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(54) **EPITOPES RELATED TO COELIAC DISEASE**

EPITOPE IM ZUSAMMENHANG MIT ZOLIAKIE

EPITOPES ASSOCIES A UNE MALADIE COELIAQUE

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(73) Proprietor: **BTG International Limited**
London EC4M 7RD (GB)

(72) Inventors:
- **ANDERSON, Robert**
  Parkville
  VIC 3050 (AU)
- **BEISSBATH, Tim**
  69120 Heidelberg (DE)
- **DIN, Jason Tye**
  Parkville
  VIC 3050 (AU)

(74) Representative: **J A Kemp**
14 South Square
Gray's Inn
London WC1R 5JJ (GB)

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WO-A-01/25793 **WO-A-02/083722**
WO-A-03/066079 **WO-A-03/104273**

- **FRASER J S ET AL: "Coeliac disease: In vivo toxicity of the putative immunodominant epitope." GUT, vol. 52, no. 12, December 2003 (2003-12), pages 1698-1702, XP002363983 ISSN: 0017-5749**

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Description

[0001] The invention relates to epitopes useful in the diagnosis and therapy of coeliac disease, including diagnostics, therapeutics, kits, and methods of using the foregoing.

[0002] Coeliac disease is caused by an immune mediated hypersensitivity to dietary gluten. Gluten proteins in wheat, rye, barley and in some cases oats are toxic in coeliac disease. Gluten is composed of alpha/beta, gamma and omega gliadins, and low and high molecular weight (LMW and HMW) glutenins in wheat, hordeins in barley, secalins in rye and avenins in oats. Hordeins and secalins are homologous to gamma and omega gliadins and low and high molecular weight glutenins in wheat. Avenins are phylogenetically more distant than hordeins and secalins from wheat gluten.

[0003] The goal of research in coeliac disease has been to define the toxic components of gluten by defining the peptides that stimulate gluten-specific T-cells. Precise definition of gluten epitopes permits development of new diagnostics, therapeutics, tests for gluten contamination in food and non-toxic grains that retain the cooking/baking qualities of traditional gluten. Many of these applications require a comprehensive understanding of all rather than the most common toxic peptides in gluten.

[0004] Genes encoding HLA-DQ2 and/or HLA-DQ8 are present in over 99% of individuals with coeliac disease compared to approximately 35% of the general Caucasian population. Gluten-derived peptides (epitopes) bound to HLA-DQ2 or HLA-DQ8 stimulate specific T-cells. HLA-DQ2 and DQ8-restricted epitopes include a "core" 9 amino acid sequence that directly interacts with the peptide binding groove of HLA-DQ2 or DQ8 and with cognate T-cell receptors. In general, libraries of overlapping peptides (usually 15 to 20mers) containing all unique 10 or 12mer peptides in an antigen have been used to map HLA class II-restricted T-cells epitopes.

[0005] A series of gluten peptides are known to activate gluten specific T-cells in coeliac disease. Previous studies have identified gluten peptides from selected gluten proteins or gluten digests. T-cell clones and lines isolated from intestinal biopsies have been used to screen these gluten components.

[0006] Modification of gluten by the enzyme, tissue transglutaminase (tTG) present in intestinal tissue, substantially increases gluten's stimulatory capacity on gluten specific T-cells. Most of the known epitopes for gluten-specific T-cells correspond to tTG-deamidated gluten peptides. Transglutaminase mediates deamidation of specific glutamine residues (to glutamate) in gluten. Glutamine-containing sequences susceptible to deamidation by tTG generally conform to a motif: QXPX or QXX (FYMILVW) (see Vader W. et al 2002 J. Exp. Med. 195:643-649, PCT WO 03/066079, and Fleckenstein B. 2002. J Biol Chem 277:34109-16). The motif for peptides that bind to HLA-DQ2 and that are susceptible to deamidation by tTG has been used to predict certain gluten epitopes (Vader et al J Exp Med 2002 J. Exp. Med. 195:643-649, PCT WO 03/066079).

[0007] However, other groups have identified epitopes for gluten-specific intestinal T-cell clones and lines using panels of eleven recombinant alpha/beta (11) and five gamma gliadins (Arentz-Hansen H. 2000. J. Exp. Med. 191:603-612, Arentz-Hansen H. 2002. Gastroenterology 123:803-809, PCT WO 02/083722), and lysates of purified gluten proteins (Sjostrom H. et al 1998. Scand. J. Immunol. 48,111-115; van de Wal, Y. et al 1998. J. Immunol. 161(4):1585-1588; van de Wal, Y. et al 1999. Eur. J. Immunol. 29:3133-3139; Vader W. et al 2002. Gastroenterology 122:1729-1737.).

[0008] Our work has exploited the observation that gluten challenge *in vivo* induces HLA-DQ2 restricted CD4+ gluten-specific T-cells in peripheral blood expressing a gut-homing integrin (alpha4beta7). This technique allowed the mapping of the dominant epitope in A-gliadin (57-73 QE65) (Anderson, RP et al 2000. Nat. Med. 6:337-342., WO 01/25793). A-gliadin 57-73 QE65 corresponds to two overlapping epitopes identified using intestinal T-cell clones (Arentz-Hansen H. et al 2000. J. Exp. Med. 191:603-612, Arentz-Hansen H. et al 2002. Gastroenterology 123:803-809). The advantage of *in vivo* gluten challenge to induce gluten specific T-cells is that any food can be consumed and the resulting T-cells induced in blood (quantified in peripheral blood using a simple overnight interferon gamma ELISPOT assay) will have been stimulated *in vivo* by endogenously presented epitopes, rather than primed *in vitro* by a synthetic or purified antigen. Overnight assays of fresh polyclonal peripheral blood T-cells also avoid the potential for artefacts associated with the lengthy purification of T-cell clones.

[0009] Interestingly, T-cell clones and lines specific for several gamma-gliadin epitopes (Arentz-Hansen H. 2002. Gastroenterology 123:803-809, PCT WO 02/083722) cross-react with the originally defined A-gliadin epitope 57-73 QE65.

[0010] Although there is substantial homology within the alpha/beta gliadins, earlier work (see WO 03/104273) has shown that the dominant epitope recognized in HLA-DQ2-associated coeliac disease, "A-gliadin 57-73 QE65", is encoded by a minority of the alpha/beta gliadins present in Genbank.

SUMMARY OF THE INVENTION

[0011] The current study set out to develop a method that would allow mapping of all T-cell epitopes in gluten. Consumption of wheat bread (200g daily for 3 days) or oats (100g daily for 3 days) was used to induce gluten or avenin-specific T-cells in peripheral blood (collected 6 days after beginning the challenge). Peripheral blood mononuclear cells

(PBMC) were assessed in overnight interferon gamma ELISPOT assays using a library of gluten and avenin peptides including all unique 12mer sequences included in every Genbank entry for wheat gluten and/or oat avenins. This goal was achieved by establishing an algorithm to design peptides spanning all potential epitopes in gluten proteins in Genbank (2922 20mers included all 14 964 unique 9mers - potential T-cell epitopes), adapting the interferon-gamma ELISPOT assay to a high throughput assay capable of screening over 1000 peptides with a single individual's blood and developing bioinformatics tools to analyse and interpret the data generated.

[0012] A series of 41 "superfamilies" of wheat gluten peptides were identified as putative T-cell epitopes. Superfamilies shared motifs in which a limited level of redundancy was allowed. Many of the most potent families include known T-cell epitopes including the previously described dominant epitope, A-gliadin 57-73.

[0013] Through comprehensive mapping of gluten epitopes using PBMC after gluten challenge, the inventors have found a series of novel gliadin, LMW and HMW glutenin, and avenin epitopes for coeliac disease associated with HLA-DQ2 and HLA-DQ8. Novel epitopes were identified for HLA-DQ2 and HLA-DQ8-associated coeliac disease. HLA-DQ2 and HLA-DQ8 associated coeliac disease are genetically and functionally distinct in terms of the range of T-cell epitopes that are recognized. In addition, three peptides present in avenin proteins of oats also activated peripheral blood mononuclear cells (PBMC) following oats challenge in HLA-DQ2+ coeliac subjects, the first time oats epitopes have been defined. Identification of avenin peptides recognized by T-cells following oats challenge *in vivo* provides a molecular basis for the observed occasional relapse of coeliacs following oat exposure (Lundin KEA et al. 2003 Gut 52:1649-52) and may provide a basis for a predictive diagnostic or genetic de-toxification of oats.

[0014] The data presented here will provide a comprehensive basis for definition of both common "dominant" and occasional "weak" T-cell epitopes in coeliac disease. This information is the platform for functional applications such as diagnostics, food tests, immunotherapeutics and prophylactics, and for design of non-toxic gluten proteins useful in modified grains.

[0015] In particular, through comprehensive mapping of gluten T cell epitopes, the inventors have found epitopes bioactive in coeliac disease in HLA-DQ2+ patients in wheat gliadins and glutenins, having similar core sequences (e.g., SEQ ID NOS: 1-199) and similar extended sequences (e.g., SEQ ID NOS:200-1554, 1555-1655,1656-1671, and 1830-1903). The inventors have also found epitopes bioactive in coeliac disease in HLA-DQ2+ patients in: oat avenins having similar core sequences (e.g., SEQ ID NOS: 1684-1695) and similar extended sequences (e.g., SEQ ID NO: 1672-1683, 1696-1698, and 1764-1768); rye secalins (SEQ ID NOS: 1769-1786) ; and barley hordeins (SEQ ID NOS: 1787-1829). Additionally, epitopes bioactive in coeliac disease in HLA-DQ8+ patients have been identified in wheat gliadins having similar core sequences (e.g., SEQ ID NOS: 1699-1721) and similar extended sequences (e.g., SEQ ID NOS: 1722-1763 and 1908-1927). This comprehensive mapping thus provides dominant epitopes recognized by T cells in coeliac patients. Thus, the methods described herein may be performed using any of these identified epitopes, and analogues and equivalents thereof. Additionally, combinations of epitopes, i.e., "combitopes" or single peptides comprising two or more epitopes, have been shown to induce equivalent responses as the individual epitopes, indicating that several epitopes may be utilized for therapeutic, diagnostic, and other uses of the invention. Such combitopes may be in the form of, e.g., SEQ ID NO: 1906.

[0016] Agents described herein include one or more of the epitopes having the sequences listed recited in SEQ ID NOS: 1578-1579, 1582-1583,1587-1593, 1600-1620,1623-1655, 1656-1671,1672-1698, 1699-1763,1764-1768, 1769-1786,1787-1829, 1895-1903,1906, and 1908-1927 and analogues and equivalents thereof as defined herein.

[0017] Specifically, the present invention provides:

[1] An agent selected from:

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length;

[2] A pharmaceutical composition comprising an agent according to [1] and a pharmaceutically acceptable carrier or diluent;

[3] A composition for use in a method of preventing or treating coeliac disease comprising at least one agent selected from;

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length;

[4] An agent selected from:

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200;
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length; and
(c) an analogue of (a) which is an isolated peptide that binds an antibody, which antibody binds to an epitope comprising SEQ ID NO:200;

for use in a method of treating or preventing coeliac disease in an individual by tolerising the individual to prevent the production of such an antibody;

[5] A method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising:

(a) contacting a sample from the host, *in vitro*, with at least one agent selected from:

(i) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(ii) an analogue of (i) which is an isolated peptide capable of being recognised by a T cell receptor that recognises (i) and which is not more than 50 amino acids in length; and

(b) determining *in vitro* whether T cells in the sample recognise the agent; recognition by the T cells indicating that the individual has, or is susceptible to, coeliac disease;

[6] A composition, for use in a method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising an agent selected from

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length,

said method comprising determining whether T cells of the individual recognise the agent, recognition by the T cells indicating that the individual has, or is susceptible to, coeliac disease;

[7] A method for identifying an analogue of a peptide comprising at least one epitope that comprises SEQ ID NO:200 said method comprising determining, *in vitro,* whether a candidate peptide is recognised by a T cell receptor that recognises an epitope that comprises SEQ ID NO:200, recognition of the candidate peptide indicating that the candidate peptide is an analogue said analogue being of not more than 50 amino acids in length;

[8] A method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising determining, *in vitro* the presence of an antibody that binds to an epitope of a peptide sequence selected from:

(i) a peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(ii) a peptide analogue of (i) which is capable of being recognised by a T cell receptor that recognises (i) and which is not more than 50 amino acids in length;

in a sample from the individual, the presence of the antibody indicating that the individual has, or is susceptible to, coeliac disease;

[9] An *in vitro* method of determining whether a composition is capable of causing coeliac disease comprising determining whether a protein sequence capable of being modified by a transglutaminase to a peptide comprising at least one epitope that comprises SEQ ID NO:200 is present in the composition, the presence of the oligopeptide sequence indicating that the composition is capable of causing coeliac disease;

[10] A kit for carrying out a method according to [5] comprising an agent selected from

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length,

and a means to detect the recognition of the peptide by the T cell; and

[11] An antibody or fragment thereof, specific for SEQ ID NO:200.

[0018] The term "gluten protein" encompasses alpha/beta, gamma and omega gliadins, and low and high molecular weight (LMW and HMW) glutenins in wheat, hordeins in barley, secalins in rye, and avenins in oats. The invention is particularly concerned with gliadins and avenins.

[0019] The agent can be used for the preparation of a diagnostic means for use in a method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual, said method comprising determining whether T cells of the individual recognise the agent, recognition by the T cells indicating that the individual has, or is susceptible to, coeliac disease.

[0020] The finding of epitopes which are modified by transglutaminase also allows diagnosis of coeliac disease based on determining whether other types of immune response to these epitopes are present.

BRIEF DESCRIPTION OF THE DRAWINGS

[0021]

Figure 1 shows a method to generate all possible peptide epitopes from a group of proteins.

Figure 2 shows Genbank accession numbers for gluten gene products present in the Genbank database on 16th June 2003.

Figure 3A shows an expectation maximization (EM) algorithm to analyze data from ELISpot. Figure 3B shows a test on a dataset of patients with coeliac disease.

Figure 4 shows an iterative procedure to find minimal set of responsive epitopes.

Figure 5 shows gliadin and glutenin sequences (SEQ ID NOS: 1-1554). In the "consensus" column, letters in lower case use the standard one letter amino acid code, but letters in upper case have a different meaning: E=[e or q], F=[f or y or w], I=[i or 1 or v], S=[s or t], R=[r or k or h]. The "sequence" column uses the standard one letter amino acid code.

Figure 6 shows gluten peptides that stimulate gamma interferon in PBMC collected 6 days after gluten challenge in HLA-DQ2+ coeliac disease volunteers (SEQ ID NOS: 1555-1655). The indicated 9mers are common to 200 groups of bioactive "structurally" related 20mer peptides. The gluten sequences are ranked according to the bioactivity X proportion of subjects responding.

Figure 7 shows the results of a wheat challenge experiment (SEQ ID NOS: 1656-1671). These peptides gave high quality responses (indicated 'Y') in ten subjects (A-J) after wheat challenge.

Figure 8 shows Avenin peptides (+/-deamidation by tTG) that stimulate interferon-γ in PBMC collected 6 days after gluten challenge in HLA-DQ2+ coeliac disease volunteers (SEQ ID NOS: 1672-1698). Those marked with a * are optimal unique 20mers inducing IFN- y after oats challenge.

Figure 9 shows the most potent 40 20mers (SEQ ID NOS: 1699-1763) in two HLA-DQ8 (not HLA-DQ2) subjects grouped according to shared core sequences. The core sequence of group 6 (QGSFQPSQQ) corresponds to the alpha-gliadin epitope described by van de Wal et al (J. Immunol. 1998, 161(4):1585-1588). The maximum response in Subject A was 271 SFC (medium alone, no peptide response: 4 SFC), and in B it was 26 SFC (medium alone, no peptide response: 1 SFC).

Figure 10 shows the amino acid sequence of A-gliadin (SEQ ID NO: 1928) based on amino acid sequencing.

DETAILED DESCRIPTION OF THE INVENTION

[0022] The term "coeliac disease" encompasses a spectrum of conditions caused by varying degrees of gluten sensitivity, including a severe form characterised by a flat small intestinal mucosa (hyperplastic villous atrophy) and other forms characterised by milder symptoms.

[0023] The individual mentioned above (in the context of diagnosis or therapy) is human. They may have coeliac disease (symptomatic or asymptomatic) or be suspected of having it. They may be on a gluten free diet. They may be

in an acute phase response (for example they may have coeliac disease, but have only ingested gluten in the last 24 hours before which they had been on a gluten free diet for 14 to 28 days).

[0024] The individual may be susceptible to coeliac disease, such as a genetic susceptibility (determined for example by the individual having relatives with coeliac disease or possessing genes which cause predisposition to coeliac disease).

The agent

[0025] The agent is a peptide, for example of length 7 to 50 amino acids, such as 10 to 40, 12 to 35 or 15 to 30 amino acids in length.

[0026] The agent may be the peptide represented by SEQ ID NO: 200 or an epitope comprising sequence that comprises SEQ ID NO: 200 which is an isolated oligopeptide derived from a gluten protein; or an equivalent of these sequences from a naturally occurring gluten protein.

[0027] Thus the epitope may be a derivative of a naturally occurring gluten protein, particularly from a wheat gluten. Such a derivative is typically a fragment of the gluten protein, or a mutated derivative of the whole protein or fragment. Therefore the epitope of the invention does not include the naturally occurring whole gluten protein, and does not include other whole naturally occurring gluten proteins.

[0028] Typically such fragments will be at least 7 amino acids in length (e. g., at least 7, 8, 9, 10,11, 12, 13, 14 or 15 amino acids in length).

[0029] Typically such fragments will be recognised by T cells to at least the same extent that the agents from which they are derived are recognised in any of the assays described herein using samples from coeliac disease patients.

[0030] The agent may be the peptide represented by SEQ ID NO: 200 or a protein comprising a sequence corresponding to SEQ ID NO: 200 (such as fragments of a gluten protein comprising SEQ ID NO: 200, for example after the gluten protein has been treated with transglutaminase). Bioactive fragments of such sequences are also agents of the invention. Typically such fragments will be at least 7 amino acids in length (e. g., at least 7,8, 9,10, 11,12, 13,14 or 15 amino acids in length). Analogues of these sequences, as defined herein, are also agents of the invention.

[0031] In the case where the epitope comprises a sequence equivalent to the above epitopes (including fragments) from another gluten protein (e. g. any of the gluten proteins mentioned herein or any gluten proteins which cause coeliac disease), such equivalent sequences will correspond to a fragment of a gluten protein typically treated (partially or fully) with transglutaminase. Such equivalent peptides can be determined by aligning the sequences of other gluten proteins with the gluten protein from which the original epitope derives (for example using any of the programs mentioned herein). Transglutaminase is commercially available (e. g. Sigma T-5398).

[0032] The agent which is an analogue is capable of being recognised by a TCR which recognises (i). Therefore generally when the analogue is added to T cells in the presence of (i), typically also in the presence of an antigen presenting cell (APC) (such as any of the APCs mentioned herein), the analogue inhibits the recognition of (i), i.e. the analogue is able to compete with (i) in such a system.

[0033] The analogue may be one which is capable of binding the TCR which recognises (i). Such binding can be tested by standard techniques. Such TCRs can be isolated from T cells which have been shown to recognise (i) (e. g. using the method of the invention). Demonstration of the binding of the analogue to the TCRs can then shown by determining whether the TCRs inhibit the binding of the analogue to a substance that binds the analogue, e. g. an antibody to the analogue. Typically the analogue is bound to a class II MHC molecule (e.g. HLA-DQ2) in such an inhibition of binding assay.

[0034] Typically the analogue inhibits the binding of (i) to a TCR. In this case the amount of (i) which can bind the TCR in the presence of the analogue is decreased. This is because the analogue is able to bind the TCR and therefore competes with (i) for binding to the TCR.

[0035] T cells for use in the above binding experiments can be isolated from patients with coeliac disease, for example with the aid of the method of the invention. Other binding characteristics of the analogue may also be the same as (i), and thus typically the analogue binds to the same MHC class II molecule to which the peptide binds (HLA-DQ2 or -DQ8). The analogue typically binds to antibodies specific for (i), and thus inhibits binding of (i) such antibodies.

[0036] The analogue is a peptide. It may have homology with (i), typically at least 70% homology, preferably at least 80, 90%, 95%, 97% or 99% homology with (i), for example over a region of at least 7,8, 9,10, 11,12, 13,14, 15 or more (such as the entire length of the analogue and/or (i), or across the region which contacts the TCR or binds the MHC molecule) contiguous amino acids. Methods of measuring protein homology are well known in the art and it will be understood by those of skill in the art that in the present context, homology is calculated on the basis of amino acid identity (sometimes referred to as "hard homology").

[0037] For example the UWGCG Package provides the BESTFIT program which can be used to calculate homology (for example used on its default settings) (Devereux et al (1984) Nucleic Acids Research 12, p387-395). The PILEUP and BLAST algorithms can be used to calculate homology or align sequences (typically on their default settings), for example as described in Altschul S. F. (1993) J Mol Evol 36:290-300; Altschul, S, F et al (1990) J Mol Biol 215:403-10.

[0038] Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information on the world wide web through the Internet at, for example, "www.ncbi.nlm.nih.gov/". This algorithm involves first identifying high scoring sequence pair (HSPs) by identifying short words of length W in the query sequence that either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighbourhood word score threshold (Altschul *et al,* supra). These initial neighbourhood word hits act as seeds for initiating searches to find HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Extensions for the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLAST program uses as defaults a word length (W) of 11, the BLOSUM62 scoring matrix (see Henikoff and Henikoff(1992) Pro. Natl. Acad. Sci. USA 89: 10915-10919) alignments (B) of 50, expectation (E) of 10, M=5, N=4, and a comparison of both strands.

[0039] The BLAST algorithm performs a statistical analysis of the similarity between two sequences; see e.g., Karlin and Altschul (1993) Proc. Natl. Acad Sci. USA 90: 5873-5787. One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a sequence is considered similar to another sequence if the smallest sum probability in comparison of the first sequence to the second sequence is less than about 1, preferably less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

[0040] The homologous peptide analogues typically differ from (i) by 1, 2, 3, 4, 5, 6, 7, 8 or more mutations (which may be substitutions, deletions or insertions). These mutations may be measured across any of the regions mentioned above in relation to calculating homology. The substitutions are preferably 'conservative'. These are defined according to the following Table. Amino acids in the same block in the second column and preferably in the same line in the third column may be substituted for each other:

ALIPHATIC	Non-polar	G A P
		I L V
	Polar - uncharged	C S T M
		N Q
	Polar - charged	D E
		K R
AROMATIC		H F W Y

[0041] Typically the amino acids in the analogue at the equivalent positions to amino acids in (i) that contribute to binding the MHC molecule or are responsible for the recognition by the TCR, are the same or are conserved.

[0042] Typically the analogue peptide comprises one or more modifications, which may be natural post-translation modifications or artificial modifications. The modification may provide a chemical moiety (typically by substitution of a hydrogen, e. g. of a C-H bond), such as an amino, acetyl, hydroxy or halogen (e. g. fluorine) group or carbohydrate group. Typically the modification is present on the N or C terminus.

[0043] The analogue may comprise one or more non-natural amino acids, for example amino acids with a side chain different from natural amino acids. Generally, the non-natural amino acid will have an N terminus and/or a C terminus. The non-natural amino acid may be an L- or a D- amino acid.

[0044] The analogue typically has a shape, size, flexibility or electronic configuration that is substantially similar to (i). It is typically a derivative of (i).

[0045] In one embodiment the agent is bound to a MHC class II molecule (or a fragment thereof capable of binding the agent). 2, 3, 4 or more of such complexes may be associated or bound to each other, for example using a biotin/streptavidin based system, in which typically 2, 3 or 4 biotin labelled MHC molecules bind to a streptavidin moiety. This bound agent typically inhibits the binding of the (i)/MHC Class II complex to a TCR or antibody which is specific for the complex.

[0046] The analogue is typically designed by computational means and then synthesised using methods known in the art. Alternatively the analogue can be selected from a library of compounds. The library may be a combinatorial library or a display library, such as a phage display library. The library of compounds may be expressed in the display library in the form of being bound to a MHC class II molecule, such as HLA-DQ2 or -DQ8. Analogues are generally selected from the library based on their ability to mimic the binding characteristics (i). Thus they may be selected based on ability

to bind a TCR or antibody which recognises (i).

[0047] Typically analogues will be recognised by T cells to at least the same extent as any of the agents (i), for example at least to the same extent as the equivalent epitope is recognised in any of the assays described herein, typically using T cells from coeliac disease patients. Analogues may be recognised to these extents in vivo and thus may be able to induce coeliac disease symptoms to at least the same extent as any of the agents mentioned herein (e. g. in a human patient or animal model).

[0048] Analogues may be identified in a method comprising determining whether a candidate substance is recognised by a T cell receptor that recognises an epitope of the invention, recognition of the substance indicating that the substance is an analogue. Such TCRs may be any of the TCRs mentioned herein, and may be present on T cells. Any suitable assay mentioned herein can be used to identify the analogue. In one embodiment this method is carried out in vivo. As mentioned above preferred analogues are recognised to at least the same extent as the equivalent epitope, and so the method may be used to identify analogues which are recognised to this extent.

[0049] In one embodiment the method comprises determining whether a candidate substance is able to inhibit the recognition of an epitope of the invention, inhibition of recognition indicating that the substance is an analogue.

[0050] The agent may be a product comprising at least 2,5, 10 or 20 agents as defined by (i) or (ii). Typically the composition comprises epitopes of the invention (or equivalent analogues) from different gluten proteins, such as any of the species or variety of or types of gluten protein mentioned herein. Preferred compositions comprise at least one epitope of the invention, or equivalent analogue, from all of the glutens present in any of the species or variety mentioned herein, or from 2,3, 4 or more of the species mentioned herein (such as from the panel of species consisting of wheat, rye, barley, oats and triticale). Thus, the agent may be monovalent or multivalent.

[0051] According to certain embodiments of the invention, the agent does not have or is not based on a sequence disclosed in WO 02/083722 and/or WO 01/25793 and/or W003/104273 and/or recited in any of SEQ ID NOS: 1555-1577,1580-1581, 1584-1586,1594-1599, 1621-1622 and/or is not an agent derived from A-gliadin, the sequence of which is given in Figure 10.

Diagnosis

[0052] As mentioned above the method of diagnosis of the invention may be based on the detection of T cells that bind the agent or on the detection of antibodies that recognise the agent.

[0053] The T cells that recognise the agent in the method (which includes the use mentioned above) are generally T cells that have been pre-sensitised *in vivo* to one or more gluten proteins. As mentioned above such antigen-experienced T cells have been found to be present in the peripheral blood.

[0054] In the method the T cells can be contacted with the agent *in vitro* or *in vivo*, and determining whether the T cells recognise the agent can be performed *in vitro* or *in vivo*. Thus the invention provides the agent for use in a method of diagnosis practiced on the human body. Different agents are provided for simultaneous, separate or sequential use in such a method.

[0055] The *in vitro* method is typically carried out in aqueous solution into which the agent is added. The solution will also comprise the T cells (and in certain embodiments the APCs discussed below). The term 'contacting' as used herein includes adding the particular substance to the solution.

[0056] Determination of whether the T cells recognise the agent is generally accomplished by detecting a change in the state of the T cells in the presence of the agent or determining whether the T cells bind the agent. The change in state is generally caused by antigen specific functional activity of the T cell after the TCR binds the agent. The change of state may be measured inside (e.g. change in intracellular expression of proteins) or outside (e.g. detection of secreted substances) the T cells.

[0057] The change in state of the T cell may be the start of or increase in secretion of a substance from the T cell, such as a cytokine, especially IFN-γ, IL-2 or TNF-α. Determination of IFN-γ secretion is particularly preferred. The substance can typically be detected by allowing it to bind to a specific binding agent and then measuring the presence of the specific binding agent/substance complex. The specific binding agent is typically an antibody, such as polyclonal or monoclonal antibodies. Antibodies to cytokines are commercially available, or can be made using standard techniques.

[0058] Typically the specific binding agent is immobilised on a solid support. After the substance is allowed to bind the solid support can optionally be washed to remove material which is not specifically bound to the agent. The agent/substance complex may be detected by using a second binding agent that will bind the complex. Typically the second agent binds the substance at a site which is different from the site which binds the first agent. The second agent is preferably an antibody and is labelled directly or indirectly by a detectable label.

[0059] Thus the second agent may be detected by a third agent that is typically labelled directly or indirectly by a detectable label. For example the second agent may comprise a biotin moiety, allowing detection by a third agent which comprises a streptavidin moiety and typically alkaline phosphatase as a detectable label.

[0060] In one embodiment the detection system which is used is the *ex-vivo* ELISPOT assay described in WO 98/23960.

In that assay IFN-γ secreted from the T cell is bound by a first IFN-γ specific antibody that is immobilised on a solid support. The bound IFN-γ is then detected using a second IFN-γ specific antibody which is labelled with a detectable label. Such a labelled antibody can be obtained from MABTECH (Stockholm, Sweden). Other detectable labels which can be used are discussed below.

[0061] The change in state of the T cell that can be measured may be the increase in the uptake of substances by the T cell, such as the uptake of thymidine. The change in state may be an increase in the size of the T cells, or proliferation of the T cells, or a change in cell surface markers on the T cell.

[0062] In one embodiment the change of state is detected by measuring the change in the intracellular expression of proteins, for example the increase in intracellular expression of any of the cytokines mentioned above. Such intracellular changes may be detected by contacting the inside of the T cell with a moiety that binds the expressed proteins in a specific manner and which allows sorting of the T cells by flow cytometry.

[0063] In one embodiment when binding the TCR the agent is bound to an MHC class II molecule (typically HLA-DQ2 or -DQ8), which is typically present on the surface of an antigen presenting cell (APC). However as mentioned herein other agents can bind a TCR without the need to also bind an MHC molecule.

[0064] Generally the T cells which are contacted in the method are taken from the individual in a blood sample, although other types of samples which contain T cells can be used. The sample may be added directly to the assay or may be processed first. Typically the processing may comprise diluting of the sample, for example with water or buffer. Typically the sample is diluted from 1.5 to 100 fold, for example 2 to 50 or 5 to 10 fold.

[0065] The processing may comprise separation of components of the sample. Typically mononuclear cells (MCs) are separated from the samples. The MCs will comprise the T cells and APCs. Thus in the method the APCs present in the separated MCs can present the peptide to the T cells. In another embodiment only T cells, such as only CD4 T cells, can be purified from the sample. PBMCs, MCs and T cells can be separated from the sample using techniques known in the art, such as those described in Lalvani et al (1997) J. Exp. Med. 186, p859-865.

[0066] In one embodiment, the T cells used in the assay are in the form of unprocessed or diluted samples, or are freshly isolated T cells (such as in the form of freshly isolated MCs or PBMCs) which are used directly *ex vivo,* i.e. they are not cultured before being used in the method. Thus the T cells have not been restimulated in an antigen specific manner *in vitro.* However the T cells can be cultured before use, for example in the presence of one or more of the agents, and generally also exogenous growth promoting cytokines. During culturing the agent(s) are typically present on the surface of APCs, such as the APC used in the method. Pre-culturing of the T cells may lead to an increase in the sensitivity of the method. Thus the T cells can be converted into cell lines, such as short term cell lines (for example as described in Ota et al (1990) Nature 346, p183-187).

[0067] The APC that is typically present in the method may be from the same individual as the T cell or from a different host. The APC may be a naturally occurring APC or an artificial APC. The APC is a cell that is capable of presenting the peptide to a T cell. It is typically a B cell, dendritic cell or macrophage. It is typically separated from the same sample as the T cell and is typically co-purified with the T cell. Thus the APC may be present in MCs or PBMCs. The APC is typically a freshly isolated *ex vivo* cell or a cultured cell. It may be in the form of a cell line, such as a short term or immortalised cell line. The APC may express empty MHC class II molecules on its surface.

[0068] In the method one or more (different) agents may be used. Typically the T cells derived from the sample can be placed into an assay with all the agents which it is intended to test or the T cells can be divided and placed into separate assays each of which contain one or more of the agents.

[0069] The invention also provides the agents such as two or more of any of the agents mentioned herein (e.g. the combinations of agents which are present in the composition agent discussed above) for simultaneous separate or sequential use (eg. for *in vivo* use).

[0070] In one embodiment agent *per se* is added directly to an assay comprising T cells and APCs. As discussed above the T cells and APCs in such an assay could be in the form of MCs. When agents that can be recognised by the T cell without the need for presentation by APCs are used then APCs are not required. Analogues which mimic the original (i) bound to a MHC molecule are an example of such an agent.

[0071] In one embodiment the agent is provided to the APC in the absence of the T cell. The APC is then provided to the T cell, typically after being allowed to present the agent on its surface. The peptide may have been taken up inside the APC and presented, or simply be taken up onto the surface without entering inside the APC.

[0072] The duration for which the agent is contacted with the T cells will vary depending on the method used for determining recognition of the peptide. Typically 10^5 to 10^7, preferably $5x10^5$ to 10^6 PBMCs are added to each assay. In the case where agent is added directly to the assay its concentration is from 10^{-1} to 10^3μg/ml, preferably 0.5 to 50μg/ml or 1 to 10μg/ml.

[0073] Typically the length of time for which the T cells are incubated with the agent is from 4 to 24 hours, preferably 6 to 16 hours. When using *ex vivo* PBMCs it has been found that $0.3x10^6$ PBMCs can be incubated in 10μg/ml of peptide for 12 hours at 37°C.

[0074] The determination of the recognition of the agent by the T cells may be done by measuring the binding of the

agent to the T cells (this can be carried out using any suitable binding assay format discussed herein). Typically T cells which bind the agent can be sorted based on this binding, for example using a FACS machine. The presence of T cells that recognise the agent will be deemed to occur if the frequency of cells sorted using the agent is above a "control" value. The frequency of antigen-experienced T cells is generally 1 in 10^6 to 1 in 10^3, and therefore whether or not the sorted cells are antigen-experienced T cells can be determined.

[0075] The determination of the recognition of the agent by the T cells may be measured *in vivo.* Typically the agent is administered to the host and then a response which indicates recognition of the agent may be measured. The agent is typically administered intradermally or epidermally. The agent is typically administered by contacting with the outside of the skin, and may be retained at the site with the aid of a plaster or dressing. Alternatively the agent may be administered by needle, such as by injection, but can also be administered by other methods such as ballistics (e.g. the ballistics techniques which have been used to deliver nucleic acids). EP-A-0693119 describes techniques that can typically be used to administer the agent. Typically from 0.001 to 1000 μg, for example from 0.01 to 100 μg or 0.1 to 10 μg of agent is administered.

[0076] In one embodiment a product can be administered which is capable of providing the agent *in vivo.* Thus a polynucleotide capable of expressing the agent can be administered, typically in any of the ways described above for the administration of the agent. The polynucleotide typically has any of the characteristics of the polynucleotide provided by the invention which is discussed below. The agent is expressed from the polynucleotide *in vivo.* Typically from 0.001 to 1000 μg, for example from 0.01 to 100 μg or 0.1 to 10 μg of polynucleotide is administered.

[0077] Recognition of the agent administered to the skin is typically indicated by the occurrence of inflammation (e.g. induration, erythema or oedema) at the site of administration. This is generally measured by visual examination of the site.

[0078] The method of diagnosis based on the detection of an antibody that binds the agent is typically carried out by contacting a sample from the individual (such as any of the samples mentioned here, optionally processed in any manner mentioned herein) with the agent and determining whether an antibody in the sample binds the agent, such a binding indicating that the individual has, or is susceptible to coeliac disease. Any suitable format of binding assay may be used, such as any such format mentioned herein.

Therapy

[0079] The identification of the immunodominant epitope and other epitopes described herein allows therapeutic products to be made which target the T cells which recognise this epitope (such T cells being ones which participate in the immune response against gluten proteins). These findings also allow the prevention or treatment of coeliac disease by suppressing (by tolerisation) an antibody or T cell response to the epitope(s).

[0080] Certain agents of the invention bind the TCR that recognises the epitope of the invention (as measured using any of the binding assays discussed above) and cause tolerisation of the T cell that carries the TCR. Such agents, optionally in association with a carrier, can therefore be used to prevent or treat coeliac disease.

[0081] Generally tolerisation can be caused by the same peptides which can (after being recognised by the TCR) cause antigen specific functional activity of the T cell (such as any such activity mentioned herein, e.g. secretion of cytokines). Such agents cause tolerisation when they are presented to the immune system in a 'tolerising' context.

[0082] Tolerisation leads to a decrease in the recognition of a T cell or antibody epitope by the immune system. In the case of a T cell epitope this can be caused by the deletion or anergising of T cells that recognise the epitope. Thus T cell activity (for example as measured in suitable assays mentioned herein) in response to the epitope is decreased. Tolerisation of an antibody response means that a decreased amount of specific antibody to the epitope is produced when the epitope is administered.

[0083] Methods of presenting antigens to the immune system in such a context are known and are described for example in Yoshida et al. Clin. Immunol. Immunopathol. 82, 207-215 (1997), Thurau et al. Clin. Exp. Immunol. 109, 370-6 (1997), and Weiner et al. Res. Immunol. 148, 528-33 (1997). In particular certain routes of administration can cause tolerisation, such as oral, nasal or intraperitoneal. Tolerisation may also be accomplished via dendritic cells and tetramers presenting peptide. Particular products which cause tolerisation may be administered (e. g. in a composition that also comprises the agent) to the individual. Such products include cytokines, such as cytokines that favour a Th2 response (e. g. IL-4, TGF-β; or IL-10). Products or agent may be administered at a dose that causes tolerisation.

[0084] In one embodiment, the tolerising (T cell and antibody tolerising) agents are present in a composition comprising at least 2,4, 6 or more agents which tolerise to different epitopes of the invention, for example to the combinations of epitopes discussed above in relation to the agents which are a product comprising more than one substance.

Tasting whether a Composition is capable of causing coeliac disease

[0085] As mentioned above the invention provides a method of determining whether a composition is capable of causing coeliac disease comprising detecting the presence of a protein sequence which is capable of being modified

by a transglutaminase to as sequence comprising the agent or epitope of the invention (such transglutaminase activity may be a human intestinal transglutaminase activity).

[0086] Typically this is performed by using a binding assay in which a moiety which binds to the sequence in a specific manner is contacted with the composition and the formation of sequence/moiety complex is detected and used to ascertain the presence of the agent. Such a moiety may be any suitable substance (or type of substance) mentioned herein, and is typically a specific antibody. Any suitable format of binding assay can be used (such as those mentioned herein).

[0087] In one embodiment, the composition is contacted with at least 2,5, 10 or more antibodies which are specific for epitopes of the invention from different gluten proteins, for example a panel of antibodies capable of recognising the combinations of epitopes discussed above in relation to agents of the invention which are a product comprising more than one substance.

[0088] The composition typically comprises material from a plant that expresses a gluten protein which is capable of causing coeliac disease (for example any of the gluten proteins or plants mentioned herein). Such material may be a plant part, such as a harvested product (e. g. seed). The material may be processed products of the plant material (e.g. any such product mentioned herein), such as a flour or food that comprises the gluten protein. The processing of food material and testing in suitable binding assays is routine, for example as mentioned in Kricka LJ, J. Biolumin. Chemilumin. 13, 189-93 (1998).

Binding essays

[0089] The determination of binding between any two substances mentioned herein may be done by measuring a characteristic of either or both substances that changes upon binding, such as a spectroscopic change.

[0090] The binding assay format may be a 'band shift' system. This involves determining whether the presence of one substance (such as a candidate substance) advances or retards the progress of the other substance during gel electrophoresis.

[0091] The format may be a competitive binding method which determines whether the one substance is able to inhibit the binding of the other substance to an agent which is known to bind the other substance, such as a specific antibody.

Kits

[0092] The invention also provides a kit for carrying out the method comprising one or more agents and a means to detect the recognition of the agent by the T cell. Typically the different agents are provided for simultaneous, separate or sequential use. Typically the means to detect recognition allows or aids detection based on the techniques discussed above.

[0093] Thus the means may allow detection of a substance secreted by the T cells after recognition. The kit may thus additionally include a specific binding moiety for the substance, such as an antibody. The moiety is typically specific for IFN-γ. The moiety is typically immobilised on a solid support. This means that after binding the moiety the substance will remain in the vicinity of the T cell which secreted it. Thus "spots" of substance/moiety complex are formed on the support, each spot representing a T cell which is secreting the substance. Quantifying the spots, and typically comparing against a control, allows determination of recognition of the agent.

[0094] The kit may also comprise a means to detect the substance/moiety complex. A detectable change may occur in the moiety itself after binding the substance, such as a colour change. Alternatively a second moiety directly or indirectly labelled for detection may be allowed to bind the substance/moiety complex to allow the determination of the spots. As discussed above the second moiety may be specific for the substance, but binds a different site on the substance than the first moiety.

[0095] The immobilised support may be a plate with wells, such as a microtitre plate. Each assay can therefore be carried out in a separate well in the plate.

[0096] The kit may additionally comprise medium for the T cells, detection moieties or washing buffers to be used in the detection steps. The kit may additionally comprise reagents suitable for the separation from the sample, such as the separation of PBMCs or T cells from the sample. The kit may be designed to allow detection of the T cells directly in the sample without requiring any separation of the components of the sample.

[0097] The kit may comprise an instrument which allows administration of the agent, such as intradermal or epidermal administration. Typically such an instrument comprises plaster, dressing or one or more needles. The instrument may allow ballistic delivery of the agent. The agent in the kit may be in the form of a pharmaceutical composition.

[0098] The kit may also comprise controls, such as positive or negative controls. The positive control may allow the detection system to be tested. Thus the positive control typically mimics recognition of the agent in any of the above methods. Typically in the kits designed to determine recognition in vitro the positive control is a cytokine. In the kit designed to detect *in vivo* recognition of the agent the positive control may be antigen to which most individuals should

response.

[0099] The kit may also comprise a means to take a sample containing T cells from the host, such as a blood sample. The kit may comprise a means to separate mononuclear cells or T cells from a sample from the host.

Antibodies

[0100] The invention also provides monoclonal or polyclonal antibodies which specifically recognise the agents (such as the epitope of the invention) of the invention, and methods of making such antibodies. Antibodies of the invention bind specifically to these substances of the invention.

[0101] For the purposes of this invention, the term "antibody" includes antibody fragments such as Fv, F(ab) F(ab')2 as well as single-chain antibodies.

[0102] A method for producing a polyclonal antibody comprises immunising a suitable host animal, for example an experimental animal, with the immunogen and isolating immunoglobulins from the serum. The animal may therefore be inoculated with the immunogen, blood subsequently removed from the animal and the IgG fraction purified. A method for producing a monoclonal antibody comprises immortalising cells which produce the desired antibody. Hybridoma cells may be produced by fusing spleen cells from an inoculated experimental animal with tumour cells (Kohler and Milstein (1975) Nature 256, 495-497).

[0103] An immortalized cell producing the desired antibody may be selected by a conventional procedure. The hybridomas may be grown in culture or injected intraperitoneally for formation of ascites fluid or into the blood stream of an allogenic host or immunocompromised host. Human antibody may be prepared by in vitro immunisation of human lymphocytes, followed by transformation of the lymphocytes with Epstein-Barr virus.

[0104] For the production of both monoclonal and polyclonal antibodies, the experimental animal is suitably a goat, rabbit, rat or mouse. If desired, the immunogen may be administered as a conjugate in which the immunogen is coupled, for example via a side chain of one of the amino acid residues, to a suitable carrier. The carrier molecule is typically a physiologically acceptable carrier. The antibody obtained may be isolated and, if desired, purified.

[0105] The agent or antibody of the invention, may carry a detectable label. Detectable labels which allow detection of the secreted substance by visual inspection, optionally with the aid of an optical magnifying means, are preferred. Such a system is typically based on an enzyme label which causes colour change in a substrate, for example alkaline phosphatase causing a colour change in a substrate. Such substrates are commercially available, e. g. from BioRad. Other suitable labels include other enzymes such as peroxidase, or protein labels, such as biotin; or radioisotopes, such as ^{32}P or ^{35}S. The above labels may be detected using known techniques.

[0106] Agents or antibodies of the invention may be in substantially purified form. They may be in substantially isolated form, in which case they will generally comprise at least 80% e. g. at least 90, 95, 97 or 99% of the peptide, antibody or dry mass in the preparation. The agent or antibody is typically substantially free of other cellular components. The agent or antibody may be used in such a substantially isolated, purified or free form in the method or be present in such forms in the kit.

[0107] The invention provides therapeutic (including prophylactic) agents or diagnostic substances (the agents of the invention). These substances are formulated for clinical administration by mixing them with a pharmaceutically acceptable carrier or diluent. For example they can be formulated for topical, parenteral, intravenous, intramuscular, subcutaneous, intraocular, intradermal, epidermal or transdermal administration. The substances may be mixed with any vehicle which is pharmaceutically acceptable and appropriate for the desired route of administration. The pharmaceutically carrier or diluent for injection may be, for example, a sterile or isotonic solution such as Water for Injection or physiological saline, or a carrier particle for ballistic delivery.

[0108] The dose of the substances may be adjusted according to various parameters, especially according to the agent used; the age, weight and condition of the patient to be treated; the mode of administration used; the severity of the condition to be treated; and the required clinical regimen. As a guide, the amount of substance administered by injection is suitably from 0.01 mg/kg to 30 mg/kg, preferably from 0.1 mg/kg to 10 mg/kg.

[0109] The routes of administration and dosages described are intended only as a guide since a skilled practitioner will be able to determine readily the optimum route of administration and dosage for any particular patient and condition.

[0110] The substances of the invention may thus be used in a method of treatment of the human or animal body, or in a diagnostic method practised on the human body. In particular they may be used in a method of treating or preventing coeliac disease. The invention also provide the agents for use in a method of manufacture of a medicament for treating or preventing coeliac disease. Thus the invention provides for a method of preventing or treating coeliac disease comprising administering to a human in need thereof a substance of the invention (typically a non-toxic effective amount thereof).

[0111] The agent of the invention can be made using standard synthetic chemistry techniques, such as by use of an automated synthesizer. The agent may be made from a longer polypeptide e.g. a fusion protein, which polypeptide typically comprises the sequence of the peptide. The peptide may be derived from the polypeptide by for example

hydrolysing the polypeptide, such as using a protease; or by physically breaking the polypeptide. The polynucleotide of the invention can be made using standard techniques, such as by using a synthesiser.

Deamidation

[0112] Where a sequence described herein includes a Gln residue, the invention also provides that sequence where the Gln residue has been deamidated to a Glu residue. One or more (e.g., 1, 2, 3, 4, 5, etc.) Gln residue(s) per sequence may be deamidated, but when there is more than one Gln residue, not all of them must be deamidated. Preferably, the Gln residues that are deamidated are those susceptible to deamidation by transglutaminase.

[0113] Examples where Gln may be deamidated are given in the sequence listing. For example, residue 4 of SEQ ID NO:1 can be a Gln residue or a Glu residue, residue 6 of SEQ ID NO:2 can be a Gin residue or a Glu residue, residues 4 and 7 of SEQ ID NO: 6 can each independently be Gln or Glu residues, etc. The Gln residues that are susceptible to deamidation, and their deamidated Glu counterparts, are referred to as "Glx" residues.

[0114] Where the agent includes more than one Glx residue, these may be arranged in any configuration. For example, the Glx residues may be consecutive residues, and/or may be separated by one or more (e. g., 1, 2, 3, 4, 5, 6, 7, 8, etc.) other residues. As mentioned above, for HLA-DQ8 epitopes, the agent preferably comprises a Glx residue that is separated by seven residues from another Glx residue. Preferred agents of the invention are deamidated agents, i.e., the agent comprises the one or more Glx residues in the Glu form. This can be achieved in various ways, e.g., by including Glu residues during production, or by converting Gln residues to Glu by deamidation. Conversion of Gln to Glu can be achieved by treating an agent that contains Gln residues that are susceptible to deamidation with a deamidating agent. The one or more Gln residues are preferably deamidated to Glu by transglutaminase, for example as described in the examples.

[0115] The skilled person will be able to determine which particular Gln residues in the agent are susceptible to deamidation and thus which residues should be Glu residues arising from deamidation of a Gln residue. For example, Gln-containing sequences susceptible to deamidation by transglutaminase generally conform to a motif: e. g., QXPX, QXPF(Y), QXX(FYMILVW), QXPF, QXX(FY), PQ(QL)P(FY)P. For example, the sequence PQ(QL)P(FY) P facilitates deamidation of the underlined Q at position 2 by transglutaminase.

[0116] In particular, agents comprising the deamidated version of SEQ ID NO: 200 are preferred (where such sequences are not already deamidated). Most preferably, the agents of the invention comprise the transglutaminase-deamidated version of SEQ ID NO: 200 (again, where not already deamidated). Analogues of these agents, as defined herein, are also encompassed within the scope of the invention.

[0117] **EXAMPLES** (Reference Examples except insofar as they relate to the agent of the present invention) - The invention is illustrated by the following nonlimiting Examples:

Initial gliadin epitope screening library

[0118] In initial experiments involving 29 HLA-DQ2+ individuals with coeliac disease on long-term gluten free diet, interferon-gamma ELISPOT assays were used to screen a previous Pepset (described in WO 03/104273) initially as pools of peptides and then in 15 subjects as individual peptides with and without deamidation by tTG. This Pepset library consisted of 652 20mer gliadin peptides spanning all unique 12mers contained within all Genbank entries described as wheat gliadins found in September 2001. This Pepset library was designed "manually" from gene-derived protein sequences aligned using ClustalW software (MegAlign) arranged into phylogenetic groupings.

[0119] Approximately 0.6 micromole of each of 652 of the 20mers was provided. Two marker 20mer peptides were included in each set of 96 (VLQQHNIAHGSSQVLQESTY - peptide 161, and IKDFHVYFRESRDALWKGPG) and were characterized by reverse phase-HPLC and amino acid sequence analysis. Average purities of these marker peptides were 19% and 50%, respectively. Peptides were initially dissolved in acetonitrile (10%) and Hepes 100mM to 10mg/ml. The final concentration of individual peptides incubated with PBMC for the IFNγ ELISpot assays was 20 mcg/ml. These peptides were deamidated by incubation with guinea pig tissue tTG (Sigma T5398) in the ratio 100:32 mcg/ml for two hours at 37°C. Peptides solutions were stored at -20°C and freshly thawed prior to use. These studies were conducted in Oxford, UK. ELIspot assays were performed as described for those conducted in Melbourne, Australia (all other studies described herein). "Oxford" data regarding subject responses to individual peptides was pooled with "Melbourne" data for subsequent "minimal" epitope analysis in the "EM algorithm" (see below).

Second round gliadin epitope screening library

[0120] A second round gliadin epitope library was designed according the bioactive sequences identified from the initial gliadin epitope screening library of 652 20mers. Gliadin 20mers with mean bioactivity equivalent to >5% of the most potent gliadin 20mer (91: PQPFPPQLPYPQPQLPYPQP) in 15 HLA-DQ2+ subjects assessed with all 652 deam-

idated 20mers were defined. Since earlier studies (see WO 03/104273) indicated that deamidated pools of this Pepset were more potent than without deamidation, glutamine residues within bioactive 20mers potentially deamidated by tTG were identified according to the motif QXPX, QXZ (FYWILVM) where X is any amino acid except proline, and P is proline, Z is any amino acid, and FYWILVM represent hydrophobic amino acids (consistent with the motifs for tTG-mediated deamidation published by Vader W. et al J Exp Med 2002 J. Exp. Med. 195:643-649, PCT WO 03/066079, and Fleckenstein B. 2002. J Biol Chem 277:34109-16).

[0121] 12mer peptides were then identified in which each potential deamidation site could be in position 4, 6 or 7 in the 9mer located within HLA-DQ2 binding groove (HLA-DQ2 anchors at these positions show a preference for glutamate). Candidate 12mer core epitope sequences were then flanked with glycine followed by the N-terminal residue present in the parent gliadin polypeptide and at the C-terminal by the C-terminal residue present in the parent gliadin polypeptide followed by glycine (i.e. GXXXXXXXQXXXXXXG).

[0122] Peptides were synthesised with glutamine or glutamate in position 9. Peptides (100mcg/ml) (+/- deamidation by tTG) were then assessed in interferon gamma ELISPOT assays using PBMC from 15 HLA-DQ2+ coeliac volunteers after gluten challenge. Results of these assays were analysed according to the EM algorithm (see below). In addition, the most potent distinct peptides were synthesised and purified to >80% (Mimotopes) and assessed in interferon gamma ELISPOT assays using PBMC from 15 HLA-DQ2+ coeliac volunteers after wheat gluten challenge.

Complete gluten epitope screening library

[0123] To make practical the design of a substantially larger peptide library spanning all wheat gliadin and glutenin, rye, barley, and oat gluten-like proteins (prolamines), and to confirm data from the previous gliadin peptide library, an iterative algorithm was developed to automate design of a minimal set of 20mers including all unique 12mers (excluding signal peptide sequences) in gluten proteins. The ScanSet algorithm is shown in Figure 1.

[0124] The method tests for all possible peptide epitopes from a group of proteins whether they are potential antigens in a range of patients. T-cell epitopes range in size between 9 and 15 AA. To test all possible 12mers in a set of proteins, becomes quickly unfeasible because of the high numbers.

[0125] Here we use the fact that, for example, a 20mer peptide can cover up to 9 different 12mers. We therefore developed a combinatorial approach to cover all possible 12mers represented in a family of proteins.

[0126] 20 amino acid (20mer) long peptides are generated that are tested as antigens, and that cover all 12mer peptide sequences that exist in the group of proteins. We define the length of peptides to generate as *L* (e.g. 20) and the length of the epitopes we want to cover as *S*. We developed a computer program that generates all uniquely occurring *L*mers from a set of proteins. Further, we generate all uniquely occurring *S*mers from this set of proteins. Next we select a set of *N L*mers that contains all sequences of *L*mers. Figure 1 outlines how this algorithm works.

[0127] On 16 June 2003, Genbank contained accession numbers for 53 alpha/beta, 53 gamma and 2 omega gliadins, and 77 LMW and 55 HMW glutenins from T. aestivum, 59 hordeins, 14 secalins, and 20 avenins (see Figure 2). In total, ScanSet identified 18117 unique 12mers contained in the 225 gluten gene products.

[0128] All unique gluten 12mers could be subsumed in 2922 20mers. These 20mers were synthesised in a Pepset peptide library (Mimotopes Inc., Melbourne, Australia). Pepset peptides were synthesized in batches of 96 (Mimotopes Inc., Melbourne Australia). Approximately 0.7 to 1.3 micromole of each of 2922 20mers was provided. Two marker 20mer peptides were included in each set of 96 (one representative peptide from the 94 other peptides on each particular plate, and IKDFHVYFRESRDALWKGPG) and were characterized by reverse phase-HPLC and mass spectroscopy. Average purities of these marker peptides were 36% (range: 5-68%) and 64% (range:55-71%), respectively.

[0129] Peptides were initially dissolved in aqueous acetonitrile (50%). Peptides in aqueous acetontrile were transferred to sterile 96-well plates and diluted in sterile PBS with 1mM calcium (250 mcg/ml) and then incubated with tTG (25 mcg/ml) (Sigma T5398) for 6h 37°C and then stored frozen (-20°C) until use.

[0130] Subjects all had biopsy-proven coeliac disease and had followed a strict gluten free diet for at least 6 months. All subjects possessed *HLA-DQB01*02* (HLA-DQ2) alone (n=100) or *HLA-DQA1*03* and *HLA-DQB1*0302* (HLA-DQ8) alone (n=5). In all cases, tTG-IgA was assessed before gluten challenge and was in the normal range (30% of initial volunteers were found to have elevated tTG-IgA and were excluded since chronic gluten exposure is associated with failure to induce peripheral blood gluten-specific T-cells by short-term gluten challenge). Volunteers consumed Baker's Delight "white bread block loaf" (200g daily for three days) or Uncle Toby's oats (100g daily for three days). All but three subjects completed the three day challenge (one withdrew after first mouthful of bread, and the other two vomited after initial two slices of bread. Data from the latter two were included in subsequent analysis). Blood (300ml) was drawn six days after commencing gluten challenge. Gluten peptide-specific IFNγ ELISpot responses have not been found in our previous studies, and so "pre-challenge" blood was not assessed in this set of experiments (Anderson, RP et al 2000. Nat. Med. 6:337-342., WO 01/25793, WO 03/104273).

[0131] IFNγ ELISpot assays (Mabtech, Sweden) were performed in 96-well plates (MAIP S-45, Millipore) in which each well contained 25mcl of peptide solution and 100mcl of PBMC (2-8x10^5/well) in RPMI containing 10% heat inac-

tivated human AB serum. After development and drying, IFNγ ELISpot plates were assessed using the MAIP automated ELISpot plate counter. Data was then analysed according to a novel algorithm (Expectation Maximization: EM) to define and quantify interferon-gamma responses to 9mer sequences contained within the peptide library (see Figure 3 and below). 9mer peptides were then rationalised according to an algorithm that assumes redundancy in T-cell recognition, the "IterativeCluster" algorithm (see Figure 4 and below), by allowing groups of amino acids with similar chemical properties at any one position in the 9mer, or for glutamate to replace glutamine at any position (assuming deamidation may have occurred).

[0132] Since there were data sets from only two HLA-DQ8+ individuals who were not also HLA-DQ2+, and these were utilizing only the 721 wheat gliadin 20mers from the "Complete gluten epitope screening library", bioactive peptides were identified by taking the average rank of peptide-specific IFNγ ELISPOT responses in the two subjects. For prediction of likely HLA-DQ8-restricted gliadin epitopes, it was assumed that a glutamine residue susceptible to tTG-mediated deamidation occupied either position 1 or 9 in potential 9mer core regions of epitopes, consistent with the HLA-DQ8 binding motif and the findings of van de Wal et al (van de Wal, Y. et al 1998. J. Immunol. 161 (4):1585-1588).

Expectation Maximization (EM) algorithm to analyze data from ELISpot:

[0133] Figure 3 shows an algorithm to analyze data coming from an assay using the ELISpot. T-cell responses to different peptides are measured in 96 well plates using T-cell assays. Assays are performed on many patients using many different peptide antigens. The result of the T-cell assays can be summarized in a table where the rows represent peptides and the columns patients and the individual measurements (counts) are in the table (e.g., see Figure 3B). The purpose of the EM algorithm is to differentiate between response and non response of a patient to a peptide and to estimate a mean rate of response and a proportion of people responding for each peptide.

[0134] Responses are measured for a number of different patients (i will be used to indicate the patient) and for many different peptides (j will be used to indicate the peptide). Each measurement (*yij*) represents a count of T-cells from patient *i* responding to peptide *j.* In order to estimate, whether a measurement for a certain peptide in a patient can be called a response or whether it is more likely to be coming from a background distribution, we propose a model for an incomplete data problem, with *yij* being the observed count of spots and *zij* an unobserved indicator, whether person *i* responds to peptide *j*.

[0135] The observed number of counts *yij* are modelled to come from independent Poisson distributions: poisson(αi, λj), if patient *i* is responding to peptide *j,* i.e. *zij* = 1, and poisson(αi, $\lambda 0$), if patient *i* is not responding to peptide *j*, i.e. *zij* = 0.

- Complete data: *yij* (observed counts), *zij* (response indicator, not observed).
- Parameters: $\theta = (\alpha i, \lambda j, \lambda 0, pj)$
    - αi: Patients overall responsiveness.
    - λj: Peptide induced rate of response.
    - $\lambda 0$: Background rate of response.
    - *pj*: Proportion of people responding to peptide *j.*

EM algorithm:

[0136]

- Set variables initially to random values
- E-step: compute likelihood
- M-step: maximize likelihood function
- Iterate E- and M-step

Iterative procedure to find minimal set of responsive epitopes

[0137] A program to compute a minimal set of peptides for use in a vaccine based on the T-cell responses estimated in the EM algorithm was developed. We measured T-Cell responses to Lmers from a group of proteins. The peptides were generated to cover all possible *S*mers. We estimated the following parameters for the response by an EM algorithm: rate of response, number of people responding, proportion of people responding. The proportion of people responding multiplied with the estimated rate of response is used as a criterion to define epitopes which are good antigens. Many of the measured Lmers contain the same *S*mer epitopes. In order to find the epitopes (Smers) which can explain all the responses in *L*mers we select the *S*mer which is contained in Lmers that in the mean have the highest responses. Then we remove all Lmers that contain this *S*mer from our measurements. Next we select the *S*mer with the highest responses

in the remaining *L*mers. We iterate this procedure until no *S*mers with responses higher than a specified cutoff exist. We use several iterations with different cutoffs. This process is sketched in Figure 4. The such defined list of clustered Lmers can be used as a basis to define the optimal epitopes and select peptides that function as good antigens.

HLA-DQ2 epitopes in wheat gluten

[0138] HLA-DQ2 epitopes in wheat gliadins and glutenins were identified using PBMC collected on day 6 after commencing gluten challenge in a total of 76 HLA-DQ2+ individuals in gamma-interferon ELISPOT assays (Initial gliadin epitope library: n=15, Second round gliadin epitope screening library: n=15, Complete gluten epitope screening library: n= 46). All data relating to individual peptide responses in coeliac subjects was pooled and analysed by the EM algorithm.

[0139] A series of 9mer sequences were identified and ordered according to the intensity of gamma-interferon responses and the proportion of individuals responding (see Figure 5). Many of the sequences identified could be grouped in "superfamilies" allowing for several different amino acids with similar chemical properties to be present at any one position in the putative epitope (see Figure 6). For example, in "Sequence 1" of Figure 6 (SEQ ID NO: 1555) P(QR)P(QE)LP(FY)PQ, glutamine (Q) or arginine (R) are both accepted at position 2 except that Q generates a substantially more bioactive epitope.

[0140] By reviewing the 110 most "active" 9mer sequences identified by the EM algorithm, the "list" of 9mer motifs could be condensed to 41 9mers, many of which overlapped (for example "Sequence 1" and "2" (SEQ ID NOS: 1555 and 1558 respectively) overlap by 7 residues and are both present in A-gliadin 57-73 QE65). In selected cases, high-grade peptides were synthesised and confirmed the bioactivity of peptides identified by the EM algorithm (see Figure 7).

HLA-DQ2 epitopes in oats avenins

[0141] Avenin peptides were assessed after challenge with oats (n=30 subjects) or after wheat bread (n=8) in HLA-DQ2+ coeliac subjects. ELISPOT responses were found for the peptides found in Figure 8. One of the reactive avenin peptides was homologous to a sequence in wheat gluten (SEQ ID NO: 1590).

Oats (avenin) high quality peptide studies

[0142] High grade avenin peptides were assessed 3 days after completing oats challenge with pure wheat-free oats, 100g/d for 3 days ("day 6" PBMC interferon gamma ELISPOT responses). These peptides were designed upon peptides previously defined using the screening grade ("first round") avenin peptide library and on potential deamidation sites. There were 25 peptides (as 16mers) with purity verified by HPLC as >80%, and sequences confirmed by mass spectroscopy.

[0143] Interferon gamma ELISPOT responses to the high grade avenin peptides following deamidation by tTG were compared in 18 subjects with DQ2+ coeliac disease.

[0144] The dominant (>70% maximal response) peptides after oats challenge included: EQQFGQNIFSGFSVQL (SEQ ID NO: 1764) (11/18 subjects), QLRCPAIHSVVQAIIL (SEQ ID NO: 1765) (4/18 subjects), and QYQPYPEQEQPILQQQ (SEQ ID NO: 1766) (3/18 subjects). 2/18 subjects did not have avenin specific responses (defined by SFU (spot forming units) > 3 X blank) and 6/18 subjects mean maximal SFU were less than 10. Two additional peptides elicited positive responses: QIPEQLRCPAIHSVVQ (SEQ ID NO: 1767) (3/18 subjects) and EQYQPEQQPFMQPL (SEQ ID NO: 1768) (>40% maximal peptide response in 5/18 subjects). The panel of 25 peptides included several peptides similar to peptide 1490 (SEQYQPYPEQQEPFVQ) reported in Arentz-Hansen, PLoS Medicine (Oct. 2004, vol. 1, issue 1 (84-92), however, that peptide induced a strong positive response in only one subject, and far weaker response in 5 subjects.

[0145] Interferon gamma ELISPOT responses to high grade avenin peptides were absent prior to gluten challenge, and were blocked by pre-treatment of PBMC with anti-HLA DQ but not anti-HLA DR antibody.

Rye and barley screening peptide libraries

[0146] Secalin and hordein 20mer first round peptide libraries were assessed 3 days after completing rye (bread, 100g/d for 3 days) or barley (boiled, 100g/d for 3 days) challenge ("day 6" PBMC interferon gamma ELISPOT responses). Although iterative analysis using 2nd and 3rd round peptide libraries to define epitopes has not yet been performed, the 20mers pre-treated with tTG found to induce "potent" responses shared substantial structural similarity to the bioactive peptides identified after wheat challenge. However, the dominant peptide sequences after rye or barley challenge did not include peptides with the PQPQLPY sequence found to be dominant after wheat challenge. The dominant (>70% maximal response) 20mer after rye challenge was usually PQQLFPLPQQPFPQPQQPFP (SEQ ID NO: 1769) (8/14 subjects), or occasionally QPFPQPQQPTPIQPQQPFPQ (SEQ ID NO: 1770) (4/14), QQPQQLFPQTQQSSPQQPQQ (SEQ ID NO: 1771) (1/14), PQTQQPQQPFPQPQQPQQLF (SEQ ID NO: 1772) (1/14) and/or QEQREGV-

QILLPQSHQQLVG (SEQ ID NO: 1773) (1/14). Additional peptides noted for greater than 40% maximal response in at least 1 subject include:

FPQQPQQPFPQPQQQLPLQP (SEQ ID NO: 1774) (3/14, 2 > 70%)
PQQPFPQQPEQIIPQQPQQP (SEQ ID NO: 1775) (5/14, 3 > 70%)
QQLPLQPQQPFPQPQQPIPQ (SEQ ID NO: 1776) (6/14, 2 > 70%)
QQPQQPFPLQPQQPVPQQPQ (SEQ ID NO: 1777) (3/14, 1 > 70%)
SIPQPQQPFPQPQQPFPQSQ (SEQ ID NO: 1778) (4/14, 1 > 70%)
QTQQSIPQPQQPFPQPQQPF (SEQ ID NO: 1779) (3/14, 1 > 70%)
NMQVGPSGQVEWPQQQPLPQ (SEQ ID NO: 1780) (2/14, 1 > 70%)
VGPSGQVSWPQQQPLPQPQQ (SEQ ID NO: 1781) (2/14, 2 > 70%)
QQPFLLQPQQPFSQPQQPFL (SEQ ID NO: 1782) (1/14, 1 > 70%)
FPLQPQQPFPQQPEQIISQQ (SEQ ID NO: 1783) (5/14, 1 > 70%)
PQQPQRPFAQQPEQIISQQP (SEQ ID NO: 1784) (3/14, 1 > 70%)
SPQQPQLPFPQPQQPFVVVV (SEQ ID NO: 1785) (4/14, 1 > 70%)
QQPSIQLSLQQQLNPCKNVL (SEQ ID NO: 1786) (1/14, 1 > 70%)

[0147] Typically, the dominant peptides after barley challenge included one of six peptide motifs, or were one of eight other individual 20mers "dominant" in only one of 17 subjects after barley challenge. The six motifs identified:

QQPIPQQPQPY (SEQ ID NO: 1787)
PFPQPQQPFPW (SEQ ID NO: 1788)
LQPQQPFPQ (SEQ ID NO: 1789)
PQPQQASPL (SEQ ID NO: 1790)
IIPQQPQQPF (SEQ ID NO: 1791)
YPEQPQQPF (SEQ ID NO: 1792)

[0148] The barley hordein peptides showing at least 40% maximal peptide response in at least one subject include the following, wherein an asterisk indicates the eight individual peptides showing maximal response in a single individual:

QQQPFPQQPIPQQPQPYPQQ (SEQ ID NO: 1793) (8/17, 2 > 70%)
QQPQPFSQQPIPQQPQPYPQ (SEQ ID NO: 1794) (9/17, 8 > 70%)
PQQPVPQQPQPYPQQPQPFP (SEQ ID NO: 1795) (5/17, 1 > 70%)
PQPFPQQPIPQQPQPYPQQP (SEQ ID NO: 1796) (6/17,2 > 70%)
YPQQPQPFPQQPIPQQPQPY (SEQ ID NO: 1797) (6/17, 2 > 70%)
QPQPYPQQPQPYPQQPFQPQ (SEQ ID NO: 1798) (7/17, 2 > 70%)
QPQQPQPFPQQPVPQQPQPY (SEQ ID NO: 1799) (5/17, 2 > 70%)
PQPYPQQPQPFPQQPPFCQQ (SEQ ID NO: 1800) (1/17, 1 > 70%)*
QPFPQPQQPFPWQPQQPFPQ (SEQ ID NO: 1801) (10/17, 2 > 70%)
PFPQQPQQPFPQPQQPFRQQ (SEQ ID NO: 1802) (6/17, 3 > 70%)
WQPQQPFPQPQQPFPLQPQQ (SEQ ID NO: 1803) (9/17, 5 > 70%)*
PWQPQQPFPQPQEPIPQQPQ (SEQ ID NO: 1804) (1/17, 1 > 70%)
QQPFPQPQQPIPYQPQQPFN (SEQ ID NO: 1805) (5/17, 1 > 70%)
PQQPQQPFPQPQQPFSWQPQ (SEQ ID NO: 1806) (6/17, 2 > 70%)*
QPQQPFPQPQQPIPYQPQQP (SEQ ID NO: 1807) (4/17, 1 > 70%)*
QSQQQFPQPQQPFPQQPQQP (SEQ ID NO: 1808) (1/17, 0 > 70%)
PFPQPQQPFSWQPQQPFLQP (SEQ ID NO: 1809) (1/17, 0 > 70%)
FPQPQEPFPQQPQQPFPLQP (SEQ ID NO: 1810) (1/17, 0 > 70%)
PFPQPQQPFPWQPQQPFPQP (SEQ ID NO: 1811) (6/17, 0 > 70%)
FPQYQIPTPLQPQQPFPQQP (SEQ ID NO: 1812) (2/17, 1 > 70%)
FPLQPQQPFPQQPQQPFPQQ (SEQ ID NO: 1813) (1/17, 0 > 70%)
QQPFPLQPQQPFPQPQPFPQ (SEQ ID NO: 1814) (1/17, 0 > 70%)
SPLQPQQPFPQGSEQIIPQQ (SEQ ID NO: 1815) (1/17, 0 > 70%)
PQQASPLQPQPQQASPLQPQ (SEQ ID NO: 1816) (1/17, 1 > 70%)
PQQPPFWPQQPFPQQPPFGL (SEQ ID NO: 1817) (1/17, 1 > 70%)*
PVLSQQQPCTQDQTPLLQEQ (SEQ ID NO: 1818) (1/17, 1 > 70%)
RQLPKYIIPQQPQQPFLLQP (SEQ IDNO: 1819) (1/17, 1 > 70%)
QGSEQIIPQQPQQPFPLQPH (SEQ ID NO: 1820) (7/17, 3 > 70%)*

PQGSEQIIPQQPFPLQPQPF (SEQ ID NO: 1821) (2/17, 1 > 70%)
QPFPTPQQFFPYLPQQTFPP (SEQ ID NO: 1822) (4/17, 1 > 70%)
PFPQPPQQKYPEQPQQPFPW (SEQ ID NO: 1823) (1/17, 1 > 70%)
QKYPEQPQQPFPWQQPTIQL (SEQ ID NO: 1824) (1/17, 1 > 70%)
FQQPQQSYPVQPQQPFPQPQ (SEQ ID NO: 1825) (3/17, 1 > 70%)
QIPYVHPSILQQLNPCKVFL (SEQ ID NO: 1826) (1/17, 1 > 70%)
LAAQLPAMCRLEGGGGLLAS (SEQ ID NO: 1827) (1/17, 1 > 70%)
PYLPEELSPQYQIPTPLQPQ (SEQ ID NO: 1828) (1/17, 1 > 70%)*
VSPHPGQQTTVSPHQGQQTT (SEQ ID NO: 1829) (1/17, 1 > 70%)*

Second and third round wheat glutenin and gliadin peptide libraries

[0149] The second round wheat gliadin and glutenin library was designed upon the sequences of 20mer wheat gliadin and glutenin peptides that induced at least 5% of the response (interferon gamma ELISPOT) stimulated by the most active transglutaminase (tTG) pre-treated (enzymatically deamidated) 20mer peptide in any subject. All 2nd round 16mer peptides were assessed in at least 18 subjects. The 2nd round library generated from the "Oxford" gliadin 20mer library had been assessed in ten subjects - this data was merged with data generated from the 18 subjects used to assess the new 2nd round (expanded) gliadin/glutenin library. Hence, individual 16mer peptides pre-treated with transglutaminase were assessed in either 18 (novel gliadin/glutenin sequences based on "Melbourne" 20mer library) or 28 subjects (gliadin sequences based on "Oxford" 20mer library). All 16mers identified for the second round Oxford library also fulfilled the selection criteria for the Melbourne second round library.

[0150] The second round peptide library data was analysed according to the "dominance" of peptide responses in the interferon gamma ELISPOT in individual subjects i.e. the percent response of an individual's PBMC to a specific peptide normalized against that individual's maximal peptide-induced response. Sequences of peptides that stimulated at least 40% of the maximal peptide-specific response in at least one subject are shown in Table 1 below. The dataset supports the consistency and "dominance" of peptides conforming to the sequences identified using the first round 20mer peptide library using the Expectation Maximization (EM) algorithm described above.

Table 1: Peptides confirmed in Second Round Library as at least 40% as active as the peptide with maximal activity in any one subject: Ranked according to potency of peptide family

Peptide	SEQ ID NO:	>70%	40-70%	10-40%	<10%
G-QLPYPQPQLPYPQP-G	1830	18/28	4/28	3/28	3/28
G-LQPFPQPQLPYPQP-G	1831	14/28	8/28	Nil	6/28
G-LQPFPQPQLPFPQP-G	1832	4/28	8/28	5/28	10/28
G-LQPFPQPQLPYLQP-G	1833	1/28	1/28	12/28	14/28
G-LQPFPQPQLPYSQP-G	1834	2/28	3/28	12/28	11/28
G-LQPFPQPQLSYSQP-G	1835	Nil	1/28	2/28	25/28
G-**QQPFPQPQQPFPWQ**-G	1837	9/28	8/28	5/28	6/28
G-QQPFPQPQQPIPVQ-G	1838	8/28	5/28	7/28	8/28
G-QQPFPQPQQPFSQQ-G	1839	4/28	7/28	8/28	9/28
G-QQPFPQPQQPFCQQ-G	1840	2/28	2/28	13/28	11/28
G-**GLERPWQQQPLPPQ**-G	1841	2/18	1/18	Nil	15/18
G-**QTFPHQPQQAFPQP**-G	1842	1/28	2/28	Nil	25/28
LQQQCSPVAMPQRLAR	1843	1/28	1/28	11/28	15/28
QGQQGYYPISPQQSGQ	1844	1/18	1/18	1/18	15/18
PGQGQSGYYPTSPQQS	1845	1/18	1/18	Nil	16/18
QGQPGYYPTSPQQIGQ	1846	1/18	1/18	1/18	15/18
GQGQSGYYPTSPQQSG	1847	1/18	Nil	2/18	15/18
QQGYYPTSPQQSGQGQ	1848	Nil	1/18	Nil	17/

(continued)

Peptide	SEQ ID NO:	>70%	40-70%	10-40%	<10%
QGQQGYYPTSPQQPPQ	1849	Nil	1/18	Nil	Nil
QQGYYPISPQQLGQGQ	1850	Nil	1/18	Nil	Nil
YVPPDCSTINVPYANI	1851	1/18	1/18	1/18	15/18
IIMQQEQQEQRQGVQI	1852	1/28	Nil	8/28	19/28
VAHAIIMHQQQQQQQE	1853	Nil	1/28	2/28	25/28
G-**QPIPQQPQQPFPLQ**-G	1854	1/28	Nil	5/28	
G-**FPQLQQPQQPFPQQ**-G	1855	1/28	Nil	1/28	26/28
G-FPQTQQPQQPFPQQ-G	1856	Nil	1/28	2/28	25/28
G-QPLSQQPQQTFPQQ-G	1857	Nil	1/28	Nil	27/28
G-QQPQQQPQQPFPQQ-G	1858	Nil	1/28	5/28	22/28
G-FPQPQQPQQPFPQQ-G	1859	Nil	1/28	3/28	25/28
G-FPQPQQPQQSFPQQ-G	1860	Nil	1/28	1/28	26/28
G-**QPQQTFPQQPQLPF-**G	1861	1/18	Nil	2/18	15/18
G-**MQVDPSGQVQWPQQ-**G	1862	1/18	Nil	Nil	17/18
G-IQVDPSGQVQWPQQ-G	1863	1/18	Nil	Nil	17/18
G-MQADPSGQVQWPQQ-G	1864	1/18	Nil	Nil	17/18
G-MQVDPSSQVQWPQQ-G	1865	1/18	Nil	Nil	17/18
G-**QQEQQILQQILQQQ**-G	1866	1/18	Nil	Nil	17/18
VPLYRTTTSVPFGVGT	1867	1/18	Nil	Nil	17/18
LQTLPSMCNVYIPPYC	1868	1/18	Nil	Nil	17/18
LALQTLPAMCNVYIPP	1869	1/18	Nil	Nil	17/18
DAIRAIIYSIVLQEQQ	1870	1/18	Nil	Nil	17/18
G-**QQQFSQPQQQFPQP**-G	1871	Nil	5/28	7/28	16/28
G-FFPQPQQQFPQPQQ-G	1872	Nil	1/28	10/28	17/28
G-FPQQPQQQFPQPQQ-G	1873	Nil	1/28	Nil	27/28
G-QQPFPQPQQQFPQP-G	1874	Nil	1/28	12/28	15/28
G-QPQPFLPQLPYPQP-G	1875	Nil	4/28	9/28	15/28
G-**QQPFPQPQQQLPQP**-G	1876	Nil	3/28	6/28	19/28
G-LPFPQQPQQPLPQP-G	1877	Nil	2/18	4/18	12/18
G-**QQAFPQPQQTFPHQ-**G	1878	Nil	3/28	8/28	17/28
G-QQPFTQPQQPTPIQ-G	1879	Nil	1/28	4/28	23/28
G-QQIFPQPQQTFPHQ-G	1880	Nil	1/28	10/28	17/28
G-**QQQFIQPQQPFPQQ-**G	1881	Nil	2/28	11/28	15/28
G-QPFPLQPQQPFPQQ-G	1882	Nil	2/28	7/28	19/28
G-QPFPWQPQQPFPQQ-G	1883	Nil	2/28	8/28	18/28
G-QPTPIQPQQPFPQQ-G	1884	Nil	2/28	5/28	21/28
G-**QVSFQQPQQQYPSP**-G	1885	Nil	2/28	4/28	22/28
G-FFQQPQQQYPSSQQ-G	1886	Nil	1/28	1/28	26/28

(continued)

Peptide	SEQ ID NO:	>70%	40-70%	10-40%	<10%
G-**GKSQVLQQSTYQLL**-G	1887	Nil	2/18	1/18	15/18
GQVVNNHGQTVFNDIG	1888	Nil	1/18	4/18	
G-**QPQLPFPQQPQQQF**-G	1889	Nil	1/28	2/28	25/28
G-QPFPQPQQAQLPFP-G	1890	Nil	1/28	2/28	25/28
G-**HQQPGQRQQGYYPT**-G	1891	Nil	1/18	1/18	16/18
G-**HQQFPQQQIPWQP**-G	1892	Nil	1/18	1/18	16/18
LEAVTSIALRTLPTMC	1893	Nil	1/18	1/18	
G-**QQPQFSQQQQIPVI**-G	1894	Nil	1/18	Nil	17/18

[0151] The third round peptide library consisted of 74 peptides based upon structurally distinct sequences in the second round library found to induce at least 10% of the maximal response to any peptide in any subject. These peptides corresponded to wild-type (non-deamidated) sequences virtually identical to those used in the second round library. The distinct feature of this library was that it consisted of peptides with purity verified by HPLC as >80%, and with sequences confirmed by mass spectroscopy.

[0152] Interferon gamma ELSIPOT responses to the 3rd round library peptides following deamidation by tTG were compared in 14 subjects. Once again, sequences including the PQPQLPY motif were "dominant" in 9/14 subjects. However PFPQPQQPFPW (SEQ ID NO: 1895) stimulated >70% of maximal response in 1/14 subjects, PFPQQPQQP-FPQ (SEQ ID NO: 1896) in 1/14, PQPFLPQLPYPQP (SEQ ID NO: 1897) in 1/14, QPFPQPQQPQQP (SEQ ID NO: 1898) in 4/14 (including 3 subjects in whom PQPQLPY peptides were not potent epitopes) SGQGVSQSQQQSQQQ (SEQ ID NO: 1899) in 2/14 (including one in which PQPQLPY peptides were not potent), QYEVIRSLVLRTLPNM (SEQ ID NO: 1900) and GLARSQMLQQSICHVG (SEQ ID NO: 1901) each in one (the same) subject in whom PQPQLPY peptides were not potent epitopes, RTTTSVPFGVGTGVGA (SEQ ID NO: 1902) in 1/14 subjects and AIHTVIHSI-IMQQEQQ (SEQ ID NO: 1903) in 1/14 subjects.

[0153] Many of the sequences tested in third round were structurally related and individual subject's responses were present or absent according to the "relatedness" of certain sequences, suggesting redundancy of peptides recognized by gluten specific T cells induced by in vivo gluten challenge.

[0154] Interferon gamma ELISPOT responses to 3rd round peptides were absent before gluten challenge, and were blocked by pre-treatment of PBMC with anti-HLA DQ but not HLA DR antibody.

Combitopes

[0155] The issue of epitope redundancy and the potential utility in diagnostics and therapeutics of peptides designed to combine "unique" dominant epitopes was addressed by comparing interferon gamma ELISPOT responses after wheat (n=16 HLA DQ2 coeliac disease subjects), rye (n=17) or barley (n=13) challenge to the sequences: QLQPF-PQPELPYPQPQL (SEQ ID NO: 1904) ("P04724E"), QPEQPFPQPEQPFPWQP (SEQ ID NO: 1905) ("626fEE"), and QLQPFPQPELPYPQPFPQQPEQPFPQPEQPFPWQP (SEQ ID NO: 1906) ("Combitope"). After rye and barley challenge the sum of the median ELISPOT responses (spot forming units) to P04724E and 626fEE were almost identical (99%, and 102%, respectively) to the response to a similar (optimal) concentration of the Combitope. However, after wheat challenge (n=16 subjects), median P04724E response was 89% of that to Combitope, and median 626fEE responses was 70% of the response to Combitope. These findings would be consistent with substantial redundancy of these related epitope sequences, P04724E and 626fEE, after wheat challenge but not after rye or barley, and that combining dominant epitope sequences within longer peptides does not reduce their biological availability. Hence, combitopes derived from selected potent epitopes may be efficient delivery devices for T cell epitope-based therapeutics and diagnostics in coeliac disease.

Epitopes in wheat gluten associated with HLA-DQ8+ coeliac disease

[0156] Epitopes in wheat gliadins were identified using PBMC after gluten challenge in two individuals, one HLA-DQ8 homozygous, and one HLA-DQ8 heterozygote. Induced T-cell responses in other HLA-DQ8 (not DQ2) coeliac individuals responded weakly to gluten challenge and their data did not allow detailed analysis.

[0157] Deamidated 20mers including the core sequence: QGSFQPSQQ (SEQ ID NO: 1907), corresponding to the

known HLA-DQ8-restricted alpha-gliadin epitope (in which Q1 and Q9 are deamidated by tTG for optimal activity), induced moderately strong peptide responses. However, a series of "core" peptides were associated with more potent responses in 20mers derived from gamma and omega gliadins (see Figure 9). The most potent peptides possessed glutamine in a sequence that would suggest susceptibility to deamidation separated by seven residues from a second glutamine also susceptible to deamidation (as found in QGSFQPSQQ (SEQ ID NO: 1907)) suggesting that these deamidated sequences would become high affinity binders for HLA-DQ8 following deamidation by tTG. (The binding motif for HLA-DQ8 favours glutamate at positions 1 and 9.) A further group of 20mers possessed glutamine residues susceptible to deamidation but not separated by seven residues from a second glutamine susceptible to tTG-mediated deamidation.

HLA DQ8 coeliac disease gliadin and glutenin epitopes

[0158] Five subjects with coeliac disease that possess HLA DQ2 and HLA DQ8 alleles underwent wheat gluten challenge. PBMC from two subjects initially challenged were used to screen the first round "Melbourne" wheat gliadin 20mer library. The 20mer sequences identified using PBMC from these two HLA DQ8 CD subjects were dissected further by screening, in five HLA DQ8+ DQ2- CD subjects including the two original subjects, a second round library based on reactive 20mers in the 1st round library. The 2nd round library consisted of *screening grade* overlapping 16mers, and 13mers predicted to correspond to tTG-mediated deamidation products of epitopes with the potential for deamidation of glutamine at position 1 and/or position 9 (consistent with the HLA DQ8 peptide binding motif). In addition, the 1400 glutenin (HMW and LMW) tTG-pretreated 20mers in the "Melbourne" wheat gluten library were also screened in these five subjects.

[0159] The most potent and consistently dominant gliadin 16mers were the related sequences VYIPPYCTIAPFGIFG (SEQ ID NO: 1908) (3/5 subjects >70% response to maximal gliadin 16mer) and AMCNVYIPPYCAMAPF (SEQ ID NO: 1908) also dominant in 3/5 subjects (4/5 subjects produced dominant responses to one or both of these peptides). In addition, a series of peptides derived from previously identified bioactive 20mers whose responses in the ELISPOT were enhanced or permissive to specific glutamine residues being deamidated were identified: (QE) QPTPIQP (QE) (SEQ ID NO: 1909), (QE) QPFPLQP (QE) (SEQ ID NO: 1910), (QE)QPIPVQP(QE) (SEQ ID NO: 1911), (QE) QPQQPFP (QE) (SEQ ID NO: 1912), (QE) QP (QE) LPFP (QE) (SEQ ID NO: 1913), (QE) GSFQPSQ (QE) (SEQ ID NO: 1914) (previously published HLA DQ8 epitope, van der Wal 1998), (QE) LPFP (QE) QP (QE) (SEQ ID NO: 1915), and (QE) QPFP (QE) QP (QE) (SEQ ID NO: 1916).

[0160] Screening the glutenin 20mer library identified a further series of sequences that were dominant in at least one of the five subjects. Dominant 20mer peptides shared the motifs or had the sequences: PQQQQQQLVQQQ (SEQ ID NO: 1917), QGIFLQPH (LQ) I (AS) QLEV (SEQ ID NO: 1918), QPGQGQQG(HY) Y (SEQ ID NO: 1919), QSRYEAIRAII(FY) S (SEQ ID NO: 1920), RTTTSVPFD (SEQ ID NO: 1921), QPPFWRQQP (SEQ ID NO: 1922), Q (PS) (PS) (FI) (PS) QQQQ (SEQ ID NO: 1923), (QPLR) GYYPTSPQ (SEQ ID NO: 1924) (previously identified HLA DQ8 epitope, van der Wal 2001), QGSYYPGQASPQ (SEQ ID NO: 1925), GYYPTSSLQPEQGQQGYYPT (SEQ ID NO: 1926), and QGQQLAQGQQGQQPAQVQQG (SEQ ID NO: 1927). Glutenin peptides were assessed after pre-treatment with transglutaminase. Hence, the requirement for deamidation for these epitopes is not known.

[0161] A comprehensive library of "uncharacterised" screening grade peptides including all unique 12mer sequences encoded by genes present in Genbank defined as (bread making) wheat (Triticum aestivum), rye, barley, or oats gluten, gliadin, glutenin, secalin, hordein, or avenin have been assessed using T cells from HLA DQ2+ (and in some cases HLA DQ8+) coeliac disease volunteers six days after commencing in vivo gluten challenge. A relatively consistent pattern of epitope hierarchy has been identified in HLA DQ2 coeliac disease that is similar to but not identical after consumption of other grains toxic in coeliac disease. Peptides with the sequence PQPQLPY are dominant after wheat challenge in at least two thirds of HLA DQ2+ coeliac disease, but other epitopes are occasionally dominant while PQLPY peptides are essentially inactive in fewer than one in six HLA DQ2+ subjects with coeliac disease. The contribution of rare dominant epitopes will be better assessed after screening large numbers (e.g. >30) subjects (in progress). Epitope hierarchy after rye and barley consumption is similar to that after wheat with the exception that deamidated peptides similar to the gliadin/hordein/secalin sequences PQPQQPFP or PFPQQPQQP are usually dominant rather than PQPQLPY (a sequence unique to wheat alpha-gliadins). Combitopes that comprise serial and partially overlapping gluten epitopes are as active or more active than single epitopes alone and offer a means of efficiently delivering multiple gluten epitopes for T cell recognition. Such combitopes are therefore useful in design and delivery of peptide therapies in coeliac disease that target multiple unique T cell epitopes.

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Claims

1. An agent selected from:

(a) a isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length.

2. An agent according to claim 1 which is HLA-DQ2-restricted.

3. An agent according to claim 1 which is HLA-DQ8-restricted.

4. An agent according to any of claims 1 to 3 which is an isolated peptide of up to 50 amino acids.

5. An agent according to any of claims 1 to 3 bound to (i) an HLA molecule, or (ii) a fragment of an HLA molecule capable of binding (a) or (b).

6. A pharmaceutical composition comprising an agent according to any of claims 1 to 5 and a pharmaceutically acceptable carrier or diluent.

7. A pharmaceutical composition according to claim 6 comprising one peptide which is HLA-DQ2-restricted and a second peptide which is HLA-DQ8-restricted.

8. A pharmaceutical composition according to claim 6 wherein the peptide comprises a wheat epitope.

9. A pharmaceutical composition according to claim 6 wherein the peptide comprises an oat epitope.

10. A pharmaceutical composition according to claim 6 comprising one peptide which comprises a wheat epitope and one peptide which comprises an oat epitope.

11. A composition for use in a method of preventing or treating coeliac disease comprising at least one agent selected from:

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO: 200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length.

12. A composition for use according to claim 11, said method comprising tolerising an individual to a gluten protein to suppress the production of a T cell or antibody response to an agent as defined in claim 11.

13. An agent selected from:

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO: 200;
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length; and
(c) an analogue of (a) which is an isolated peptide that binds an antibody, which antibody binds to an epitope comprising SEQ ID NO:200,

for use in a method of treating or preventing coeliac disease in an individual by tolerising the individual to prevent the production of such an antibody.

14. A method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising:

(a) contacting a sample from the host, *in vitro*, with at least one agent selected from:

(i) an isolated peptide comprising at least one epitope that comprises SEQ ID NO: 200; and
(ii) an analogue of (i) which is an isolated peptide capable of being recognised by a T cell receptor that recognises (i) and which is not more than 50 amino acids in length; and

(b) determining *in vitro* whether T cells in the sample recognise the agent; recognition by the T cells indicating that the individual has, or is susceptible to, coeliac disease.

15. A composition for use in a method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising an agent selected from

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO: 200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognised by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length,

said method comprising determining whether T cells of the individual recognise the agent, recognition by the T cells indicating that the individual has, or is susceptible to, coeliac disease.

16. A method according to claim 14 wherein the agent comprises (i) or (ii) bound to (a) an HLA molecule, or (b) a fragment of an HLA molecule capable of binding (i) or (ii).

17. A method according to claim 16 wherein the HLA molecule or fragment is in a complex comprising four HLA molecules or fragments of HLA molecules.

18. A composition for use according to claim 15 wherein the method comprises administering the agent to the skin of an individual and detecting the presence of inflammation at the site of administration, the detection of inflammation indicating that the T cells of the individual recognise the agent.

19. A method according to claim 14 wherein the sample is blood sample.

20. A method according to claim 19 wherein the T cells are not restimulated in an antigen specific manner *in vitro* before the said determining.

21. A method according to claim 20 in which the recognition of the agent by the T cells is determined by detecting the secretion of a cytokine from the T cells.

22. A method according to claim 21 in which the cytokine is IPN-γ.

23. A method according to claim 21 or claim 22 in which the cytokine is detected by allowing the cytokine to bind to an immobilised antibody specific to the cytokine and then detecting the presence of the antibody/cytokine complex.

24. A method according to claim 14 wherein said determining is done by measuring whether the agent binds the T cell receptor.

25. A method for identifying an analogue of a peptide comprising at least one epitope that comprises SEQ ID NO:200 said method comprising determining, in vitro, whether a candidate peptide is recognised by a T cell receptor that recognises an epitope that comprises SEQ ID NO: 200, recognition of the candidate peptide indicating that the candidate peptide is an analogue said analogue being of not more than 50 amino acids in length.

26. A method of diagnosing coeliac disease, or susceptibility to coeliac disease, in an individual comprising determining, *in vitro* the presence of an antibody that binds to an epitope of a peptide sequence selected from:

(i) a peptide comprising at least one epitope that comprises SEQ ID NO: 200; and
(ii) a peptide analogue of (i) which is capable of being recognised by a T cell receptor that recognises (i) and which is not more than 50 amino acids in length;

in a sample from the individual, the presence of the antibody indicating that the individual has, or is susceptible to, coeliac disease.

27. An *in vitro* method of determining whether a composition is capable of causing coeliac disease comprising determining whether a protein sequence capable of being modified by a transglutaminase to a peptide comprising at least one epitope that comprises SEQ ID NO:200 is present in the composition, the presence of the protein sequence indicating that the composition is capable of causing coeliac disease.

28. A method according to claim 27 wherein said determining is done by contacting the composition with an antibody specific for the protein sequence which is capable of being so modified, binding of the antibody to a protein sequence in the composition indicating that the composition is capable of causing coeliac disease.

29. A kit for carrying out a method according to claims 14, 16, 17 or 19 to 24 comprising an agent selected from

(a) an isolated peptide comprising at least one epitope that comprises SEQ ID NO:200; and
(b) an analogue of (a) which is an isolated peptide capable of being recognized by a T cell receptor that recognises the peptide of (a) and which is not more than 50 amino acids in length,

and a means to detect the recognition of the peptide by the T cell.

30. A kit according to claim 29 wherein the means to detect recognition comprises an antibody to IFN-γ.

31. A kit according to claim 30 wherein the antibody is immobilised on a solid support and optionally the kit also comprises a means to detect the antibody/IFN-γ complex.

32. An antibody or fragment thereof, specific for SEQ ID NO:200

Patentansprüche

1. Mittel, das ausgewählt ist aus:

(a) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und
(b) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der das Peptid aus (a) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt.

2. Mittel nach Anspruch 1, das HLA-DQ2-beschränkt ist.

3. Mittel nach Anspruch 1, das HLA-DQ8-beschränkt ist.

4. Mittel nach einem der Ansprüche 1 bis 3, bei dem es sich um ein isoliertes Peptid aus bis zu 50 Aminosäuren handelt.

5. Mittel nach einem der Ansprüche 1 bis 3, das an (i) ein HLA-Molekül, oder (ii) ein Fragment eines HLA-Moleküls gebunden ist, das in der Lage ist, (a) oder (b) zu binden.

6. Pharmazeutische Zusammensetzung, die ein Mittel nach einem der Ansprüche 1 bis 5 und einen pharmazeutisch unbedenklichen Träger oder Verdünner umfasst.

7. Pharmazeutische Zusammensetzung nach Anspruch 6, umfassend ein Peptid, das HLA-DQ2-beschränkt ist, und ein zweites Peptid, das HLA-DQ8-beschränkt ist.

8. Pharmazeutische Zusammensetzung nach Anspruch 6, wobei das Peptid ein Weizenepitop umfasst.

9. Pharmazeutische Zusammensetzung nach Anspruch 6, wobei das Peptid ein Haferepitop umfasst.

10. Pharmazeutische Zusammensetzung nach Anspruch 6, umfassend ein Peptid, das ein Weizenepitop umfasst, und ein Peptid, das ein Haferepitop umfasst.

11. Zusammensetzung zur Verwendung in einem Verfahren zum Vorbeugen oder Behandeln von Zöliakie, umfassend mindestens ein Mittel, das ausgewählt ist aus:

(a) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und
(b) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der das Peptid aus (a) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt.

12. Zusammensetzung zur Verwendung nach Anspruch 11, wobei das Verfahren das Herbeiführen einer Verträglichkeit für Glutenprotein bei einem Individuum umfasst, um die Erzeugung einer T-Zell- oder Antikörperreaktion auf ein Mittel entsprechend der Definition in Anspruch 11 zu unterdrücken.

13. Mittel, das ausgewählt ist aus:

(a) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst;
(b) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der das Peptid aus (a) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt; und
(c) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das einen Antikörper bindet, wobei der Antikörper an ein Epitop bindet, das SEQ ID NO:200 umfasst,

zur Verwendung in einem Verfahren zum Vorbeugen oder Behandeln von Zöliakie bei einem Individuum durch Herbeiführen einer Verträglichkeit bei dem Individuum, um der Produktion eines derartigen Antikörpers vorzubeugen.

14. Verfahren zum Diagnostizieren von Zöliakie oder Anfälligkeit für Zöliakie bei einem Individuum, umfassend:

(a) In-vitro-Inkontaktbringen einer Probe von dem Wirt mit mindestens einem Mittel, das ausgewählt ist aus:

(i) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und
(ii) einem Analogon von (i), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der (i) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt; und

(b) In-vitro-Bestimmen, ob T-Zellen in der Probe das Mittel erkennen; wobei das Erkennen durch die T-Zellen darauf hinweist, dass das Individuum Zöliakie hat oder anfällig dafür ist.

15. Zusammensetzung zur Verwendung in einem Verfahren zum Diagnostizieren von Zöliakie oder Anfälligkeit für Zöliakie bei einem Individuum, umfassend ein Mittel, das ausgewählt ist aus

(a) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und

(b) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der das Peptid aus (a) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt;

wobei das Verfahren das Bestimmen umfasst, ob T-Zellen des Individuums das Mittel erkennen, wobei das Erkennen durch die T-Zellen darauf hinweist, dass das Individuum Zöliakie hat oder anfällig dafür ist.

16. Verfahren nach Anspruch 14, wobei das Mittel (i) oder (ii) umfasst, das an (a) ein HLA-Molekül oder (b) ein Fragment eines HLA-Moleküls gebunden ist, das in der Lage ist, (i) oder (ii) zu binden.

17. Verfahren nach Anspruch 16, wobei das HLA-Molekül oder das Fragment sich in einem Komplex befindet, der vier HLA-Moleküle oder Fragmente von HLA-Molekülen umfasst.

18. Zusammensetzung zur Verwendung nach Anspruch 15, wobei das Verfahren das Verabreichen des Mittels an die Haut eines Individuums und das Detektieren des Vorhandenseins einer Entzündung an der Verabreichungsstelle umfasst, wobei das Detektieren einer Entzündung darauf hinweist, dass die T-Zellen es Individuums das Mittel erkennen.

19. Verfahren nach Anspruch 14, wobei es sich bei der Probe um eine Blutprobe handelt.

20. Verfahren nach Anspruch 19, wobei die T-Zellen vor dem Bestimmen nicht auf antigenspezifische Weise in vitro restimuliert werden.

21. Verfahren nach Anspruch 20, in dem das Erkennen des Mittels durch die T-Zellen durch das Detektieren der Ausscheidung eines Zytokins aus den T-Zellen bestimmt wird.

22. Verfahren nach Anspruch 21, in dem es sich bei dem Zytokin um IFN-γ handelt.

23. Verfahren nach Anspruch 21 oder Anspruch 22, in dem das Zytokin dadurch erkannt wird, dass es dem Zytokin gestattet wird, an einen immobilisierten, für das Zytokin spezifischen Antikörper zu binden und anschließend das Vorhandensein des Antikörper/Zytokin-Komplexes zu detektieren.

24. Verfahren nach Anspruch 14, wobei das Bestimmen durch das Messen, ob das Mittel den T-Zell-Rezeptor bindet, erfolgt.

25. Verfahren zum Identifizieren eines Analogons eines Peptids, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst, wobei das Verfahren das In-Vitro-Bestimmen umfasst, ob ein Peptidkandidat von einem T-Zell-Rezeptor erkannt wird, der ein Epitop erkennt, das SEQ ID NO:200 umfasst, wobei das Erkennen des Peptidkandidaten darauf hinweist, dass es sich bei dem Peptidkandidaten um ein Analogon handelt, wobei die Länge des Analogons nicht mehr als 50 Aminosäuren beträgt.

26. Verfahren zum Diagnostizieren von Zöliakie oder Anfälligkeit für Zöliakie bei einem Individuum, umfassend das In-vitro-Bestimmen des Vorhandenseins eines Antikörpers, der an ein Epitop einer Peptidsequenz bindet, die ausgewählt ist aus:

(i) einem Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und
(ii) einem Peptidanalogon von (i), das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der (i) erkennt und dessen Länge nicht mehr als 50 Aminosäuren beträgt;

in einer Probe von dem Individuum, wobei das Vorhandensein des Antikörpers darauf hinweist, dass das Individuum Zöliakie hat oder anfällig dafür ist.

27. In-vitro-Verfahren zum Bestimmen, ob eine Zusammensetzung in der Lage ist, Zöliakie hervorzurufen, umfassend das Bestimmen, ob eine Proteinsequenz, die in der Lage ist, von einer Transglutaminase zu einem Peptid modifiziert zu werden, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst, in der Zusammensetzung vorhanden ist, wobei das Vorhandensein der Proteinsequenz darauf hinweist, dass die Zusammensetzung in der Lage ist, Zöliakie hervorzurufen.

28. Verfahren nach Anspruch 27, wobei das Bestimmen durch das Inkontaktbringen der Zusammensetzung mit einem Antikörper erfolgt, der für die Proteinsequenz spezifisch ist, die in der Lage ist, auf diese Weise modifiziert zu werden, wobei das Binden des Antikörpers an eine Proteinsequenz in der Zusammensetzung darauf hinweist, dass die Zusammensetzung in der Lage ist, Zöliakie hervorzurufen.

29. Kit zum Ausführen eines Verfahren nach den Ansprüchen 14, 16, 17 oder 19 bis 24, umfassend ein Mittel, das ausgewählt ist aus

(a) einem isolierten Peptid, das mindestens ein Epitop umfasst, das SEQ ID NO:200 umfasst; und
(b) einem Analogon von (a), bei dem es sich um ein isoliertes Peptid handelt, das in der Lage ist, von einem T-Zell-Rezeptor erkannt zu werden, der das Peptid aus (a) erkennt, und dessen Länge nicht mehr als 50 Aminosäuren beträgt,

und einem Mittel zum Detektieren der Erkennung des Peptids durch die T-Zelle.

30. Kit nach Anspruch 29, wobei das Mittel zum Detektieren des Erkennens einen Antikörper für IFN-γ umfasst.

31. Kit nach Anspruch 30, wobei der Antikörper auf einem festen Träger immobilisiert wird und das Kit gegebenenfalls auch ein Mittel zum Detektieren des Antikörper/IFN-γ-Komplexes umfasst.

32. Antikörper oder Fragment davon, der für SEQ ID NO:200 spezifisch ist.

Revendications

1. Agent sélectionné parmi :

(a) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(b) un analogue de (a) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui reconnaît le peptide de (a) et dont la longueur ne dépasse pas 50 acides aminés.

2. Agent selon la revendication 1, à restriction HLA-DQ2.

3. Agent selon la revendication 1, à restriction HLA-DQ8.

4. Agent selon l'une quelconque des revendications 1 à 3, qui est un peptide isolé d'au plus 50 acides aminés.

5. Agent selon l'une quelconque des revendications 1 à 3, lié à (i) une molécule HLA, ou (ii) un fragment d'une molécule HLA capable de se lier à (a) ou (b).

6. Composition pharmaceutique comprenant un agent selon l'une quelconque des revendications 1 à 5, et un véhicule ou un diluant pharmaceutiquement acceptables.

7. Composition pharmaceutique selon la revendication 6, comprenant un peptide à restriction HLA-DQ2 et un second peptide à restriction HLA-DQ8.

8. Composition pharmaceutique selon la revendication 6, dans laquelle le peptide comprend un épitope du blé.

9. Composition pharmaceutique selon la revendication 6, dans laquelle le peptide comprend un épitope de l'avoine.

10. Composition pharmaceutique selon la revendication 6, comprenant un peptide qui comprend un épitope du blé et un peptide qui comprend un épitope de l'avoine.

11. Composition pour son utilisation dans un procédé de prévention ou de traitement de la maladie coeliaque comprenant au moins un agent sélectionné parmi :

(a) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(b) un analogue de (a) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui

reconnaît le peptide de (a) et dont la longueur ne dépasse pas 50 acides aminés.

12. Composition pour son utilisation selon la revendication 11, ledit procédé comprenant la tolérisation d'un individu à une protéine du gluten pour supprimer la production d'une réponse des lymphocytes T ou des anticorps à un agent selon la revendication 11.

13. Agent sélectionné parmi :

(a) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ;
(b) un analogue de (a) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui reconnaît le peptide de (a) et dont la longueur ne dépasse pas 50 acides aminés ; et
(c) un analogue de (a) qui est un peptide isolé qui se lie à un anticorps, ledit anticorps se lie à un épitope comprenant SEQ ID NO : 200,

pour son utilisation dans un procédé de traitement ou de prévention de la maladie coeliaque chez un individu par tolérisation de l'individu pour empêcher la production d'un tel anticorps.

14. Procédé de diagnostic de la maladie coeliaque, ou de la susceptibilité à la maladie coeliaque, chez un individu, comprenant :

(a) la mise en contact d'un échantillon de l'hôte, in vitro, avec au moins un agent sélectionné parmi :

(i) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(ii) un analogue de (i) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui reconnaît (i) et dont la longueur ne dépasse pas 50 acides aminés ; et

(b) la détermination in vitro du fait que les lymphocytes T dans l'échantillon reconnaissent ou non l'agent ; la reconnaissance par les lymphocytes T indiquant que l'individu a, ou est susceptible à, la maladie coeliaque.

15. Composition pour son utilisation dans un procédé de diagnostic de la maladie coeliaque, ou de la susceptibilité à la maladie coeliaque, chez un individu comprenant un agent sélectionné parmi

(a) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(b) un analogue de (a) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui reconnaît le peptide de (a) et dont la longueur ne dépasse pas 50 acides aminés,

ledit procédé comprenant la détermination du fait que les lymphocytes T reconnaissent ou non l'agent, la reconnaissance par les lymphocytes T indiquant que l'individu a, ou est susceptible à, la maladie coeliaque.

16. Procédé selon la revendication 14, dans lequel l'agent comprend (i) ou (ii) lié à (a) une molécule HLA, ou (b) un fragment d'une molécule HLA capable de se lier à (i) ou (ii).

17. Procédé selon la revendication 16, dans lequel la molécule HLA ou le fragment est dans un complexe comprenant quatre molécules HLA ou fragments de molécules HLA.

18. Composition pour son utilisation selon la revendication 15, dans laquelle le procédé comprend l'administration de l'agent à la peau d'un individu et la détection de la présence d'une inflammation au niveau du site d'administration, la détection de l'inflammation indiquant que les lymphocytes T de l'individu reconnaissent l'agent.

19. Procédé selon la revendication 14, dans lequel l'échantillon biologique est un échantillon de sang.

20. Procédé selon la revendication 19, dans lequel les lymphocytes T ne sont pas restimulés de manière spécifique des antigènes in vitro avant ladite détermination.

21. Procédé selon la revendication 20, dans lequel la reconnaissance de l'agent par les lymphocytes T est déterminée par détection de la sécrétion d'une cytokine par les lymphocytes T.

22. Procédé selon la revendication 21, dans lequel la cytokine est l'IFN-γ.

23. Procédé selon la revendication 21 ou la revendication 22, dans lequel la cytokine est détectée en laissant la cytokine se lier à un anticorps immobilisé spécifique de la cytokine puis en détectant la présence du complexe anticorps/cytokine.

24. Procédé selon la revendication 14, dans lequel ladite détermination est réalisée par mesure du fait que l'agent se lie ou non au récepteur des lymphocytes T.

25. Procédé d'identification d'un analogue d'un peptide comprenant au moins un épitope qui comprend SEQ ID NO : 200, ledit procédé comprenant la détermination, in vitro, du fait qu'un peptide candidat est reconnu ou non par un récepteur des lymphocytes T qui reconnaît un épitope qui comprend SEQ ID NO : 200, la reconnaissance du peptide candidat indiquant que le peptide candidat est un analogue, la longueur dudit analogue ne dépassant pas 50 acides aminés.

26. Procédé de diagnostic de la maladie coeliaque, ou de la susceptibilité à la maladie coeliaque, chez un individu, comprenant la détermination, in vitro, de la présence d'un anticorps qui se lie à un épitope d'une séquence peptidique sélectionnée parmi :

(i) un peptide comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(ii) un analogue peptidique de (i) qui peut être reconnu par un récepteur des lymphocytes T qui reconnaît (i) et dont la longueur ne dépasse pas 50 acides aminés ;

dans un échantillon de l'individu, la présence de l'anticorps indiquant que l'individu a, ou est susceptible à, la maladie coeliaque.

27. Procédé in vitro de détermination du fait qu'une composition est capable de provoquer la maladie coeliaque comprenant la détermination du fait qu'une séquence protéique pouvant être modifiée par une transglutaminase en un peptide comprenant au moins un épitope qui comprend SEQ ID NO : 200 est ou non présente dans la composition, la présence de la séquence protéique indiquant que la composition est capable de provoquer la maladie coeliaque.

28. Procédé selon la revendication 27, dans lequel ladite détermination est réalisée par mise en contact de la composition avec un anticorps spécifique de la séquence protéique qui peut être ainsi modifiée, la liaison de l'anticorps à une séquence protéique dans la composition indiquant que la composition est capable de provoquer la maladie coeliaque.

29. Kit pour la réalisation d'un procédé selon les revendications 14, 16, 17 ou 19 à 24 comprenant un agent sélectionné parmi

(a) un peptide isolé comprenant au moins un épitope qui comprend SEQ ID NO : 200 ; et
(b) un analogue de (a) qui est un peptide isolé pouvant être reconnu par un récepteur des lymphocytes T qui reconnaît le peptide de (a) et dont la longueur ne dépasse pas 50 acides aminés,

et un moyen de détection de la reconnaissance du peptide par le lymphocyte T.

30. Kit selon la revendication 29, dans lequel le moyen de détection de la reconnaissance comprend un anticorps contre l'IFN-γ.

31. Kit selon la revendication 30, dans lequel l'anticorps est immobilisé sur un support solide et facultativement le kit comprend également un moyen de détection du complexe anticorps/IFN-γ.

32. Anticorps ou fragment de celui-ci, spécifique de SEQ ID NO : 200.

Figure 1

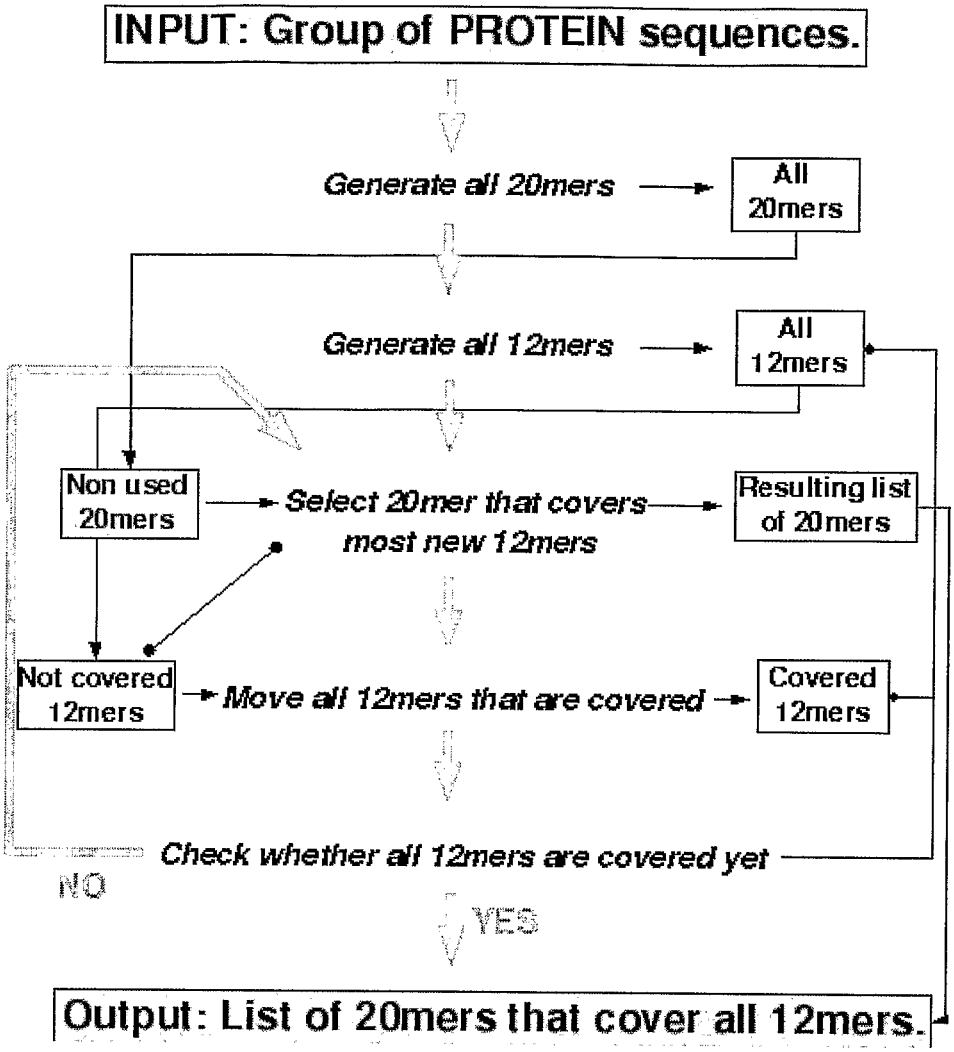

INPUT: Group of PROTEIN sequences.
Generate all 20mers
All 20mers
Generate all 12mers
All 12mers
Non used 20mers
Select 20mer that covers most new 12mers
Resulting list of 20mers
Not covered 12mers
Move all 12mers that are covered
Covered 12mers
Check whether all 12mers are covered yet
NO
YES
Output: List of 20mers that cover all 12mers.

Figure 2

Alpha/beta gliadins (n=53)	Gamma gliadins (n=53)	LMW glutenins (n=77)	HMW glutenins (n=55)	Hordeins (n=59)	Secalins (n=14)	Avenins (n=20)
S07361	AAA34272	AAB35353	A03353	A24095.1	A23277	S29209
P18573	S07398	AAB48474	A24266	A25677	AAB37403	S29208
P04728	PS0094	AAB48475	A30843	AAA32942	AAB37404	S29207
P04727	P21292	AAB48476	AAA62315	AAA32943	AAB37405	S06455
P04726	P08453	AAB48477	AAB02788	AAA32944	AAB37406	P80356
P04725	P08079	AAB48478	AAB23624	AAA32955	AAB37407	P27919
P04724	P06659	AAB48479	AAB23625	AAA32967	AAB58403	P14812
P04723	P04730	BAA22613	AAB23626	AAA92333	AAG35598	JQ1048
P04722	P04729	BAA22614	AAB23627	AAB28161	CAA26449	JQ1047
P04721	JS0402	BAA23162	AAB23628	AAB71678	S18235	JQ1046
P02863	JA0153	BAB78737	AAD32223	AAB71679	S18236	JG0015
EEWTA	EEWTG	BAB78738	AAF23506	B24095	S70327	B36433
E22364	CAC94871	BAB78739	AAF23507	B25677	S70328	AAB32025
D22364	CAC94870	BAB78740	B30843	BAA11642	S70329	AAB23365
CAB76964	CAC94869	BAB78741	CAA26847	CAA25509		AAA32716
CAB76963	CAC94868	BAB78742	CAA27052	CAA25912		AAA32715
CAB76962	CAC11089	BAB78743	CAA31395	CAA25913		AAA32714
CAB76961	CAC11088	BAB78744	CAA31396	CAA25914		AAA32713
CAB76960	CAC11087	BAB78745	CAA32115	CAA26889		1502200A
CAB76959	CAC11080	BAB78746	CAA43331	CAA31861		1411172A
CAB76958	CAC11079	BAB78747	CAA43361	CAA37729		
CAB76957	CAC11078	BAB78748	CAA59340	CAA42642		
CAB76956	CAC11065	BAB78749	CAC40684	CAA48209		
CAB76955	CAC11064	BAB78750	CAC40685	CAA51204		
CAB76954	CAC11057	BAB78751	CAC40686	CAA59104		
CAA35238	CAC11056	BAB78752	CAC40687	CAA60681		
CAA26385	CAC11055	BAB78753	CAC83002	P02864		
CAA26384	CAB75404	BAB78754	CAC83003	P06470		
CAA26383	BAA11251	BAB78755	CAC83018	P06471		
CAA10257	AAN32705	BAB78756	CAC84118	P06472		
C22364	AAK84880	BAB78757	CAC84119	P17990		
BAA12318	AAK84780	BAB78758	CAC84120	P17991		
B22364	AAK84779	BAB78759	CAC84121	P17992		
AAN32704	AAK84778	BAB78760	CAC84122	P80198		
AAB23109	AAK84777	BAB78761	EEWTHW	S07189		
AAB23108	AAK84776	BAB78762	JC2099	S07365		
AAA96525	AAK84775	BAB78763	JC4966	S07975		
AAA96524	AAK84774	BAB78764	JN0689	S07976		
AAA96523	AAK84773	CAA30570	JN0690	S08312		
AAA96522	AAK84772	CAA31685	P02861	S18350		
AAA96276	AAF42989	CAA59313	P02862	S20519		
AAA34283	AAD30556	CAA59338	P08488	S52390.1		
AAA34282	AAD30440	CAA59339	P08489	T04369		
AAA34281	AAB31090	CAA59340	P10387	T04473		
AAA34280	AAA34289	CAA76890	P10388	T04474		
AAA34279	AAA34288	EEWT1	S02262	T05718		
AAA34278	AAA34287	P10385	S04832	T05737		
AAA34277	AAA34286	P10386	S15720	T06211		
AAA34276	AAA34285	P16315	S18733	1103203A		

AAA17741	AAA34274	S01992	S29176	1103203B		
A27319	1802407A	S04325	S29177	1103203C		
A22364	1507333A	S57645	S29178	1210226A		
1307187B	1209306A	S57654	S29179	1307151A		
	Omega gliadins (n=2)	S57655	AAN78346	1307151B		
	A59156	S57656	AAO74630	1604464A		
	AAG17702	T05910		A24095.2		
		T05923		AAP31051		
		T06505.1		CAA48209.2		
		T06505.2		S52390.2		
		T06506				
		T06508				
		T06980				
		T06981				
		T06982				
		AAP44992				
		AAP44991				
		AAP44989				
		AAO53259				
		CAD58622				
		CAD58619				
		CAD58621				
		A03353				
		AAO53264				
		AAO53265				
		AAO53266				
		AAO53267				
		CAA76890				
TOTAL:	All gliadins					
20mers	721	645	786	416	155	199
12mers	4465	3945	4799	2672	957	1279
9mers	3739	3164	3630	2413	811	1207

FIGURE 3

Figure 3A

M-Step:

$$\frac{dQ(\theta,\theta^{(c)})}{dp_j^{(c)}}=0 \qquad \rightarrow p_j^{(c+1)}=\frac{\sum_i \hat{z}_{ij}^{(c)}}{n}$$

$$\frac{dQ(\theta,\theta^{(c)})}{d\alpha_i^{(c)}}=0 \qquad \rightarrow \alpha_i^{(c+1)}=\frac{\sum_j[\hat{z}_{ij}^{(c)}y_{ij}+(1-\hat{z}_{ij}^{(c)})y_{ij}]}{\sum_j[\lambda_j^{(c)}\hat{z}_{ij}^{(c)}+\lambda_0^{(c)}(1-\hat{z}_{ij}^{(c)})]}$$

$$\frac{dQ(\theta,\theta^{(c)})}{d\lambda_j^{(c)}}=0 \qquad \rightarrow \lambda_j^{(c+1)}=\frac{\sum_i \hat{z}_{ij}^{(c)}y_{ij}}{\sum_i \alpha_i^{(c)}\hat{z}_{ij}^{(c)}}$$

$$\frac{dQ(\theta,\theta^{(c)})}{d\lambda_0^{(c)}}=0 \qquad \rightarrow \lambda_0^{(c+1)}=\frac{\sum_{ij}(1-\hat{z}_{ij}^{(c)})y_{ij}}{\sum_{ij}\alpha_i^{(c)}(1-\hat{z}_{ij}^{(c)})}$$

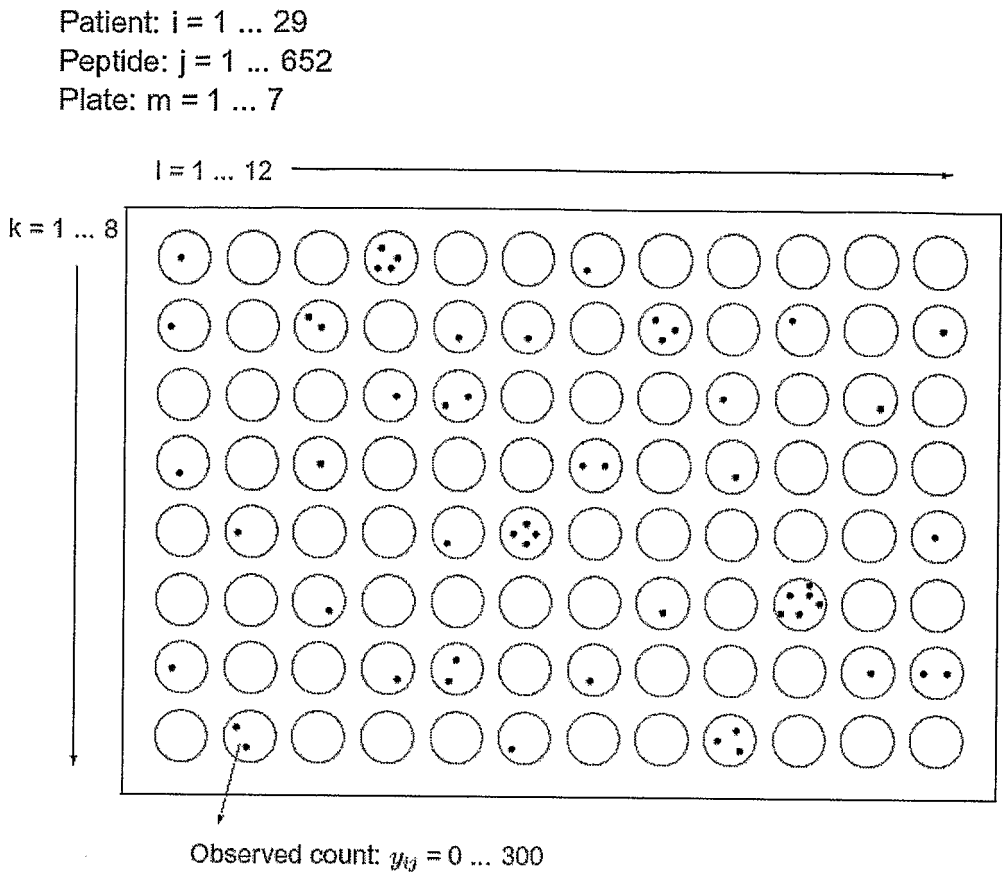
E-Step:

$$\hat{z}_{ij}^{(c+1)}=E(z_{ij}|y_{ij})=pr(z_{ij}=1|y_{ij})=\frac{p_j^{(c)}pr(y_{ij}|\alpha_i\lambda_j^{(c)})}{p_j^{(c)}pr(y_{ij}|\alpha_i\lambda_j^{(c)})+(1-p_j^{(c)})pr(y_{ij}|\alpha_i\lambda_0^{(c)})}$$

Compute:

$$Q(\theta,\theta^{(c)})=\sum_{ij}\{\hat{z}_{ij}^{(c)}\log p_j+(1-\hat{z}_{ij}^{(c)})\log(1-p_j)+y_{ij}\hat{z}_{ij}^{(c)}\log(\alpha_i\lambda_j)$$
$$+y_{ij}(1-\hat{z}_{ij}^{(c)})\log(\alpha_i\lambda_0)-\hat{z}_{ij}^{(c)}\alpha_i\lambda_j-(1-\hat{z}_{ij}^{(c)})\alpha_i\lambda_0\}$$

Figure 3B


Patient: i = 1 ... 29
Peptide: j = 1 ... 652
Plate: m = 1 ... 7
l = 1 ... 12
k = 1 ... 8
Observed count: y_{ij} = 0 ... 300

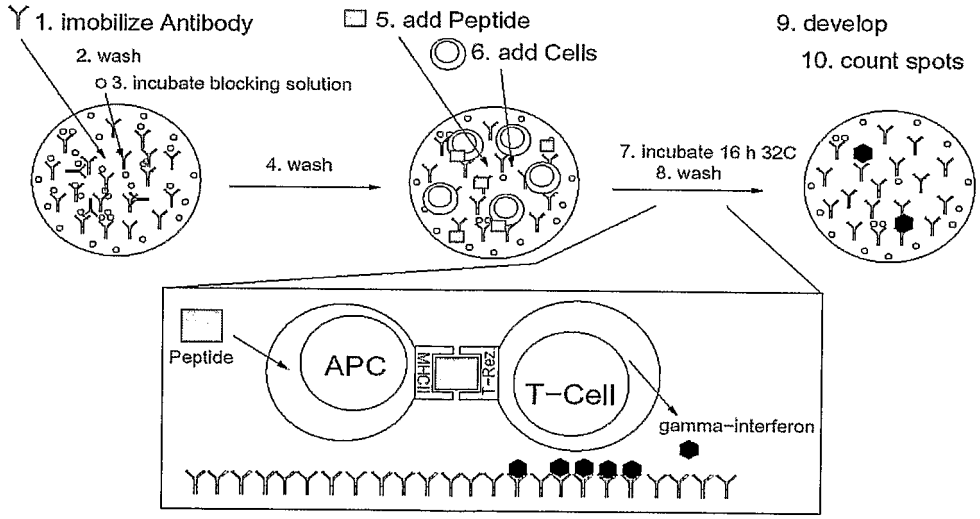

1. imobilize Antibody
2. wash
3. incubate blocking solution
4. wash
5. add Peptide
6. add Cells
7. incubate 16 h 32C
8. wash
9. develop
10. count spots
Peptide
APC
MHCII
zeta-1
T-Cell
gamma-interferon

Figure 4

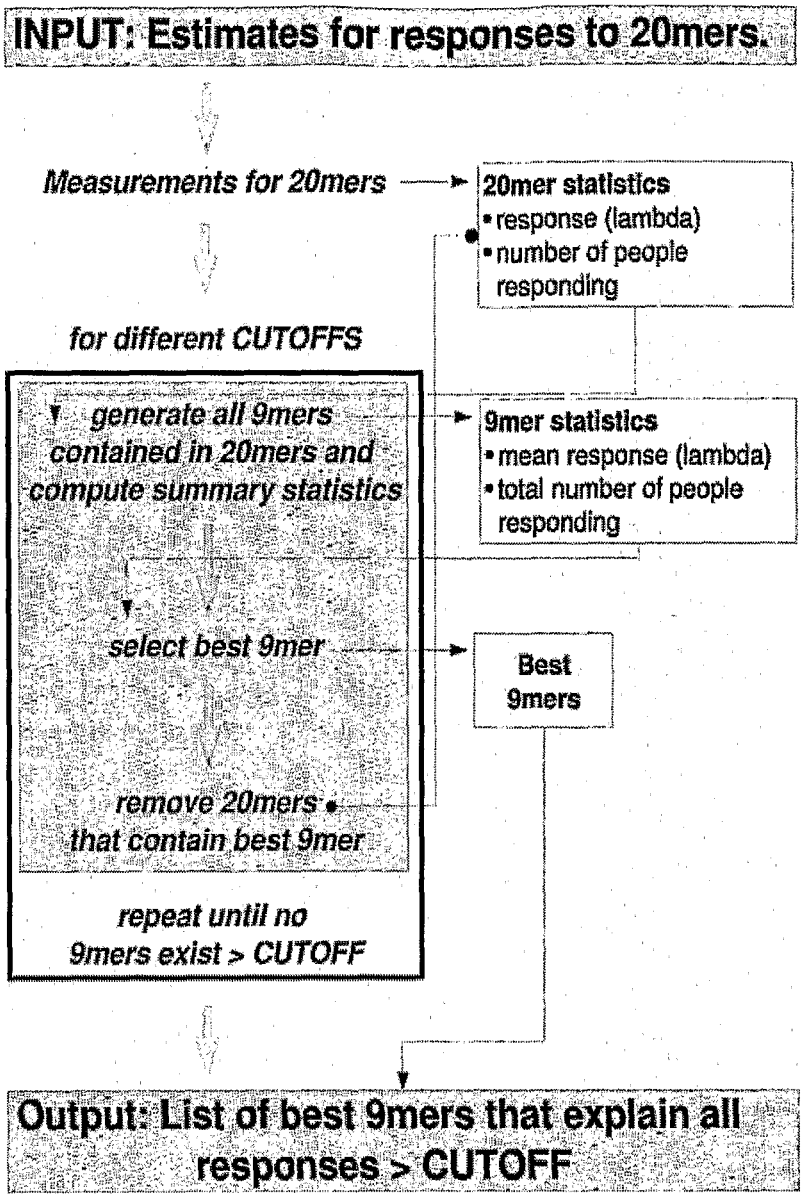

INPUT: Estimates for responses to 20mers.
Measurements for 20mers
20mer statistics
• response (lambda)
• number of people responding
for different CUTOFFS
generate all 9mers contained in 20mers and compute summary statistics
9mer statistics
• mean response (lambda)
• total number of people responding
select best 9mer
Best 9mers
remove 20mers that contain best 9mer
repeat until no 9mers exist > CUTOFF
Output: List of best 9mers that explain all responses > CUTOFF

Figure 5

Group	Consensus	SEQ ID NO:	Sequence	SEQ ID NO:
1	gEIpFpEpE	1	gqlpypqpelpypqpg	200
			gqlpypqpqlpypqpg	201
2	EIpFpEpEI	2	ylqlqpfpqpqlpypqpqlp	202
			pqpfppqlpypqpqlpypqp	203
			pymqlqpfpqpqlpypqpql	204
			qlqpfpqpqlpypqpql	205
			gqlpypqpelpypqpg	206
			gqlpypqpqlpypqpg	207
			pqpfppqlpypqpqlpypqp	208
			mqlqpfpqpqlpypqpqlpy	209
			qlqpfpqpelpypqpql	210
			pqlpypqpqlpypqpqlpyp	211
			pqlpypqpqlpypqpqpfrp	212
			lqpfpqpelpypqpelpypf	213
			lqlqpfpqpqlpypqpqlpy	214
			pqpfppqlpypqpqlpypqp	215
3	EEpFpFEpE	3	qpfpqpqqpfpwqpqqpfpq	216
			pqqpqqpfpqpqqpfpwqpq	217
			qpfpqpqqpfpwqpqqpfpq	218
			qpfpqpqqpfpwqpqqpfpq	219
5	FpEpEEpIp	4	qqpfpqqpqqpfpqpqqpip	220
			pqqpqqpfpqpqqpipvqpq	221
			pqqpqqpfpqpqqpipvqpq	222
			gqqpfpqpeqpipvqg	223
			gqqpfpqpqqpipvqg	224
			pqqpqqpfpqpqqpipvqpq	225
6	FpEpEEpFp	5	qpfpqpqqpfpwqpqqpfpq	226
			pqqpqqpfpqpqqpfpwqpq	227
			pqqpqqpqqpfpqpqqpfpw	228
			qpfpqpqqpfpwqpqqpfpq	229
			gqqpfpqpeqpfpwqg	230
			gqqpfpqpqqpfpwqg	231
			pqqpqqpqqpfpqpqqpfpw	232
			qpfpqpqqpfpwqpqqpfpq	233
			qpqqpfpqpeqpfpwqp	234
			qpeqpfpqpeqpfpwqp	235
			qpqqpfpqpqqpfpwqp	236
			qpeqpfpqpqqpfpwqp	237
7	EpEIpFpEp	6	ylqlqpfpqpqlpypqpqlp	238
			pqpfppqlpypqpqlpypqp	239
			pymqlqpfpqpqlpypqpql	240
			qlqpfpqpqlpypqpql	241
			gqlpypqpelpypqpg	242
			glqpfpqpelpypqpg	243
			lqlqpfpqpqlpypqpqpfr	244
			qlqpfpqpelpypqpqp	245

			gqlpyp**qpqlpypqp**g	246
			pqpfppqlpyp**qpqlpypqp**	247
			lqlqpfp**qpqlpypqp**qpfr	248
			lqlqpfp**qpelpypqp**elpypqpelpypqpqpf	249
			mqlqpfp**qpqlpypqp**qlpy	250
			glqpfp**qpelpfpqp**g	251
			qlqpfp**qpelpypqp**ql	252
			pqlpyp**qpqlpypqp**qlpyp	253
			qlqpfp**qpqlpypqp**qp	254
			glqpfp**qpqlpypqp**g	255
			pqlpyp**qpqlpypqp**qpfrp	256
			lqpfp**qpelpypqp**elpypf	257
			lqlqpfp**qpqlpypqp**qlpy	258
			glqpfp**qpqlpfpqp**g	259
			pqpfppqlpyp**qpqlpypqp**	260
			yp**qpqlpypqp**qpfrpqqsy	261
			gsg**qpelpypqp**gsg	262
			lqlqpfp**qpqlpypqp**qlpypqpqlpypqpqpf	263
8	EEpFpEpEI	7	qqp**qqpfpqpql**pfpqqseq	264
			fpqqp**qqpfpqpql**pfpqqs	265
9	pEEpFSEEg	8	gqqpfpq**peqpfsqqg**	266
			gqqpfpq**pqqpfsqqg**	267
10	gFpEpEIpF	9	gs**gfpqpelpy**pgsg	268
11	pEEEpEEpF	10	qpqlpfpqq**pqqqpqqpf**pq	269
			gqq**pqqqpeqpf**pqqg	270
			gqq**pqqqpqqpf**pqqg	271
12	IEEpEEpFp	11	**lqqpqqpfp**qpqqqlpqpqq	272
			lqqpqqpfpqpqqqlpqpqq	273
			qpqqpfpq**lqqpqqpfp**qpq	274
			pfpq**lqqpqqpfp**qpqqqlp	275
			gfpq**lqqpeqpfp**qqg	276
			gfpq**lqqpqqpfp**qqg	277
13	pEpEpFIpE	12	lqpf**pqpqpflpq**lpypqpq	278
			lqlqpf**pqpqpflpq**lpypq	279
14	EEISpcmdI	13	lqqilq**qqltpcmdv**vlqqh	280
			ilqqilq**qqltpcmdv**vlqq	281
15	FEEEpFpEE	14	qspqqsfs**yqqqpfpqq**pyp	282
			yqqqpfpqqpypqqpypsqq	283
			pqqsfs**yqqqpfpqq**pypqq	284
16	EpFpEpEIp	15	ylql**qpfpqpqlp**ypqpqlp	285
			pymql**qpfpqpqlp**ypqpql	286
			ql**qpfpqpqlp**ypqpql	287
			gl**qpfpqpelp**ypqpg	288
			lql**qpfpqpqlp**ypqpqpfr	289
			ql**qpfpqpelp**ypqpqp	290
			lql**qpfpqpqlp**ypqpqpfr	291
			lql**qpfpqpelp**ypqpelpypqpelpypqpqpf	292
			mql**qpfpqpqlp**ypqpqlpy	293
			gl**qpfpqpelp**fpqpg	294

			ql**qpfpqpelp**ypqpql	295
			ql**qpfpqpqlp**ypqpqp	296
			gl**qpfpqpqlp**ypqpg	297
			l**qpfpqpelp**ypqpelpypf	298
			qqpq**qpfpqpqlp**fpqqseq	299
			lql**qpfpqpqlp**ypqpqlpy	300
			fpqqpq**qpfpqpqlp**fpqqs	301
			gl**qpfpqpqlp**fpqpg	302
			gl**qpfpqpelp**ylqpg	303
			pylql**qpfpqpqlp**ysqpqp	304
			gl**qpfpqpelp**ysqpg	305
			lql**qpfpqpqlp**ylqpqpfr	306
			lql**qpfpqpqlp**ylqpqpfr	307
			lql**qpfpqpqlp**ysqpqpfr	308
			gsg**qpfpqpelp**gsg	309
			lql**qpfpqpqlp**ypqpqlpypqpqlpypqpqpf	310
			gl**qpfpqpqlp**ysqpg	311
			qpfpqpqlpysqpqqfrpqq	312
			lql**qpfpqpqlp**ysqpqqfr	313
			gl**qpfpqpqlp**ylqpg	314
			qfpsqlpylql**qpfpqpqlp**	315
			pfpsqqpymql**qpfpqpqlp**	316
			pfpsqlpylql**qpfpqpqlp**	317
			pfpsqqpylql**qpfpqpqlp**	318
			pfpsqqpylql**qpfpqpqlp**	319
17	pEEpEEEFp	16	pqqpf**pqqpqqqfp**qpqqpq	320
			gqpf**pqqpeqefp**qpg	321
			qqpqqpf**pqqpqqqfp**qpqq	322
			gqpf**pqqpeqqfp**qpg	323
			gf**pqqpeqefp**qpqqg	324
			pqqpf**pqqpqqqfp**qpqqpq	325
			gf**pqqpqqefp**qpqqg	326
			gqpf**pqqpqqqfp**qpg	327
			qqpqqpf**pqqpqqqfp**qpqq	328
			pqqpqqpf**pqqpqqqfp**qpq	329
			gf**pqqpqqqfp**qpqqg	330
18	pFSEEEEpI	17	gqp**pfsqqeqpv**lpqg	331
			lqqs**pfsqqqqpv**lpqqqpv	332
			pfsqqqqp**pfsqqqqpv**lpq	333
			qqqpilsqqp**pfsqqqqpv**l	334
			qp**pfsqqqqpv**lpqqspfsq	335
			qqqvlpqqp**pfsqqqqpv**ll	336
			qqqpilsqqp**pfsqqqqpv**l	337
			qqqp**pfsqqqqpi**lpqqppf	338
			pfsqqqp**pfsqqqqpi**lpqq	339
			qqqpilpql**pfsqqqqpv**lp	340
			qqp**pfsqqqqpv**lppqqspf	341
			gqp**pfsqqqqpv**lpqg	342
			qs**pfsqqqqpv**lpqqqpvii	343
			qp**pfsqqqqpv**lpqqspfsq	344
			qqqp**pfsqqqqpv**ipqqpsf	345

			lppfsqqlp**pfsqqqqpv**lp	346
			qqqqp**pfsqqqqpv**lpqqsp	347
			sqqqpilsqqp**pfsqqqqpv**	348
			pfsqqqqpvllqqqipfvhp	349
19	pEEpSpIEp	18	ftq**pqqptpiqp**qqpfpqqp	350
			pftq**pqqptpiqp**qqpfpqq	351
			fpeqsqqpftq**pqqptpiqp**	352
20	EpEEpIpIE	19	pqqpqqpfp**qpqqpipvq**pq	353
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			pqp**fppqlpypq**pqsfppqq	1522
			mqlqpfpqpqp**fppqlpypq**	1523
			gqpqp**fppelpypq**pg	1524
			gqpqp**fppqlpypq**pg	1525
			pfpqpqp**fppqlpypq**pppf	1526
			pfpqpqp**fppqlpypq**pqsf	1527
			pqpqpfpqpqp**fppqlpypq**	1528
			pqp**fppqlpypq**pppfspqq	1529
			gsg**fppelpypq**gsg	1530
			pqpqp**fppqlpypq**tqpfpp	1531
			pqp**fppqlpypq**tqpfppqq	1532
			qp**fppqlpypq**tqpfppqqp	1533
195	cSpIampER	194	ckvflqqq**cspvampqr**lar	1534
			lqqq**cspvampqr**lar	1535
			vflqq**cspvampqr**larsqm	1536
			lnpckvflqqq**cspvampqr**	1537
			qqq**cspvampqr**larsqmwq	1538
			cspvampqrlarsqmwqqss	1539
			q**cspvampqh**larsqmwqqs	1540
			ckvflqqq**cspvampqr**lar	1541
			qlnpckvflqq**cspvampqr**	1542
			qqq**cspvampqr**larsqmwq	1543
			vflqqq**cspvampqh**larsq	1544
			vflqqq**cspvampqr**larsq	1545
196	dEESgEgEE	195	sgqrqq**dqqsgqgqq**pgqrq	1546
197	EIEESIIFg	196	ttppq**qlqqsilwg**ipallr	1547
198	pFSEEEIpI	197	qqp**pfsqqelpi**lpqqppfs	1548
			qqqppfsqqqp**pfsqqelpi**	1549
			fsqqqp**pfsqqelpi**lpqqp	1550
199	SSRIpgIER	198	me**tshipglek**psqqqplpl	1551
			me**tsrvpglek**pwqqqplpp	1552
200	SIaIRSIpm	199	lmt**sialrtlpm**mcsvnvpv	1553
			hlevmt**sialrtlpm**mcsvn	1554

Figure 6

Sequence 1: P(QR)P(QE)LP(FY)PQ (SEQ ID NO: 1555)
Optimal responding peptide: PQLPYPQPQLPYPQPQPFRP (SEQ ID NO: 1556)
High grade peptide: QLQPFPQPELPYPQPQP (SEQ ID NO: 1557)
Groups included: 1,2,7,8,10,16,22,23,72

Sequence 2: P(FY)P(QR)P(QE)LP(FY) (SEQ ID NO: 1558)
Optimal responding peptide: PQLPYPQPQLPYPQPQPFRP (SEQ ID NO: 1559)
High grade peptide: QLQPFPQPELPYPQPQP (SEQ ID NO: 1560)
Groups included: 1,2,7,8,10,16,22,23,72, 76 PART

Sequence 3: (PIAT)FPQ(PT)(QE)Q(PTS)(FITY) (SEQ ID NO: 1561)
Optimal responding peptide: QPFPQPQQPFPWQPQQPFPQ (SEQ ID NO: 1562)
High grade peptides: GQQPFPQPEQPFPWQG (SEQ ID NO: 1563)
GQQPFPQPEQPIPVQG (SEQ ID NO: 1564)
Groups included:
3,5,6,9,20,24,29,60,69,70,73,74,77,102PT,108,112

Sequence 4: PQ(PT)(QE)Q(PTS)(FIY)(PS)(VWHQL) (SEQ ID NO: 1565)
Optimal responding peptide: QPFPQPQQPFPWQPQQPFPQ (SEQ ID NO: 1566)
High grade peptide: GQQPFPQPEQPFPWQG (SEQ ID NO: 1567)
GQQPFPQPEQPIPVQG (SEQ ID NO: 1568)
Groups included: 3,5,6,9,20,60,66,69,73,74,77,PART 86
(3 AND 4 OVERLAP)
Status: Similar to Sequence 3

Sequence 5: (KQW)(QR)P(QE)Q(SPIT)(FLY)PQ (SEQ ID NO: 1569)
Optimal responding peptide: QPQLPFPQQPQQQPQQPFPQ (SEQ ID NO: 1570)
High grade peptide: GFPQTQQPEQPFPQQG (SEQ ID NO: 1571)
Groups included: 11,12,32,54,55,56,59,61,62,65

Sequence 6: P(FIYL)(PS)(QE)(QR)P(QE)Q(PT) (SEQ ID NO: 1572)
Optimal responding peptide: PLQPQQPFPQQPQQPFPQPQ (SEQ ID NO: 1573)
High grade peptide: GQPFPEQPQQPFPQQG (SEQ ID NO: 1574)
Groups included: 32PT,54,55,56,57,61,62,63PT,75,85.94PT

Sequence 7: FLP(QE)LPYPQ (SEQ ID NO: 1575)
Optimal responding peptide: LQLQPFPQPQPFLPQLPYPQ (SEQ ID NO: 1576)
High grade peptide: PQPQPFLPELPYPQPQS (SEQ ID NO: 1577)
Groups included: 13,27,71 PART

Sequence 8: LQQIL(QE)QQL (SEQ ID NO: 1578)
Optimal responding peptide: LQQILQQQLTPCMDVVLQQH (SEQ ID NO: 1579)
High grade peptide: NO
Groups included: 14,89

Sequence 9: FSYQ(EQ)QPFPQQ (SEQ ID NO: 1580)

Optimal responding peptide: PQQSFSYQQQPFPQQPYPQQ (SEQ ID NO: 1581)
High grade peptide: NO
Groups included: 15,96PT

Sequence 10: FPS(QE)(LQ)PY(LM)Q (SEQ ID NO: 1582)
Optimal responding peptide: PFPSQQPYLQLQPFPQPQLP (SEQ ID NO: 1583)
High grade peptide: NO
Groups included: 16PT,35,38,71PT, 76PT,92PT,93PT

Sequence 11: (PQSH)QP(QE)Q(QE)(LF)(PS)Q (SEQ ID NO: 1584)
Optimal responding peptide: QQPQQPFPQQPQQQFPQPQQ (SEQ ID NO: 1585)
High grade peptide: GFFPQPEQEFPQPQQG (SEQ ID NO: 1586)

Groups included: 17,25,36,40,41,80,88

Sequence 12: P(FW)(SP)(EQ)Q(EQT)QP(VILSF) (SEQ ID NO: 1587)
Optimal responding peptide: QQQQPPFSQQQQPVLPQQSP (SEQ ID NO: 1588)
(similar to but more active than PFSQQQQSF in WO 02/083722)
PSGQVQWPQQQQPFPQPQQP (SEQ ID NO: 1589)
High grade peptide: GQPPFSEQEQPVLPQG (SEQ ID NO: 1590)
Groups included: 18,79,84PT,97,102PT,103PT,115

Sequence 13: (IL)QP(QE)QPFPQ (SEQ ID NO: 1591)
Optimal responding peptide: FTQPQQPTPIQPQQPFPQQP (SEQ ID NO: 1592)
(similar to but more active than IIQPQQPAQ in WO 02/083722)
High grade peptide: GQQQFIQPEQPFPQQG (SEQ ID NO: 1593)
Groups included: 19 (PART),26,30,58

Sequence 14: QQP(EQ)LPFPQ (SEQ ID NO: 1594)
Optimal responding peptide: QQPQQPFPQPQQPQLPFPQQ (SEQ ID NO: 1595)
High grade peptide: NO
Groups included: 21,64

Sequence 15: QPQQP(EQ)LPF (SEQ ID NO: 1596)
Optimal responding peptide: QQPQQPFPQPQQPQLPFPQQ (SEQ ID NO: 1597)
High grade peptide: NO
Groups included: 21

Sequence 16: PQP(EQ)QP(EQ)LP (SEQ ID NO: 1598)
Optimal responding peptide: QQPQQPFPQPQQPQLPFPQQ (SEQ ID NO: 1599)
High grade peptide: NO
Groups included: 21

Sequence 17: VFLQQQCSPV (SEQ ID NO: 1600)
Optimal responding peptides:
GROUP 28: SVLQQLNPCKVFLQQQCSHV (SEQ ID NO: 1601)
GROUP 122: CKVFLQQQCSPVAMPQRLAR (SEQ ID NO: 1602)
GROUP 114: KVFLQQQCSPVAIPYRLARS (SEQ ID NO: 1603)
High grade peptide: NO
Groups included: 28,114,122

Sequence 18: (M)(WL)(QW)QSSCHVMQ (SEQ ID NO: 1604)
Optimal responding peptides:
GROUP 39: PQRLARSQMWQQSSCHVMQQ (SEQ ID NO: 1605)
GROUP 110: ARSQTLWQSSCHVMQQCCR (SEQ ID NO: 1606)
High grade peptide: NO
Groups included: 39,110

Sequence 19: QPQQQQLAH (SEQ ID NO: 1607)
Optimal responding peptide: QQPQQQQLAHGTFLQPHQIA (SEQ ID NO: 1608)
High grade peptide: NO
Groups included: 31

Sequence 20: FPLQPQQP(FL)PQ (SEQ ID NO: 1609)
Optimal responding peptide: PQQLQQPFPLQPQQPFPQQP (SEQ ID NO: 1610)
High grade peptide: NO
Groups included: 33

Sequence 21: FPP(QE)(LQ)PYPQ (SEQ ID NO: 1611)
Optimal responding peptide: PQPFPPELPYPQPQPFRPQQ (SEQ ID NO: 1612)
High grade peptide: PQPQPFPPQLPYPQPQS, (SEQ ID NO: 1613)
GQQQPFPPEQPYPQQG (SEQ ID NO: 1614)
Groups included: 37,52,71,92PT,93PT,105

Sequence 22: LCC(QE)(HQR)L(PW)(QE)IP (SEQ ID NO: 1615)
Optimal responding peptide: QSTYQPLQQLCCQQLWQIPE (SEQ ID NO: 1616)
High grade peptide: NO
Groups included: 39PT,100,104,107,178

Sequence 23: P(WLS)(QL)(QE)QPL(PQ)(PQ) (SEQ ID NO: 1617)
Optimal responding peptide: ERPWQEQPLPPQHTLFPQQQ (SEQ ID NO: 1618)
High grade peptide: NO
Groups included: 44,78,90,95PT,98,116,117,113

Sequence 24: QPLP(QE)QPSF (SEQ ID NO: 1619)
Optimal responding peptide: PPFSQQQQQPLPQQPSFSQQ (SEQ ID NO: 1620)
High grade peptide: NO
Groups included: 45,95PT

Sequence 25: PF(SP)QQQQQP(LVI) (SEQ ID NO: 1621)
Optimal responding peptide: PPFSQQQQQPLPQQPSFSQQ (SEQ ID NO: 1622)
High grade peptide: NO
Groups included: 45 PART

Sequence 26: IVYSTILQE (SEQ ID NO: 1623)
Optimal responding peptide: QQSRYEAIRAIVYSTILQEQ (SEQ ID NO: 1624)
High grade peptide: NO
Groups included: 46

Sequence 27: PFSQ(QE)QP(IS)(LF)S (SEQ ID NO: 1625)
Optimal responding peptide: FSQQQPPFSQQQPILSQQPP (SEQ ID NO: 1626)
High grade peptide: NO
Groups included: 47 PART,68

Sequence 28: QGIQILRPL (SEQ ID NO: 1627)
Optimal responding peptide: QEQQQGIQILRPLFQLVQGQ (SEQ ID NO: 1628)
High grade peptide: NO
Groups included: 48

Sequence 29: PFSSVVAGI (SEQ ID NO: 1629)
Optimal responding peptide: CSIIKAPFSSVVAGIGGQYR (SEQ ID NO: 1630)
High grade peptide: NO
Groups included: 49

Sequence 30: YCSTTIAPV (SEQ ID NO: 1631)
Optimal responding peptide: YIPPYCSTTIAPVGIFGTN (SEQ ID NO: 1632)
High grade peptide: NO
Groups included: 50

Sequence 31: HVAMSQRLA (SEQ ID NO: 1633)
Optimal responding peptides: QQCSHVAMSQRLARSQMWQQ (SEQ ID NO: 1634)
High grade peptide: NO
Groups included: 51

Sequence 32: YSIILQ(QE)(QE)(QE)QGF (SEQ ID NO: 1635)
Optimal responding peptide: RYDAICAITYSIILQEQQQG (SEQ ID NO: 1636)
High grade peptide: NO
Groups included: 53

Sequence 33: FPHQPQEQAFPQ (SEQ ID NO: 1637)
Optimal responding peptide: QQIFPQPQQTFPHQPQQAFP (SEQ ID NO: 1638)
High grade peptide: GQTFPHQPEQAFPQPG (SEQ ID NO: 1639)
Groups included: 67

Sequence 34: PS(GS)(QE)V(QE)WPQ (SEQ ID NO: 1640)
Optimal responding peptide: ATANMQADPSGQVQWPQQQP (SEQ ID NO: 1641)
High grade peptide: NO
Groups included: 81,102PT

Sequence 35: GALCSSLSN (SEQ ID NO: 1642)
Optimal responding peptide: FDEEKNSTGALCSSLSNQAS (SEQ ID NO: 1643)
High grade peptide: NO
Groups included: 82

Sequence 36: QFP(QE)Q(QE)IPV (SEQ ID NO: 1644)
Optimal responding peptide: QPPFPQQHQQFPQQQIPVVQ (SEQ ID NO: 1645)
High grade peptide: NO
Groups included: 83

Sequence 37: P(FY)P(QE)QP(YF)PQ (SEQ ID NO: 1646)
Optimal responding peptide: QQPFPQQPYPQQPYPSQQPY (SEQ ID NO: 1647)
High grade peptide: NO
Groups included: 96

Sequence 38: QAGQG(QE)(QE)GY (SEQ ID NO: 1648)
Optimal responding peptide: GQQAGQGQQGYYPTSPQQLG (SEQ ID NO: 1649)
High grade peptide: NO
Groups included: 101

Sequence 39: TLPSMCNVY (SEQ ID NO: 1650)
Optimal responding peptide: FEEIRNLALQTLPSMCNVYI (SEQ ID NO: 1651)
High grade peptide: NO
Groups included: 106

Sequence 40: FQPSQ(QE)NPQ (SEQ ID NO: 1652)
Optimal responding peptide: QQYPSGQGFFQPSQQNPQAQ (SEQ ID NO: 1653)
High grade peptide: NO
Groups included: 109

Sequence 41: IRSLVLKTL (SEQ ID NO: 1654)
Optimal responding peptide: QPQQPAQLEGIRSLVLKTLP (SEQ ID NO: 1655)
High grade peptide: NO
Groups included: 119,120,121

Figure 7

Gliadin Sequence	SEQ ID NO:	High quality peptide	A	B	C	D	E	F	G	H	I	J
1,2	1656	QLQPFPQPQLPYPQPQP	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
3,4	1657	GQQPFPQPEQPFPWQG	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
3,4	1658	GQQPFPQPEQPIPVQG	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
3,4	1659	GQQPFPQPEQPFPWQG	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
5	1660	GFPQTQQPEQPFPQQG		Y	Y				Y	Y		Y
5	1661	GFPQTQQPEQPFPQQG		Y	Y				Y	Y		Y
6	1662	GQPFPEQPQQPFPQQG								Y		
7	1663	PQPQPFLPQLPYPQPQS	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
11	1664	GFFPQPEQEFPQPQQG		Y	Y	Y	Y		Y			Y
11	1665	GFFPQPEQEFPQPQQG		Y	Y	Y	Y		Y			Y
12	1666	GQPPFSEQEQPVLPQG					Y		Y		Y	
12	1667	GQPPFSEQEQPVLPQG					Y		Y		Y	
13	1668	GQQQFIQPEQPFPQQG		Y	Y	Y		Y	Y	Y		Y
21	1669	PQPQPFPPQLPYPQPQS	Y	Y	Y	Y	Y	Y	Y	Y	Y	Y
21	1670	GQQQPFPPEQPYPQQG		Y	Y						Y	Y
33	1671	GQTFPHQPEQAFPQPG		Y	Y				Y		Y	Y

Figure 8

Avenin 20mer	SEQ ID NO:	Predicted core sequence(s)	SEQ ID NO:	Lambda	Proportion
TTTVQYDPSEQYQPYPEQQQ	1672	DPSEQYQPY	1684	33.58	0.145
QFDPSEQYQPYPEQQQPILQ	1673	PYPEQQQPI DPSEQYQPYP	1685 1686	122.34	0.103
*VQYDPSEQYQPYPEQQQPFV	1674	PYPEQQQPF DPSEQYQPYP	1687 1688	122.34	0.105
TVQYNPSEQYQPYPEQQEPF	1675	PYPEQQEPF DPSEQYQPYP	1689 1690	146.42	0.051
YQPYPEQQQPILQQQQMLLQ	1676	PYPEQQQPI	1691	52.56	0.126
EQYQPYPEQQQPFVQQQPPF	1677	PYPEQQQPF	1692	90.39	0.053
SEQYQPYPEQQPFMQPLLQQ	1678	PYPEQQPFM	1693	19.07	0.226
*CRRLEQIPEQLRCPAIHSVV	1679	RRLEQIPEQ	1694	61.47	0.069
*QIPEQLRCPAIHSVVQAIIL	1680			94.84	0.033
*NNKREQQFGQNIFSGFSVQL	1681			104.78	0.077
*QILRQAICQVTRQQCCRQLA	1682			34.01	0.055
*VPFLRSQILRQSTCHVMRRQ	1683			92.47	0.067

Sequence 1: PYPEQ(QE)QP(IF)(VLM) (SEQ ID NO:1695)
Optimal responding peptide: QFDPSEQYQPYPEQQQPILQ (SEQ ID NO:1696)
High grade peptide: NO
POTENCY: 6/30 RESPONDERS, RESPONSE RATE 122.3
(HOMOLOGOUS TO WHEAT Sequence 12: GQPPFSEQEQPVLPQG (SEQ ID NO:1590))

Sequence 2: CRRLEQIPEQLRCPAIHSVV (SEQ ID NO:1697)

Sequence 3: QFGQNIFSGFSVQLLSEALG (SEQ ID NO:1698)
POTENCY: 12/30 RESPONDERS, RESPONSE RATE 11.2

Figure 10

VRVPVPQLQP QNPSQQQPQE QVPLVQQQQF PGQQQQFPPQ
QPYPQPQPFP SQQPYLQLQP FPQPQLPYPQ PQSFPPQQPY
PQPQPQYSQP QQPISQQQAQ QQQQQQQQQQ QQQILQQILQ
QQLIPCMDVV LQQHNIAHAR SQVLQQSTYQ LLQELCCQHL
WQIPEQSQCQ AIHNVVHAII LHQQQKQQQQ PSSQVSFQQP
LQQYPLGQGS FRPSQQNPQA QGSVQPQQLP QFEEIRNLAL
QTLPAMCNVY IAPYCTIAPF GIFGTN (SEQ ID NO: 1928)

Figure 9

Group	Predicted epitope "core(s)"	SEQ ID NO:	Sequences	SEQ ID NO:	Response (SFC) A	B
1	QQPTPIQPQ	1699	PFTQPQQPTPIQPQQPFPQQ	1722	70	18
2	QQPFPQQPQ or QQPFPWQPQ or QQPFPQSQQ	1700	QQQFIQPQQPFPQQPQQTYP	1723	57	21
		1701	PFPQQPQQPFPQQPQQSFPQ	1724	53	23
		1702	TPIQPQQPFPQQPQQPQQPF	1725	64	16
			QTQQPQQPFPQQPQQPFPQT	1726	52	17
			QQFLQPQQPFPQQPQQPYPQ	1727	44	16
			PQQPFPQQPQQPQQPFPQPQ	1728	51	14
			QQPFPQQPQQPFPQPQQPIP	1729	81	10
			PQQPFLQPQQPFPQQPQQPF	1730	45	9
			QPFPQPQQPFPWQPQQPFPQ	1731	19	8
			QPQQPFPQSQQPQQPFPQPQ	1732	41	15
			PQTQQPQQPFPQSQQPQQPF	1733	36	9
3	QQPFPLQPQ or QQPFPQLQQ	1703	SEQIIPQQLQQPFPLQPQQP	1734	12	3
		1704	PFPQTQQPQQPFPQLQQPQQ	1735	25	13
4	QQPIPVQPQ or QQPIPQQPQ or QQPYPQQPQ	1705	QPQQPIPVQPQQSFPQQSQQ	1736	60	12
		1706	FPELQQPIPQQPQQPFPLQP	1737	22	7
		1707	FPQQPQQPYPQQPQQPFPQT	1738	19	4
			PRQPFPQQPQQPYPQQPQQP	1739	19	13
5	PQQPQQSFPQQQ or PQQPQQPFPQQQ	1708	PFPQPQQPQQSFPQQQQPLI	1740	29	15
			LPQPQQPQQSFPQQQRPFIQ	1741	37	12
		1709	QPQQPQQPFPQQQQPLIQPY	1742	49	17
6	QGSFQPSQQ	1710	QPQQQYPSGQGSFQPSQQNP	1743	54	5
			QYPSSQGSFQPSQQNPQAQG	1744	32	7
			GQGFFQPSQQNPQAQGSFQP	1745	7	3
7	PQQPFPQPQQ or PQQPFPQTQQ or	1711	PQTQQPQQPFPQPQQTFPQQ	1746	21	9
		1712	QQPQQPFPQPQLPFPQQSEQ	1747	15	2
			FPWQPQQPFPQTQQSFPLQP	1748	25	7
8	QQPQQPFPQ or QQPQQPYPQ	1713	PFPQTQQPQQPFPQLQQPQQ	1749	25	13
		1714	QQPLPQPQQPQQPFPQSQQP	1750	54	20
			PFPQLQQPQQPFPQPQQQLP	1751	36	5
			PRQPFPQQPQQPYPQQPQQP	1752	19	13
9	PFPQPQQPQ or PFPQSQQPQ or PFPQPQQAQ or QFPQTQQPQ	1715	PQQPFPQPQQPQQPFPQLQQ	1753	37	8
		1716	PQTQQPQQPFPQSQQPQQPF	1754	36	9
		1717	QPQQPFPQSQQPQQPFPQPQ	1755	41	15
		1718	PQQPQQPFPQPQQAQLPFPQ	1756	16	7
			HQPQQQFPQTQQPQQPFPQP	1757	30	14
10	PQQQFIQPQ	1719	FSQPQQPQQQFIQPQQPFPQ	1758	45	8
11	QQPQLPFPQ or LQPQQPFPQ	1720	PQPQQPQLPFPQQPQQPFPQ	1759	79	7
		1721	PQQQFLQPQQPFPQQPRQPY	1760	35	14
12			QTLPAMCNVYIPPHCSTTIA	1761	35	7
13			NPSQQQPQEQVPLVQEQQFQ	1762	5	16
14			HHFRSNSNHHFHSNNNQFYR	1763	14	3

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摘要(译)

本文公开的发明涉及可用于诊断，治疗和预防乳糜泻的方法中的表位。提供了包含至少一个表位的治疗组合物。

Figure 1