Gly	Gln	Asn 195	Ser	Ala	Leu	Glu	Ly s 200	Arg	Leu	Gln	Ala	Leu 205	Glu	Thr	Lys	
Gln	Gln 210	Glu	Glu	Leu	Ala	Ser 215	Ile	Leu	Ser	Lys	L y s 220	Ala	Lys	Leu	Leu	
Asn 225	Thr	Leu	Ser	Arg	Gln 230	Ser	Ala	Ala	Leu	Thr 235	Asn	Ile	Glu	Arg	Gly 240	
Leu	Arg	Gly	Val	Arg 245	His	Asn	Ser	Ser	Leu 250	Leu	Gln	Asp	Gln	Gln 255	His	
Ser	Leu	Arg	Gln 260	Leu	Leu	Val	Leu	Leu 265	Arg	His	Leu	Val	Gln 270	Glu	Arg	

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Gln Asp Cys 290	Ala Glu	Ile Gln 295		Gly Ala	Ser Ala 300	Ser Gl	y Val
Tyr Thr Ile	e Gln Val	Ser Asn 310	Ala Thr	Lys Pro 315	Arg Lys	Val Ph	e Cys 320
Asp Leu Glr	Ser Ser 325		Arg Trp	Thr Leu 330	Ile Gln	Arg Ar	
Asn Gly Th	Val Asn 340	Phe Gln	Arg Asn 345	Trp Lys	Asp Tyr	Lys Gl: 350	n Gly
Phe Gly Asp 355		Gly Glu	His Trp 360	Leu Gly	Asn Glu 365	Val Va	l His
Gln Leu Thi	Arg Arg	Ala Ala 375		Leu Arg	Val Glu 380	Leu Gl:	n Asp
Trp Glu Gly 385	His Glu	Ala Tyr 390	Ala Gln	Tyr Glu 395	His Phe	His Le	u Gly 400
Ser Glu Asr	Gln Leu 405		Leu Ser	Val Val 410	Gly Tyr	Ser Gl	
Ala Gly Arc	Gln Ser 420	Ser Leu	Val Leu 425	Gln Asn	Thr Ser	Phe Se	r Thr
Leu Asp Ser 435		Asp His	Cys Leu 440	Cys Lys	Cys Ala 445	Gln Va	l Met
Ser Gly Gly 450	Trp Trp	Phe Asp 455		Gly Leu	Ser Asn 460	Leu As:	n Gly
Val Tyr Tyr 465	His Ala	Pro Asp 470	Asn Lys	Tyr Lys 475	Met Asp	Gly Il	e Arg 480
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						aat Asn											336
						aat Asn											384
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						aat Asn											528
						aat Asn	-		_	_			_			_	576
						atc Ile											624
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	g (atc Ile 230											720
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						att Ile											1008
						aaa Lys											1056
						gga Gly											1104
	n A					aaa Lys											1152

	tac															
	Tyr				gaa Glu 390											1200
	agg Arg															1248
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Leu Leu Glu His Lys Ile Leu Glu Met Glu Gly Lys His Lys Glu Glu 195 200 205

Leu	Asp 210	Thr	Leu	Lys	Glu	Glu 215	Lys	Glu	Asn	Leu	Gln 220	Gly	Leu	Val	Thr	
Arg 225	Gln	Thr	Tyr	Ile	Ile 230	Gln	Glu	Leu	Glu	L y s 235	Gln	Leu	Asn	Arg	Ala 240	
Thr	Thr	Asn	Asn	Ser 245	Val	Leu	Gln	Lys	Gln 250	Gln	Leu	Glu	Leu	Met 255	Asp	
Thr	Val	His	Asn 260	Leu	Val	Asn	Leu	Cys 265	Thr	Lys	Glu	Gly	Val 270	Leu	Leu	
Lys	Gly	Gly 275	Lys	Arg	Glu	Glu	Glu 280	Lys	Pro	Phe	Arg	Asp 285	Cys	Ala	Glu	
Val	Phe 290	Lys	Ser	Gly	His	Thr 295	Thr	Asn	Gly	Ile	Ty r 300	Thr	Leu	Thr	Phe	
Pro 305	Asn	Ser	Thr	Glu	Glu 310	Ile	Lys	Ala	Tyr	Cys 315	Asp	Met	Glu	Ala	Gly 320	
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Gly	Glu	Ty r 355	Trp	Leu	Gly	Asn	Glu 360	Phe	Val	Ser	Gln	Leu 365	Thr	Asn	Gln	
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	cag Gln														240
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	cag Gln														384
	aac Asn 130														432
	gaa Glu														480
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_	aag L y s			_	_	_	_	_					_		624
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	atc Ile														720
	tca Ser	-			_				-		_		-		768
	tta Leu														816
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Leu	Glu	Asn	Ty r	Ile	Gln	Asp	Asn	Met 105	Lys	Lys	Glu	Met	Val 110	Glu	Ile	
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Leu	Glu	His	Ser	Leu 165	Ser	Thr	Asn	Lys	Leu 170	Glu	Lys	Gln	Ile	Leu 175	Asp
Gln	Thr	Ser	Glu 180	Ile	Asn	Lys	Leu	Gln 185	Asp	Lys	Asn	Ser	Phe 190	Leu	Glu
Lys	Lys	Val 195	Leu	Ala	Met	Glu	Asp 200	Lys	His	Ile	Ile	Gln 205	Leu	Gln	Ser
Ile	L y s 210	Glu	Glu	Lys	Asp	Gln 215	Leu	Gln	Val	Leu	Val 220	Ser	Lys	Gln	Asn
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Ile	Leu	His	Gly 420	Ala	Asp	Phe	Ser	Thr 425	Lys	Asp	Ala	Asp	Asn 430	Asp	Asn
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Ala	Cys 450	Gly	Pro	Ser	Asn	Leu 455	Asn	Gly	Met	Phe	Ty r 460	Thr	Ala	Gly	Gln
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Asn Ala Leu Gln Arg Asp Ala Pro His Val Glu Pro Asp Asp Ser Val

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Val	Glu	Ile 115	Gln	Gln	Asn	Ala	Val 120	Gln	Asn	Gln	Thr	Ala 125	Val	Met	Ile
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Leu	Gln	Leu	Leu	Glu 165	His	Ser	Leu	Ser	Thr 170	Asn	Lys	Leu	Glu	L y s 175	Gln
Ile	Leu	Asp	Gln 180	Thr	Ser	Glu	Ile	Asn 185	Lys	Leu	Gln	Asp	L y s 190	Asn	Ser
Phe	Leu	Glu 195	Lys	Lys	Val	Leu	Ala 200	Met	Glu	Asp	Lys	His 205	Ile	Ile	Gln
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Thr	Val	Asn	Asn 260	Leu	Leu	Thr	Met	Met 265	Ser	Thr	Ser	Asn	Ser 270	Ala	Lys
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Phe 305	Pro	Asn	Ser	Thr	Glu 310	Glu	Ile	Lys	Ala	Ty r 315	Cys	Asp	Met	Glu	Ala 320
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Asp	Phe	Gln	Arg 340	Thr	Trp	Lys	Glu	Tyr 345	Lys	Val	Gly	Phe	Gly 350	Asn	Pro
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gca gcc tat aac aac ttt cgg aag agc atg gac agc ata gga aag aag 9 Ala Ala Tyr Asn Asn Phe Arg Lys Ser Met Asp Ser Ile Gly Lys Lys 20 25 30	96								
caa tat cag gtc cag cat ggg tcc tgc agc tac act ttc ctc ctg cca 14 Gln Tyr Gln Val Gln His Gly Ser Cys Ser Tyr Thr Phe Leu Leu Pro 35 40 45	44								
gag atg gac aac tgc cgc tct tcc tcc agc ccc tac gtg tcc aat gct 19 Glu Met Asp Asn Cys Arg Ser Ser Ser Ser Pro Tyr Val Ser Asn Ala 50 55 60	92								
gtg cag agg gac gcg ccg ctc gaa tac gat ttc tct tcc cag aaa ctt 24 Val Gln Arg Asp Ala Pro Leu Glu Tyr Asp Phe Ser Ser Gln Lys Leu 65 70 75 80	40								
caa cat ctg gaa cat gtg atg gaa aat tat act cag tgg ctg caa aaa 28 Gln His Leu Glu His Val Met Glu Asn Tyr Thr Gln Trp Leu Gln Lys 85 90 95	38								
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cag cag aat gca gtt cag aac cac acg gct acc atg ctg gag ata gga 38 Gln Gln Asn Ala Val Gln Asn His Thr Ala Thr Met Leu Glu Ile Gly 115 120 125	34								
acc agc ctc ctc tct cag act gca gag cag acc aga aag ctg aca gat 43 Thr Ser Leu Leu Ser Gln Thr Ala Glu Gln Thr Arg Lys Leu Thr Asp 130 135 140	32								
gtt gag acc cag gta cta aat caa act tct cga ctt gag ata cag ctg 48 Val Glu Thr Gln Val Leu Asn Gln Thr Ser Arg Leu Glu Ile Gln Leu 145 150 150	80								
ctg gag aat tca tta tcc acc tac aag cta gag aag caa ctt ctt caa 52 Leu Glu Asn Ser Leu Ser Thr Tyr Lys Leu Glu Lys Gln Leu Leu Gln 165 170 175	28								
cag aca aat gaa atc ttg aag atc cat gaa aaa aac agt tta tta gaa 57 Gln Thr Asn Glu Ile Leu Lys Ile His Glu Lys Asn Ser Leu Leu Glu 180 185 190	76								
cat aaa atc tta gaa atg gaa gga aaa cac aag gaa gag ttg gac acc 62 His Lys Ile Leu Glu Met Glu Gly Lys His Lys Glu Glu Leu Asp Thr 195 200 205	24								
tta aag gaa gag aaa gag aac ctt caa ggc ttg gtt act cgt caa aca 67 Leu Lys Glu Glu Lys Glu Asn Leu Gln Gly Leu Val Thr Arg Gln Thr 210 215 220	72								
tat ata atc cag gag ctg gaa aag caa tta aac aga gct acc acc aac 72 Tyr Ile Ile Gln Glu Leu Glu Lys Gln Leu Asn Arg Ala Thr Thr Asn 225 230 235 240	20								

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	agt Ser															768	
	ctt Leu															816	
	aga Arg															864	
	ggt Gly 290															912	
	gaa Glu															960	
	act Thr	_				_	_	-		_		-			-	1008	
	tgg Trp															1056	
	ctg Leu															1104	
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	tat Tyr															1200	
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	cac His															1296	
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Glu	Met 50	Asp	Asn	Сув	Arg	Ser 55	Ser	Ser	Ser	Pro	Ty r 60	Val	Ser	Asn	Ala
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Thr	Ser 130	Leu	Leu	Ser	Gln	Thr 135	Ala	Glu	Gln	Thr	Arg 140	Lys	Leu	Thr	Asp
Val 145	Glu	Thr	Gln	Val	Leu 150	Asn	Gln	Thr	Ser	Arg 155	Leu	Glu	Ile	Gln	Leu 160
Leu	Glu	Asn	Ser	Leu 165	Ser	Thr	Tyr	Lys	Leu 170	Glu	Lys	Gln	Leu	Leu 175	Gln
Gln	Thr	Asn	Glu 180	Ile	Leu	Lys	Ile	His 185	Glu	Lys	Asn	Ser	Leu 190	Leu	Glu
His	Lys	Ile 195	Leu	Glu	Met	Glu	Gly 200	Lys	His	Lys	Glu	Glu 205	Leu	Asp	Thr
Leu	L y s 210	Glu	Glu	Lys	Glu	Asn 215	Leu	Gln	Gly	Leu	Val 220	Thr	Arg	Gln	Thr
Ty r 225	Ile	Ile	Gln	Glu	Leu 230	Glu	Lys	Gln	Leu	Asn 235	Arg	Ala	Thr	Thr	Asn 240
Asn	Ser	Val	Leu	Gln 245	Lys	Gln	Gln	Leu	Glu 250	Leu	Met	Asp	Thr	Val 255	His
Asn	Leu	Val	Asn 260	Leu	Cys	Thr	Lys	Glu 265	Gly	Val	Leu	Leu	L y s 270	Gly	Gly
Lys	Arg	Glu 275	Glu	Glu	Lys	Pro	Phe 280	Arg	Asp	Сув	Ala	Asp 285	Val	Tyr	Gln
Ala	Gl y 290	Phe	Asn	Lys	Ser	Gly 295	Ile	Tyr	Thr	Ile	Ty r 300	Ile	Asn	Asn	Met
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Trp	Thr	Val	Ile	Gln 325	His	Arg	Glu	Asp	Gly 330	Ser	Leu	Asp	Phe	Gln 335	Arg
Gly	Trp	Lys	Glu 340	Tyr	Lys	Met	Gly	Phe 345	Gly	Asn	Pro	Ser	Gly 350	Glu	Tyr
Trp	Leu	Gly 355	Asn	Glu	Phe	Ile	Phe 360	Ala	Ile	Thr	Ser	Gln 365	Arg	Gln	Tyr
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Tyr	Leu	Lys	Gly	His 405	Thr	Gly	Thr	Ala	Gly 410	Lys	Gln	Ser	Ser	Leu 415	Ile
Leu	His	Gly	Ala 420	Asp	Phe	Ser	Thr	L y s 425	Asp	Ala	Asp	Asn	Asp 430	Asn	Cys
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-continued

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435
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What is claimed is:

- 1. An isolated nucleic acid molecule encoding a fusion protein, wherein said fusion protein comprises a modified TIE-2 ligand 2 protein and human immunoglobulin gamma-1 constant region (IgG1 Fc), wherein TIE-2 ligand 2 comprises an N-terminal domain, a coiled-coil domain, and C-terminal fibrinogen-like domain, and the modified TIE-2 ligand protein has the N-terminal and coiled-coil domains deleted and the fibrinogen-like domain comprising amino acids 281–496 of SEQ ID NO: 6.
- 2. A vector comprising the nucleic acid molecule of claim
- 3. The vector of claim 2, operatively linked to an expression control sequence capable of directing its expression in a host cell.

- 4. The vector of claim 3, which is a plasmid.
- 5. An isolated host-vector system for the production of a modified TIE-2 ligand 2, comprising the vector of claim 2 in a host cell.
 - 6. The isolated host-vector system of claim 5, wherein the host cell is a bacterial, yeast, insect, or mammalian cell.
 - 7. A method for producing a modified TIE-2 ligand 2 protein, comprising growing the isolated host-vector system of claim 6 under conditions permitting production of a modified TIE2-ligand 2 protein, and recovering the polypeptide so produced.

* * * * *



专利名称(译)	表达配体 - 血管细胞间信号分子										
公开(公告)号	<u>US6825008</u>	公开(公告)日	2004-11-30								
申请号	US10/225060	申请日	2002-08-21								
[标]申请(专利权)人(译)	DAVIS SAMUEL YANCOPOULOS GEORGEÐ										
申请(专利权)人(译)	DAVIS SAMUEL YANCOPOULOS GEORGE D.										
当前申请(专利权)人(译)	REGENERON制药公司.										
[标]发明人	DAVIS SAMUEL YANCOPOULOS GEORGE D										
发明人	DAVIS, SAMUEL YANCOPOULOS, GEORGE D.										
IPC分类号		A61P7/02 A61P9/00 A61P17/0 C12N1/21 C12N5/02 C12N5/1	2 A61P27/00 A61P29/00 A61P35/00 0 C12N15/09 C12N15/12 C12N15/62								
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审查员(译)	陈,CHRISTINA										
其他公开文献	US20030092891A1										
外部链接	Espacenet USPTO										

摘要(译)

本发明提供了修饰的TIE-2配体,其通过添加,缺失或取代一个或多个氨基酸或通过标记,例如人IgG-1的Fc部分而改变,但保留了它结合TIE-2受体的能力。本发明进一步提供了修饰的TIE-2配体,其是嵌合TIE-2配体,其包含至少一部分第一TIE-2配体和一部分第二TIE-2配体,其不同于第一配体。在一个具体实施方案中,本发明还提供嵌合TIE配体,其包含至少一部分TIE-2配体-1和一部分TIE-2配体-2。此外,本发明提供了编码所述修饰的TIE-2配体的分离的核酸分子。本发明还提供治疗组合物以及阻断血管生长的方法,促进新血管形成的方法,促进表达TIE受体的细胞生长或分化的方法,阻断生长或分化的方法。表达TIE受体的细胞和减轻或预防人体肿瘤生长的方法。

