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(54) **N-GLYCAN CORE  
BETA-GALACTOSYLTRANSFERASE AND  
USES THEREOF**

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(57) **ABSTRACT**

§ 371 (c)(1),  
(2), (4) Date: **Nov. 25, 2011**

The present invention relates to new galactosyltransferases, nucleic acids encoding them, as well as recombinant vectors, host cells, antibodies, uses and methods relating thereto.

Fig. 1

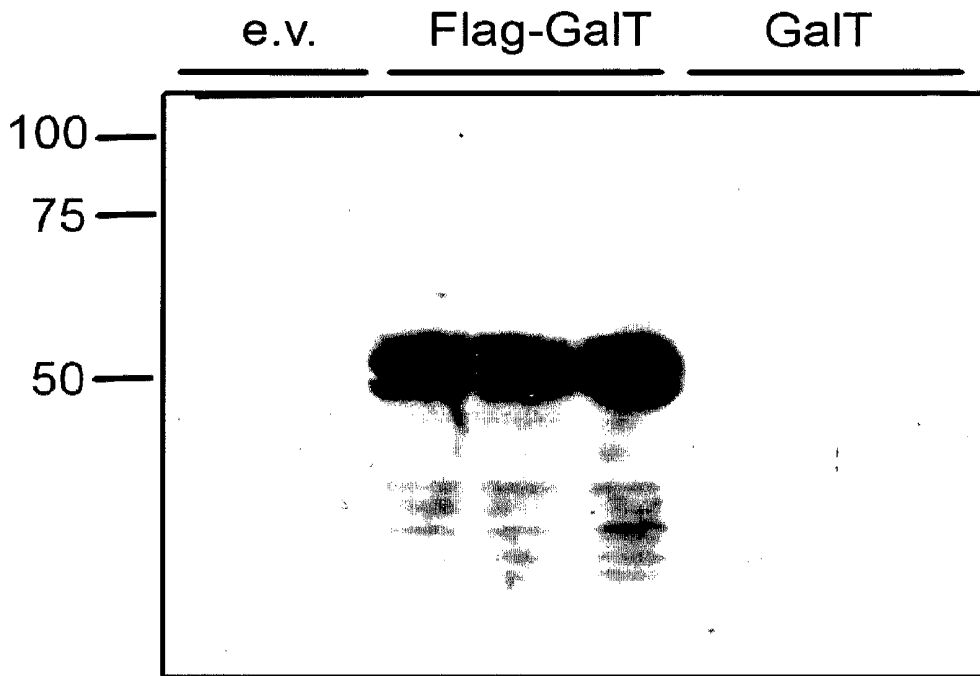


Fig. 2

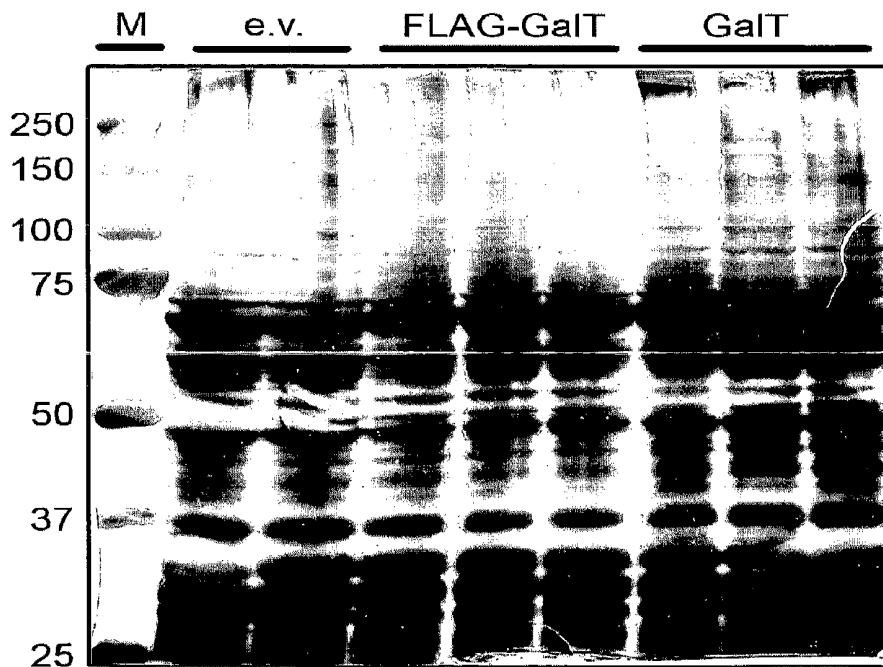


Fig. 3

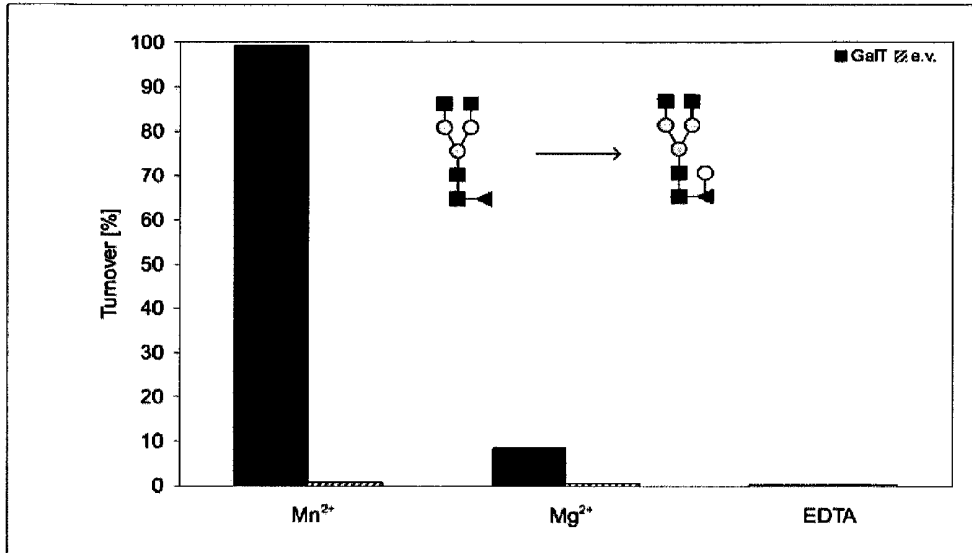


Fig. 4

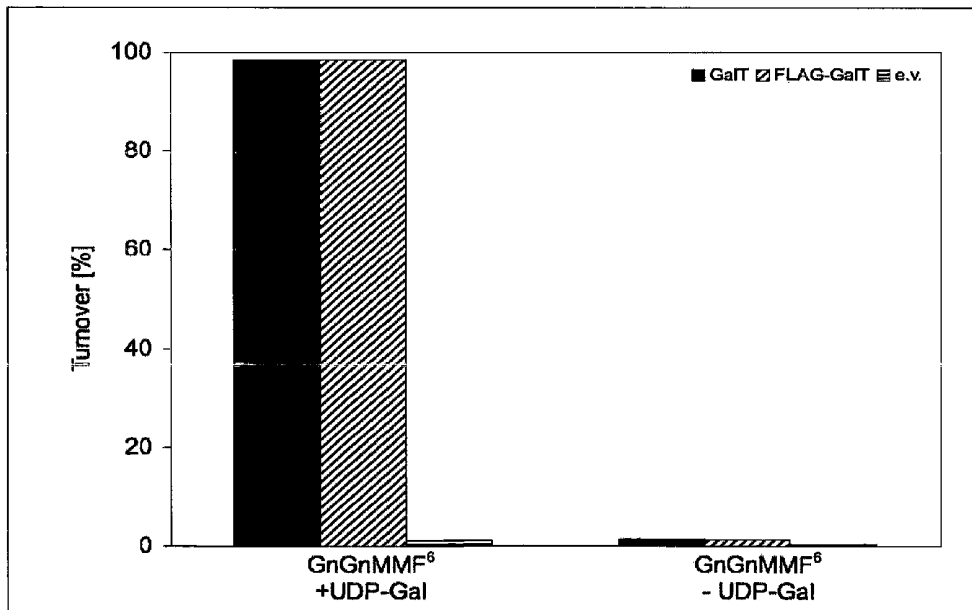


Fig. 5

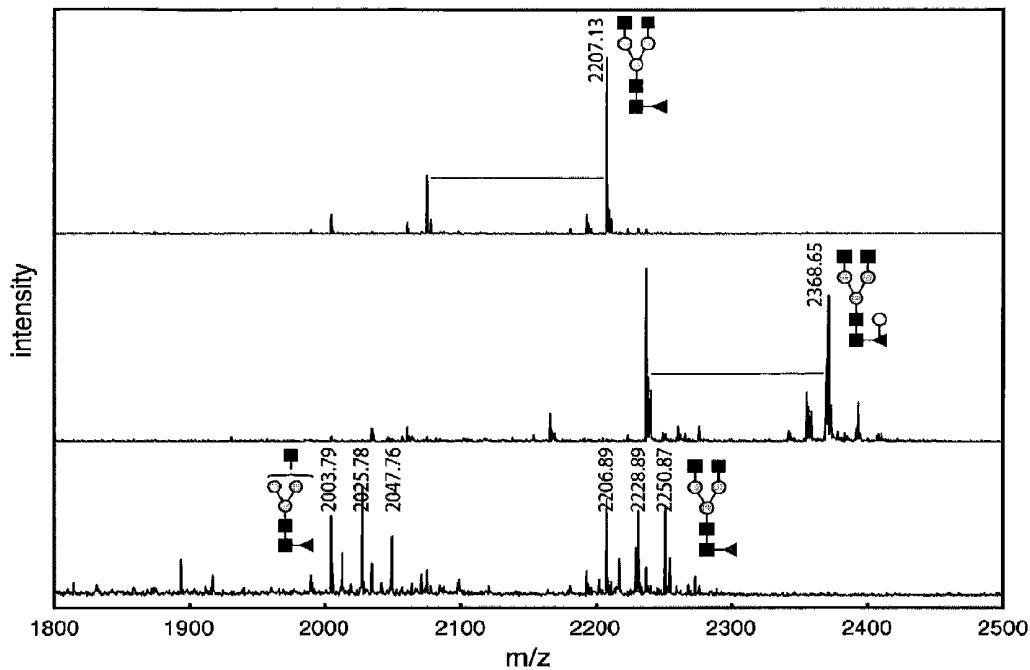


Fig. 6

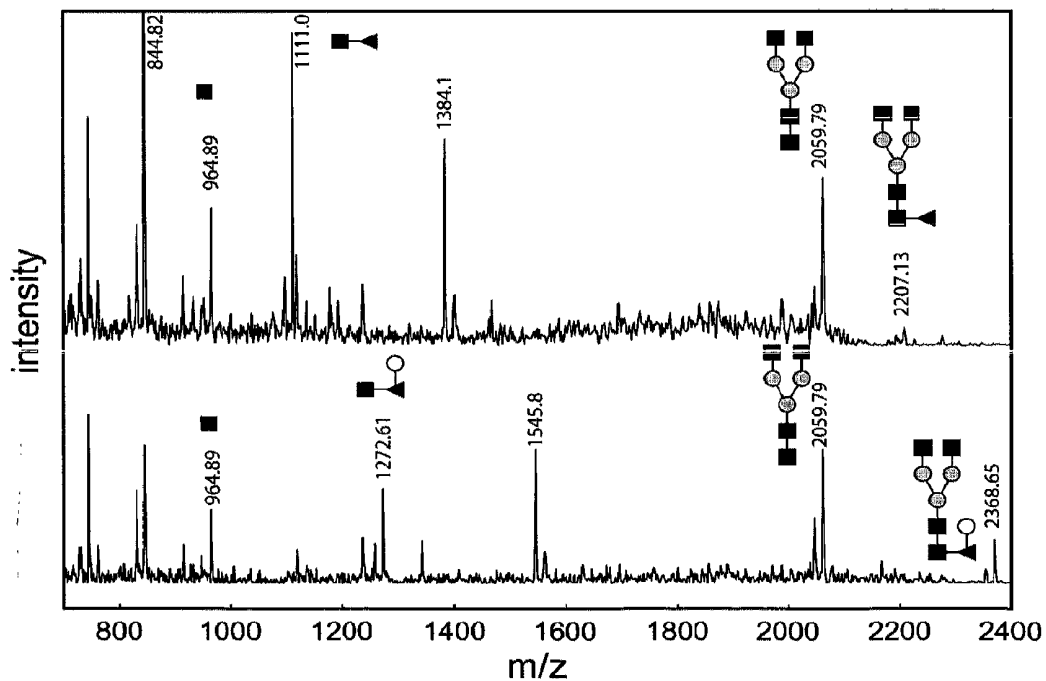


Fig. 7

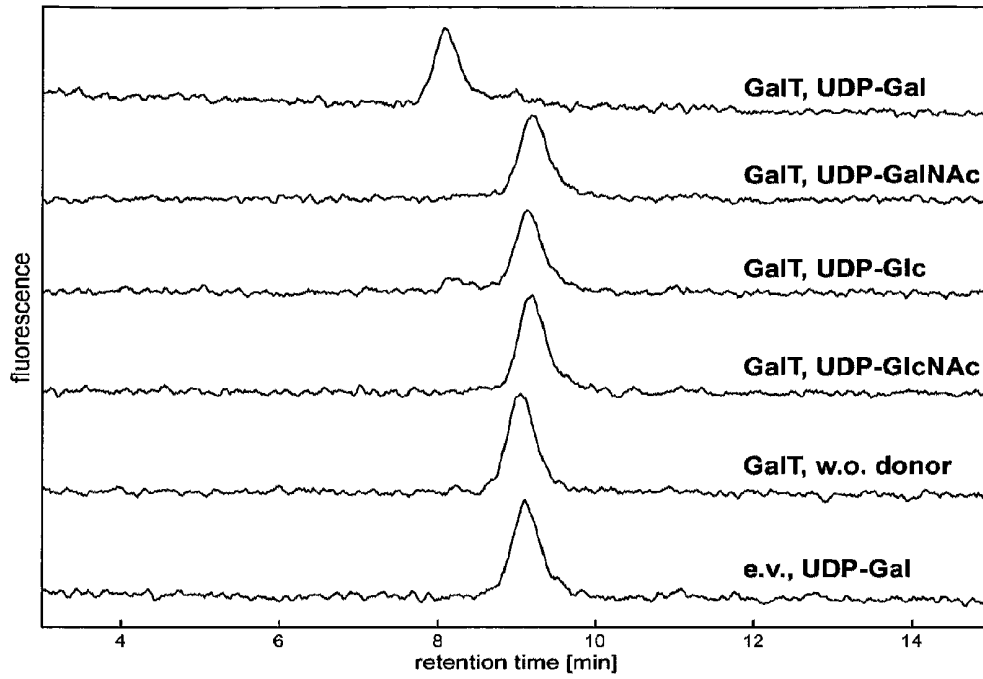


Fig. 8

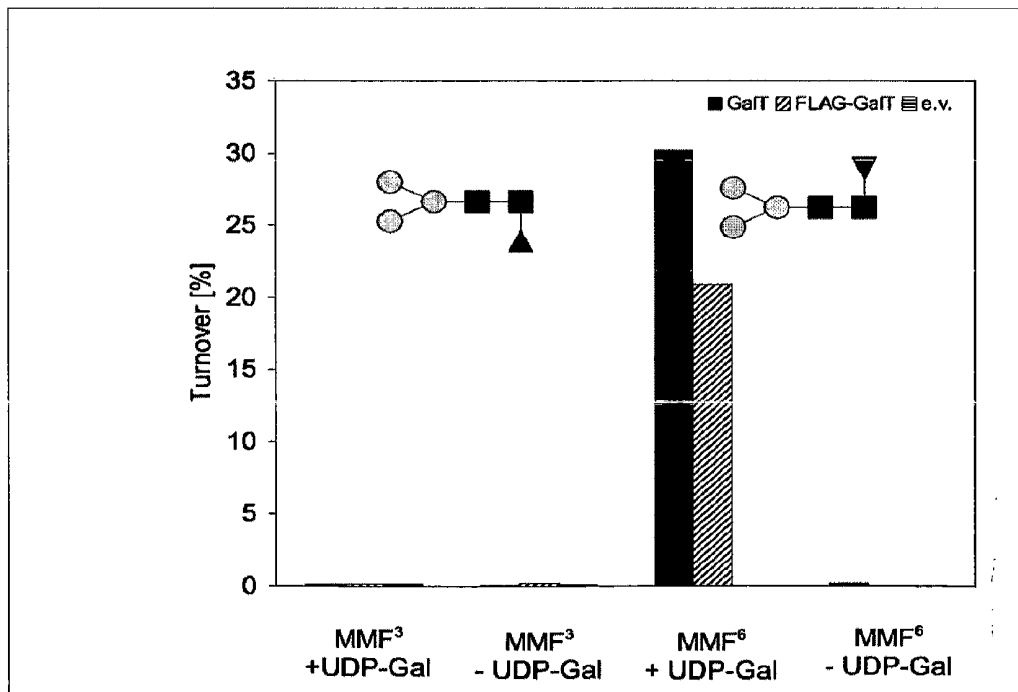


Fig. 9a

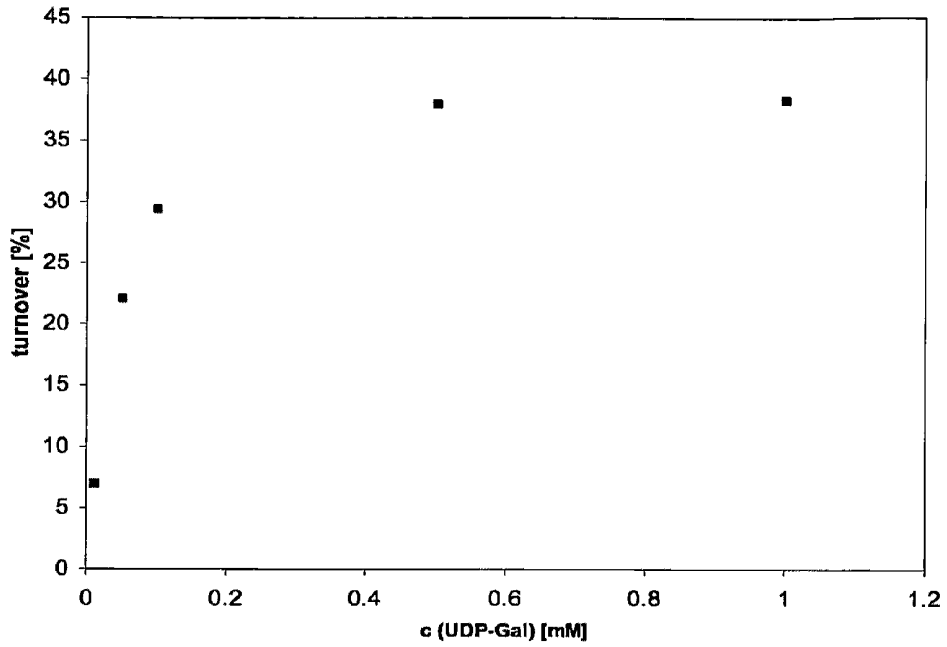


Fig. 9b

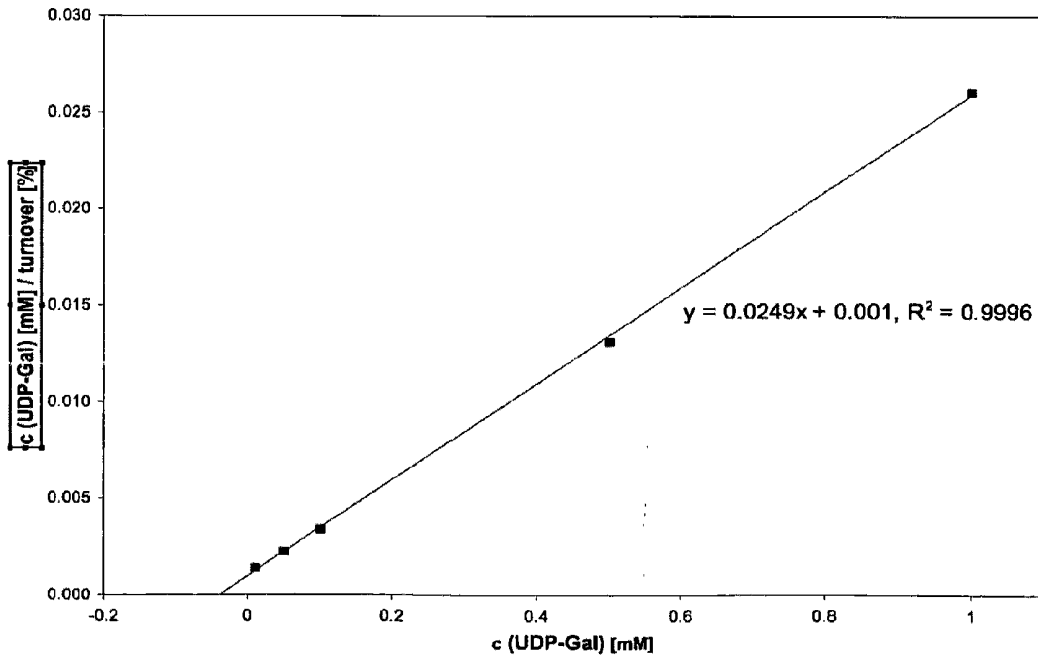


Fig. 10

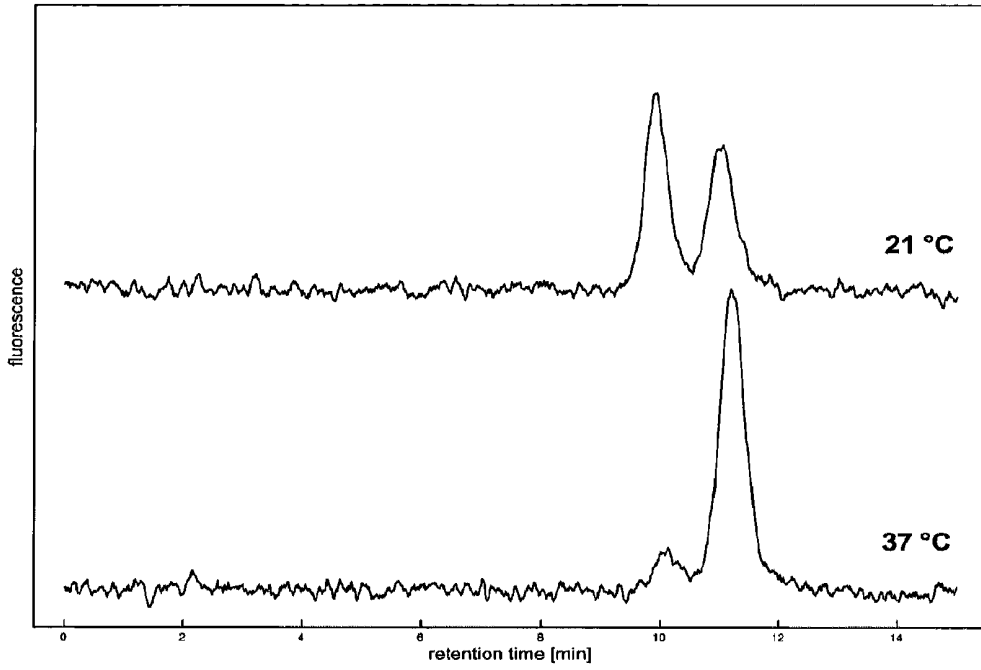


Fig. 11

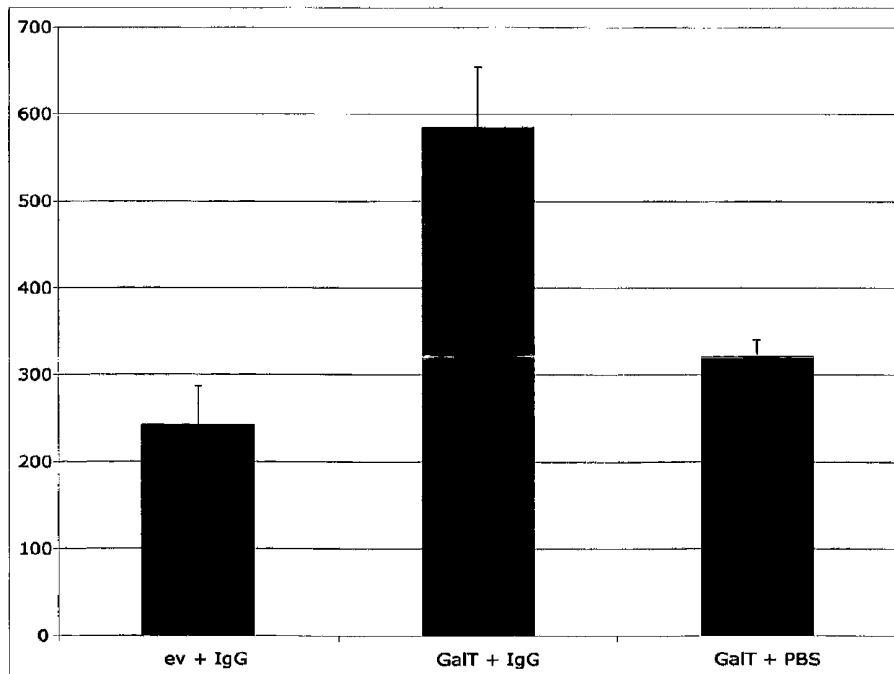
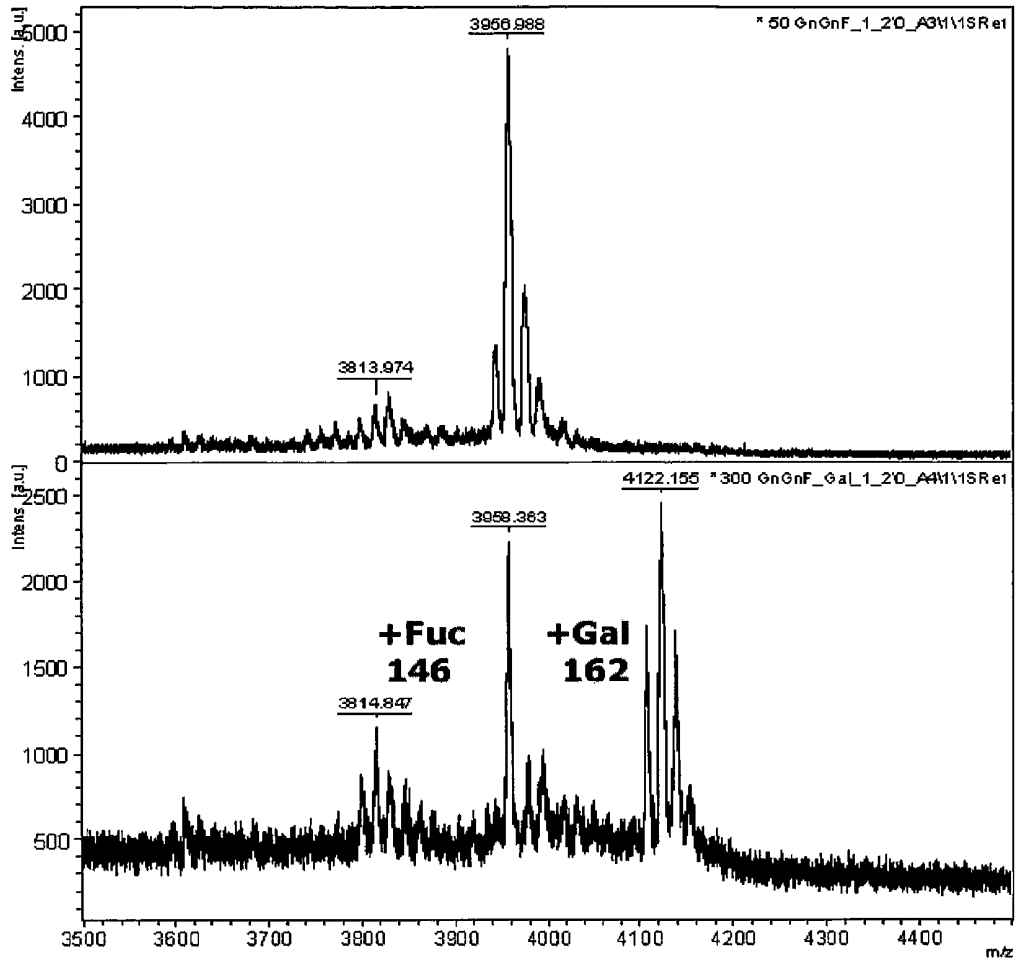


Fig. 12



**N-GLYCAN CORE  
BETA-GALACTOSYLTRANSFERASE AND  
USES THEREOF**

**[0001]** The present invention relates to new galactosyltransferases, nucleic acids encoding them, as well as recombinant vectors, host cells, antibodies, uses and methods relating thereto.

**[0002]** The “roundworms” or “nematodes” are the most diverse phylum of pseudocoelomates and one of the most diverse of all animals. Nematode species are difficult to distinguish; over 80,000 have been described, of which over 15,000 are parasitic. It has been estimated that the total number of roundworm species might be more than 500,000. Nematodes are ubiquitous in freshwater, marine and terrestrial environments. The many parasitic forms include pathogens in most plants, animals and also in humans.

**[0003]** *Caenorhabditis elegans* is a model nematode and is unsegmented, vermiform, bilaterally symmetrical, with a cuticle integument, four main epidermal cords and a fluid-filled pseudocoelomate cavity. In the wild, it feeds on bacteria that develop on decaying vegetable matter. Hannemann et al. (Glycobiology, 16, 874, 2006) isolated and structurally characterized D-galactopyranosyl- $\beta$ -1,4-L-fucopyranosyl- $\alpha$ -1,6-D-GlcNAc (Gal-Fuc) epitopes at the core of N-glycans from *Caenorhabditis elegans*. The N-glycosylation pattern of *Caenorhabditis elegans* was recently reviewed in Paschinger et al. (Carbohydrate Res., 343, 2041, 2008).

**[0004]** It is the object of the present invention to provide new means for the recombinant production of Gal-Fuc-containing (poly/oligo)saccharides and Gal-Fuc-containing glycoconjugates. An additional object is to provide new uses for Gal-Fuc-containing poly/oligosaccharides and Gal-Fuc-containing glycoconjugates.

**[0005]** In a first aspect, the object is solved by an isolated and purified nucleic acid selected from the group consisting of:

**[0006]** (i) a nucleic acid comprising at least a nucleic acid sequence selected from the group consisting of nucleic acid sequences listed in SEQ ID NOs: 1, 3, 5, 7 and 9, preferably SEQ ID NO 1;

**[0007]** (ii) a nucleic acid having a sequence of at least 60, 65, 70 or 75% identity, preferably at least 80, 85 or 90% identity, more preferred at least 95% identity, most preferred at least 98% identity with a nucleic acid sequence selected from the group consisting of nucleic acid sequences listed in SEQ ID NOs 1, 3, 5 and 7, preferably SEQ ID NO: 1;

**[0008]** (iii) a nucleic acid that hybridizes to a nucleic acid of (i) or (ii);

**[0009]** (iv) a nucleic acid, wherein said nucleic acid is derivable by substitution, addition and/or deletion of one of the nucleic acids of (i), (ii) or (iii);

**[0010]** (v) a fragment of any of the nucleic acids of (i) to (iv), that hybridizes to a nucleic acid of (i).

**[0011]** In a preferred aspect the isolated and purified nucleic acid selected from the group consisting of:

**[0012]** (i) a nucleic acid comprising at least a nucleic acid sequence selected from the group consisting of nucleic acid sequences listed in SEQ ID NOs: 1, 3, 7 and 9 as well as the first 1428 nucleic acids of SEQ ID NO: 5, preferably SEQ ID NO 1;

**[0013]** (ii) a nucleic acid having a sequence of at least 60, 65, 70 or 75% identity, preferably at least 80, 85 or 90% identity, more preferred at least 95% identity, most preferred at least 98% identity with a nucleic acid sequence selected from the group consisting of nucleic acid sequences listed in SEQ ID NOs 1, 3 and 7 as well as the first 1428 nucleic acids of SEQ ID NO: 5, preferably SEQ ID NO: 1;

**[0014]** (iii) a nucleic acid that hybridizes to a nucleic acid of (i) or (ii);

**[0015]** (iv) a nucleic acid, wherein said nucleic acid is derivable by substitution, addition and/or deletion of one of the nucleic acids of (i), (ii) or (iii);

**[0016]** (v) a fragment of any of the nucleic acids of (i) to (iv), that hybridizes to a nucleic acid of (i).

**[0017]** Preferably, the above nucleic acids encode a polypeptide of the invention, preferably one having an enzymatic galactosyltransferase activity, more preferably one having a  $\beta$ -1,4-galactosyltransferase activity, preferably one with L-fucoside-, more preferably one with  $\alpha$ -L-fucoside-, more preferably one with Fuc- $\alpha$ -1,6-GlcNAc— and most preferably one with GnGnF<sup>6</sup>— (nomenclature according to Schachter, Biochem. Cell. Biol. 64(3), 163-181, 1986) containing poly/oligosaccharides or glycoconjugates as acceptor substrates.

**[0018]** Galactosyltransferase activity, as used herein, is meant to describe an enzymatic transfer of a galactose residue from an activated donor form (i.e. nucleotide-activated galactose, preferably UDP-Gal) to an acceptor.  $\beta$ -1,4-Galactosyltransferase activity, as used herein, is meant to describe the specificity of the galactosyltransferase activity, i.e. the transfer of galactose in a beta 1,4-configuration onto an acceptor molecule.  $\beta$ -1,4-Galactosyltransferase activity on L-fucosides as acceptor substrate, as used herein, is meant to describe the specificity of the galactosyltransferase activity in a beta-linked 1,4-transfer onto L-fucosides as the acceptor substrate. L-fucosides, as meant herein, are meant to describe poly/oligosaccharides or glycoconjugates as acceptor substrates containing terminal L-fucose in alpha, most preferably in alpha-1,6 configuration, e.g. as part of MMF6 or GnGnF<sup>6</sup> (Schachter, Biochem. Cell. Biol. 64(3), 163-181, 1986).

**[0019]** In a most preferred embodiment, the encoded polypeptide comprises a polypeptide sequence selected from the group consisting of polypeptide sequences listed in SEQ ID NOs 2, 4, 6, 8 and 10, preferably SEQ ID NO: 2, or a functional fragment or functional derivative of any of these.

SEQ ID NO: 1 is the nucleic acid sequence coding for SEQ ID NO 2:  
(also listed in NCBI as Ref Seq NM\_072144.4 and in Wormbase as M03F8.4; coding for galactosyltransferase [referred to as GalT in the Examples section] from *Caenorhabditis elegans*)

ATGCCTCGAATCACCGCCAGTAAATAGTCTTCTAATTGCATTATCATTTTGTATTA

CTGTTATTTATCACTTTCCAATAGCAACGAGAAGCAGTAAGGAGTACGATGAATATG

-continued

GAAATGAATATGAAAACGTTGCATCGATAGAGTCGGATATAAAAAATGTACGTCGAT  
TACTTGACGAGGTACCGGATCCCTCACAAAACCGTCTACAATTCCTGAAACTTGATG  
AGCATGCTTTTGCATTCTCGGCCCTACACAGACGATCGAAATGGAAATATGGGGTAC  
AAATATGTCCGAGTCCTGATGTTTATCACGTACAAGACAACCTTTTCCTGTGAAATAA  
ACGGGAGAAAGTCCACAGATGTATCACTTTACGAGTTCCTCGGAAAAATCACAAAATGA  
AGTGGCAAATGTTTATTTGAATGTAAACTACCCGATGGTATAGATTTCAATAATGT  
TAGCTCTGTAAGGTCATAAGAAGCACAACCAAGCAGTTTGTGATGTGCCGATTCTG  
GTATAGAATTCAAGATGAGAAAAATAATTACGCCAGACGAATATGACTATAAAATGTCA  
ATTTGTGTTCCAGCATTGTTIGGAAATGGATATGATGCAAAGCGAATTGTTGAGITTA  
TTGAGCTGAATACTTTGCAAGGAATCGAGAAAATATACATTTACACTAATCAAAAAGA  
GCTTGATGGATCCATGAAGAAAACGTTGAAATACTATTTCGGATAATCACAAAATAAC  
ATTAATTGATTACACATTACCATTACAGAGAGGATGGTGTGTTGGTATCACGGGCAATT  
GGCAACTGTTACTGATTGTTTACTGAGAAACTGGAATCACAAAATACACATTTTTTC  
AATGATTTTGATGAGTTCTTCGTCCTCCCGTTATCAAAAGTCGGACTCTCTTTGAAACAA  
TCAGTGGGCTTTTTGAAGATCCCACTATTGGATCGCAACGAACAGCTTTGAAGTATA  
TAAATGCAAAAATCAAGAGCGCTCCGTATTCAGTAAAAATATGTTTCCGAAAAAC  
GAATTGAAACAAGATTCACGAAATGTGTAGTTCGACCGGAAATGGTTTTTGAACAGG  
GTATTCATCATACGAGTAGAGTGATTCAAGACAACATAAAAACGGTTTCCCATGGCG  
GATCCCTTCTACGGGTTTATCATTACAAGGATAAAAAGTATTGTTGCGAAGACGAGA  
GCCTCTTGAAAAACGGCATGGAGATCAACTTCGGGAAAAATTCGATTCAGTTGTTG  
GTCTTTTAGACTTG TAG

SEQ ID NO: 2 (also listed in NCBI Ref Seq NP\_504545.2)  
MPRITASKIVLLIALSFCTVIYHFPIATRSSKEYDEYNEYENVASIESDIKNVRRLLD

EVPDPSONRQLQFLKLEHAFAPFAYTDDRNGNMGYKYVRVLMFITSQDNFSCEINGRK  
STDVSLYEFSENHMKWQMPILNCKLPDGDIDFNVS SVKVIIRSTTKQFVDVPIRYIQDE  
KIITPDEYDYKMSICVPALFNGYDAKRIVEFIELNTLQIEKIYIYTNQKELDGSMMKTLK  
YYSNHIKITLIDYTLFPREDGVVYHQLATVTDCLLRNTGITKYTFNDFEFFVPVIKSR  
TLFETISGLFEDPTIGSQRTALKYINAKIKSAPYSLKNIVSEKRIETRFKCVVRPEMVFEO  
GIHHTSRVIQDNYKTVSHGGSLLRVYHYKDKKYCCEDSLKKRHGDQLREKFD SVVG  
LLDL

SEQ ID NO: 3 is the nucleic acid sequence coding for SEQ ID NO: 4:  
(also listed in NCBI Ref Seq XM\_001674213.1; coding for galactosyltransferase from  
*Caenorhabditis briggsae*)

ATGCCACGAA TAACGGCAAG CAAAATAGTG TTATTATCTG TATTATCCTT  
ACTAACAGTT TTCTATCTGA ATACATTTTC GTCTATTAAA ATTGAAAACG  
ATCTCGACGG GACTGATTAC GACTTGGATT ACATAGAATC TGATATCAAA  
AAGACGCGTC GATTACTCAA TGAATCCCT GATCCATCTC AAAACCGAGT  
TCAATTTTTT AAACGATG ATAATGGATA TGCATTCTCA GCATATACAG  
ATAATAGGAA AGGAAATATG GGTACAAAAT ATGTCAGAAT ATTAGTGTTC  
CTAACTAAAT TTGATGATTT TTCTTGGCAA ATTAACGCGA AGAAATCCTA  
TGTGTTTACA CTCTACGAGC TATCAGAAA TCACAATATG AAGTGGAAAA

-continued

TGTATATTTT GAATTGTTTA CTTCCCGATG GAATCACTTT CAACGATGTG  
 AATTCTGTAA AAATATCTAG AAGTTCTTCA AAACCTTCAG TCCAAATCCC  
 GATCAGATAT AGAATTC AAG ATGAGAAAAT GATGACTCCA GATGAATACG  
 ATTATAAGTT GTCGATTTGT GTTCCTGCAC TTTTGGAAA CGTTTATTAT  
 CCAAGGAGGA TTATTGAATT TGTGGAAC TAACAGCTTGC AAGACATCGA  
 CAAAATCTAC ATCTACTACA ATCCTTTAGA AATGACAGAT GAGGCCACAG  
 AAAGGACTTT GAAGTTTTAT TCCAATAATG GGAAAATCAA TTTAATAGAA  
 TTCATTCTCC CATTCTCTAC TCGAGATGTT TGGTATTATG GGCAATTGGC  
 CACCGTTACA GATTGTCTTC TCCGTAACAC TGAATAACT CAATACACAT  
 TTTCAATGA TTTGGATGAA TTTTTCGTGC CAGTACTGGA CAACCAAAT  
 CTCTCTGAAA CTGTGTCAGG ATTATTTGAA AATCGAAAAA TTGCCTCTCA  
 GAGAACGGCC TTGAAATTTA TTAGTACAAA AATCAATCGA TCTCTGTAA  
 CTCTCAATAA TATTGTGTCT TCTAAAAAAT TTGAAACGAG ATTCACAAAA  
 TGCCTCGTAC GGCCGGAAAT GGTTTTTGAG CAGGGCATTG ACCATACGAG  
 TAGAGTAATA CAAGACGACT ACGAAACCCC ATCCCATGAT GGATCACTTT  
 TGCCTGTGTA TCACTACAGA GAACCAAGAT ATTGCTGCGA AAACGAGAAT  
 CTTCTAAAAC AAAGATACGA TAAGAAGCTT CAAGAAGTTT TTGATGCTGT  
 AGTTCTTATA TTGCATGTCA CATTGATGT ATGGATATAT CACCTGAAAA  
 ACACCTCTA A

SEQ ID NO: 4 (also listed in NCBI Ref Seq XP\_001674265.1)  
 MPRI TASKIV LLSVLSLLTV FYLNTFSSIK IENDLDGTDY DLDYIESDIK KTRRLLEIIP

DPSQNRVQFF KLDNDGYAPS AYTDNRKGNM GHKYVRILVF LTKFDFDSCF  
 INSKSYVVT LYELSENHNM KWKMYILNCL LPDGITFNDV NSVKISRSSS  
 KLSVQIPYRIY RIQDEKMMTP DEYDYKLSIC VPALFGNVVY PRRIIEFVEL NSLQDIDKIY  
 IYYNPLEMTD EATERTLKFY SNNKINLIE FILPFSTRDV WYQGQLATVT  
 DCLLRNTGIT QYTFNDLDE FFVPVLDNQT LSETVSGLFE NRKIASQRTA  
 LKFISTKINR SPVTLNIVS SKNFETRFTK CVVRPEMVPE QGIHHTSRVI  
 QDDYETPSHD GSLLRVYHYR EPRYCCENEN LLKQRYDKKL QEVFPAVLI  
 LHVTFDVWIY HLNKTL

SEQ ID NO: 5 is the nucleic acid sequence coding for SEQ ID NO: 6 (1428 nucleic acids) followed by a stop codon and further 68 nucleotides: (also listed in NCBI Ref Seq XM\_001629141.1; coding for galactosyltransferase from *Nematostella vectensis*)  
 ATGCGATGCT ATATTACAA ATTGAGGTG TCCGTTTGTG TGTTGTAGT

GCTCTTACACA GCACTGCTTT TCATCACCTA TTTAAACCAC TCAGAGCTTG  
 AATCAGCAGA GAAAAGTAGC GGAAAAGGA AGACGCGACA TCGTAAACGA  
 ACACGTTTAC GCAAACAACA CGAGAGCCAT TTTAGAAAAG CTCGACTACA  
 AGAAAAGAGAA CTAGTATTAA GATCTACAGC GCCACCAACA TTACGAAGAG  
 AAGTACAAGC GCATCGATTA GGGCAGATCC GTGGCAAGAA CACGGACCAG  
 GGGATAACTG GAAAGTTCAC AGAGATCGCT AAAGACACGC ATATTTATTC  
 AGCGTTTTAC GACGATGCCA AGTCAAATCC ATTCATTCTG CTTATCATCC

-continued

TCTCGGGAAA ACACTACCAG CCTGGATTAT CTTGCCAATT TTGCGAACCT  
 TTGTCGCCCA GTTGTAGTTT TGCGGACTCT AAAGCTGAAT ACTACACGAC  
 CAACGAGAAC CATGGGAGAG TATTTGGCGG GTTCATTGCG AGTTGCCCTCG  
 TGCCTGATGG ATTCAATGCA GTGCCATGT TTGTTGACAT AACGGCCGAT  
 GTTAAGGGGG AGAAAAGCAA GGCACGGGTA CCTGTGGTGT CTAATGCACA  
 TCTCTACTAC CCTATTAAAT ACGCAATCTG CGTCCCACCC CTCCGATCAG  
 AGAAACTAAC AGCGAAAAGA CTCATAGAGT TTGTCGAGCT AACCAAATT  
 TTAGGCGCTA ACCATTTTAC TTTTATGAC TTCAAACCG ACCCGGAAGT  
 CAATAACGTT TTAAGATATT ACCAGGAGAC ACAAGTAGCA AATGTTCTGC  
 CATGGAATCT ACCTTCAAAT TTGGTATCCA GGCCGAACGA TATTTGGTAC  
 TTTGGTCAGG TTTTGGCTAT TCTAGATGCG TTGTATCGCT ACAAGAACAG  
 GGCAAAATTT GTAGCCTTCA ATGACGTAGA TGAGTTTATC GTTCCGCTAA  
 GGAACAGCTC GATAGTGGAA ATACTAAACG CGTTTCACCG GCCATACCAC  
 TGTGGACATT GCTTTCAGAG CGTGGTGTTC AGCTCAAACG CGAGATTTCC  
 CAGGCAAAAA AGCGAGTTAG TTTCTCAGCG GTTCTTCCAC AGGACCCAGG  
 AAACCATCCC TCTCCTCTCG AAATGCATTG TGGATCCTTT GAGAGTGTTC  
 GAGATGGGGA TTCACCACAT AAGCAAGGCT ACAGGTCTGC GGTATTCCGT  
 CAACTCAGTA CACGAGAGTG ACGCGGTAT CTTCATTAC AGGACTTGCA  
 CTACGTGATT TGGTATACGT CATCAGTGCA TGAACCTAGT GCATGATGGG  
 ACCATGGCCA AATATGAAA ACGACTTCAG AAAATGTTTA GAAAGTTGT  
 AAATGATTTA AAACCTTTGG CACCAACGTA GCTATTTCTG AACACTTAC  
 ACTTTCATTG TTATAACAGA ATACAGAATA AATTAATGAT TGTGTGCC  
 SEQ ID NO: 6 (also listed in NCBI Ref Seq XP\_001629191)  
 MRCYIYKLR L SVCLFVVLFT ALLFITYLNH SELESAEKSS GKRKTRHRKR  
 TRSRKQHESH FQKARLQERE LVLRSTAPPT LRREVQAHRL GQIRGKNTDQ  
 GITGKFTEIA KDTHIYSAPY DDAKSNPFIR LIILSGKHYQ PGLSCQFCEP  
 LSASCSFADS KAEYYTTNEN HGRVFGGFIA SCLVPDFNA VPLFVDITAD  
 VKGEKSKARV PVSNAHLYY PIKYAICVPP LRSEKLTAKR  
 LIEFVELTKL LGANHFTFYD FKTDPEVNNV LRYQETQVA NVLPWNLPSN  
 LVSRPNDIYW FGQVLAAILDC LYRYKNRAKF VAFNDVDEFI VPLRNSIVE  
 ILNAPHRPYH CGHCFQSVVF SSNARFPRQK SELVSQRFFH RTQETIPLLS  
 KCIVDPLRVF EMGIHHISKA TGLRYSVNSV HESDAVIFHY RTCTTSPGIR  
 HQCMNLVHDG TMAKYGKRLQ KMRKVVNDL KLLAPT  
 SEQ ID NO: 7 is the nucleic acid sequence coding for SEQ ID NO: 8:  
 (also listed in NCBI Ref Seq XM\_002189335, coding for galactosyltransferase from  
*Taeniopygia guttata*)  
 ATGACTGTAA CTTAATGCT TGTGGTTTCT TATCTGAGAT TACAGAGACT  
 TTCTCATCAG CCAAAAGTAA TTCAAGAAAG TAGAAGATGT AGAGGGAAAA  
 TTGCCCTTAG CACAATAACA GCATTGGAAG GTAACAAAAC TGATATTATA  
 TCCCATACT TTGATGACAG AGAAAACAAA ATCACTCGTC TGATTGGGAT  
 TGTTACCCAT AAAGATGTAA AACAACTGTT CTGCTGGTTC TGCTGTCAAG

-continued

CCAATGGAAA GATATATGTA TCAAAAGCAG AAATAGATGT TCACTCGGAT  
 AGATTGGAT TCCCTTATGG TGCAGCAGAT ATAATTTGTT TGGAACCTGA  
 AAACTGTGAT CCAACACATG TATCAATTCA TCAGTCTCCA TATGGAAATA  
 TTGACCAGCT GCCGAGGTTT GAAATTAATA ATCGCAGGCC TGAGACCTTT  
 TCTGTTGACT TCACCGTGTG CATTCTGCC ATGTTTGAA ACTACAACAA  
 TGTCTTGCAG TTTGTACAGA GTATGGAAAT GTATAAGATT CTTGGAGTAC  
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 TTGAAATTTT ATATAGAAGA AGGAACTGTT GAGGTAATTC CCTGGCCAAT  
 AGACTCACAC CTCAGGGTTT CTTCTAAATG GCGCTCATG GAAGACGGGA  
 CACACATTGG CTACTATGGA CAAATCACAG CTCTAAATGA CTGTATATA  
 CGCAACATGG AAAGACCAA GTTTGTGGTC CTTAATGACG CTGATGAAAT  
 AATTCTTCCC CTTAAACACC CAGACTGGAA AACAAATGATG AACAGTCTTC  
 AGGAGCAAAA CCCAGGGACT AGTGTTTTCC TTTTfGAGAA CCATATCTTC  
 CCAGAACTG TATTTTCTCC CATGTTCAAC ATTTTCATCTT GGAATACTGT  
 GCCAGGTGTT AACATATTGC AGCATGTGTA CAGAGAGCCT GACAGGAAAC  
 ATGTAATCAA TCCCAGGAAA ATGATAGTTG ATCCACGAAA GGTGATTGAG  
 ACTTCAGTCC ATTCTGTCTC ACGTGCTTAT GGAAGAGCG TGAATGTTCC  
 CATGGAAGTT GCCCTCATTT ATCACTGTGC GAAGGCCCTT CAAGGAAACC  
 TCCCAGAGA ATCTCTCATC AGGGATACAA CACTGTGGAG ATATAACTCA  
 TCATTAATCA TGAATGTTAA CAAGTTCTA TCTCAAACCA TGCTGCAAAC  
 TCAAAATGA

SEQ ID NO: 8 (also listed in NCBI Ref Seq XP\_002189371)  
 MTVTLMLVVS YLRLQLSHQ PKVIQESRRC RGKIALSTIT ALEGNKTDII  
 SPYFDDRENK ITRLIGIVHH KDVKQLFCWF CCQANGKIYV SKAEIDVHSD  
 RFGFPYGAAD IICLEPENCD PTHVSIHQSP YGNIDQLPRF EIKNRRPETF  
 SVDFTVCISA MFGNYNNVLO FVQSMEMYKI LGVQKVVIYK NNCSHLMEKV  
 LKFYIEEGTV EVIPWPIDSH LRVSSKWRPM EDGTHIGYYG QITALNDCIY  
 RNMERTKFVV LNDADIILP LKHPDWKTM NSLQEQNPQT SVFLFENHIF  
 PETVFSMPFN ISSWNTVPGV NILQHVYREP DRKHVINPRK MIVDPRKVIQ  
 TSVHSLRAY GKSVNVPMEV ALIYHCRKAL QGNLPRESLI RDTTLWRVNS  
 SLIMNVNKVL SQTMLQTQN

SEQ ID NO: 9 is the nucleic acid sequence coding for SEQ ID NO: 10:  
 (also listed in NCBI Ref Seq XM\_626032, coding for galactosyltransferase from  
*Cryptosporidium parvum*)  
 ATGCAAAGTA AAGTCATTT TAGGATCTTG GTATTGATCA TTCGGGTGAT  
 TGGATCCTTA TACTCAATAA TTCAATTAAT GCTAAAGGAG CTATCAAGTA  
 ACAAAAATAT TCAAGAGGTT AGTCATTCOA GGAGGCTAAT AAGTGAACTT  
 TACAGTGAAA GTATTAATGA ACAAATGAT CAAGATTGGA AAGAACTAAA  
 GCTAATAATT CCAAATCATT CTCAAATTA CCAGCAGGAA AAAAATGGTA  
 ATTTGATTGA GTTTAAAGTT TATATATACT CAGCATATTA TGATTGGAGA

-continued

ATAGATAGGA TACGAATAAA TTCACTTATC CCATCGAATT TTTATGATCG  
 AATAGAAATG GAATGTGCAA TAATCTTGA CAAAAATATT TACACAGGAA  
 CTATTAATAA AGTGATTCAT AAGGAGCACC ATAATAAAGA ATATGTATCA  
 TCGACTTTAC TCTGCGAAAT TGCAAAAAAT GAAATTAAT TTGAGGATAT  
 TTCAAGGAAA GTTTTGATAA CAATTTTGA AAATGGAAAC AGCACAAATA  
 AATCAGAAAT ATGGATAACT CTAAAAAAA TTCCAAAAA TAGCTCTAAT  
 AATCATGAGC TGACTGTTG TGTGAGACCT TGGTGGGGAG AGCCAATAAA  
 GAATGGAAAC TTGGGAAATA AACAAAAATT TAACAATTCA GGGTTAATGC  
 TTGAATTTAT TAATTCATAT TTATCTTAG GAGCAAATAA ATTTATTTA  
 TATCAAAATT ACTTGGACAT TGACGAAGAT GTAAGAAATA TAATAAATTA  
 TTATCTAAT ATCAAAAATG TTTTGGAAAT TATCCATAC TCATTACCAA  
 TAATTCATT TAAACAAGTT TGGGATTTG CACAAACAAC AATGATACAG  
 GACTGCCTAC TAAGAAATAT TGGAAAAACA AAATACTTGT TATTCGTAGA  
 TACCGATGAA TTTGTATTT CAAACTTGAA AAATTATAAC TTAATGGATT  
 TTTTAAATTT ATTAGAAGCC AACAAATCCTT ATTATAAAAA CAAAGTCGGG  
 GCAATGTGGA TTCCAATGTA TTTTCATTTT TTAGAGTGGG AATCTGATAA  
 AAATAATTTG AAGAAATATT CAACAATTGA GAAAAAATT AAGAAAAAGA  
 TGGCAAATAT TGAGTTTGT CTATATCGTA AAACATGTAG AATGTTAAGT  
 TCTGGAACAA AAAAAAGTGA CAAGACGAGA AGAAAAGTTA TTATTAGACC  
 TGAAGAGT TTGTATATGG GTATACATGA AACAGAAGAG ATGCTAAGCA  
 AAAAAATTTCA TTTCATTAGA GCTCCTGTAA TTAATGTGGG TGGAGGAAAC  
 GAAC TAAGTA TATATTTACA TCATTATAGA AAAGCAAAG GTATTGTA  
 CAATGATCCC AAACAAAGAG AACTTGTGAA TATGTATTTA GAAAATGTTT  
 GTTCAGATAA GCTGTTAGAT TCAGGGGGAG ATTCCATTCA AGATGGAGTA  
 ATTGTCGACA ATACTGTTG GGAGATATTT GGAACACACT TATACCAGAT  
 AATTTTGTAG CATATTAAG AAATCCAAGA TATGTACACA AATAAGGAAA  
 TAATTAATGG AAATAAAAA TTAAGTGTG AAGAATTACA TAATTA  
 SEQ ID NO: 10 (also listed in NCBI Ref Seq XP\_626032)  
 MQSKVIFRIL VLIISVIGSL YSIIQLMLKE LSSNKNIQEV SHSRRLISEP YSESINEQND  
 QDWKELKLI PNHSQINQQE KGNLIEFKV YIYSAYDWR IDRIRINSLI PSNFYDRIEM  
 ECAILDKNI YGTIKKVIH KEHKNKEYVS STLLCEIAKN EIKFEDISRK VLITILENGN  
 STNKSEIWIT LKKIPKNSN NHELTVCVRP WWGEPKNGN LGNKQKPNNS  
 GLMLEFINSY LFLGANKFYL YQNYLDDED VRNIINYYSN IKNVLEIIPY SLPIIPFKQV  
 WDFQATTMIQ DCLLRNIGKT KYLLFVDTDE FVFPNLKNYN LMDPLNLLEA  
 NNPPYKKNVGV AMWIPMYFHF LEWESDKNNL KKYSTIEKKI KKKMANIEFV  
 LYRKTCRMLS SGTKSDKTR RKVIIRPERV LYMGIHETEE MLSKPFHFR  
 APVINVGGGN ELSIYLHHYR KAKGIVNNDP KQRELVNMYL ENVCSDKLLD  
 SGGDSIQDGV IVDNTVWEIP GTHLYQIIFE HIKEIQDMYT NKEIINGNKN  
 LSVEELHN

**[0020]** The term “nucleic acid encoding a polypeptide” as it is used in the context of the present invention is meant to include allelic variations and redundancies in the genetic code.

**[0021]** The term “% (percent) identity” as known to the skilled artisan and used herein indicates the degree of relatedness among two or more nucleic acid molecules that is determined by agreement among the sequences. The percentage of “identity” is the result of the percentage of identical regions in two or more sequences while taking into consideration the gaps and other sequence peculiarities.

**[0022]** The identity of related nucleic acid molecules can be determined with the assistance of known methods. In general, special computer programs are employed that use algorithms adapted to accommodate the specific needs of this task. Preferred methods for determining identity begin with the generation of the largest degree of identity among the sequences to be compared. Preferred computer programs for determining the identity among two nucleic acid sequences comprise, but are not limited to, BLASTN (Altschul et al., *J. Mol. Biol.*, 215, 403-410, 1990) and LALIGN (Huang and Miller, *Adv. Appl. Math.*, 12, 337-357, 1991). The BLAST programs can be obtained from the National Center for Biotechnology Information (NCBI) and from other sources (BLAST handbook, Altschul et al., NCB NLM NIH Bethesda, Md. 20894).

**[0023]** The nucleic acid molecules according to the invention may be prepared synthetically by methods well-known to the skilled person, but also may be isolated from suitable DNA libraries and other publicly available sources of nucleic acids and subsequently may optionally be mutated. The preparation of such libraries or mutations is well-known to the person skilled in the art.

**[0024]** In a preferred embodiment, the nucleic acid molecules of the invention are cDNA, genomic DNA, synthetic DNA, RNA or PNA, either double-stranded or single-stranded (i.e. either a sense or an anti-sense strand). The nucleic acid molecules and fragments thereof, which are encompassed within the scope of the invention, may be produced by, for example, polymerase chain reaction (PCR) or generated synthetically using DNA synthesis or by reverse transcription using mRNA from *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* or *Cryptosporidium parvum*.

**[0025]** In some instances the present invention also provides novel nucleic acids encoding the polypeptides of the present invention characterized in that they have the ability to hybridize to a specifically referenced nucleic acid sequence, preferably under stringent conditions. Next to common and/or standard protocols in the prior art for determining the ability to hybridize to a specifically referenced nucleic acid sequence under stringent conditions (e.g. Sambrook and Russell, *Molecular cloning: A laboratory manual* (3 volumes), 2001), it is preferred to analyze and determine the ability to hybridize to a specifically referenced nucleic acid sequence under stringent conditions by comparing the nucleotide sequences, which may be found in gene databases (e.g. <http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=nucleotide>) with alignment tools, such as e.g. the above-mentioned BLASTN (Altschul et al., *J. Mol. Biol.*, 215, 403-410, 1990) and LALIGN alignment tools.

**[0026]** Most preferably the ability of a nucleic acid of the present invention to hybridize to a nucleic acid, e.g. those listed in any of SEQ ID NOS 1, 3, 5, 7 and/or 9, is confirmed in a Southern blot assay under the following conditions: 6x

sodium chloride/sodium citrate (SSC) at 45° C. followed by a wash in 0.2xSSC, 0.1% SDS at 65° C.

**[0027]** The nucleic acid of the present invention is preferably operably linked to a promoter that governs expression in suitable vectors and/or host cells producing the polypeptides of the present invention in vitro or in vivo.

**[0028]** Suitable promoters for operable linkage to the isolated and purified nucleic acid are known in the art. In a preferred embodiment the nucleic acid of the present invention is one that is operably linked to a promoter selected from the group consisting of the *Pichia pastoris* AOX1 or GAP promoter (see for example *Pichia* Expression Kit Instruction Manual, Invitrogen Corporation, Carlsbad, Calif.), the *Saccharomyces cerevisiae* GAL1, ADH1, ADH2, MET25, GPD or TEF promoter (see for example *Methods in Enzymology*, 350, 248, 2002), the Baculovirus polyhedrin p10 or ie1 promoter (see for example *Bac-to-Bac Expression Kit Handbook*, Invitrogen Corporation, Carlsbad, Calif., and *Novagen Insect Cell Expression Manual*, Merck Chemicals Ltd., Nottingham, UK), the *E. coli* T7, araBAD, rhaP BAD, tetA, lac, trc, tac or pL promoter (see *Applied Microbiology and Biotechnology*, 72, 211, 2006), the plant CaMV35S, ocs, nos, Adh-1, Tet promoters (see e.g. Lau and Sun, *Biotechnol Adv.* 27, 1015-1022, 2009) or inducible promoters for mammalian cells as described in Sambrook and Russell (2001).

**[0029]** Preferably, the isolated and purified nucleic acid is in the form of a recombinant vector, such as an episomal or viral vector. The selection of a suitable vector and expression control sequences as well as vector construction are within the ordinary skill in the art. Preferably, the viral vector is a baculovirus vector (see for example *Bac-to-Bac Expression Kit Handbook*, Invitrogen Corporation, Carlsbad, Calif.). Vector construction, including the operable linkage of a coding sequence with a promoter and other expression control sequences, is within the ordinary skill in the art.

**[0030]** Hence and in a further aspect, the present invention relates to a recombinant vector, comprising a nucleic acid of the invention.

**[0031]** A further aspect of the present invention is directed to a host cell comprising a nucleic acid and/or a vector of the invention and preferably producing polypeptides of the invention. Preferred host cells for producing the polypeptide of the invention are selected from the group consisting of yeast cells, preferably *Saccharomyces cerevisiae* (see for example *Methods in Enzymology*, 350, 248, 2002), *Pichia pastoris* cells (see for example *Pichia* Expression Kit Instruction Manual, Invitrogen Corporation, Carlsbad, Calif.), *E. coli* cells (BL21(DE3), K-12 and derivatives) (see for example *Applied Microbiology and Biotechnology*, 72, 211, 2006), plant cells, preferably *Nicotiana tabacum* or *Physcomitrella patens* (see e.g. Lau and Sun, *Biotechnol Adv.* 27, 1015-1022, 2009), NIH-3T3 mammalian cells (see for example Sambrook and Russell, 2001) and insect cells, preferably sf9 insect cells (see for example *Bac-to-Bac Expression Kit Handbook*, Invitrogen Corporation, Carlsbad, Calif.)

**[0032]** Another important aspect of the invention is directed to an isolated and purified polypeptide selected from the group consisting of

**[0033]** (a) polypeptides having an amino acid sequence selected from the group consisting of SEQ ID NOS: 2, 4, 6, 8 and 10, preferably SEQ ID NO: 2,

**[0034]** (b) polypeptides encoded by a nucleic acid of the present invention,

**[0035]** (c) polypeptides having an amino acid sequence identity of at least 25, 30 or 40%, preferably at least 50 or 60%, more preferably at least 70 or 80%, most preferably at least 90 or 95% with the polypeptides of (a) and/or (b),

**[0036]** (d) a fragment and/or functional derivative of (a), (b) or (c).

**[0037]** The identity of related amino acid molecules can be determined with the assistance of known methods. In general, special computer programs are employed that use algorithms adapted to accommodate the specific needs of this task. Preferred methods for determining identity begin with the generation of the largest degree of identity among the sequences to be compared. Preferred computer programs for determining the identity among two amino acid sequences comprise, but are not limited to, TBLASTN, BLASTP, BLASTX or TBLASTX (Altschul et al., *J. Mol. Biol.*, 215, 403-410, 1990). The BLAST programs can be obtained from the National Center for Biotechnology Information (NCBI) and from other sources (BLAST handbook, Altschul et al., NCB NLM NIH Bethesda, Md. 20894).

**[0038]** Preferably, said polypeptides are encoded by an above-mentioned nucleic acid of the invention.

**[0039]** In a preferred embodiment, the polypeptide, fragment and/or derivative of the invention is functional, i.e. has enzymatic galactosyltransferase activity, preferably an enzymatic  $\beta$ -1,4-galactosyltransferase activity, more preferably an enzymatic  $\beta$ -1,4-galactosyltransferase activity, preferably with L-fucoside-, more preferably with  $\alpha$ -L-fucoside-, more preferably with Fuc- $\alpha$ -1,6-GlcNAc—and most preferably with GnGnF<sup>o</sup>—(nomenclature according to Schachter, *Biochem. Cell. Biol.* 64(3), 163-181, 1986) containing poly/oligosaccharides or glycoconjugates as acceptor substrates.

**[0040]** For example, a preferred assay for determining the functionality, i.e. enzymatic activity, of the polypeptides, fragments and derivatives thereof according to the present invention is provided in example 4 below.

**[0041]** The term “functional derivative” of a polypeptide of the present invention is meant to include any polypeptide or fragment thereof that has been chemically or genetically modified in its amino acid sequence, e.g. by addition, substitution and/or deletion of amino acid residue(s) and/or has been chemically modified in at least one of its atoms and/or functional chemical groups, e.g. by additions, deletions, rearrangement, oxidation, reduction, etc. as long as the derivative still has at least one of the above enzymatic activities to a measurable extent, e.g. of at least about 1 to 10% of the original unmodified polypeptide.

**[0042]** In this context a functional fragment of the invention is one that forms part of a polypeptide or derivative of the invention and still has at least one of the above enzymatic activities in a measurable extent, e.g. of at least about 1 to 10% of the complete protein.

**[0043]** The term “isolated and purified polypeptide” as used herein refers to a polypeptide or a peptide fragment which either has no naturally-occurring counterpart (e.g., a peptide-mimetic), or has been separated or purified from components which naturally accompany it, e.g. in *Caenorhabditis elegans* tissue or a fraction thereof. Preferably, a polypeptide is considered “isolated and purified” when it makes up for at least 60% (w/w) of a dry preparation, thus being free from most naturally-occurring polypeptides and/or organic molecules with which it is naturally associated. Preferably, a polypeptide of the invention makes up for at least

80%, more preferably at 90%, and most preferably at least 99% (w/w) of a dry preparation. More preferred are polypeptides according to the invention that make up for at least 80%, more preferably at least 90%, and most preferably at least 99% (w/w) of a dry polypeptide preparation. Chemically synthesized polypeptides are by nature “isolated and purified” within the above context.

**[0044]** An isolated polypeptide of the invention may be obtained, for example, by extraction from a natural source, e.g. *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* or *Cryptosporidium parvum*; by expression of a recombinant nucleic acid encoding the polypeptide in a host, preferably a heterologous host; or by chemical synthesis. A polypeptide that is produced in a cellular system being different from the source from which it naturally originates is “isolated and purified”, because it is separated from components which naturally accompany it. The extent of isolation and/or purity can be measured by any appropriate method, e.g., column chromatography, polyacrylamide gel electrophoresis, HPLC analysis, NMR spectroscopy, gas liquid chromatography, or mass spectrometry.

**[0045]** Furthermore, in one aspect the present invention relates to antibodies, functional fragments and functional derivatives thereof that specifically bind a polypeptide of the invention. These are routinely available by hybridoma technology (Kohler and Milstein, *Nature*, 256, 495-497, 1975), antibody phage display (Winter et al., *Annu. Rev. Immunol.* 12, 433-455, 1994), ribosome display (Schaffitzel et al., *J. Immunol. Methods*, 231, 119-135, 1999) and iterative colony filter screening (Giovannoni et al., *Nucleic Acids Res.* 29, E27, 2001) once the target antigen is available. Typical proteases for fragmenting anti-bodies into functional products are well-known. Other fragmentation techniques can be used as well as long as the resulting fragment has a specific high affinity and, preferably a dissociation constant in the micromolar to picomolar range.

**[0046]** A very convenient antibody fragment for targeting applications is the single-chain Fv fragment, in which a variable heavy and a variable light domain are joined together by a polypeptide linker. Other antibody fragments for identifying the polypeptide of the present invention include Fab fragments, Fab<sub>2</sub> fragments, miniantibodies (also called small immune proteins), tandem scFv-scFv fusions as well as scFv fusions with suitable domains (e.g. with the Fc portion of an immunoglobulin). For a review on certain antibody formats, see Holliger and Hudson, *Biotechnol.*, 23(9), 1126-36, 2005.

**[0047]** The term “functional derivative” of an antibody for use in the present invention is meant to include any antibody or fragment thereof that has been chemically or genetically modified in its amino acid sequence, e.g. by addition, substitution and/or deletion of amino acid residue(s) and/or has been chemically modified in at least one of its atoms and/or functional chemical groups, e.g. by additions, deletions, rearrangement, oxidation, reduction, etc. as long as the derivative has substantially the same binding affinity as to its original antigen and, preferably, has a dissociation constant in the micro-, nano- or picomolar range.

**[0048]** In a preferred embodiment, the antibody, fragment or functional derivative thereof for use in the invention is one that is selected from the group consisting of polyclonal antibodies, monoclonal antibodies, chimeric antibodies, humanized antibodies, CDR-grafted antibodies, Fv-fragments, Fab-

fragments and Fab<sub>2</sub>-fragments and antibody-like binding proteins, e.g. affilines, anticalines and aptamers.

**[0049]** For a review of antibody-like binding proteins see Binz et al. on engineering binding proteins from non-immunoglobulin domains in *Nature Biotechnol.*, 23(10), 1257-1268, 2005. The term "aptamer" describes nucleic acids that bind to a polypeptide with high affinity. Aptamers can be isolated from a large pool of different single-stranded RNA molecules by selection methods such as SELEX (see, e.g., Jayasena, *Clin. Chem.*, 45, 1628-1650, 1999; Klug and Famulok, *Mol. Biol. Rep.*, 20, 97-107, 1994; U.S. Pat. No. 5,582,981). Aptamers can also be synthesized and selected in their mirror form, for example, as the L-ribonucleotide (Nolte et al., *Nat. Biotechnol.*, 14, 1116-1119, 1996; Klussmann et al., *Nat. Biotechnol.*, 14, 1112-1115, 1996). Forms isolated in this way have the advantage that they are not degraded by naturally occurring ribonucleases and, therefore, have a greater stability.

**[0050]** Another antibody-like binding protein and alternative to classical antibodies are the so-called "protein scaffolds", for example, anticalines, that are based on lipocaline (Beste et al., *Proc. Natl. Acad. Sci. USA*, 96, 1898-1903, 1999). The natural ligand binding sites of lipocalines, for example, of the retinol-binding protein or bilin-binding protein, can be changed, for example, by employing a "combinatorial protein design" approach, and in such a way that they bind selected haptens (Skerra, *Biochem. Biophys. Acta*, 1482, pp. 337-350, 2000). For other protein scaffolds it is also known that they are alternatives for antibodies (Skerra, *J. Mol. Recognition*, 13, 167-287, 2000; Hey, *Trends in Biotechnology*, 23, 514-522, 2005).

**[0051]** In summary, the term functional antibody derivative is meant to include the above protein-derived alternatives for antibodies, i.e. antibody-like binding proteins, e.g. affilines, anticalines and aptamers, that specifically recognize a polypeptide, fragment or derivative thereof.

**[0052]** A further aspect relates to a hybridoma cell line, expressing a monoclonal antibody according to the invention.

**[0053]** The nucleic acids, vectors, host cells, polypeptides and antibodies of the present invention have a number of new applications.

**[0054]** In one aspect the present invention relates to the use of a polypeptide, a cell extract comprising a polypeptide of the invention, preferably a nematode extract, more preferably an extract of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* or *Cryptosporidium parvum*, and/or a host cell of the present invention for producing galactoside-containing oligo/polysaccharides and/or glycoconjugates, preferably galactosyl-fucoside-containing oligo/polysaccharides and glycoconjugates, more preferably D-galactopyranosyl- $\beta$ -1,4-L-fucopyranosyl- $\alpha$ -1,6-GlcNAc-containing oligo/polysaccharides and glycoconjugates, most preferably G<sub>n</sub>G<sub>n</sub>F<sup>6</sup>Gal- or MMF<sup>6</sup>Gal-containing oligo/polysaccharides and glycoconjugates.

**[0055]** It is understood that the term glycoconjugate, as used herein is non-limiting with respect to the nature of the non-sugar component. Preferably the non-sugar component of the glycoconjugate is a poly/oligopeptide.

**[0056]** The enzymatic synthesis of galactosyl-fucosyl-specific oligosaccharides and glycoconjugates is highly specific, controlled and environment-friendly and the products can serve as highly parasite-specific (this epitope is only known to also exist in octopus [Zhang et al., *Glycobiology*, 7, 1153-

1158, 1997], squid [Takahashi et al., *Eur. J. Biochem.*, 270, 2627-2632, 2003] and limpets [Wuhrer et al., *Biochem. J.*, 378, 625-632, 2004]) vaccine components for the treatment and prevention of parasitic, preferably nematode and apicomplexa infections in a subject, such as a human or other mammal, in need thereof.

**[0057]** Exemplary and preferred galactosyl-fucosyl-specific oligosaccharides and glycoconjugates are selected from the group consisting of N-linked glycans, N-glycoproteins, glycolipids and lipid-linked oligosaccharides (LOS). The term "glycoconjugate" as used herein, is meant to include any type of conjugate, preferably but not necessarily a covalently bonded one, for example bonded by a covalent linker, of an oligosaccharide- and a non-saccharide component, e.g. a polypeptide or any other type of organic or inorganic carrier that is physiologically acceptable and might even have a desired physiological function, e.g. as an immune stimulating adjuvant, imparting nematode toxicity, etc.

**[0058]** For example, raw extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* or *Cryptosporidium parvum* or recombinant insect cells producing a polypeptide of the invention can produce Gal-Fuc-containing conjugates, e.g. free Gal-Fuc glycans, Gal-Fuc-peptides, Gal-Fuc-polypeptides, Gal-Fuc-folded proteins. Alpha-1,6-linked fucosides are strongly preferred over alpha-1,3-linked fucosides.

**[0059]** Another aspect of the present invention is directed to a method for producing galactosyl-fucosyl derivatives, comprising the following steps:

- [0060]** (i) providing at least one polypeptide of the invention,
- [0061]** (ii) providing at least one fucosylated acceptor substrate,
- [0062]** (iii) incubating (i) and (ii) in the presence of at least one suitable divalent metal cation cofactor, preferably selected from manganese (II), cobalt (II) and/or iron (II) ions, more preferably manganese (II), and at least one activated sugar substrate, preferably uridine diphosphate (UDP)-galactose under conditions suitable for enzymatic activity of the polypeptide of the invention,
- [0063]** (iv) optionally isolating the galactosyl-fucose derivatives.

**[0064]** The polypeptide of the invention may be provided as an isolated polypeptide, in dry or soluble form, in a buffer, a host cell, a cell extract or any other system that will sustain its enzymatic activity and allow access to its substrate and activated sugar substrate. The fucosylated acceptor substrate is any kind of fucosyl-containing substrate, optionally in isolated form or as a component of a system that can be enzymatically modified by the polypeptide of the invention. The activated sugar substrate is preferably UDP-galactose but can also be any other type of activated, preferably phosphate-activated galactosyl derivative that can be transferred to a fucosylated acceptor substrate. The method of the invention preferably leads to galactopyranosyl- $\beta$ -1,4-L-fucopyranosyl-derivatives, more preferably D-galactopyranosyl- $\beta$ -1,4-L-fucopyranosyl- $\alpha$ -1,6- $\beta$ -GlcNAc (Gal-Fuc) derivatives.

**[0065]** The polypeptides of the present invention have a broad substrate specificity as long as the substrate features a suitable fucosyl-moiety. Galactosyl-transferase activity was demonstrated for substrates such as, e.g. fucosyl-saccharides, fucosyl-peptides, fucosyl-polypeptides and even complex and folded fucosyl-polypeptides. For example, galactosyl-

transferase activity was demonstrated for human IgG1, a glycoprotein having GnGnF<sup>6</sup> carbohydrate structures as prevalent epitopes. These IgG1 glycans are known to be accessible for PNGaseF digest. Glycosylation of human IgG1 was demonstrated with the crude sf9 insect cell extract containing the core galactosyltransferase of *Caenorhabditis elegans*. Incubation of human IgG1 with radioactively labelled UDP-Gal in the presence of enzyme extract from *Caenorhabditis elegans* led to substrate galactosylation. In addition, galactosylation was demonstrated on remodelled human transferrin carrying GnGnF<sup>6</sup> carbohydrate structures as prevalent epitopes. For this purpose human apotransferrin was sequentially treated with sialidase (Iskratsch et al, Anal. Biochem., 368, 133-146, 2009),  $\beta$ 1,4-galactosidase from *Aspergillus oryzae* and recombinant *Anopheles* core  $\alpha$ 1,6-FucT expressed in *Pichia pastoris* to produce a glycoprotein having GnGnF<sup>6</sup> carbohydrate structures as prevalent epitopes. Incubation with a crude sf9 insect cell extract containing the core galactosyltransferase of *Caenorhabditis elegans* led to galactosylation which was monitored by dot blotting with the fucose-specific *Aleuria aurantia* lectin and by MALDI-TOF MS of tryptic peptides of the various neoglycoforms.

[0066] It has very recently been shown that the serum content of core fucosylated alpha feto-protein (AFP) is highly specific for hepatocellular carcinomas (HCC), because benign liver diseases such as chronic hepatitis and liver cirrhosis do not lead to core-fucosylated AFP in mammals, in particular humans (see Tateno et al., Glycobiology, 19(5), 527-536, 2009).

[0067] Therefore, in a further aspect the polypeptides of the invention, host cells comprising polypeptides of the invention and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* can be used for covalently binding galactosyl compounds to core-fucosylated alpha-fetoprotein (AFP), preferably for detecting and/or quantifying hepatocellular carcinoma (HCC) cells, preferably by selectively labelling core-fucosylated alpha-fetoprotein (AFP) from the blood of HCC patients, because core-fucosylated AFP is selectively suitable as an acceptor substrate for the polypeptides of the present invention.

[0068] Hence, the present invention relates to polypeptides of the invention, host cells comprising polypeptides of the invention and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* for preparing diagnostic means for detecting core-fucosylated AFP, i.e. for detecting and/or quantifying hepatocellular carcinoma (HCC) cells.

[0069] Also, the polypeptides of the invention, host cells comprising polypeptides of the invention and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* are useful for preparing diagnostic means for detecting further core-fucosylated marker glycoproteins whose appearance correlates with other types of carcinoma cells.

[0070] In a preferred embodiment, the invention relates to a method of diagnosis, comprising the following steps:

- (i) providing blood or a fraction thereof, that comprises AFP, preferably serum,
- (ii) incubating said blood or said fraction thereof with (a) a polypeptide of the invention, a host cell of the invention

and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* and (b) an activated galactosyl derivative, preferably a labelled galactosyl derivative, preferably labelled UDP-galactose, under conditions that allow for the galactosyltransfer of activated galactose to core-fucosylated AFP (AFP-L3),

(iii) and detecting the galactose-labelled and hence core-fucosylated AFP (AFP-L3).

[0071] Labels for activated galactosyl derivatives for practicing the above method are selected from the group consisting of isotopes e.g. <sup>14</sup>C, chemical modifications e.g. halogen substitutions and other selectively detectable modifications e.g. biotin, azide etc. Preferably, all of the steps (i) to (iii) are performed outside the living body, i.e. in vitro.

[0072] A further aspect of the invention is directed to the use of antibodies specifically binding a polypeptide of the invention, preferably a polypeptide having a sequence selected from any of SEQ ID NOs: 2, 4, 6, 8 and/or 10, for identifying and/or quantifying nematodes and apicomplexa, preferably *Caenorhabditis elegans*, *Caenorhabditis briggsae*, and *Cryptosporidium parvum*, respectively, in a sample of interest, for example a human or mammalian sample, preferably in a cell fraction or extract sample. The design and development of typical antibody assays, e.g. ELISAs, is within the ordinary skill in the art and need not be further elaborated.

[0073] The invention has been described with the emphasis upon preferred embodiments and illustrative examples. However, it will be obvious to those of ordinary skill in the art that variations of the preferred embodiments may be used and that it is intended that the invention may be practiced otherwise than as specifically described herein. Moreover, as the foregoing examples are included for purely illustrative purposes, they should not be constructed to limit the scope of the invention in any respect. Accordingly, this invention includes all modifications encompassed within the spirit and scope of the invention as defined by the claims appended hereto.

## FIGURES

[0074] FIG. 1 is an anti-FLAG immunoblotting of baculovirus-infected sf9 whole cell extracts. Different clones of baculoviruses containing empty vector control (e.v.), N-terminally FLAG-tagged M03F8.4 (FLAG-GalT) and untagged M03F8.4 (GalT). Loading ca. 150 kcells/slot, SDS-PAGE 12%,  $\alpha$ -FLAG (1:2000, SIGMA),  $\alpha$ -mouse-HRP (1:2000, Santa Cruz Biotechnology), ECL (Pierce, 2 s exposure).

[0075] FIG. 2 is an SDS-PAGE analysis of baculovirus-infected sf9 whole cell extracts. Different clones of baculoviruses containing empty vector control (e.v.), N-terminally FLAG-tagged M03F8.4 (FLAG-GalT) and untagged M03F8.4 (GalT). Loading ca. 150 kcells/slot, SDS-PAGE 12%, detection by silver staining. (Protein is expressed in low amounts, not detectable by silver staining with respect to the empty vector construct in crude extracts.)

[0076] FIG. 3 is a column chart showing the galactosylation turnover of a GnGnF<sup>6</sup> acceptor substrate (dabsyl-GEN [GnGnF<sup>6</sup>]R) in the presence of Mn<sup>2+</sup>, Mg<sup>2+</sup> and EDTA demonstrating metal ion dependency; MES, pH 6, r.t., 2.5 h, turnover determined by ratio of MALDI-MS peak intensity ( $[m/z\ 2369/(m/z\ 2207+m/z\ 2369)]*100$ ) from crude reaction mixture.

**[0077]** FIG. 4 is a column chart showing the galactosylation of a GnGnF<sup>6</sup> acceptor substrate (dabsyl-GEN[GnGnF<sup>6</sup>]R)—functionality of the tagged and non-tagged construct; MES, pH 6, r.t., 2.5 h, turnover determined by ratio of MALDI-MS peak intensity ( $[(m/z\ 2369)/(m/z\ 2207+m/z\ 2369)]*100$ ) from crude reaction mixture.

**[0078]** FIG. 5 shows the galactosylation of a GnGnF<sup>6</sup> acceptor substrate (dabsyl-GEN[GnGnF<sup>6</sup>]R)—functionality of the tagged and non-tagged construct (MES pH 6, r.t., 2.5 h) by way of MS analysis. Upper spectrum: reaction without UDP-Gal, central spectrum: with UDP-Gal, bottom spectrum: digest of the product from the central spectrum with *Aspergillus*  $\beta$ -galactosidase (citrate buffer, pH 5, r.t., 2 d). The enzyme clearly adds a galactose to this acceptor substrate which can be digested with (3-galactosidase, and therefore shows a  $\beta$ -linked Gal residue incorporated by the GalT. Additional GlcNAc removal takes place after prolonged reaction times (>2 d) due to presence of hexosaminidase in the insect cell crude extract.

**[0079]** FIG. 6 is a comparison of MS/MS spectra of acceptor (upper spectrum) and galactosylated reaction product (lower spectrum) of FIG. 5. The MS/MS analysis clearly shows the galactose being linked to the core fucose, as observed from secondary ion 1272.61 corresponding to a Hex-dHex-HexNAc motif linked to the dabsylated GENR peptide.

**[0080]** FIG. 7 is a comparative analysis of the donor specificity of the galactosyl transferase (dansyl-N[GnGnF<sup>6</sup>]ST, MES pH 6.5, Mn<sup>2+</sup>, r.t., 13 h). The enzyme seems to have a high specificity for UDP-Gal, with a negligible residual activity on UDP-Glc.

**[0081]** FIG. 8 is column chart of an analysis of the acceptor specificity: *Caenorhabditis elegans* GalT galactosylates selectively  $\alpha$ -1,6 linked over  $\alpha$ -1,3-linked fucose; dabsyl-GEN-[MMF<sup>6/3</sup>]R, MES pH 6.5, r.t., 2.5 h, turnover determined by ratio of MALDI-MS peak intensity ( $[(m/z\ 1963)/(m/z\ 1801+m/z\ 1963)]*100$ ) from crude reaction mixture.

**[0082]** FIG. 9 shows the graphic determination of the  $K_m$  (app) of the untagged galactosyl transferase for UDP-Gal:  $K_m$  (app, UDP-Gal)=ca. 40  $\mu$ M.

**[0083]** FIG. 10 is an analysis of the temperature dependency of the galactosyltransferase of the invention (dansyl-N[GnGnF<sup>6</sup>]ST, UDP-Gal, MES pH 6.5, Mn<sup>2+</sup>, 2.5 h).

**[0084]** FIG. 11 is a column chart demonstrating the glycosylation of human IgG1 (possessing GnGnF<sup>6</sup> epitopes) with the polypeptide of the invention, i.e. *Caenorhabditis elegans* core galactosyltransferase.

**[0085]** FIG. 12 is a MALDI-TOF MS spectrum demonstrating the glycosylation of remodelled human transferrin (possessing GnGnF<sup>6</sup> epitopes) with a polypeptide of the invention, i.e. *Caenorhabditis elegans* core galactosyltransferase. The indicated mk values correspond to peptide 622-642 carrying GnGn (3813), GnGnF<sup>6</sup> (3957) and GnGnF<sup>6</sup>Gal (4119), respectively.

## EXAMPLES

### Experimental Procedures

#### Chemicals and Suppliers

**[0086]** UDP-Gal (VWR International and Sigma), UDP-Glc, UDP-GlcNAc, UDP-GalNAc (all SIGMA), UDP-<sup>14</sup>C-Gal (GE Healthcare), GlcNAc- $\beta$ -1,2-Man- $\alpha$ -1,6-[GlcNAc- $\beta$ -1,2-Man- $\alpha$ -1,3-]-Man, Man- $\beta$ -1,4-GlcNAc- $\beta$ -1,4-[ $\alpha$ -1,6-Fuc]-GlcNAc, MMF6, GnGnF<sup>6</sup> (all Dextra Laboratories,

UK), Fuc- $\alpha$ -1,6-GlcNAc (Carbosynth Ltd., UK), dabsyl-GEN[GnGnF<sup>6</sup>]R (Paschinger et al., *Glycobiology*, 15(5), 463-474, 2005), dabsyl-GEN[MMF6]R (Fabini et al., *J. Biol. Chem.* 276(30), 28058-28067, 2001), dabsyl-GEN[MMF3]R (Fabini et al., *J. Biol. Chem.* 276(30), 28058-28067, 2001) and dansyl-N[GnGnF<sup>6</sup>]ST (Roitingner et al., *Glycoconj. J.*, 15(1), 89-91, 1998) were obtained according to previously published methods.

### Example 1

#### Isolation of *Caenorhabditis elegans* cDNA and Cloning of M03F8.4 into Expression Vectors

##### Nematode Strains:

**[0087]** Methods for culturing *Caenorhabditis elegans* are described in Brenner, S. (*Genetics* 77(1), 71-94, 1974). The wild type Bristol N2 strain was grown at 20° C. on standard NGM agar plates seeded with *Escherichia coli* OP50.

##### Isolation of *Caenorhabditis elegans* M03F8.4 cDNA:

A *Caenorhabditis elegans* mixed culture was harvested from one standard NGM agar plate and washed twice in sterile M9 buffer (22 mM KH<sub>2</sub>PO<sub>4</sub>, 42 mM Na<sub>2</sub>HPO<sub>4</sub>, 85 mM NaCl, 1 mM MgSO<sub>4</sub>). Total RNA was extracted using the NucleoSpin® RNA II RNA isolation kit (MACHEREY-NA-GEL AG). cDNA synthesis was performed with 0.5  $\mu$ g total RNA using the First-strand cDNA synthesis step of the Super-Script™ III Platinum Two-Step qRT-PCR Kit (Invitrogen AG).

Construction of the pFastBac1 Donor Plasmid for Recombinant Gene Expression in sf9 Insect Cells:

**[0088]** M03F8.4 cDNA was isolated from a previously prepared cDNA library by PCR using Phusion High-Fidelity DNA Polymerase (Finnzymes) according to the manual supplied. For construction of an untagged version, the following forward and reverse primers, flanked with SalI and XbaI restrictions sites, respectively, were used: 5'-TTTGTGCA-CACTCTGAATGCCTCG-3' (SEQ ID NO: 11) and 5'-TTTTCTAGACTACAAGTCTAA-AAGACCAAC-3' (SEQ ID NO: 12). The resulting fragment was digested with the appropriate restriction enzymes and cloned into the pFastBac1 donor plasmid (Invitrogen). For construction of an N-terminally FLAG tagged version, a forward primer lacking the start codon was used: 5'-TTTGTGACCCCTCGAAT-CACCGCC-3' (SEQ ID NO: 13). The resulting fragment was cloned into a pFastBac1 donor plasmid containing an N-terminal FLAG sequence (Muller et al., *J. Biol. Chem.* 277(36), 32417-32420, 2002) (both vectors kindly provided by Thierry Henet, Institute of Physiology, University of Zurich).

### Example 2

#### Expression of Recombinant Proteins

**[0089]** Recombinant baculoviruses containing the *Caenorhabditis elegans* core beta-1,4-GalT candidate cDNA (with and without N-terminal FLAG-tag) and an empty vector control were generated according to the manufacturers instructions (Invitrogen). After infection of  $2 \times 10^6$  *S. frugiperda* (sf9) adherent insect cells with recombinant baculoviruses and incubation for 72 h at 28° C., the cells were lysed with shaking (4° C., 15 min) in 150  $\mu$ L tris-buffered saline (pH 7.4) containing 2% (v/v) Triton-X100 and protease inhibitor cocktail (Roche, complete EDTA-free). The lysis

mixtures were centrifuged (2000×g, 5 min) and the post-nuclear supernatant was recovered and used for all further enzymatic studies.

#### Example 3

##### Denaturing Gel Electrophoretic Analysis and Immunoblotting

**[0090]** Infected sf9 cells ( $2 \times 10^6$  cells, see above) were vortexed in 200  $\mu$ L Laemmli buffer and proteins denatured by heating (95° C., 5 min). After cooling to r.t. the samples were centrifuged (16 krpm, 5 min) and the supernatant was used for further analysis. The samples were separated by SDS-PAGE (12% acrylamide, 120 V). The resulting gels were either analyzed by silver-staining or by blotting onto a nitrocellulose membrane. After blocking the membrane (5% BSA in PBST) immuno-detection was performed by incubation with anti-FLAG antibody M2 (SIGMA, dilution 1:2000 in PBST+1% BSA) followed by anti-mouse-HRP (Santa Cruz Biotechnology, dilution 1:10000 in PBST+1% BSA) after extensive washing (PBST) and final detection using ECL (Pierce) and exposure to photographic film.

#### Example 4

##### Glycosyltransferase Assays

**[0091]** Enzymatic activity towards appropriate carbohydrates or glycoconjugates was assessed using 0.5  $\mu$ L of raw extract of sf9 cells (containing either an empty vector control bacmid, a putative GalT expressing bacmid or a putative FLAG-tagged GalT expressing bacmid) in 2.5  $\mu$ L final volume of MES buffer (pH 6.5, 40  $\mu$ M) containing manganese (II) chloride (10  $\mu$ M), UDP-galactose (1 mM) and the acceptor fucoside (glycan or glyco(poly)peptide, 40  $\mu$ M). Glycosylation reactions were typically run for 2 h at room temperature, unless noted otherwise. For donor specificity analysis UDP-galactose was replaced by equal concentrations of UDP-Glc, UDP-GlcNAc or UDP-GalNAc (Sigma) respectively. For co-factor-specificity analysis  $MnCl_2$  was replaced by equal concentrations of the various metal chlorides or  $Na_2EDTA$ . To quantify the incorporation of galactose into the acceptor glycans total UDP-Gal concentration was doped with 10% UDP- $^{14}C$ -Gal (25 nCi, GE Healthcare). Excess radioactivity (UDP- $^{14}C$ -Gal) was removed by loading the reaction mixture (quenched with 100  $\mu$ L  $H_2O$ ) onto a column of anion exchange resin (AG1-X8,  $Cl^-$  form, Bio-Rad Laboratories, 200 mg) and elution of the uncharged products ( $H_2O$ , 900  $\mu$ L).

**[0092]** Glycosylation of human IgG1 (5  $\mu$ L of 3 g/L, Calbiochem) was performed in 50  $\mu$ L total volume using the same buffer, salt and enzyme conditions as described above, except the absence of non-radioactive UDP-Gal, which was replaced by UDP- $^{14}C$ -Gal (75 nCi). The reaction was performed at r.t. over night. A suspension of sepharose-protein G beads (Amersham Biosciences, 10  $\mu$ L) in PBS (200  $\mu$ L) was added and binding of IgG1 to the beads was done with shaking (4° C., 1 h). The beads were washed with PBS (5×200  $\mu$ L) and IgG1 was eluted with 20 mM aqueous HCl (3×100  $\mu$ L). Analysis (vide infra) of the reaction products was performed either by direct MALDI-TOF mass spectrometry, HPLC analysis of fluorescently labelled glycopeptides for donor specificity or scintillation counting of radio-labelled assays.

**[0093]** Stepwise remodelling of human asialotransferrin N-glycans was performed as follows: Asialotransferrin (Gal-

Gal) was previously prepared by sialidase treatment of human apotransferrin (Iskratsch et al, Anal. Biochem., 368, 133-146, 2009).

**[0094]** To produce asialoagalactotransferrin (GnGn),  $\beta$ 1,4-galactosidase (3U, from *Aspergillus oryzae*) was added to about 1 mg of GalGal and the sample was incubated for 48 hours at 37° C. (total volume 50  $\mu$ L).

**[0095]** To obtain GnGnF<sup>6</sup>, the sample was brought to a neutral pH with 0.5  $\mu$ L 1M NaOH, before 50 nmol of GDP-fucose and 15  $\mu$ L of a preparation of recombinant *Anopheles* core  $\alpha$ 1,6-FucT, expressed in *Pichia pastoris*, were added. The preparation was incubated overnight before another 50 nmol of GDP-fucose and a further 15  $\mu$ L enzyme (FucT) were added and again incubated overnight at 37° C. In total, approximately 1 mg of GnGnF<sup>6</sup> was obtained.

**[0096]** To prepare GalFuc-transferrin, 1  $\mu$ L of a preparation of recombinant *Caenorhabditis elegans* GalT, 0.2 mmol of  $MnCl_2$  and 20 nmol of UDP-galactose were added to an aliquot of GnGnF<sup>6</sup> (300  $\mu$ g) and incubated overnight at 30° C. Again, the desired glycan structure was boosted with a second incubation overnight after the addition of further substrate (UDP-galactose) and enzyme (GalT).

**[0097]** The degree of modification of the transferrin was monitored by dot blotting with the fucose-specific *Aleuria aurantia* lectin and by MALDI-TOF MS of tryptic peptides of the various neoglycoforms.

#### Example 5

##### Structural Analysis

**[0098]** After exposing dansyl-GEN[GnGnF<sup>6</sup>]R to galactosylation conditions, the resulting crude mixture was adjusted to 50 mM sodium citrate and pH 4.5, digested with *Aspergillus oryzae*  $\beta$ -galactosidase (27 mU) (see Gutterneigg et al., *J. Biol. Chem.* 282(38), 27825-27840, 2007) for 2 days at 30° C. The samples were analyzed by MALDI-TOF mass spectrometry (vide infra).

**[0099]** HPLC Analysis:

**[0100]** Both, for analysis of donor specificity and the reaction rate dependence on donor concentration, the dansyl-N[GnGnF<sup>6</sup>]ST acceptor substrate was separated from the galactosylated reaction product using an isocratic solvent system (0.7 mL/min, 9% MeCN (95%, (v/v)) in 0.05% aqueous TFA (v/v)) on a reversed phase Hypersil ODS C18 column (4×250 mm, 5  $\mu$ m) and fluorescence detection (excitation at 315 nm, emission detected at 550 nm) at room temperature. The Shimadzu HPLC system consisted of a SCL-10A controller, two LC10AP pumps and a RF-10AXL fluorescence detector controlled by a personal computer using Class-VP software (V6.13SP2). Dansyl-N[GnGnF<sup>6</sup>]ST eluted at a retention time of 9.09 min and the galactosylated reaction product at 8.06 min.

Mass Spectrometry:

**[0101]** Glycans were analyzed by MALDI-TOF mass spectrometry on a BRUKER Ultraflex TOF/TOF machine using a  $\alpha$ -cyano-4-hydroxy cinnamic acid matrix. A peptide standard mixture (Bruker) was used for external calibration.

Scintillation Counting:

**[0102]** The eluates of the anion exchange resin column and protein G beads were thoroughly mixed with scintillation

fluid (Irga-Safe Plus, Packard, 4 mL) and measured with a Perkin Elmer Tri-Carb 2800TR.

Abbreviations for Carbohydrates:

**[0103]** Fuc—L-fucose, Gal—D-galactose, GalNAc—D-N-acetylgalactosamine, Glc—D-glucose, GlcNAc—D-N-acetylglucosamine, Man—D-mannose

**[0104]** Abbreviations for complex glycans (according to the Schachter nomenclature [Biochem Cell Biol 64(3), 163-181, 1986]):

**[0105]** GalGal Gal-β-1,4-GlcNAc-β-1,2-Man-α-1,6-[Gal-β-1,4-GlcNAc-β-1,2-Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-GlcNAc

**[0106]** GnGn GlcNAc-β-1,2-Man-α-1,6-[GlcNAc-β-1,2-Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-GlcNAc

**[0107]** GnGnF<sup>6</sup> GlcNAc-β-1,2-Man-α-1,6-[GlcNAc-β-1,2-Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-[α-1,6-Fuc]-GlcNAc

**[0108]** GnGnF<sup>6</sup>Gal GlcNAc-β-1,2-Man-α-1,6-[GlcNAc-β-1,2-Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-[Gal-β-1,4-Fuc-α-1,6]-GlcNAc

**[0109]** MMF<sup>6</sup> Man-α-1,6-[Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-[α-1,6-Fuc]-GlcNAc

**[0110]** MMF<sup>6</sup>Gal Man-α-1,6-[Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-[Gal-β-1,4-Fuc-α-1,6]-GlcNAc

**[0111]** MMF<sup>3</sup> Man-α-1,6-[Man-α-1,3-]-Man-β-1,4-GlcNAc-β-1,4-[α-1,3-Fuc]-GlcNAc

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tttggtcagg ttttgctat tctagattgc ttgtatcgct acaagaacag ggcaaaattt    960
gtagccttca atgacgtaga tgagtttata gttccgctaa ggaacagctc gatagtggaa   1020
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agctcaaacg cgatatttcc caggcaaaaa agcaggttag tttctcagcg gttcttccac   1140
aggaccagg aaacctccc tctcctctcg aaatgcattg tggatccttt gagagtgttc   1200
gagatgggga ttcaccacat aagcaaggct acaggtctgc ggtattccgt caactcagta   1260
cacgagagtg acgcggttat cttccattac aggacttgca ctacgtcatt tggatatacgt   1320
catcagtgca tgaacctagt gcatgatggg accatggcca aatatggaaa acgacttcag   1380
aaaatgttta gaaaggttgt aaatgattta aaacttttgg caccaacgta gctatttcgt   1440
aacacttcac actttcattg ttataacaga atacagaata aattaatgat tgttgtgcc   1499

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<210> SEQ ID NO 6  
<211> LENGTH: 476  
<212> TYPE: PRT  
<213> ORGANISM: Nematostella vectensis

<400> SEQUENCE: 6

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Met Arg Cys Tyr Ile Tyr Lys Leu Arg Leu Ser Val Cys Leu Phe Val
 1           5           10           15

Val Leu Phe Thr Ala Leu Leu Phe Ile Thr Tyr Leu Asn His Ser Glu
 20           25           30

Leu Glu Ser Ala Glu Lys Ser Ser Gly Lys Arg Lys Thr Arg His Arg

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Lys	Arg	Thr	Arg	Ser	Arg	Lys	Gln	His	Glu	Ser	His	Phe	Gln	Lys	Ala
50					55					60					
Arg	Leu	Gln	Glu	Arg	Glu	Leu	Val	Leu	Arg	Ser	Thr	Ala	Pro	Pro	Thr
65					70					75					80
Leu	Arg	Arg	Glu	Val	Gln	Ala	His	Arg	Leu	Gly	Gln	Ile	Arg	Gly	Lys
			85						90					95	
Asn	Thr	Asp	Gln	Gly	Ile	Thr	Gly	Lys	Phe	Thr	Glu	Ile	Ala	Lys	Asp
		100						105					110		
Thr	His	Ile	Tyr	Ser	Ala	Phe	Tyr	Asp	Asp	Ala	Lys	Ser	Asn	Pro	Phe
		115					120						125		
Ile	Arg	Leu	Ile	Ile	Leu	Ser	Gly	Lys	His	Tyr	Gln	Pro	Gly	Leu	Ser
	130					135					140				
Cys	Gln	Phe	Cys	Glu	Pro	Leu	Ser	Ala	Ser	Cys	Ser	Phe	Ala	Asp	Ser
145					150					155					160
Lys	Ala	Glu	Tyr	Tyr	Thr	Thr	Asn	Glu	Asn	His	Gly	Arg	Val	Phe	Gly
			165						170					175	
Gly	Phe	Ile	Ala	Ser	Cys	Leu	Val	Pro	Asp	Gly	Phe	Asn	Ala	Val	Pro
			180					185					190		
Leu	Phe	Val	Asp	Ile	Thr	Ala	Asp	Val	Lys	Gly	Glu	Lys	Ser	Lys	Ala
		195					200						205		
Arg	Val	Pro	Val	Val	Ser	Asn	Ala	His	Leu	Tyr	Tyr	Pro	Ile	Lys	Tyr
	210					215						220			
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225					230					235					240
Leu	Ile	Glu	Phe	Val	Glu	Leu	Thr	Lys	Leu	Leu	Gly	Ala	Asn	His	Phe
			245					250						255	
Thr	Phe	Tyr	Asp	Phe	Lys	Thr	Asp	Pro	Glu	Val	Asn	Asn	Val	Leu	Arg
			260				265						270		
Tyr	Tyr	Gln	Glu	Thr	Gln	Val	Ala	Asn	Val	Leu	Pro	Trp	Asn	Leu	Pro
		275					280					285			
Ser	Asn	Leu	Val	Ser	Arg	Pro	Asn	Asp	Ile	Trp	Tyr	Phe	Gly	Gln	Val
	290					295					300				
Leu	Ala	Ile	Leu	Asp	Cys	Leu	Tyr	Arg	Tyr	Lys	Asn	Arg	Ala	Lys	Phe
305					310					315					320
Val	Ala	Phe	Asn	Asp	Val	Asp	Glu	Phe	Ile	Val	Pro	Leu	Arg	Asn	Ser
			325						330					335	
Ser	Ile	Val	Glu	Ile	Leu	Asn	Ala	Phe	His	Arg	Pro	Tyr	His	Cys	Gly
		340						345					350		
His	Cys	Phe	Gln	Ser	Val	Val	Phe	Ser	Ser	Asn	Ala	Arg	Phe	Pro	Arg
		355					360					365			
Gln	Lys	Ser	Glu	Leu	Val	Ser	Gln	Arg	Phe	Phe	His	Arg	Thr	Gln	Glu
	370					375						380			
Thr	Ile	Pro	Leu	Leu	Ser	Lys	Cys	Ile	Val	Asp	Pro	Leu	Arg	Val	Phe
	385					390					395				400
Glu	Met	Gly	Ile	His	His	Ile	Ser	Lys	Ala	Thr	Gly	Leu	Arg	Tyr	Ser
				405					410					415	
Val	Asn	Ser	Val	His	Glu	Ser	Asp	Ala	Val	Ile	Phe	His	Tyr	Arg	Thr
			420					425					430		
Cys	Thr	Thr	Ser	Phe	Gly	Ile	Arg	His	Gln	Cys	Met	Asn	Leu	Val	His
		435					440					445			

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Asp Gly Thr Met Ala Lys Tyr Gly Lys Arg Leu Gln Lys Met Phe Arg  
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Lys Val Val Asn Asp Leu Lys Leu Leu Ala Pro Thr  
 465 470 475

<210> SEQ ID NO 7  
 <211> LENGTH: 1260  
 <212> TYPE: DNA  
 <213> ORGANISM: Taeniopygia guttata

<400> SEQUENCE: 7

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 gcattggaag gtaacaaaa tgatattata tcccatact ttgatgacag agaaaacaaa 180  
 atcactcgtc tgattgggat tgttcaccat aaagatgtaa aacaactggt ctgctgggtc 240  
 tgctgtcaag ccaatggaag gatatatgta tcaaaagcag aaatagatgt tcaactcgat 300  
 agatttggat tcccttatgg tgcagcagat ataatttgtt tggaacctga aaactgtgat 360  
 ccaacacatg tatcaattca tcagctccca tatggaaata ttgaccagct gccgaggttt 420  
 gaaattaaaa atcgcaggcc tgagaccttt tctgttgact tcaccgtgtg catttctgcc 480  
 atgtttgga actacaacaa tgtcttgac tttgtacaga gtatggaaat gtataagatt 540  
 cttggagtac agaaagtgg gatctataag aacaactgca gccatctgat ggagaaagt 600  
 ttgaaatfff atatagaaga aggaactggt gaggtaattc cctggccaat agactcacac 660  
 ctcaagggttt cttctaatag gcgcttcacg gaagacggga cacacattgg ctactatgga 720  
 caaatcacag ctctaataga ctgtatatac cgcaacatgg aaaggaccaa gtttgtggtc 780  
 cttaatgacg ctgatgaaat aattcttccc cttaaacacc cagactggaa aacaatgatg 840  
 aacagtcttc aggagcaaaa cccagggact agtgttttcc tttttgagaa ccatatcttc 900  
 ccagaaactg tattttctcc catgttcaac atttcatctt ggaatactgt gccaggtgtt 960  
 aacatattgc agcatgtgta cagagagcct gacagaaaac atgtaatcaa tcccaggaaa 1020  
 atgatagttg atccacgaaa ggtgattcag acttcagtec attctgtcct acgtgettat 1080  
 gggagagcgc tgaatgttcc catggaagtt gcctcattt atcactgtcg gaaggccctt 1140  
 caaggaaacc tcccagaga atctctcatc agggatacaa cactgtggag atataactca 1200  
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<210> SEQ ID NO 8  
 <211> LENGTH: 419  
 <212> TYPE: PRT  
 <213> ORGANISM: Taeniopygia guttata

<400> SEQUENCE: 8

Met Thr Val Thr Leu Met Leu Val Val Ser Tyr Leu Arg Leu Gln Arg  
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Leu Ser His Gln Pro Lys Val Ile Gln Glu Ser Arg Arg Cys Arg Gly  
 20 25 30

Lys Ile Ala Leu Ser Thr Ile Thr Ala Leu Glu Gly Asn Lys Thr Asp  
 35 40 45

Ile Ile Ser Pro Tyr Phe Asp Asp Arg Glu Asn Lys Ile Thr Arg Leu  
 50 55 60

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Ile Gly Ile Val His His Lys Asp Val Lys Gln Leu Phe Cys Trp Phe  
 65 70 75 80  
 Cys Cys Gln Ala Asn Gly Lys Ile Tyr Val Ser Lys Ala Glu Ile Asp  
 85 90 95  
 Val His Ser Asp Arg Phe Gly Phe Pro Tyr Gly Ala Ala Asp Ile Ile  
 100 105 110  
 Cys Leu Glu Pro Glu Asn Cys Asp Pro Thr His Val Ser Ile His Gln  
 115 120 125  
 Ser Pro Tyr Gly Asn Ile Asp Gln Leu Pro Arg Phe Glu Ile Lys Asn  
 130 135 140  
 Arg Arg Pro Glu Thr Phe Ser Val Asp Phe Thr Val Cys Ile Ser Ala  
 145 150 155 160  
 Met Phe Gly Asn Tyr Asn Asn Val Leu Gln Phe Val Gln Ser Met Glu  
 165 170 175  
 Met Tyr Lys Ile Leu Gly Val Gln Lys Val Val Ile Tyr Lys Asn Asn  
 180 185 190  
 Cys Ser His Leu Met Glu Lys Val Leu Lys Phe Tyr Ile Glu Glu Gly  
 195 200 205  
 Thr Val Glu Val Ile Pro Trp Pro Ile Asp Ser His Leu Arg Val Ser  
 210 215 220  
 Ser Lys Trp Arg Phe Met Glu Asp Gly Thr His Ile Gly Tyr Tyr Gly  
 225 230 235 240  
 Gln Ile Thr Ala Leu Asn Asp Cys Ile Tyr Arg Asn Met Glu Arg Thr  
 245 250 255  
 Lys Phe Val Val Leu Asn Asp Ala Asp Glu Ile Ile Leu Pro Leu Lys  
 260 265 270  
 His Pro Asp Trp Lys Thr Met Met Asn Ser Leu Gln Glu Gln Asn Pro  
 275 280 285  
 Gly Thr Ser Val Phe Leu Phe Glu Asn His Ile Phe Pro Glu Thr Val  
 290 295 300  
 Phe Ser Pro Met Phe Asn Ile Ser Ser Trp Asn Thr Val Pro Gly Val  
 305 310 315 320  
 Asn Ile Leu Gln His Val Tyr Arg Glu Pro Asp Arg Lys His Val Ile  
 325 330 335  
 Asn Pro Arg Lys Met Ile Val Asp Pro Arg Lys Val Ile Gln Thr Ser  
 340 345 350  
 Val His Ser Val Leu Arg Ala Tyr Gly Lys Ser Val Asn Val Pro Met  
 355 360 365  
 Glu Val Ala Leu Ile Tyr His Cys Arg Lys Ala Leu Gln Gly Asn Leu  
 370 375 380  
 Pro Arg Glu Ser Leu Ile Arg Asp Thr Thr Leu Trp Arg Tyr Asn Ser  
 385 390 395 400  
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<210> SEQ ID NO 9  
 <211> LENGTH: 1647  
 <212> TYPE: DNA  
 <213> ORGANISM: Cryptosporidium parvum  
 <400> SEQUENCE: 9

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agtcattcaa ggaggcta at aagtgaacct tacagtgaat gtattaatga acaaaatgat    180
caagattgga aagaactaaa gctaataatt ccaaatcatt ctcaaattaa ccagcaggaa    240
aaaaatggta atttgattga gtttaaagtt tatatatact cagcatatta tgattggaga    300
atagatagga tacgaataaa ttcacttata ccatcgaatt tttatgatcg aatagaaatg    360
gaatgtgcaa taactcttga caaaaatatt tacacaggaa ctattaaaaa agtgattcat    420
aaggagcacc ataataaaga atatgtatca tcgactttac tctcgogaaat tgcaaaaaat    480
gaaattaaat ttgaggatat ttcaaggaaa gttttgataa caattttgga aaatggaaac    540
agcacaataa aatcagaaa at tggataact ctaaaaaaaa ttcaaaaaaa tagctcta at    600
aatcatgagc tgactgtttg tgtgagacct tgggtggggag agccaataaa gaatggaaac    660
ttgggaaata acaaaaaatt taacaattca gggttaatgc ttgaatttat taattcatal    720
ttattcttag gagcaataaa attttattta tatcaaaatt acttggacat tgacgaagat    780
gtaagaaata taataaatta ttattcta atcaaaaatg ttttgaaat tattccatac    840
tcattaccaa taattccatt taaacaagtt tgggatttcg cacaaacaac aatgatacag    900
gactgcctac taagaaatat tggaaaaaca aaatacttgt tattcgtaga taccgatgaa    960
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aacaatcctt attataaaaa caaagtcggg gcaatgtgga ttccaatgta ttttcatttt   1080
ttagagtggg aatctgataa aaataattg aagaaatatt caacaattga gaaaaaatt   1140
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tctggaacaa aaaaaagtga caagacgaga agaaaagtta ttattagacc tgaagagtt   1260
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gctcctgtaa ttaatgtggg tggaggaaac gaactaagta tatatttaca tcattataga   1380
aaagcaaaag gtattgtaa caatgatccc aaacaagag aacttgtgaa tatgtattta   1440
gaaaaatgtt gttcagataa gctgttagat tcagggggag attccattca agatggagta   1500
attgtcgaca atactgtttg ggagatattt ggaacacact tataccagat aatttttgag   1560
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ttaagtgttg aagaattaca taattaa                                     1647

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<210> SEQ ID NO 10  
<211> LENGTH: 548  
<212> TYPE: PRT  
<213> ORGANISM: Cryptosporidium parvum

<400> SEQUENCE: 10

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                20           25           30
Ser Asn Lys Asn Ile Gln Glu Val Ser His Ser Arg Arg Leu Ile Ser
            35           40           45
Glu Pro Tyr Ser Glu Ser Ile Asn Glu Gln Asn Asp Gln Asp Trp Lys
50           55           60

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Glu Leu Lys Leu Ile Ile Pro Asn His Ser Gln Ile Asn Gln Gln Glu  
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 Lys Asn Gly Asn Leu Ile Glu Phe Lys Val Tyr Ile Tyr Ser Ala Tyr  
 85 90 95  
 Tyr Asp Trp Arg Ile Asp Arg Ile Arg Ile Asn Ser Leu Ile Pro Ser  
 100 105 110  
 Asn Phe Tyr Asp Arg Ile Glu Met Glu Cys Ala Ile Ile Leu Asp Lys  
 115 120 125  
 Asn Ile Tyr Thr Gly Thr Ile Lys Lys Val Ile His Lys Glu His His  
 130 135 140  
 Asn Lys Glu Tyr Val Ser Ser Thr Leu Leu Cys Glu Ile Ala Lys Asn  
 145 150 155 160  
 Glu Ile Lys Phe Glu Asp Ile Ser Arg Lys Val Leu Ile Thr Ile Leu  
 165 170 175  
 Glu Asn Gly Asn Ser Thr Asn Lys Ser Glu Ile Trp Ile Thr Leu Lys  
 180 185 190  
 Lys Ile Pro Lys Asn Ser Ser Asn Asn His Glu Leu Thr Val Cys Val  
 195 200 205  
 Arg Pro Trp Trp Gly Glu Pro Ile Lys Asn Gly Asn Leu Gly Asn Lys  
 210 215 220  
 Gln Lys Phe Asn Asn Ser Gly Leu Met Leu Glu Phe Ile Asn Ser Tyr  
 225 230 235 240  
 Leu Phe Leu Gly Ala Asn Lys Phe Tyr Leu Tyr Gln Asn Tyr Leu Asp  
 245 250 255  
 Ile Asp Glu Asp Val Arg Asn Ile Ile Asn Tyr Tyr Ser Asn Ile Lys  
 260 265 270  
 Asn Val Leu Glu Ile Ile Pro Tyr Ser Leu Pro Ile Ile Pro Phe Lys  
 275 280 285  
 Gln Val Trp Asp Phe Ala Gln Thr Thr Met Ile Gln Asp Cys Leu Leu  
 290 295 300  
 Arg Asn Ile Gly Lys Thr Lys Tyr Leu Leu Phe Val Asp Thr Asp Glu  
 305 310 315 320  
 Phe Val Phe Pro Asn Leu Lys Asn Tyr Asn Leu Met Asp Phe Leu Asn  
 325 330 335  
 Leu Leu Glu Ala Asn Asn Pro Tyr Tyr Lys Asn Lys Val Gly Ala Met  
 340 345 350  
 Trp Ile Pro Met Tyr Phe His Phe Leu Glu Trp Glu Ser Asp Lys Asn  
 355 360 365  
 Asn Leu Lys Lys Tyr Ser Thr Ile Glu Lys Lys Ile Lys Lys Lys Met  
 370 375 380  
 Ala Asn Ile Glu Phe Val Leu Tyr Arg Lys Thr Cys Arg Met Leu Ser  
 385 390 395 400  
 Ser Gly Thr Lys Lys Ser Asp Lys Thr Arg Arg Lys Val Ile Ile Arg  
 405 410 415  
 Pro Glu Arg Val Leu Tyr Met Gly Ile His Glu Thr Glu Glu Met Leu  
 420 425 430

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Ser Lys Lys Phe His Phe Ile Arg Ala Pro Val Ile Asn Val Gly Gly  
435 440 445

Gly Asn Glu Leu Ser Ile Tyr Leu His His Tyr Arg Lys Ala Lys Gly  
450 455 460

Ile Val Asn Asn Asp Pro Lys Gln Arg Glu Leu Val Asn Met Tyr Leu  
465 470 475 480

Glu Asn Val Cys Ser Asp Lys Leu Leu Asp Ser Gly Gly Asp Ser Ile  
485 490 495

Gln Asp Gly Val Ile Val Asp Asn Thr Val Trp Glu Ile Phe Gly Thr  
500 505 510

His Leu Tyr Gln Ile Ile Phe Glu His Ile Lys Glu Ile Gln Asp Met  
515 520 525

Tyr Thr Asn Lys Glu Ile Ile Asn Gly Asn Lys Asn Leu Ser Val Glu  
530 535 540

Glu Leu His Asn  
545

<210> SEQ ID NO 11  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 11

tttgctgaca cttctgaatg cctcg

25

<210> SEQ ID NO 12  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 12

tttctagac tacaagtcta aaagaccaac

30

<210> SEQ ID NO 13  
<211> LENGTH: 24  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: PCR primer

<400> SEQUENCE: 13

tttgctgacc ctggaatcac cgcc

24

1. An isolated and purified nucleic acid, wherein said nucleic acid is selected from the group consisting of:

- (i) a nucleic acid comprising at least a nucleic acid sequence selected from the group consisting of nucleic acid sequences listed in SEQ ID NOs: 1, 3, 5, 7 and 9;
- (ii) a nucleic acid having a sequence of at least 60 or 70% identity, preferably at least 80 or 90% identity, more preferred at least 95% identity, most preferred at least 98% identity with the nucleic acid sequence listed in SEQ ID NO 1;
- (iii) a nucleic acid that hybridizes to a nucleic acid of (i) or (ii);

(iv) a nucleic acid, wherein said nucleic acid is derivable by substitution, addition and/or deletion of one of the nucleic acids of (i), (ii) oder (iii);

(v) a fragment of any of the nucleic acids of (i) to (iv), that hybridizes to a nucleic acid of (i).

2. The nucleic acid according to claim 1, wherein said nucleic acid is a DNA, RNA or PNA, preferably DNA or PNA, more preferably DNA.

3. The nucleic acid according to claim 1, wherein said nucleic acid encodes a protein having galactosyltransferase activity, preferably  $\beta$ -1,4-galactosyltransferase activity, pref-

erably with L-fucoside-, more preferably with  $\alpha$ -L-fucoside-, more preferably with Fuc- $\alpha$ -1,6-GlcNAc— and most preferably with GnGnF<sup>6</sup>— containing poly/oligosaccharides or glycoconjugates as acceptor substrates.

4. An isolated and purified polypeptide selected from the group consisting of:

- (a) polypeptides having an amino acid sequence selected from the group consisting of SEQ ID NOs: 2, 4, 6, 8 and 10, preferably SEQ ID NO: 2,
- (b) polypeptides encoded by a nucleic acid of claim 1,
- (c) polypeptides having an amino acid sequence identity of at least 25, 30 or 40%, preferably at least 50 or 60%, more preferably at least 70 or 80%, most preferably at least 90 or 95% with the polypeptides of (a) and/or (b),
- (d) a fragment and/or functional derivative of (a), (b) or (c).

5. The polypeptide according to claim 4, wherein said polypeptide has galactosyltransferase activity, preferably  $\beta$ -1,4-galactosyltransferase activity, preferably with L-fucoside-, more preferably with  $\alpha$ -L-fucoside-, more preferably with Fuc- $\alpha$ -1,6-GlcNAc— and most preferably with GnGnF<sup>6</sup>-containing poly/oligosaccharides or glycoconjugates as acceptor substrates.

6. A recombinant vector comprising a nucleic acid of claim 1, preferably a viral or episomal vector, preferably a baculovirus vector.

7. A host cell comprising a nucleic acid claim 1, preferably selected from the group consisting of yeast cells, preferably *Saccharomyces cerevisiae*, *Pichia pastoris* cells, *E. coli* cells, plant cells, preferably *Nicotiana tabacum* or *Physcomitrella patens* cells, NIH-3T3 mammalian cells and insect cells, more preferably sf9 insect cells.

8. An antibody that specifically binds a polypeptide of claim 4.

9. An antibody according to claim 8, wherein said antibody is monoclonal antibody.

10. Hybridoma cell line, expressing a monoclonal antibody that specifically binds a polypeptide according to claim 5.

11. Use of a polypeptide of claim 4, a cell extract comprising a polypeptide of claim 4, preferably a *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* or *Cryptosporidium parvum* extract, and/or a host cell comprising a nucleic acid, selected from the group consisting of yeast cells, *E. Coli*, plant cells, NIH-3T3 mammalian cells and insect cells, for producing galactosyl-containing oligo/polysaccharides and/or glycoconjugates, preferably galactosyl-fucoside-containing oligo/polysaccharides and/or glycoconjugates, more preferably D-galactopyranosyl- $\beta$ -1,4-L-fucopyranosyl- $\alpha$ -1,6-GlcNAc-containing

oligo/polysaccharides and/or glycoconjugates, most preferably GnGnF<sup>6</sup>Gal- and/or MMF<sup>6</sup>Gal-containing oligosaccharides and glycoconjugates.

12. Method for producing galactosyl-fucosyl derivatives, comprising the following steps:

- (i) providing at least one polypeptide of the invention,
- (ii) providing at least one fucosylated acceptor substrate,
- (iii) incubating (i) and (ii) in the presence of at least one suitable divalent metal cation cofactor, preferably selected from manganese (II), cobalt (II) and/or iron (II) ions, more preferably manganese (II), and at least one activated sugar substrate, preferably uridine diphosphate (UDP)-galactose under conditions suitable for enzymatic activity of the polypeptide of the invention,
- (iv) optionally isolating the galactosyl-fucose derivatives.

13. Use of at least one polypeptide of claim 4, a host cell comprising a nucleic acid, selected from the group consisting of yeast cells, *E. Coli*, plant cells, NIH-3T3 mammalian cells and insect cells,

and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* for covalently binding galactosyl compounds to core-fucosylated alpha-fetoprotein (AFP), preferably for detecting and/or quantifying hepatocellular carcinoma (HCC).

14. A method of diagnosis, comprising the following steps:

- (i) providing blood or a fraction thereof, that comprises AFP, preferably serum,
- (ii) incubating said blood or said fraction thereof with (a) a polypeptide of claim 4, a host cell comprising a nucleic acid, selected from the group consisting of yeast cells, *E. Coli*, plant cells, NIH-3T3 mammalian cells and insect cells, and/or cell extracts of *Caenorhabditis elegans*, *Caenorhabditis briggsae*, *Nematostella vectensis*, *Taeniopygia guttata* and/or *Cryptosporidium parvum* and (b) an activated galactosyl derivative, preferably a labelled galactosyl derivative, preferably labelled UDP-galactose, under conditions that allow for the galactosyl-transfer of activated galactose to core-fucosylated AFP (AFP-L3),
- (iii) and detecting the galactose-labelled and hence core-fucosylated AFP (AFP-L3).

15. Use of antibodies according to claim 8 for identifying and/or quantifying nematodes or pathogens, preferably *Caenorhabditis elegans*, *Caenorhabditis briggsae* and/or *Cryptosporidium parvum* in a sample of interest, for example a human or mammalian sample, preferably in a cell fraction or extract sample.

\* \* \* \* \*

专利名称(译)	N-聚糖核心 $\beta$ -半乳糖基转移酶及其用途		
公开(公告)号	<a href="#">US20120064541A1</a>	公开(公告)日	2012-03-15
申请号	US13/322505	申请日	2010-05-28
[标]申请(专利权)人(译)	UNIV FUR BODENKULTUR WIEN 苏黎世大学		
申请(专利权)人(译)	苏黎世联邦理工学院 Universität 大学 FUR BODENKULTUR WIEN Universität 大学 ZURICH		
当前申请(专利权)人(译)	苏黎世联邦理工学院 Universität 大学 ZURICH Universität 大学 FUR BODENKULTUR WIEN		
[标]发明人	KUNZLER MARKUS AEBI MARKUS WILSON LAIN TITZ ALEXANDER WALTER HENGARTNER MICHAEL BUTSCHI ALEX		
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摘要(译)

本发明涉及新的半乳糖基转移酶，编码它们的核酸，以及重组载体，宿主细胞，抗体，用途和与其相关的方法。

