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(54) **CHIRAL INDOLE INTERMEDIATES AND THEIR FLUORESCENT CYANINE DYES CONTAINING FUNCTIONAL GROUPS**

CHIRALE INDOLE ALS ZWISCHENPRODUKTE UND DARAUS HERGESTELLTE
CYANINFLUORESCENZFARBSTOFFE MIT FUNKTIONELLEN GRUPPEN

INTERMEDIAIRES CHIRAUX D'INDOLE ET LEURS COLORANTS DE CYANINE FLUORESCENTS
CONTENANT DES GROUPES FONCTIONNELS

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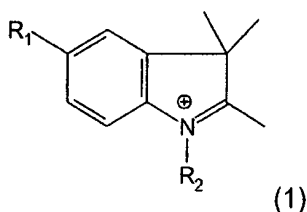
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Description

[0001] This invention relates to the functionalized cyanine dyes and more particularly, to the synthesis of chiral 3-substituted 2,3'-dimethyl-3H-indole and its derivatives as intermediates for the preparation of cyanine dyes, and the dyes so prepared.

[0002] Highly fluorescent carboxyl containing indocyanines are useful as labeling reagents for biological investigations. Functional groups on the dyes permit covalent binding to biomaterials and/or other non-biological materials for purpose of fluorescence detection, while water soluble arylsulfonate groups reduce dye-dye aggregation and enhance fluorescence brightness (see US patents by Waggoner et. al U.S. Patent Nos. 5,268,486; 5,486,616; 5,569,587; 5,852,191; 5,981,747; 5,986,093).

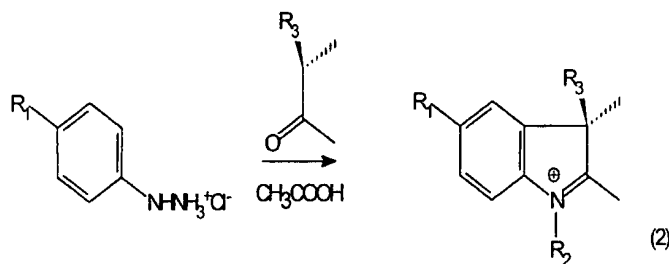
[0003] The carboxylic and arylsulfonate groups occupy key positions in the heteroaromatic bases such as R_1 and R_2 in (1), and thereby restrict the ability for dye modifications.



wherein R_1 is CH_2NH_2 , CH_2COOH , SO_3H or H and R_2 is $(\text{CH}_2)_m\text{NH}_2$, $(\text{CH}_2)_m\text{COOH}$, $(\text{CH}_2)_m\text{SO}_3\text{H}$ or $(\text{CH}_2)_m\text{OH}$, and m is an integer ranging from 1 to 6.

[0004] Thus, it is of interest to synthesize compounds with functional groups at different sites in the indole bases. These new intermediates will enable chemists to synthesize a wide range of cyanines.

[0005] Chiral 2,3-dimethyl-3H indoles (2) are expected to possess exceptional advantages as a precursor of a family of cyanine dyes for use as covalently attached fluorescent labels for biological research. Disclosed herein is a convenient synthesis of various 3-substituted 2-butanones and their conversion to cyanine dyes (Figure 1). These dyes are very similar to those described in the U.S. Patent No. 5,268,486 and other related publications of the inventor, such as, for example, listed herein which disclose luminescent mono- and polymethine- cyanine and related dyes such as merocyanine and styryl dyes which contain groups enabling these dyes to covalently attach to amine, hydroxyl, aldehyde and sulphhydryl groups on a target material. The disclosed compounds fluoresce in the green, orange, red and near infrared regions of the spectrum.



$R_1 = -\text{CH}_2\text{NH}_2$, $-\text{CH}_2\text{COOH}$, SO_3H , H, $-\text{NO}_2$ or X (Cl, Br, or I)

$R_2, R_3 = -(\text{CH}_2)_m\text{NH}_2$, $-(\text{CH}_2)_m\text{COOH}$, $-(\text{CH}_2)_m\text{SO}_3\text{H}$, $-(\text{CH}_2)_m\text{CH}$, $-\text{CH}_2\text{X}$ (Cl, Br, I), or H

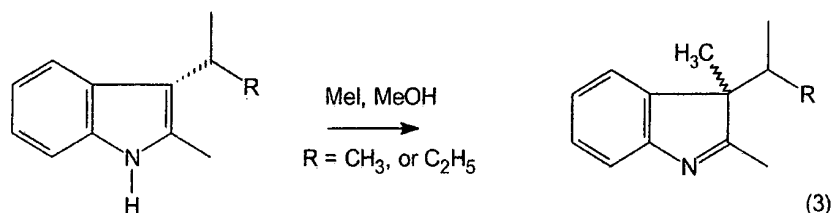
wherein m is an integer ranging from 1 to 6.

[0006] Rosenstock (Research Laboratories, The National Drug Company, NOTES (Dec. 1966 pp. 537-539) has described the synthesis of a plant growth hormone by means of Fischer indolization of levulinic acid phenylhydrazone. The indole is used for the development of various plant growth hormones.

[0007] WO 02/26891 discloses cyanine dyes incorporating similar structures.

[0008] A number of papers, such as Eggers et al (Angew. Chem. Int. Ed. Engl. (1997) 36, No. 8, 881-883), Eggers et al (Liebigs Ann. (1996), 979-983), Reichardt et al (Chem. Rev. 123, (1990), 565-581), Reichardt et al (Liebigs Ann.

(1995) 329-340), and U.S. Patent Nos. 6,190,641; 6,183,726; 6,180,087; 6,180,086 and 6,180,085 have described the synthesis of chiral indoles such as (3). Cyanine dyes described in these papers are water insoluble and have no functional groups that react with biological specimens. The chiral indoles are obtained by direct alkylation of 2,3-dimethylindole as shown below. This method has very limited applications. Attempts to alkylate 2,3-dimethylindole with various alkyl-



Brief Description of the Drawings

[0009]

20 FIGURE 1. A General scheme for the synthesis of 3-substituted-2-butanone and its cyanine dyes.

FIGURE 2: Computer generated energy minimized structure of 6-(1,2-dimethyl-7-sulfohydrobenzo[e]indolyl)hexanoic acid. The hexanoic acid chain is almost perpendicular to the benzindole ring. Such a configuration of the chain prevents dye-dye aggregation.

25 FIGURE 3: ^1H NMR spectra in CDCl_3 of 6-(2,3-dimethyl-3*H*-indol-3-yl)hexanoic acid and 5-(2,3-dimethyl-3*H*-indol-3-yl)pentan-1-ol show identical aromatic signals. Some of the methylene protons of the hexanoic acid and pentanol alkyl chains show high field shift, suggesting shielding of those protons due to ring current effect of the indole.

FIGURE 4: ^1H NMR spectra of Cy3.18 and NS Cy3 in D_2O . Some of the methylene protons of the hexanoic acid chain in NSCy3 show high field shift, suggesting shielding of those protons due to ring current effect of the indole.

30 FIGURE 5: Absorption spectra of non-sulfonated Cy3 isomers and their antibody conjugates.

FIGURE 6: Relative fluorescence intensities of non-sulfonated Cy3 isomers NCy3, Cy3.10 and Cy3.24. The absorbance for all dyes in methanol and PBS solution at 550 nm is 0.05 and the excitation wavelength is 514 nm

35 FIGURE 7: Absorption spectra of Cy7.18 free acid (dotted line) and its IgG conjugate (solid line) are shown in (A). Absorption spectra of NCy7 free acid (dotted line) and its IgG conjugates (solid line) are shown in (B). NSCy7 showed less dye-dye aggregation (indicated by low vibronic shoulder).

FIGURE 8: Antibody brightness of NCy3. The brightness is a product of number of dyes per protein, extinction coefficient of the dye and quantum yield of the dye/antibody conjugates.

40 FIGURE 9: is a graph showing the emission spectra of Cy3.18.OH and the disulfonated dye (NSCy3) in methanol and phosphate buffer solution when solutions of equal concentrations were excited at 514 nm. NSCy3 is 20-25% brighter than Cy3.18 in both solutions.

45 [0010] The Figures 2-9 emphasize improved properties of the cyanine dyes of the present invention due to the modifications at the 3, 3' positions of the indolenine. Specifically, Figure 2 shows a computer generated, energy minimized structure of 6-(1,2-dimethyl-7-sulfo-3-hydrobenz[e]indolyl)hexanoic acid. The hexanoic acid chain is perpendicular to the plane of benzindole ring. Some of alkyl protons of the hexanoic acid chain are over the plane of the ring. This is also substantiated by the NMR spectrum of 5-(2,3-dimethyl-3*H*-indol-3-yl)pentan-1-ol (Example 9) and 6-(2,3-dimethyl-3*H*-indol-3-yl)hexanoic acid (Example 3) in Figure 3. The alkyl chain protons of hexanoic acid and pentanol are shielded due to indole ring and therefore appear at high field (0.5-0.7 ppm). In Figure 4, the NMR of NSCy3 shows similar upfield protons (shown by arrow). The hexanoic acid protons for Cy3.18 appear at 1.5 -2.0 ppm. Such a configuration prevents dye-dye aggregation thereby increasing the fluorescence brightness of the dye in aqueous solutions. This is a major advantage of these new intermediates of this invention. Figure 5 shows the absorbance spectra of three non-sulfonated Cy3 dyes and their IgG conjugates. Cy3.10- IgG conjugates (Figure 5C) show considerable deviation from the base line suggesting significant dye-dye aggregation. The spectrum Cy3.24 and its IgG conjugates (Figure 5B) and NCy3 and its IgG conjugates (Figure 5A) are almost identical. But NCy3 is 20-25% brighter than Cy3.24 in aqueous solution. This is shown in Figure 6. The equal concentration of the dyes in methanol and PBS solutions were excited at 514 nm; fluorescence intensities of the dyes in methanol are almost the equal. Because of high solubility in methanol, no dye-dye aggregation is observed. However, in a phosphate buffer solution, NSCy3 is 20-25% brighter than Cy3.24. The reduction of dye-dye aggregation in an antibody conjugated dyes is also evident from Figure 7. The absorption spectra of com-

mercialized Cy7.18 and new NSCy7 are compared with their antibody conjugates. Cy7.18 aggregates more on antibody as seen from the increase in the vibronic shoulder.

[0011] Dyes according to the invention are as follows:

- 5 i) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium (Example 13);
 ii) 3-(5-carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2*H*-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium (Example 14);
 10 iii) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3*H*-indolium (Example 15);
 iv) 3-(5-carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium (Example 16);
 v) 3-(5-carboxypentyl)-3-methyl-2-((1*E*,3*E*,5*E*)-5-[3-methyl-5-sulfo-3-(4-sulfobutyl)-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium (Example 17).

15 **[0012]** The compounds or dyes of the present invention may be used to label, for example avidin, streptavidin, antibodies, DNA, RNA, nucleosides, nucleotides or lectins to detect, measure and/or quantify, for example, biotinylated materials, antigens, haptens, carbohydrate groups, DNA, RNA and complementary DNA or RNA, such as described therein.

20 **[0013]** A cyanine dye or its derivative of the present invention preferably absorbs maximally in the range of 400-950 nm, preferably in the range of 420-470 nm, 520-580 nm, 640-680 nm, 740-790 nm or 820-850 nm. In yet another embodiment, the present invention provides a method of fluorescent detection wherein the dye or derivative of the present invention is detected or when multiple fluorescent dyes are used, a dye or derivative of the present invention may be used in conjunction with dyes which do not either fluoresce at the same or similar pH conditions and/or at the wavelengths of the presently provided dyes or derivatives.

25 **[0014]** In a further embodiment, a compound of the invention may be used in a fluorescence method for the qualitative and/or quantitative detection of pH. The method comprises contacting or mixing the compound with a composition containing cells, tissues or biological fluid, and detecting the emitted fluorescence. The presence of a fluorescent signal is indicative of an acidic environment. In one embodiment, the method may be used for detecting intracellular environments, such as may be contained in subcellular compartments or structures. Compounds of the invention may be actively or passively absorbed by cells where they may be detected by fluorescence detection. One of ordinary skill will appreciate the variability of cell permeability properties of compounds according to the invention and will be able to routinely test for the same.

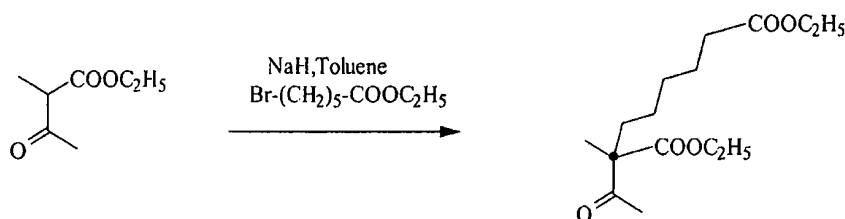
30 **[0015]** The methods according to the present invention may employ known devices for illumination and detection at separate defined wavelengths. Such devices include fluorescence spectrophotometers, fluorescence microscopes. Alternatively, changes in fluorescence intensity may be measured by means of a charge coupled device (CCD) imager (such as a scanning imager or an area imager) to image all of the wells of a microtitre plate.

[0016] The present invention is further described by the following examples.

40 EXAMPLES

1. Synthesis of diethyl 2-acetyl-2-methyloctane-1,8-dioate

45 **[0017]**



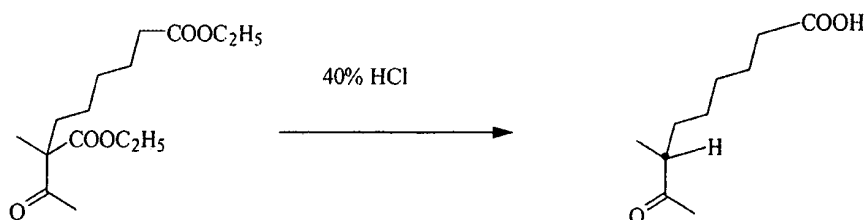
55 **[0018]** In a 1L three necked flask equipped with a mechanical stirrer and reflux condenser was placed dry toluene (300 ml). The system was flushed with argon and sodium hydride (Aldrich, 60% dispersion in mineral oil) (7.2 g, 0.18 mmol) was added. Ethyl- γ -methylacetoacetate (Aldrich, 21 g, 0.15 mole) was added with stirring over a 30 minutes. The

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resulting solution was heated under reflux for 2 hrs and cooled slightly. (Note: Reaction mixture becomes a thick paste and mechanical stirrer is essential). Ethyl 6-bromohexanoate (Aldrich, 33.5 g, 0.15 mol) was added (all at once) and the suspension was heated under reflux for 12 hrs, cooled, filtered and the solvent evaporated under pressure. The residue was distilled under vacuum to yield 20 g (46%) of colorless liquid, b.p. 110-115° C (0.1 mm). IR (neat): vcm^{-1} = 1735 (s) and 1713 (s). ^1H NMR in CDCl_3 δ , 4.1-4.2 (m, 4H, (O- CH_2) $_2$); 2.1 (t, J = 8.4Hz, 2H, CH_2COO -); 2.15(s, 3H, CH_3CO); 1.5-2.0 (m, 4H, (CH_2) $_2$); 1.2-1.4 (m, 13H, 2 CH_2 and three $-\text{CH}_3$).

2. Synthesis of 7-acetyloctanoic acid (or 7-methyl-8-oxononanoic acid)

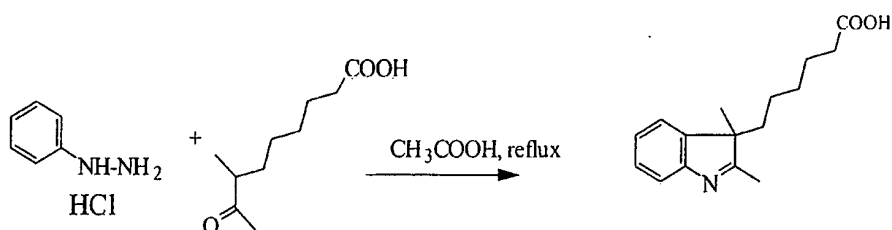
[0019]



[0020] A mixture of diester from Example 1 (15 g, 0.052 mol) in dilute hydrochloric acid (40-50%) (100 ml) was heated to reflux for 12 hrs. The reaction mixture was cooled and extracted with ethyl acetate (3 x 100 ml). The organic phase was dried over sodium sulfate, filtered, and ethyl acetate was removed. The residue was distilled under reduced pressure, to yield 7.7 g (80%), colorless liquid, b.p. 120-125 °C, IR (neat): vcm^{-1} = 1736 (s) and 1710 (s). ^1H NMR, CDCl_3 , δ , 2.45-2.55 (q, 1H, J = 6.6 Hz, 7-H); 2.35 (t, J = 7.3 Hz, 2H, $-\text{CH}_2-\text{COOH}$); 2.125 (s, 3H, COCH_3); 1.65-1.1.75 (m, 2H, $-\text{CH}_2$); 1.23-1.4 (m, 6H, ($-\text{CH}_2$) $_3$); 1.1 (d, J = 5.9Hz, 7- CH_3).

3. Synthesis of 6-(2,3-dimethyl-3H-indol-3-yl)hexanoic acid

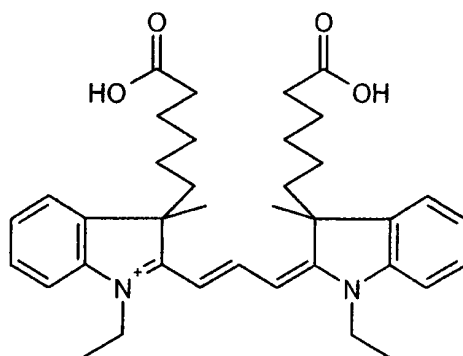
[0021]



[0022] To a stirred solution of phenylhydrazine hydrochloride (Aldrich, 7.2 g, 0.05 mol) in acetic acid (50 ml) was added 7-acetyloctanoic acid (11 g, 0.06 mol). The reaction mixture was heated under reflux for 4 hrs. The solution was cooled. (No clear precipitate was observed). Acetic acid was removed under reduced pressure. The resulting yellow liquid was chromatographed on a silica gel column using chloroform/methanol mixture as solvent. Pure 6-(2,3-dimethyl-3H-indol-3-yl)hexanoic acid (8.0 g, 61 %) was obtained as pale yellow oil which crystallized on standing, m.p. 115-118 °C; IR v cm^{-1} = 2927, 2860, 2526, and 1709. ^1H NMR, CDCl_3 , δ , 7.6 (d, 1H, J = 7 Hz, 4-H); 7.2-7.4 (m, 3H, aromatic protons); 2.25 (s, 3H, $-\text{CH}_3$); 2.1-2.2(t, $-\text{CH}_2\text{COOH}$); 1.72-1.94 (m 2H, $-\text{CH}_2$); 1.4-1.45 (m, 2H, $-\text{CH}_2$); 1.23-1.05 (m, singlet merged in m, 5H, $-\text{CH}_2$, $-\text{CH}_3$); 0.55-0.85 (m, 2H, $-\text{CH}_2$). TLC, R_f = 0.22 (silica gel, methanol-chloroform, 5:95).

4. Synthesis of 3-(5-carboxypentyl)-2-((1E,3E)-3-[3-(5-carboxypentyl)-1-ethyl-3-methyl-1,3-dihydro-2H-indol-2-ylidene]prop-1-enyl)-1-ethyl-3-methyl-3H-indolium iodide (NCy3) (not claimed)

[0023]



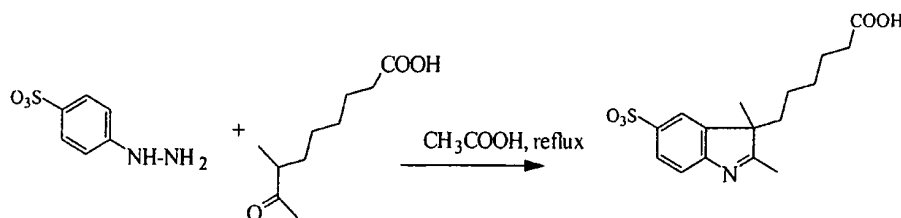
4.1 3-(5-Carboxypentyl)-1-ethyl-2,3-dimethyl-3H-indolium iodide

[0024] 6-(2,3-Dimethyl-3-hydroindole-3-yl)hexanoic acid (518 mg, 2 mmol) was suspended in acetonitrile (5 ml) and ethyl iodide (1.2g, 10 mmol) and the mixture was heated to reflux with stirring for 1 hr. More ethyl iodide (2 g) was added and heating continued for an additional 12 hrs. The mixture was then cooled, and acetonitrile and excess ethyl iodide were removed on a rotary evaporator. The sticky mass was washed several times with ice-cooled diethyl ether and dried to yield pink amorphous powder (0.45 g, 78%). The product was used for the next reaction without further purification.

[0025] 4.2 To a stirred solution of 6-(1ethyl-2,3-dimethyl-3-hydroindol-3-yl)hexanoic acid (288 mg, 0.95 mmol) in pyridine (10 ml) at 120 °C was added dropwise, triethyl orthoformate (100 mg, 67 mmol) over 30 minutes. After 2 hrs the reaction mixture was cooled and triturated with diethyl ether. The product was purified on a silica gel column using chloroform/methanol mixture as solvent. The major compound ($R_f = 0.45$, tlc, silica gel, methanol in chloroform 5%) was obtained as a pink solid (251 mg, 35%); $^1\text{H NMR}$, CDCl_3 , δ , 8.2 (t, H, $J = 7\text{Hz}$, $\beta\text{-H}$); 7.1-7.6 (m, 8H, aromatic); 6.4 (d, 2H, $J = 7\text{Hz}$, $\alpha\text{-H}$, $\alpha'\text{-H}$); 4.2 (m, 4H, 2- NCH_2); 2.2-2.3 (m, 4H, 2- CH_2COOH , 2- CH_2); 1.8 (s, 6H, ($-\text{CH}_3$) $_2$); 1.43 (t, 6H, $J = 6.5\text{Hz}$, 2- CH_3); 1.1-1.2 (m, 4H, 2- CH_2); 0.5-0.7 (m, 4H, 2- CH_2); λ_{max} : 550 nm, methanol.

5. Preparation of 6-(2,3-dimethyl-5-sulfo-3H-indol-3-yl)hexanoic acid

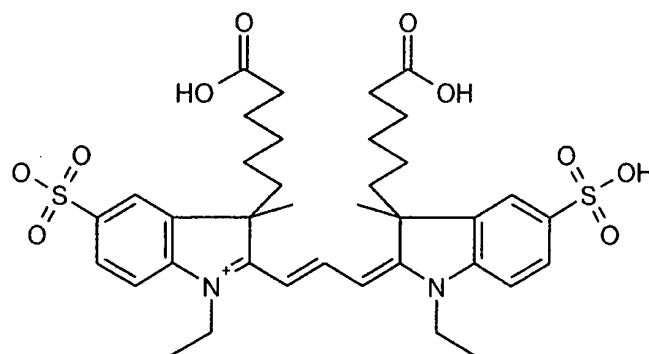
[0026]



[0027] To a stirred solution of 4-hydrazinobenzenesulfonic acid (Aldrich, 11.25 g, 0.06 mol) in acetic acid (50 ml) was added 7-acetyloctanoic acid (16.7 g, 0.09 mol). The reaction mixture was heated under reflux for 12 hrs. Acetic acid was removed under reduced pressure. The resulting solid was dissolved in methanol and reprecipitated with a saturated solution of potassium hydroxide in isopropanol. The solid was filtered, washed with isopropanol and dried, (8 g, 40%). The analytical sample was obtained by C18 reversed phase column chromatography using water/methanol mixture as solvent, m.p. 250 °C dec; IR $\nu_{\text{cm}^{-1}} = 2930, 2597, \text{ and } 1719$. $^1\text{H NMR}$, D_2O , δ , 7.8-7.9 (m, 2H, 4-H and 6-H of aromatic protons); 7.6 (d, $J = 7\text{Hz}$, 1 H, 7-H of aromatic); 2.2 (t, $J = 7\text{Hz}$, 2H, $-\text{CH}_2\text{-COOH}$); 1.9-2.1 (m, 2H, alkyl); 1.2-1.6 (a singlet merged in a multiplet, 7H, $-\text{CH}_3$, ($-\text{CH}_2$) $_2$); 0.6-0.9 (m, 2H, alkyl.).

6. Synthesis of 3-(5-carboxypentyl)-2-((1E,3E)-3-[3-(5-carboxypentyl)-1-ethyl-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]prop-1-enyl)-1-ethyl-3-methyl-3H-indolium-5-sulfonate (NSCy3) (not claimed)

[0028]

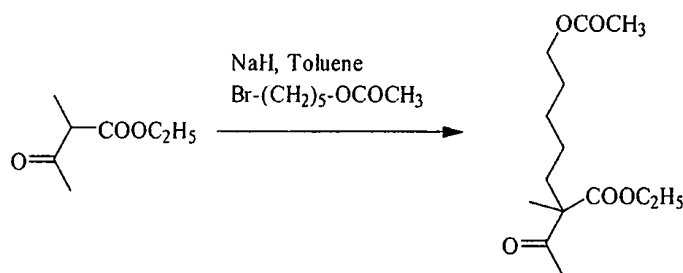


15 **[0029]** 3-(5-Carboxypentyl)1-ethyl-2,3-dimethyl-3*H*-indolium-5-sulfonate was synthesized following the procedure described in Example 4.1. 6-(2,3-Dimethyl-5-sulfo-3-hydroindole-3-yl)hexanoic acid (680 mg, 2 mmol) was suspended in acetonitrile (10 ml) and ethyl iodide (1.2g, 10 mmol) and the mixture was heated to reflux with stirring for 1hr. More ethyl iodide (2 g) was added and heating continued for additional 12 hrs. The mixture was then cooled and solvent removed on a rotary evaporator. The sticky mass was dissolved in methanol (5ml). A solution of potassium acetate in isopropanol was added until the reaction product was alkaline to pH. The solid obtained was filtered and washed several times with isopropanol to yield a gray amorphous powder (500 mg, 61 %). The product was used for the next reaction without further purification.

20 **[0030]** To a stirred solution of 3-(5-carboxypentyl)1-ethyl-2,3-dimethyl-3*H*-indolium-5-sulfonate (potassium salt) (400 mg, 0.98 mmol) in pyridine (10 ml) at 100 °C was added triethyl orthoformate (100 mg, 66 mmol) over 30 minutes. After 25 2 hours the reaction mixture was cooled and diluted with several volumes diethyl ether. A product separated as red powder from which supernatant fluid was removed by decantation. It was dissolved in methanol and reprecipitated with isopropanol containing potassium acetate. The crude dye was collected on a filter paper and dried (380 mg, 90%). It was purified by C18, column chromatography using water-methanol mixture for elution. A pure dye was obtained as pink solid (251mg), λ_{max} 555 nm; 1H NMR (D_2O) δ , 8.42 (t, 1 H, $J=13$ Hz, β -H of the bridge); 7.6-7.8 (m, 4 H, 4-H, 4'-H, 6-H, 6'-4); 7.2 (d, 2H, $J=7.7$ Hz, 7-H, 7'-H); 6.5 (d, 2H, $J=13$ Hz, α , α' -H); 4.1 (broad signal, 4H, (N-CH₂)₂); 1.8-2.2 (m, 8H, alkyl and CH₂COOH); 1.2-1.7 (m, 24H, 6-CH₂, 4-CH₃); 0.45-0.8 (m, 4H, alkyl).

7. Synthesis of ethyl 2-acetyl-7-acetyloxy-2-methylheptanoate

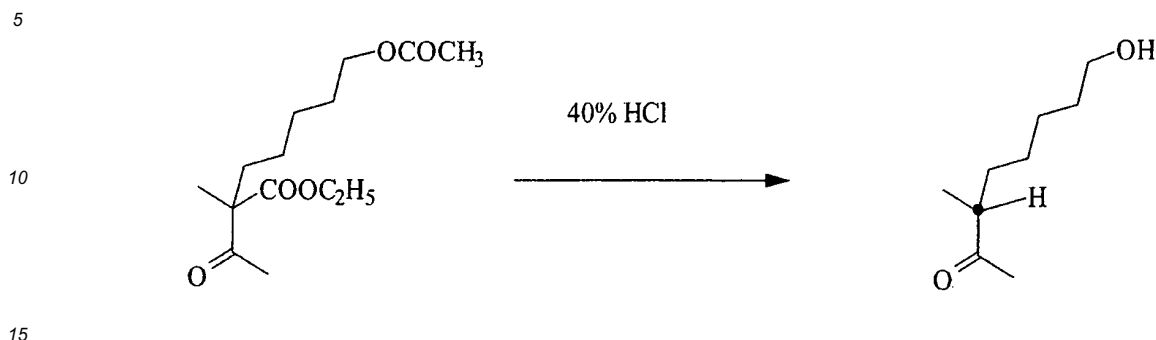
35 **[0031]**



40 **[0032]** In a 1L three necked flask equipped with a mechanical stirrer and reflux condenser was placed dry toluene (300 ml). The system was flushed with argon and sodium hydride (Aldrich, 60% dispersion in mineral oil) (7.2 g, 1.2 eq.) was added. Ethyl- α -methylacetoacetate (Aldrich, 21 g, 0.15 mole) was added with stirring over a 30 minutes. The resulting solution was heated under reflux for 2 hours and cooled slightly. (Note: Reaction mixture becomes a thick paste and a mechanical stirrer is essential). 5-Bromopentylacetate (Aldrich, 31.5 g, 15 mol) was added (all at once) and the suspension was heated under reflux for 12 hr, cooled, filtered and the solvent evaporated under reduced pressure. The residue was distilled under vacuum to yield 20 g (45%) of colorless liquid, b.p 90-92 (0.5 mm). 1H NMR in CDCl₃ δ , 4.1-4.2 (m, 2H, O-CH₂CH₃); 3.5 (t, $J = 7$ Hz, 2H, CH₂OCO-); 2.2 (s, 3H, CH₃CO); 2.1 (s, 3H, -COCH₃); 1.5-2.0 (m, 10H, 4-(CH₂)₂, C-CH₃, OCH₂CH₃).

8. Synthesis of 8-hydroxy-3-methyloctan-2-one or 6-acetyl-octanol

[0033]

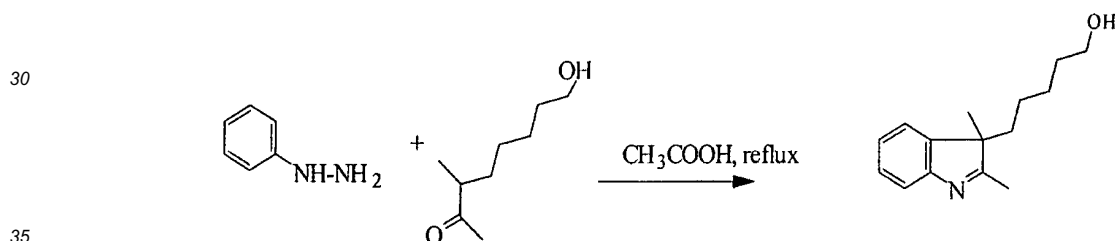


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[0034] A mixture of diester (15 g, 0.06 mol) in a dilute hydrochloric acid (40-50%) (100 ml) was heated to reflux for 12 hrs. The reaction mixture was cooled and extracted with ethyl acetate (3 x 100 ml). The organic phase was dried over sodium sulfate, filtered, and ethylacetate was removed. The residue was distilled in vacuum, to yield 7.19 g (80%) colorless liquid, b.p. 90-97 °C at 0.7 mm , IR (neat): ν cm^{-1} = 1736; ^1H NMR, CDCl_3 , δ , 3.55 (t, 2H, J = 6.6 Hz, $-\text{CH}_2\text{OH}$) 2.45 (m, 1H, 7-H); 2.15 (s, 3H, COCH_3); 1.65-1.82 (m, 4H, $(-\text{CH}_2)_2$); 1.2-1.55 (m, 4H, $(-\text{CH}_2)_2$); 1.1 (d, J = 7.3 Hz, $-\text{CH}-\text{CH}_3$).

9. Synthesis of 5-(2,3-dimethyl-3H-indol-3-yl)pentan-1-ol

25 [0035]



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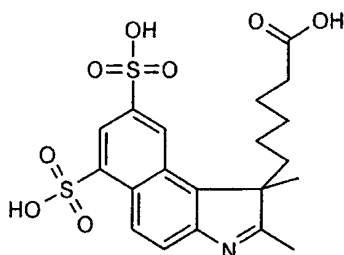
[0036] To a stirred solution of phenylhydrazine hydrochloride (Aldrich, 7.2 g, 0.05mol) in acetic acid (50 ml) was added 8-hydroxy-3-methyloctan-2-one (12 g, 0.07mol). The reaction mixture was heated under reflux for 4 hrs. The solution was cooled. (No clear precipitate was observed). Acetic acid was removed under reduced pressure. The resulting yellow liquid was chromatographed on a silica gel column using chloroform:methanol mixture as solvent. Pure 6-(2,3-dimethyl-3-hydroindol-3-yl)pentanol (8.0 g, 60%) was obtained as pale yellow oil; ^1H NMR, CDCl_3 , δ , 7.8 (d, J = 7 Hz, 4-H aromatic); 7.2-7.6 (m, 3H, aromatic protons); 3.5 (t, J = 6.5 Hz, $-\text{CH}_2\text{OH}$); 2.6 (s, 3H, 2- CH_3 of indole); 1.8-2.12 (m, 2H, alkyl), 1.3-1.7 (singlet merged in multiplets, 7H, 2 alkyl and $-\text{CH}_3$ of indole); 0.6-0.9 (m, 2H, alkyl).

10. Synthesis of 6-(1,2-dimethyl-6,8-disulfo-1H-benzo[e]indol-1-yl)hexanoic acid

[0037]

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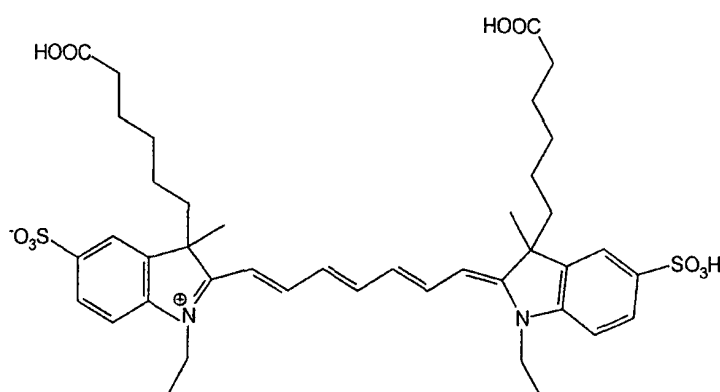
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[0038] (5,7-Disulfo-2-naphthyl)hydrazinium chloride (2.0g), 7-methyl-8-oxononanoic acid (1.5g) and acetic acid (15ml) were heated from 80-140°C for a total of 24hrs. After evaporation of the solvent under vacuum, the residue was triturated with 2-propanol to give a pink solid. This solid was collected by filtration, washed with 2-propanol, then excess diethyl ether and dried under vacuum over phosphorus pentoxide. Yield of crude product = 1.96g. This was purified as required by preparative HPLC (RPC18. Water/MeCN/TFA) to give pure product. ¹H-nmr (D₂O) δ 0.25-0.4 (1H, broad m), 0.5-0.65 (1H, broad m), 1.00 (2H, m), 1.25 (2H, m), 1.78 (3H, s), 2.06 (2H, t), 2.35-2.5 (1H, broad m), 2.65-2.8 (1H, broad m), 2.90 (3H, s), 8.05 (1H, d), 8.58 (1H, d), 8.70 (1H, d) and (8.97 (1 H, d). MS (LCMS) MH+ 470. Acc. Mass : MH+ 470.0931 (-2.6ppm for C₂₀H₂₄NO₈S₂).

11. Synthesis of 3-(5-carboxypentyl)-2-((1E,3E,5E,7E)-7-[3-(5-carboxypentyl)-1-ethyl-3-methyl-5sulfo-1,3-dihydro-2H-indol-2-ylidene]hepta-1,3,5-trienyl)-1-ethyl-3-methyl-3H-indolium-5-sulfonate(NSCy7) (not claimed)

[0039]



[0040] Glutaconaldehyde dianil hydrochloride (143 mg, 0.5 mmol) was dissolved in a heated mixture of acetic anhydride (4 ml) and pyridine (1 ml). The intermediate 6-(1-ethyl-2,3-dimethyl-5-sulfo-3-hydroindol-3-yl)hexanoic acid (368 mg, 1 mmol) was added and mixture was heated to reflux for an additional 10 min. and then cooled. The dye was precipitated with diethyl ether. The supernatant liquid was separated. Solid was redissolved in a minimum volume of methanol and reprecipitated with isopropanol. The purple solid was collected on a filter paper and dried (160 mg, yield 18%). The pyridinium salt was converted into its potassium salt and purified by C18 column chromatography using water-methanol mixture as eluent. λ max = 750 nm. ¹H NMR (D₂O) δ, 7.7 (m, 6H, 4-H, 4'-H, 6-H, 6'-H and γγ' protons of the bridge); 7.4 (t, 1H, J = 13 Hz, δ-proton of the bridge); 7.2 (d, 2H, J = 7.7 Hz, 7-H, 7'-H); 6.35 (t, 2H, J = 13 Hz, β, and β' protons of the bridge, 6.15 (d, 2H, J = 7 Hz, α, α'-H); 4.1 (broad signal, 4H, (N-CH₂)₂); 1.8-2.2 (m, 8H, alkyl and CH₂COOH); 1.2-1.7 (m, 24H, 6 -CH₂, 4 -CH₃); 0.45-0.8 (m, 4H, alkyl). Rf = 0.26 (RP-C18, water/methanol-7.5:2.5).

12. Succinimidyl Esters of Carboxyalkyl Cyanine Dyes

[0041] The following general procedure was used to prepare succinimidyl esters of Cy3.24.OH, Cy3.10.OH, NCy3, NSCy3, Cy7.18.OH, and NSCy7. A procedure for making succinimidyl active ester using disuccinimidyl carbonate (DSC) has been described by Ogura et. al. Tetrahedron Lett. 4745-4746(1979). In a typical experiment carboxyalkyl indocyanine was dissolved in mixture of dry DMF (2 mL/100 mg dye) and dry pyridine (0.1 mL/100 mg dye). Disuccinimidyl carbonate

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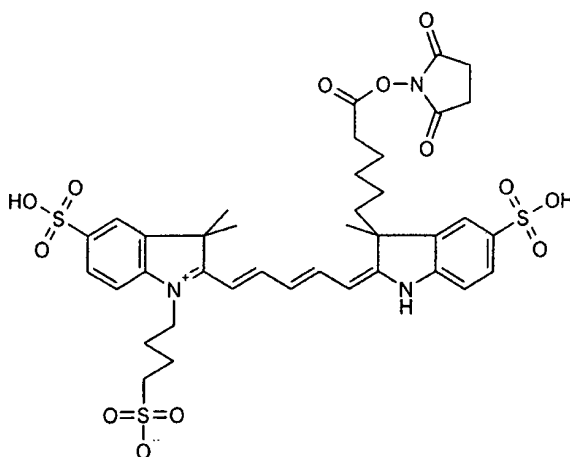
1.7-1.9 (4H, broad m), 2.60 (2H, t), 4.14 (2H, t), 6.36 (1 H, t), 6.56 (1H, d), 7.21 (1H, m), 7.3-7.5 (5H, m), 7.65 (1H, m), 7.83 (1H, s), 8.39 (1H, t), 8.83 (1H, t) and 11.7 (1H, d). MS (MALDI-TOF) M+ 504.

13.4 2-((1E,3E,5E)-5-[3-(5-Carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0047] 4-{2-[(1E,3E)-4-anilinobuta-1,3-dienyl]-3,3-dimethyl-5-sulfo-3H-indolium-1-yl}butane-1-sulfonate (250mg) and 6-(2,3-dimethyl-5-sulfo-3H-indol-3-yl)hexanoic acid (312mg) were mixed in pyridine (9ml): acetic acid (9ml) : acetic anhydride (2ml) and stirred at ambient temperature overnight. After evaporation of solvent under vacuum, the residue was then purified twice by preparative HPLC (RPC18. Water+0.1%TFA→MeCN+0.1%TFA gradient). The appropriate fractions were pooled and evaporated under vacuum. The residue was redissolved in water and freeze-dried to give the purified dye (135mg). Observed pKa in aqueous phosphate buffers (via fluorescence): 7.0. UV/Vis (Water+0.1%TFA): 645nm ($\epsilon = 2.0 \times 10^5 \text{ l mol}^{-1} \text{ cm}^{-1}$). MS (MALDI-TOF) MH+ 751. ¹H-nmr (D₂O) δ 0.5-0.65 (1H, broad), 0.75-0.9 (1H, broad), 1.1-1.2 (2H, m), 1.35 (2H, m), 1.5 (3H, s), 1.65 (6H, 2xs), 1.75-2.05 (6H, m), 2.1 (2H, t), 2.95 (2H, t), 4.05 (2H, broad t), 6.0 (1 H, d), 6.2 (1 H, d), 6.5 (1H, t), 7.2 (1 H, d), 7.3 (1 H, d) and 7.1-8.1 (6H, m).

13.5 2-((1E,3E,5E)-5-[3-(5-Carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium, N-hydroxysuccinimidyl ester

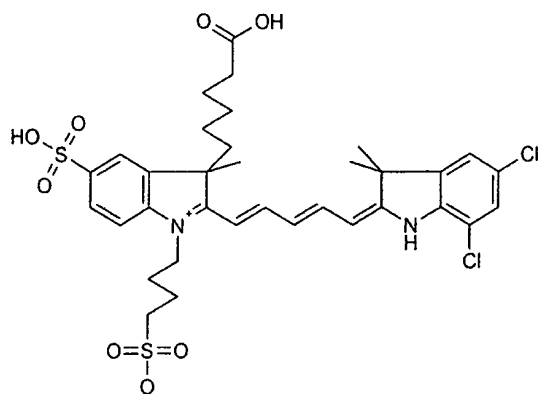
[0048]



[0049] Carboxy dye (30mg) was dissolved in anhydrous DMF (300 μ l); to this were added O-(N-succinimidyl)-N,N,N',N'-tetramethyleuronium hexafluorophosphate (HSPyU, 60mg) and diisopropylethylamine (42 μ l). The resulting solution was mixed for 2hrs, whereupon TLC (RPC18. Water/MeCN/AcOH) revealed complete reaction. The reaction mix was evaporated under vacuum and the residue was purified by preparative HPLC (Water+0.1 % AcOH → MeCN+0.1% AcOH gradient). Fractions containing the principal dye peak were pooled and evaporated under vacuum; the residue was redissolved in water and freeze-dried. UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 646nm. MS (MALDI-TOF) MH+ ~850.

14. Synthesis and activation of 3-(5-carboxypentyl)-2-((1E,3E,5E)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0050]



14.1 4-[3-(5-Carboxypentyl)-2,3-dimethyl-5-sulfo-3H-indolium-1-yl]butane-1-sulfonate

[0051] 6-(2,3-Dimethyl-5-sulfo-3H-indol-3-yl)hexanoic acid (340mg) and sodium acetate (100mg) were dissolved in methanol (5ml), then the solvent was evaporated under vacuum. The residue was treated with 1,4-butanediol (680mg) and 1,2-dichlorobenzene (5ml); this mixture was heated at 150°C for 18hrs under nitrogen. A further aliquot of 1,4-butanediol (500mg) was added and heating continued for 4hrs. After cooling, the solid product was collected by vacuum filtration, washed with dichlorobenzene and diethyl ether and dried. Crude yield 557mg. After purification by preparative HPLC (RPC18. Water+0.1%TFA → MeCN+0.1%TFA gradient), the desired product was isolated. ¹H-nmr (DMSO) δ 0.6-0.8 (2H, m), 1.05-1.12 (2H, m), 1.43 (2H, app. quin), 1.64 (3H, s), 1.95 (2H, aqq. quin), 2.05-2.42 (4H, m + 2H, t), 3.01 (2H, t), 4.60 (2H, t), 7.97 (1 H, d), 8.09 (1H, dd) and 8.13 (1 H, d).
LCMS : MH+ 476

14.2 4-[2-[(1E,3E)-4-Anilinobuta-1,3-dienyl]-3-(5-carboxypentyl)-3-methyl-5-sulfo-3H-indolium-1-yl]butane-1-sulfonate

[0052] 4-[3-(5-Carboxypentyl)-2,3-dimethyl-5-sulfo-3H-indolium-1-yl]butane-1-sulfonate (410mg), malonaldehyde bis(phenylimine) monohydrochloride (500mg), triethylamine (0.3ml) and acetic acid (3ml) were heated at 120°C for 18hrs to give a dark brown-red solution. The solvent was evaporated under vacuum and the residue semi-purified by HPLC (RPC18. Water+0.1%TFA→MeCN+0.1%TFA gradient). The product was then used directly. UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 523nm. MS (MALDI-TOF) MH+ 605.

14.3 5,7-Dichloro-2,3,3-trimethyl-3H-indole

[0053] 2,4-Dichlorophenylhydrazine.HCl (3.0g), 3-methyl-2-butanone (3.0ml) and acetic acid (25ml) were mixed and heated at 120°C for 3hrs. A dark orange solution resulted. The solvent was evaporated under vacuum; the residue was partitioned between ethyl acetate and aqueous sodium hydrogen carbonate solution. The organic phase was collected, dried (MgSO₄), filtered and the solvent evaporated to give a dark oil. Purification by flash chromatography (silica: ethyl acetate/hexane) gave the product as a light orange oil, which solidified on standing. ¹H-nmr (CDCl₃) δ 1.31 (6H, s), 2.32 (3H, s), 7.15 (1H, d), 7.26 (1H, s) and 7.32 (1H, d).

14.4 3-(5-carboxypentyl)-2-[(1E,3E,5E)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl]-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0054] 4-[2-[(1E,3E)-4-Anilinobuta-1,3-dienyl]-3-(5-carboxypentyl)-3-methyl-5-sulfo-3H-indolium-1-yl]butane-1-sulfonate (250mg) and 5,7-dichloro-2,3,3-trimethyl-3H-indole (230mg) were mixed with acetic anhydride (1.0ml) and DMF (10.0ml). The mixture was stirred under nitrogen for 18hrs, then at 50°C for 24hrs. The solvent was then evaporated under vacuum; the residue triturated with ether, dried and purified by preparative HPLC (RPC18. Water+0.1%TFA→MeCN+0.1%TFA gradient). The appropriate fractions were pooled and evaporated under vacuum to give the product dye, 84mg. Observed pKa in aqueous phosphate buffers (via fluorescence): 6.05 UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 646nm. MS (MALDI-TOF) MH+740.

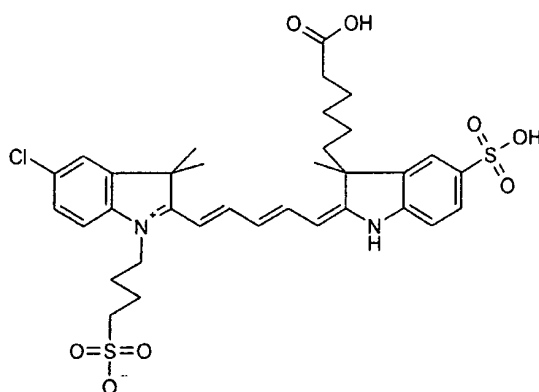
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14.5 3-(5-carboxypentyl)-2-[(1E,3E,5E)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro 2H-indol-2-ylidene)penta-1,3-dienyl]-3-methyl-5-sulfo-1-(4-sulfo-butyl)-3H-indolium, N-hydroxysuccinimidyl ester

[0055] Carboxy dye (24mg) was dissolved in anhydrous DMF (2ml) and evaporated under vacuum to ensure dryness. O-(N-Succinimidyl)-N,N,N',N'-tetramethyluronium tetrafluoroborate (TSTU, 30mg) was added, along with diisopropyl-ethylamine (25 μ l) and DMF (2ml). The red solution was allowed to stand for 1hr, whereupon TLC (RPC18. Water/MeCN/AcOH) revealed complete reaction. The reaction was quenched by addition of acetic acid (50 μ l) and the solvent evaporated under vacuum. The residue was purified by preparative HPLC (Water+0.1% AcOH \rightarrow MeCN+0.1 % AcOH gradient). Fractions containing the principal dye peak were pooled and evaporated under vacuum; the residue was redissolved in water and freeze-dried. UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 646nm. MS (MALDI-TOF) MH+ 837.

15. Synthesis and activation of 2-[(1E,3E,5E)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2ylidene]penta-1,3-dienyl]-5-chloro-3,3-dimethyl-1-(4-sulfo-butyl)-3H-indolium

[0056]



15.1 5-Chloro-2,3,3-trimethyl-3H-indole

[0057] 4-Chlorophenylhydrazine.HCl (5.4g), 3-methyl-2-butanone (6.4ml) and acetic acid (70ml) were mixed and heated to 80°C initially, to give a solution. The temperature was then raised to 120°C over 2hrs, during which time a solid separated. TLC analysis (RPC18. Water/MeCN/TFA) indicated consumption of the hydrazine starting material and generation of a single main product. After evaporation of the solvent under vacuum, the residue was partitioned between ethyl acetate and aqueous sodium hydrogen carbonate solution. The organic phase was collected, dried (MgSO₄), filtered and the solvent evaporated to give the crude indolenine. Purification by flash chromatography (silica. Ethyl acetate/hexane) gave 4.66g (80%) of pure product. MS (MALDI-TOF) M+ 193, 195. ¹H-nmr (CDCl₃) δ 1.30 (6H, s), 2.27 (3H, s), 7.23-7.29 (2H, m) and 7.44 (1H, d).

15.2 4-(5-Chloro-2,3,3-trimethyl-5-sulfo-3H-indolium-1-yl)butane-1-sulfonate

[0058] 5-Chloro-2,3,3-trimethyl-3H-indole (1.94g) and 1,4-butanedisulfone (5.0g) were mixed in 1,2-dichlorobenzene (15ml). The resulting solution was heated under nitrogen to 140°C for 18hrs, during which time a solid separated. After cooling to ambient temperature, this solid was collected by filtration, washed with dichlorobenzene followed by excess diethyl ether and then dried under vacuum to give the desired product. Yield 2.85g (86%). MS (MALDI-TOF) MH+ 330, 332. ¹H-nmr (CD₃OD) δ 1.61 (6H, s), 1.95-1.98 (2H, m), 2.08-2.18 (2H, m), 2.88 (2H, t), 4.53 (2H, t), 7.65 (1H, dd), 7.86 (1H, d) and 7.93 (1H, d)

15.3 4-[2-[(1E,3E)-4-Anilinobuta-1,3-dienyl]-5-chloro-3,3-dimethyl-3H-indolium-1-yl]butane-1-sulfonate

[0059] 4-(5-Chloro-2,3,3-trimethyl-5-sulfo-3H-indolium-1-yl)butane-1-sulfonate (0.99g), malonaldehyde bis(phenylimine).HCl (1.55g), triethylamine (1.0ml) and acetic acid (10.0ml) were mixed and heated at 120°C for 18hrs to give a dark purple solution. After evaporation of solvent, the crude reaction product was purified by flash chromatography (silica: MeOH/DCM) to give 1.12g of pure product. UV/Vis (EtOH): 525nm. MS (MALDI-TOF) M+ 458, 460.

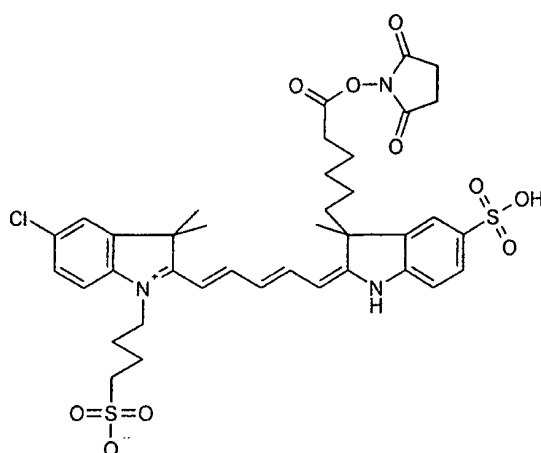
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15.4 2-((1E,3E,5E)-5-[3-(5-Carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3H-indolium

[0060] 4-{2-[(1E,3E)-4-Anilinobuta-1,3-dienyl]-5-chloro-3,3-dimethyl-3H-indolium-1-yl}butane-1-sulfonate (50mg), 6-(2,3-dimethyl-5-sulfo-3H-indol-3-yl)hexanoic acid (50mg), pyridine (2.25ml), acetic acid (2.25ml) and acetic anhydride (0.50ml) were mixed and incubated at 90°C for 1 hr, giving a deep blue solution. The solvent was then evaporated under vacuum and the residue triturated with diethyl ether. Purification by preparative HPLC (RPC18. Water/MeCN/TFA) yielded two blue components, the first-eluting component having the desired molecular mass and pH-sensitivity. Fractions containing this component were pooled and evaporated under vacuum to give the product dye, 16mg. Observed pKa in aqueous phosphate buffers (via fluorescence): 7.44. UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 645nm. MS (MALDI-TOF) M+ 704, 706.

15.5 2-((1E,3E,5E)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3H-indolium, N-hydroxysuccinimidyl ester

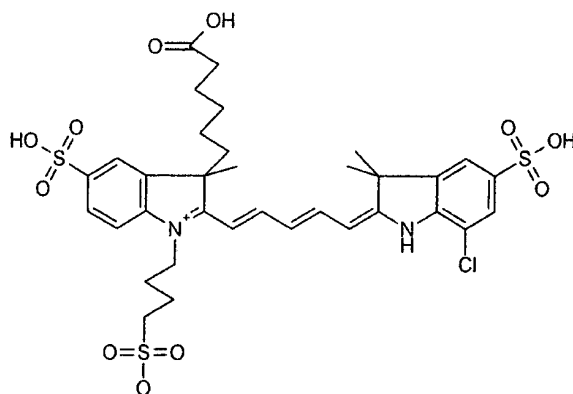
[0061]



[0062] The carboxy dye (13.5mg) was dissolved in anhydrous DMF (2ml) and evaporated under vacuum to ensure dryness. 0-(N-Succinimidyl)-N,N,N',N'-tetramethyluronium tetrafluoroborate (TSTU, 30mg) was added, together with diisopropylethylamine (25µl) and DMF (2ml). The orange solution was allowed to stand for 1hr, whereupon TLC (RPC18. Water/MeCN/AcOH) revealed complete reaction. The reaction was quenched by addition of acetic acid (50µl) and the solvent evaporated under vacuum. The residue was purified by preparative HPLC (Water+0.1% AcOH → MeCN+0.1% AcOH gradient). Fractions containing the principal dye peak were pooled and evaporated under vacuum; the residue was redissolved in water and freeze-dried. UV/Vis (Water, 50: MeCN, 50: TFA, 0.1) 645nm. MS (MALDI-TOF) M+ 801, 803.

16. Synthesis and activation of 3-(5-carboxypentyl)-2-(1E,3E,5E)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0063]



16.1 7-Chloro-2,3,3-trimethyl-3H-indole-5-sulfonic acid

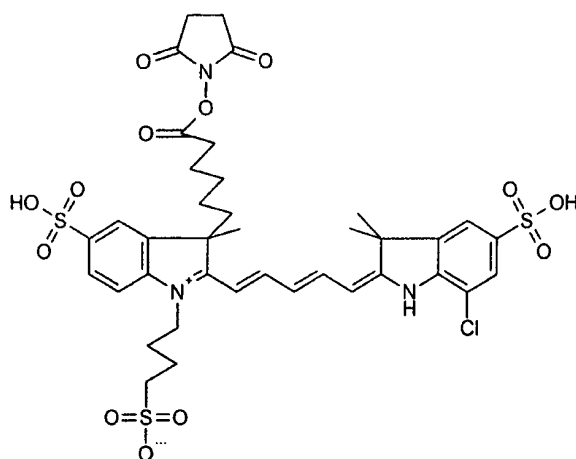
[0064] 2-Chloroaniline-4-sulfonic acid (2.5 g, 9.2 mmol) was dissolved in HCl (conc., 40 ml) and cooled to 0 °C. A solution of sodium nitrite (1.26 g, 18.3 mmol) in H₂O (10 ml) was transferred to a dropping funnel and added dropwise to the reaction vessel over 2 h. After stirring for 2 h at 0 °C, stannous chloride (8.3 g, 36.9 mmol; dissolved in 10 ml conc. HCl) was added dropwise over 2 h. The reaction mixture was then allowed to warm to room temperature and stirred overnight. The reaction mixture was then filtered and the filtrate discarded. The isolated solid was then transferred to a round-bottomed flask and treated with acetic acid (15 ml), potassium acetate (3 g) and 3-methyl-2-butanone (3 ml). After heating the vessel at 140 °C for 4 h, the reaction mixture was concentrated *in vacuo* and the resultant gum purified by RP-HPLC (Phenomenex Synergi; 10u hydro-RP 80, 250 x 21.20 mm; MeCN:H₂O; 0-100% MeCN 30 min; 10 ml/min, RT = 14 min) to isolate the desired product (550 mg; 2 mmol, 22%). MS (MALDI-TOF) MH⁺ 273.

16.23 (5-Carboxypentyl)-2-[(1E,3E,5E)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl]-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0065] To a solution of 7-chloro-2,3,3-trimethyl-3H-indole-5-sulfonic acid (50 mg; 0.18 mmol) in acetic acid/pyridine/acetic anhydride (4.5:4.5:1; 10 ml) was added (2E)-3-(5-carboxypentyl)-7-chloro-3-methyl-1-pentyl-2-[(2E,4E)-4-(phenylimino)but-2-enylidene]indoline-5-sulfonate (110 mg; 0.18 mmol). The reaction mixture was heated at 60 °C for 4 h then concentrated *in vacuo* to yield the crude dye. The resultant gum was purified by prep RP-HPLC (Phenomenex Synergi; 10u hydro-RP 80, 250 x 21.20 mm; MeCN:H₂O; 0-100% MeCN 30 min; 10 ml/min, RT = 16 min) to yield the desired product (10 mg; 0.01 mmol; 7%). MS (MALDI-TOF) MH⁺ 783

16.33 (5-Carboxypentyl)-2-[(1E,3E,5E)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl]-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium, N-hydroxysuccinimidyl ester

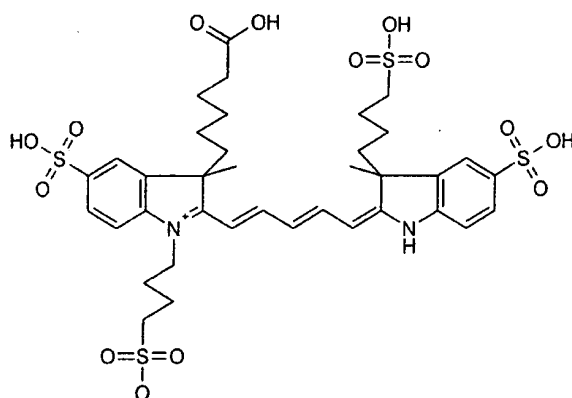
[0066]



20 **[0067]** To a stirred solution of 6-[(2E)-2-[(2E,4E)-5-(7-chloro-3,3-dimethyl-5-sulfo-3H-indol-2-yl)penta-2,4-dienylidene]-3-methyl-5-sulfo-1-(4-sulfobutyl)-2,3-dihydro-1H-indol-3-yl]hexanoic acid (9 mg; 0.01 mmol) in DMF (3 ml) was added DIPEA (2 ml) followed by O-(N-succinimidyl-N,N,N',N'-bis-tetramethylene)uronium hexafluorophosphate (14 mg; 0.05 mmol). After 4 h, the reaction mixture was concentrated *in vacuo* and the resultant gum purified by RP-HPLC (Phenomenex Synergi; 10u hydro-RP 80, 250 x 21.20 mm; MeCN:H₂O; 0-100% MeCN 30 min; 10 ml/min, RT = 17 min) to isolate the product (6 mg; 0.007 mmol; 53%). Observed pKa in aqueous phosphate buffers (via fluorescence): 6.2. MS (MALDI-TOF) M+Na⁺ 902

17. Synthesis of 3-(5-carboxypentyl)-3-methyl-2-((1E,3E,5E)-5-[3-methyl-5-sulfo-3-(4-sulfobutyl)-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-5-sulfo-1-(4-sulfobutyl)-3H-indolium

30 **[0068]**



17.1 5-Methyl-6-oxoheptane-1-sulfonic acid

50 **[0069]** Ethyl 2-methylacetoacetate (43.2g) in N,N-dimethylformamide (25ml) was added to a suspension of sodium hydride (12.0g of 60% NaH in mineral oil) in N,N-dimethylformamide (100ml), dropwise with ice-bath cooling. This mixture was allowed to warm to ambient temperature for 30mins before re-cooling. 1,4-butanediol (40.8g) in N,N-dimethylformamide (25ml) was then added dropwise. The final mixture was heated at 50 °C for 18hrs, then quenched with 50ml of water. The solvent was evaporated under vacuum; the residue was partitioned between water and diethyl ether. The aqueous layer was collected, washed with fresh diethyl ether and evaporated under vacuum; final drying was under high vacuum over phosphorus pentoxide. A yield of 103g of 5-(ethoxycarbonyl)-5-methyl-6-oxoheptane-1-sulfonic acid was obtained.

55 **[0070]** This intermediate was dissolved in concentrated hydrochloric acid (200ml) and reacted at 90 °C for 3hrs, then

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the solvent was evaporated under vacuum. The residue was purified by flash chromatography (silica. Dichloromethane → ethanol). Fractions containing the product were pooled and evaporated under vacuum to give the title compound, 49.6g. ¹H-nmr (D₂O) δ 1.05 (3H, d), 1.3-1.8 (6H, m), 2.20 (3H, s), 2.65 (1 H, m), and 2.90 (2H, m)

5 17.2 2,3-Dimethyl-3-(4-sulfobutyl)-3H-indole-5-sulfonic acid

[0071] 4-Hydrazinobenzenesulfonic acid (1.88g), 5-methyl-6-oxoheptanesulfonic acid (2.5g) and acetic acid (50ml) were mixed and heated under reflux for 6hrs. The solvent was evaporated under vacuum, then the residue was triturated with 2-propanol to yield the crude product as a yellow solid. This was purified by HPLC as required (RPC₁₈. Water + 0.1 % TFA). ¹H-nmr (D₂O) δ 0.8-1.0 (2H, m), 1.55-1.65 (5H, =3H, s + 2H, m), 2.16 (1H, ddd), 2.29 (1 H, ddd), 2.75 (2H, m), 2.81 (partially d-exchanged methyl singlet), 7.71 (1 H, d), 7.94 (1 H, d) and 8.01 (1 H, d). UV/Vis (Water+0.1%TFA): 269, 229nm. MS (LCMS) : MH⁺ 362. Acc. Mass: Found, 362.0729. MH⁺ = C₁₄H₂₀NO₆S₂ requires 362.0732 (-0.8ppm).

15 17.3 3-(5-Carboxypentyl)-3-methyl-2-((1E,3E,5E)-5-[3-methyl-5-sulfo-3-(4-sulfobutyl)-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-5-sulfo-1-(4-sulfobutyl)-3H-indolium

[0072] 4-{2-[(1E,3E)-4-Anilinobuta-1,3-dienyl]-3-(5-carboxypentyl)-3-methyl-5-sulfo-3H-indolium-1-yl}butane-1-sulfonate (made as in Example 14.2, 50mg) and 2,3-dimethyl-3-(4-sulfobutyl)-3H-indole-5-sulfonic acid (50mg) were mixed in pyridine (2.25ml), acetic acid (2.25ml) and acetic anhydride (0.50ml) and stirred at ambient temperature under nitrogen for 18hrs, then for 2hrs at 60°C. The resulting green-blue solution was evaporated under vacuum. The residue was purified by HPLC (RPC₁₈. Water / MeCN / TFA gradient). Fractions containing the principal dye peak were collected, pooled and evaporated under vacuum to give the title dye, 22mg. UV/Vis (Water+0.1%TFA): 651 nm. MS (MALDI-TOF): M+ 872.

25 18. General protein labeling procedure

[0073] The following protein labeling procedure was followed to label sheep IgG with NHS ester of Cy3.10, Cy3.24, NSCy3, Cy7.18 and NSCy7. Stock solutions of the succinimidyl active esters were made in dry DMF (0.3-1.0 mg active ester/100 μL) and are stable for days when stored at 4 °C. The active esters are also stable in distilled water for several hours provided the pH of the solution is not basic. Aqueous solutions of the dyes can be used for labeling antibodies if the use of DMF is not suitable for certain antibodies. The concentration of cyanine fluorophore in the stock solution was determined by measuring the absorbance of an aliquot of the appropriately diluted stock solution in phosphate-buffered saline (PBS) and using the extinction coefficient of the dye. The stock solution concentration of cyanine fluorophore in the active ester form was then determined by reverse-phase HPLC (typically 50-95% but sometimes as low as 35%). The antibody labeling was carried out in 0.1M carbonate-bicarbonate buffer (pH 9.4) for 15 minutes at room temperature. The sheep IgG (1 mg, 6.45 mM) was dissolved in 0.25 -1 mL buffer solution and the desired amount of dye (e.g., 20 μL of stock containing 0.35mg active ester/100 μL DMF) was added during vigorous vortex mixing. Unconjugated dye was separated from the labeled protein by gel permeation chromatography (0.7 X 20 cm column of Sephadex G-50) using pH 7 buffer solution as eluent.

40

19. Determination of dye-to-antibody ratio

[0074] A simple method for estimating dye/protein (d/p) ratios involves direct measurement of the protein absorbance at 280 nm and dye absorbance at the absorption maximum. Specifically, the dye/protein ratio is calculated using the equation below with measured values of the absorbance of the labeled dye (Cy3 at 550 nm or Cy7 at 750 nm) and the absorbance of protein at 280 nm. The extinction coefficients of Cy3 and Cy7 are approximately 150,000 and 250,000 respectively. The extinction coefficient of the IgG antibody at 280 nm was taken to be 170,000 L/mol-cm. The factor "X" in the denominator accounts for dye absorption at 280 nm which is 0.05 (for both Cy3) and 0.08 (for Cy7) of the absorption of the dye at its maximum absorption (A_{dye}).

50

$$\frac{D}{P} = \frac{A_{dye} * E_{prot}}{(A_{280} - 0.05 A_{dye}) * E_{dye}}$$

55

[0075] A more accurate method is needed when the labeling reagent shows significant spectral changes when bound to the antibody molecule. For example, this approach is needed when both dimers and monomers of dye, which have

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different absorption peaks and different extinction coefficients, are present on the protein at higher d/p ratios. In this case, the labeled protein is dissolved in formamide for absorption spectroscopy and the extinction coefficients of the dye determined independently for the calculation

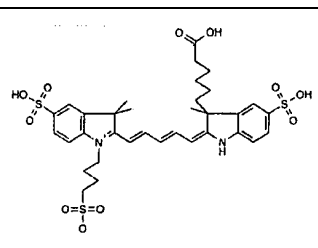
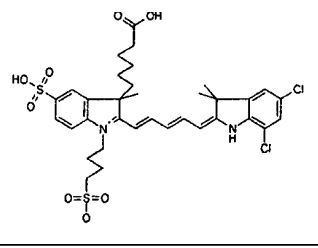
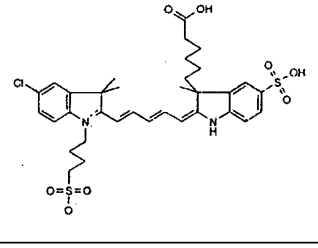
5 20. Measurement of pKa values of pH-sensitive cyanine dyes

[0076] The general method of pKa determination was performed as follows. Purified dye was dissolved in water to give a bulk stock solution (if necessary, a limited amount of acetonitrile was added to ensure solubility). By experiment, the volume of this stock solution that was required to be added to 10.0ml of water + 0.1 %v/v trifluoroacetic acid, in order to give an absorbance of 0.5 ± 0.02 at the dye peak wavelength, was determined. This volume of bulk stock solution was then added to 10.0ml of water, to be used as the working stock solution.

[0077] Aqueous phosphate buffers were prepared covering the region of pH 4-9 {see "Data for Biochemical Research", p.432, 3rd edition, R.M.C Dawson, D.C.Elliott, W.H.Elliott & K.M.Jones, 1987 Oxford University Press. Buffers outside the listed range were made by adding in orthophosphoric acid or sodium hydroxide}. Plastic fluorescence cuvettes were then charged with 3.5ml of each buffer. A fixed volume of dye working stock solution (in the range 50-250 μ l) was added to a cuvette of buffer, the solution mixed and the fluorescence signal measured immediately {Perkin Elmer LS55 fluorimeter, excitation 640nm. Emission spectrum was scanned and fluorescence intensity value at 680nm recorded}. This was repeated for all cuvettes. The graph of fluorescence intensity versus pH was plotted using PRISM, the data fitted to a sigmoidal dose-response curve and the pKa value extracted as the calculated IC50 value from the fitting.

[0078] Table 1 lists the pKa values of the carboxy dyes prepared in the above four examples, along with a selection of non-functional dyes (dyes 5-9) that were prepared using the same methodology. These extra examples help to explain the effect of substitution on the observed pKa value of the dye chromophore, and show that this figure can be tuned to any value across the physiological pH range.

Table 1

No	Name	Structure	pKa
1	2-((1E,3E,5E)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium		7.0
2	3-(5-carboxypentyl)-2-((1E,3E,5E)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2H-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium		6.05
3	2-((1E,3E,5E)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2H-indol-2-ylidene]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3H-indolium		7.44

(continued)

No	Name	Structure	pKa
4	3-(5-carboxypentyl)-2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3-methyl-5-sulfo-1-(4-sulfobutyl)-3 <i>H</i> -indolium		6.2
5	2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(3,3-dimethyl-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3 <i>H</i> -indolium		7.0
6	2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(3,3-dimethyl-5-sulfo-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3 <i>H</i> -indolium		7.0
7	2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(5-chloro-3,3-dimethyl-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3 <i>H</i> -indolium		6.65
8	2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(5-cyano-3,3-dimethyl-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3 <i>H</i> -indolium		6.65
9	5-chloro-2-[(1 <i>E</i> ,3 <i>E</i> ,5 <i>E</i>)-5-(3,3-dimethyl-5-sulfo-1,3-dihydro-2 <i>H</i> -indol-2-ylidene)penta-1,3-dienyl]-3,3-dimethyl-1-(4-sulfobutyl)-3 <i>H</i> -indolium		7.36

[0079] Comparison of dyes 5 and 6 show that substitution of H for sulfonate, in the position *para*- to the unquaternized nitrogen, has no measurable effect on dye pKa. Furthermore, dye 1 shows that addition of the functional arm onto dye 6 has no measurable effect on dye pKa.

[0080] Comparison of the above dye set with dyes 7, 4 and 2 shows the effect of incorporating an electron-withdrawing group, such as Cl, onto the unquaternized indolenine unit. Substitution of Cl *para*- to the unquaternized nitrogen lowers the dye pKa by 0.3-0.4 units (dye 1→dye 7). Substitution of Cl *ortho*-to the unquaternized nitrogen lowers the dye pKa by ~0.8 units (dye 1→dye 4). Incorporating both substitutions gives an additive effect, lowering the pKa by ~1.0 unit (dye 1→dye 2). Dye 8 shows that another electron-withdrawing group, cyano, has a similar effect to chloro. Fluoro and trifluoromethyl act similarly.

[0081] Comparison of dyes 9 and 3 with dyes 6, 7 and 1 shows the effect of incorporating an electron-withdrawing group, such as Cl, onto the quaternized indolenine unit. Substitution of Cl *para*- to the quaternized nitrogen raises the dye pKa by 0.3-0.4 units. Hence the pKa of these pH-sensitive dyes can be adjusted from that of the base structures, at pKa = 7.0, in either direction by the considered placement of electron-withdrawing groups such as chloro (fluoro,

cyano, trifluoromethyl).

Claims

- 5
1. A compound selected from the group consisting of:
- 10
- i) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium;
- 15
- ii) 3-(5-carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2*H*-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium;
- 20
- iii) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3*H*-indolium;
- 25
- iv) 3-(5-carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2*H*-indol-2-ylidene)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium; and
- 30
- v) 3-(5-carboxypentyl)-3-methyl-2-((1*E*,3*E*,5*E*)-5-[3-methyl-5-sulfo-3-(4-sulfobutyl)-1,3-dihydro-2*H*-indol-2-ylidene]penta-1,3-dienyl)-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium.
- 35
2. A method of detecting any one of an antibody, protein, peptide, enzyme substrate, hormone, lymphokine, metabolite, receptor, antigen, hapten, lectin, avidin, streptavidin, toxin, carbohydrate, oligosaccharide, polysaccharide, nucleic acid, derivatized deoxy nucleic acid, DNA fragment, RNA fragment, derivatized DNA fragment, derivatized RNA fragment, nucleoside, nucleotide, natural drug, synthetic drug, virus particle, bacterial particle, virus component, yeast component, blood cell, blood cell component, plasma component, serum component, biological cell, noncellular blood component, bacteria, bacterial component, natural or synthetic lipid vesicle, poison, environmental pollutant, polymer, polymer particle, glass particle, glass surface, plastic particle, plastic surface, polymer membrane, conductor or semiconductor comprising detecting a compound of claim 1 bound to said antibody, protein, peptide, enzyme substrate, hormone, lymphokine, metabolite, receptor, antigen, hapten, lectin, avidin, streptavidin, toxin, carbohydrate, oligosaccharide, polysaccharide, nucleic acid, derivatized deoxy nucleic acid, DNA fragment, RNA fragment, derivatized DNA fragment, derivatized RNA fragment, nucleoside, nucleotide, natural drug, synthetic drug, virus particle, bacterial particle, virus component, yeast component, blood cell, blood cell component, plasma component, serum component, biological cell, noncellular blood component, bacteria, bacterial component, natural or synthetic lipid vesicle, poison, environmental pollutant, polymer, polymer particle, glass particle, glass surface, plastic particle, plastic surface, polymer membrane, conductor or semiconductor.
- 40
3. A method for detecting acidic or basic conditions comprising contacting a compound according to claim 1 with a composition suspected of being acidic or basic and detecting the fluorescence of said compound as an indicator of said acidic or basic conditions.
- 45
4. The method according to claim 3, wherein said composition comprises an intracellular environment.
5. Use of a compound according to claim 1, or a conjugate of a component comprising the same for analysis or detection.
6. Use according to claim 5, wherein said detection is by optical means.

Patentansprüche

1. Verbindung, ausgewählt aus jener Gruppe, die besteht aus:
- 50
- i) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-yliden]penta-1,3-dienyl)-3,3-dimethyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium;
- 55
- ii) 3-(5-Carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(5,7-dichloro-3,3-dimethyl-1,3-dihydro-2*H*-indol-2-yliden)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium;
- iii) 2-((1*E*,3*E*,5*E*)-5-[3-(5-carboxypentyl)-3-methyl-5-sulfo-1,3-dihydro-2*H*-indol-2-yliden]penta-1,3-dienyl)-5-chloro-3,3-dimethyl-1-(4-sulfobutyl)-3*H*-indolium;
- iv) 3-(5-Carboxypentyl)-2-((1*E*,3*E*,5*E*)-5-(7-chloro-3,3-dimethyl-5-sulfo-1,3-dihydro-2*H*-indol-2-yliden)penta-1,3-dienyl)-3-methyl-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium; und
- v) 3-(5-Carboxypentyl)-3-methyl-2-((1*E*,3*E*,5*E*)-5-[3-methyl-5-sulfo-3-(4-sulfobutyl)-1,3-dihydro-2*H*-indol-2-yliden]penta-1,3-dienyl)-5-sulfo-1-(4-sulfobutyl)-3*H*-indolium.

den]penta-1,3-dienyl}-5-sulfo-1-(4-sulfobutyl)-3H-indolium.

2. Verfahren zum Nachweisen eines beliebigen von einer bzw. einem Antikörper, Protein, Peptid, Enzymsubstrat, Hormon, Lymphokin, Metaboliten, Rezeptor, Antigen, Hapten, Lektin, Avidin, Streptavidin, Toxin, Kohlehydrat, Oligosaccharid, Polysaccharid, Nukleinsäure, derivatisierten Desoxynukleinsäure, DNA-Fragment, RNA-Fragment, derivatisierten DNA-Fragment, derivatisierten RNA-Fragment, Nukleosid, Nukleotid, natürlichen Wirkstoff, synthetischen Wirkstoff, Viruspartikel, Bakterienpartikel, Viruskomponente, Hefekomponente, Blutzelle, Blutzellenkomponente, Plasmakomponente, Serumkomponente, biologischen Zelle, nichtzellulären Blutkomponente, Bakterien, Bakterienkomponente, natürlichen oder synthetischen Lipidvesikel, Gift, Umweltschadstoff, Polymer, Polymerpartikel, Glaspartikel, Glasfläche, Kunststoffpartikel, Kunststofffläche, Polymermembran, Leiter oder Halbleiter, umfassend das Nachweisen einer Verbindung des Anspruchs 1, die gebunden ist an den Antikörper, das Protein, das Peptid, das Enzymsubstrat, das Hormon, das Lymphokin, den Metaboliten, den Rezeptor, das Antigen, das Hapten, das Lektin, das Avidin, das Streptavidin, das Toxin, das Kohlehydrat, das Oligosaccharid, das Polysaccharid, die Nukleinsäure, die derivatisierte Desoxynukleinsäure, das DNA-Fragment, das RNA-Fragment, das derivatisierte DNA-Fragment, das derivatisierte RNA-Fragment, das Nukleosid, das Nukleotid, den natürlichen Wirkstoff, den synthetischen Wirkstoff, das Viruspartikel, das Bakterienpartikel, die Viruskomponente, die Hefekomponente, die Blutzelle, die Blutzellenkomponente, die Plasmakomponente, die Serumkomponente, die biologische Zelle, die nichtzelluläre Blutkomponente, die Bakterien, die Bakterienkomponente, das natürliche oder synthetische Lipidvesikel, das Gift, den Umweltschadstoff, das Polymer, das Polymerpartikel, das Glasfläche, das Kunststoffpartikel, die Kunststofffläche, die Polymermembran, den Leiter oder Halbleiter.
3. Verfahren zum Nachweisen saurer oder basischer Bedingungen, umfassend das In-Kontakt-Bringen einer Verbindung nach Anspruch 1 mit einer Zusammensetzung, von der angenommen wird, dass sie sauer oder basisch ist, und das Nachweisen der Fluoreszenz der Verbindung als Indikator der sauren oder basischen Bedingungen.
4. Verfahren nach Anspruch 3, wobei die Zusammensetzung eine intrazelluläre Umgebung umfasst.
5. Verwendung einer Verbindung nach Anspruch 1, oder eines Konjugats mit einer dieselbe umfassenden Komponente, zwecks Analyse oder Nachweis.
6. Verwendung nach Anspruch 5, wobei der Nachweis durch optische Mittel erfolgt.

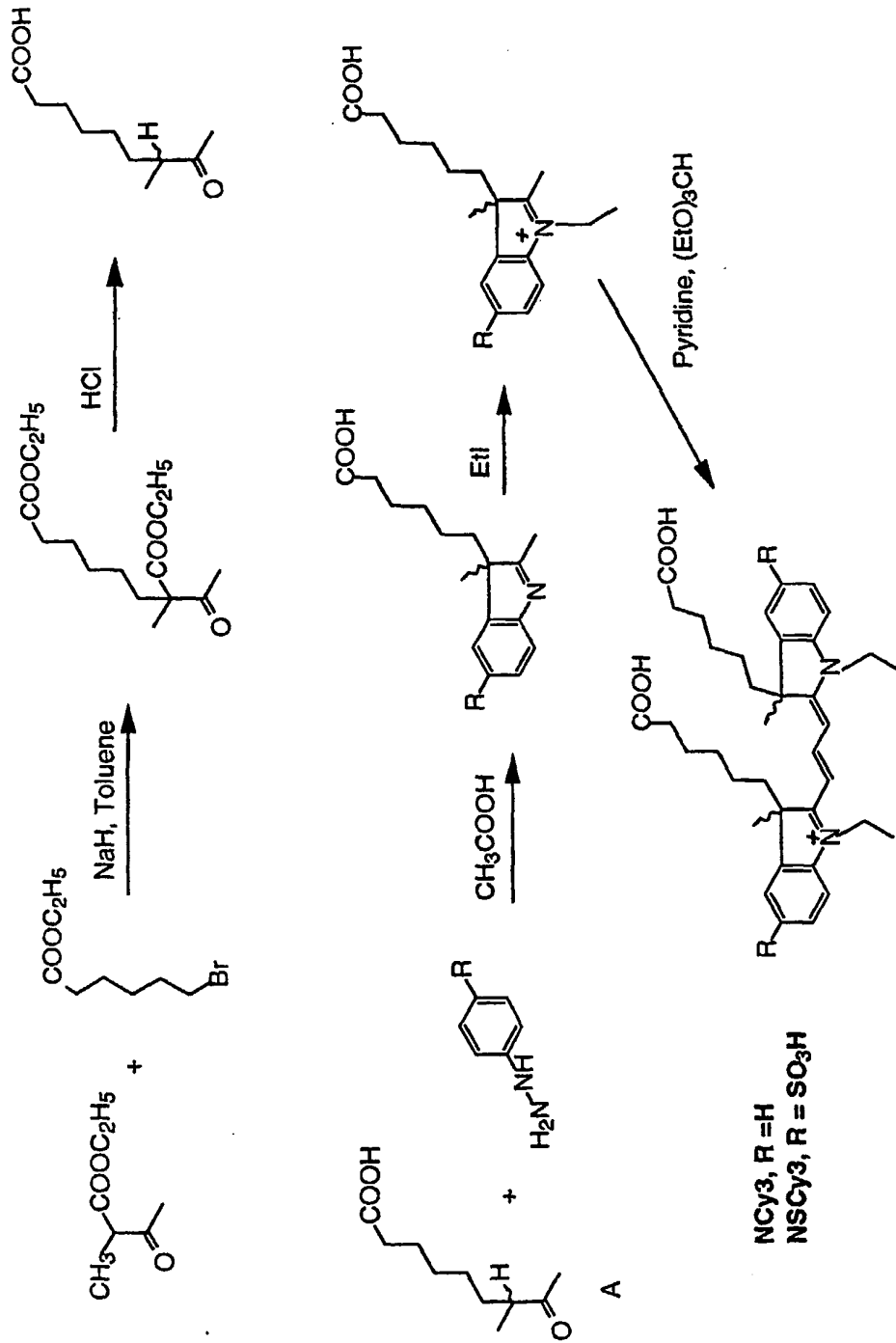
Revendications

1. Composé sélectionné à partir du groupe constitué par :
 - i) le 2-((1E, 3E, 5E)-5-[3-(5-carboxypentyl)-3-méthyl-5-sulfo-1, 3-dihydro-2H-indol-2-ylidène]penta-1, 3-diényl)-3, 3-diméthyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium ;
 - ii) le 3-(5-carboxypentyl)-2-((1E, 3E, 5E)-5-(5, 7-dichloro-3, 3-diméthyl-1, 3-dihydro-2H-indol-2-ylidène)penta-1, 3-diényl)-3-méthyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium ;
 - iii) le 2-((1E, 3E, 5E)-5-[3-(5-carboxypentyl)-3-méthyl-5-sulfo-1, 3-dihydro-2H-indol-2-ylidène]penta-1, 3-diényl)-5-chloro-3, 3-diméthyl-1-(4-sulfobutyl)-3H-indolium ;
 - iv) le 3-(5-carboxypentyl)-2-((1E, 3E, 5E)-5-(7-chloro-3, 3-diméthyl-5-sulfo-1, 3-dihydro-2H-indol-2-ylidène)penta-1, 3-diényl)-3-méthyl-5-sulfo-1-(4-sulfobutyl)-3H-indolium ; et
 - v) le 3-(5-carboxypentyl)-3-méthyl-2-(1E, 3E, 5E)-5-[3-méthyl-5-sulfo-3-(4-sulfobutyl)-1, 3-dihydro-2H-indol-2-ylidène]penta-1, 3-diényl)-5-sulfo-1-(4-sulfobutyl)-3H-indolium.
2. Procédé de détection de l'un quelconque d'un anticorps, d'une protéine, d'un peptide, d'un substrat d'enzyme, d'une hormone, d'une lymphokine, d'un métabolite, d'un récepteur, d'un antigène, d'un haptène, d'une lectine, d'une avidine, d'une streptavidine, d'une toxine, d'un hydrate de carbone, d'un oligosaccharide, d'un polysaccharide, d'un acide nucléique, d'un dérivé désoxy d'acide nucléique, d'un fragment d'ADN, d'un fragment d'ARN, d'un fragment de dérivé d'ADN, d'un fragment de dérivé d'ARN, d'un nucléoside, d'un nucléotide, d'un médicament naturel, d'un médicament de synthèse, d'une particule virale, d'une particule bactérienne, d'un composant viral, d'un composant de levure, d'une cellule sanguine, d'un composant de cellules sanguine, d'un composant de plasma, d'un composant de sérum, d'une cellule biologique, d'un composant sanguin non cellulaire, de bactéries, d'un composant bactérien, d'une vésicule de lipide naturelle ou de synthèse, d'un poison, d'un polluant environnemental, d'un polymère, d'une particule polymère, d'une particule de verre, d'une surface de verre, d'une particule de matière plastique, d'une

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5 surface de matière plastique, d'une membrane de polymère, d'un conducteur ou d'un semi-conducteur comprenant la détection d'un composé selon la revendication 1 lié audit anticorps, protéine, peptide, substrat d'enzyme, hormone, lymphokine, métabolite, récepteur, antigène, haptène, lectine, avidine, streptavidine, toxine, hydrate de carbone, oligosaccharide, polysaccharide, acide nucléique, dérivé désoxy d'acide nucléique, fragment d'ADN, fragment d'ARN, fragment de dérivé d'ADN, fragment de dérivé d'ARN, nucléoside, nucléotide, médicament naturel, médicament de synthèse, particule virale, particule bactérienne, composant viral, composant de levure, cellule sanguine, composant de cellule sanguine, composant de plasma, composant de sérum, cellule biologique, composant sanguin non cellulaire, bactéries, composant bactérien, vésicule de lipide naturelle ou de synthèse, poison, polluant environnemental, polymère, particule polymère, particule de verre, surface de verre, particule de matière plastique, surface de matière plastique, membrane de polymère, conducteur ou semi-conducteur.

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3. Procédé de détection de conditions acides ou basiques comprenant la mise en contact d'un composé selon la revendication 1 avec une composition suspectée d'être acide ou basique et de détection de la fluorescence dudit composé en tant qu'indicateur desdites conditions acides ou basiques.
 4. Procédé selon la revendication 3, dans lequel ladite composition comprend un environnement intracellulaire.
 5. Utilisation d'un composé selon la revendication 1, ou d'un conjugué d'un composant comprenant ce dernier à des fins d'analyse ou de détection.
 6. Utilisation selon la revendication 5, dans lequel ladite détection est assurée par des moyens optiques.



General scheme for the synthesis of functionalized ketone (A) and its cyanine dyes

FIGURE 1

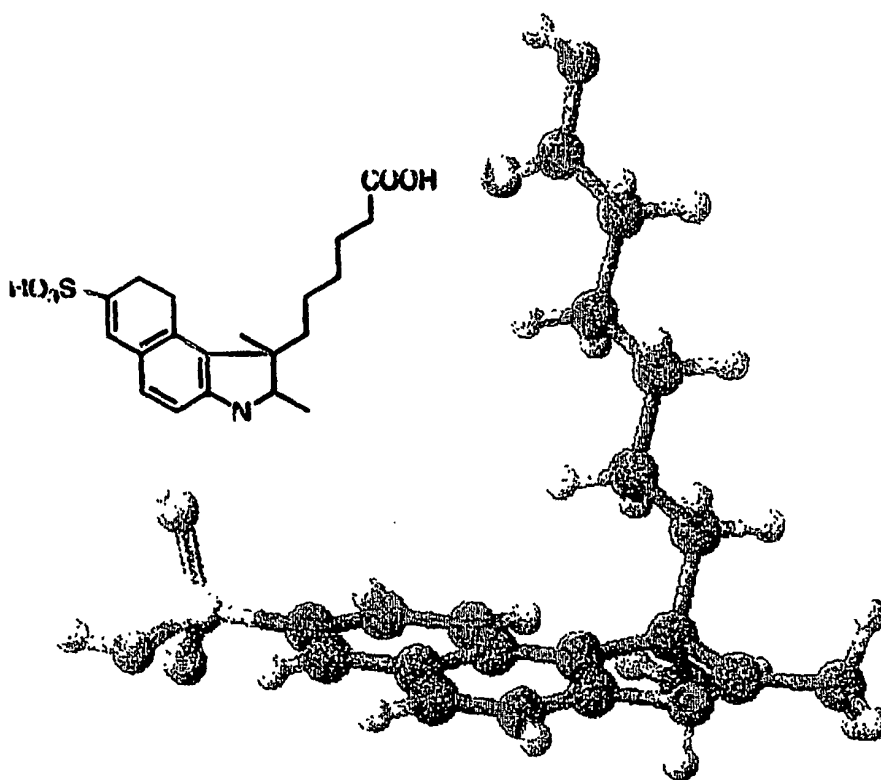


FIGURE 2

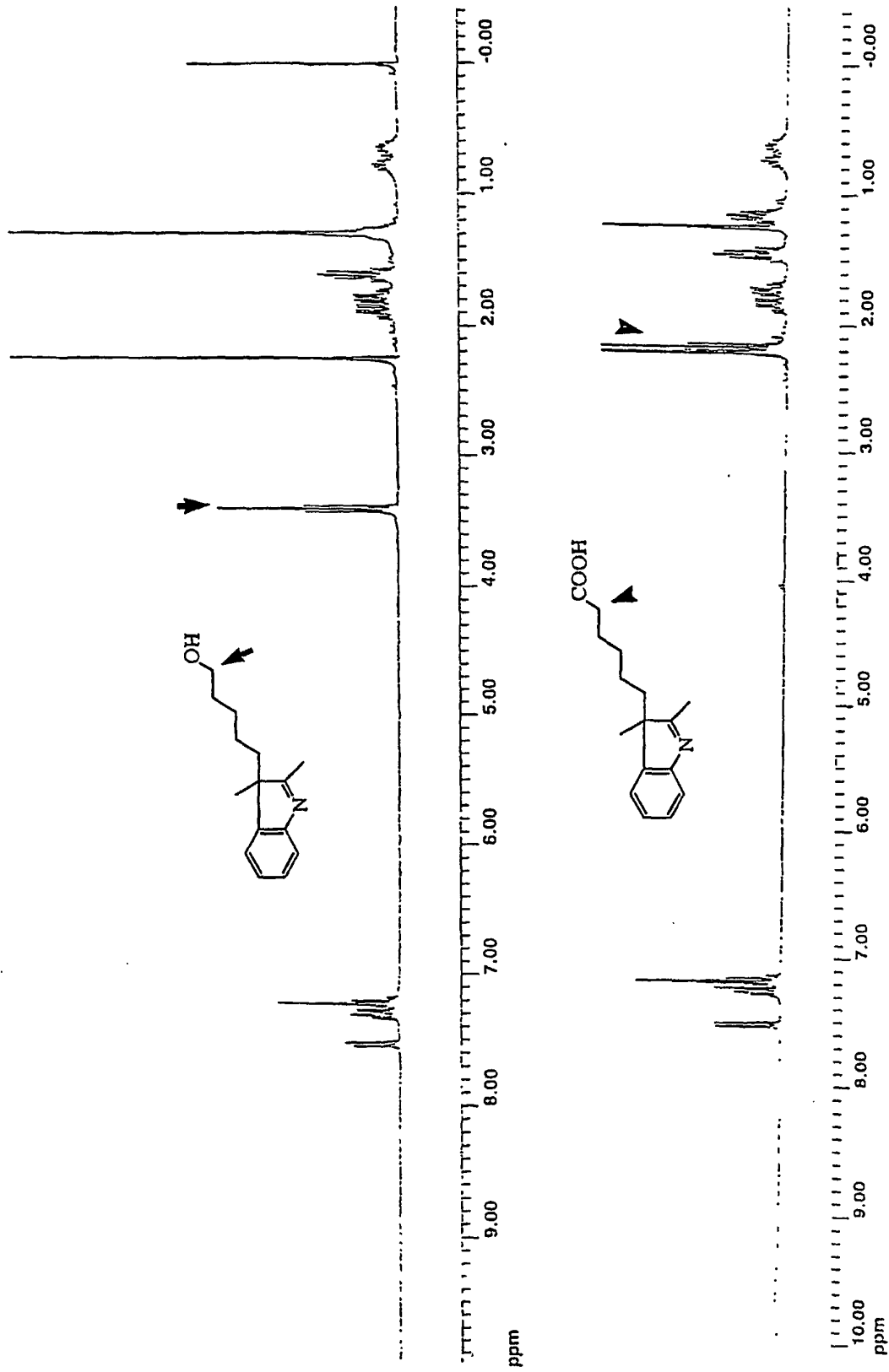
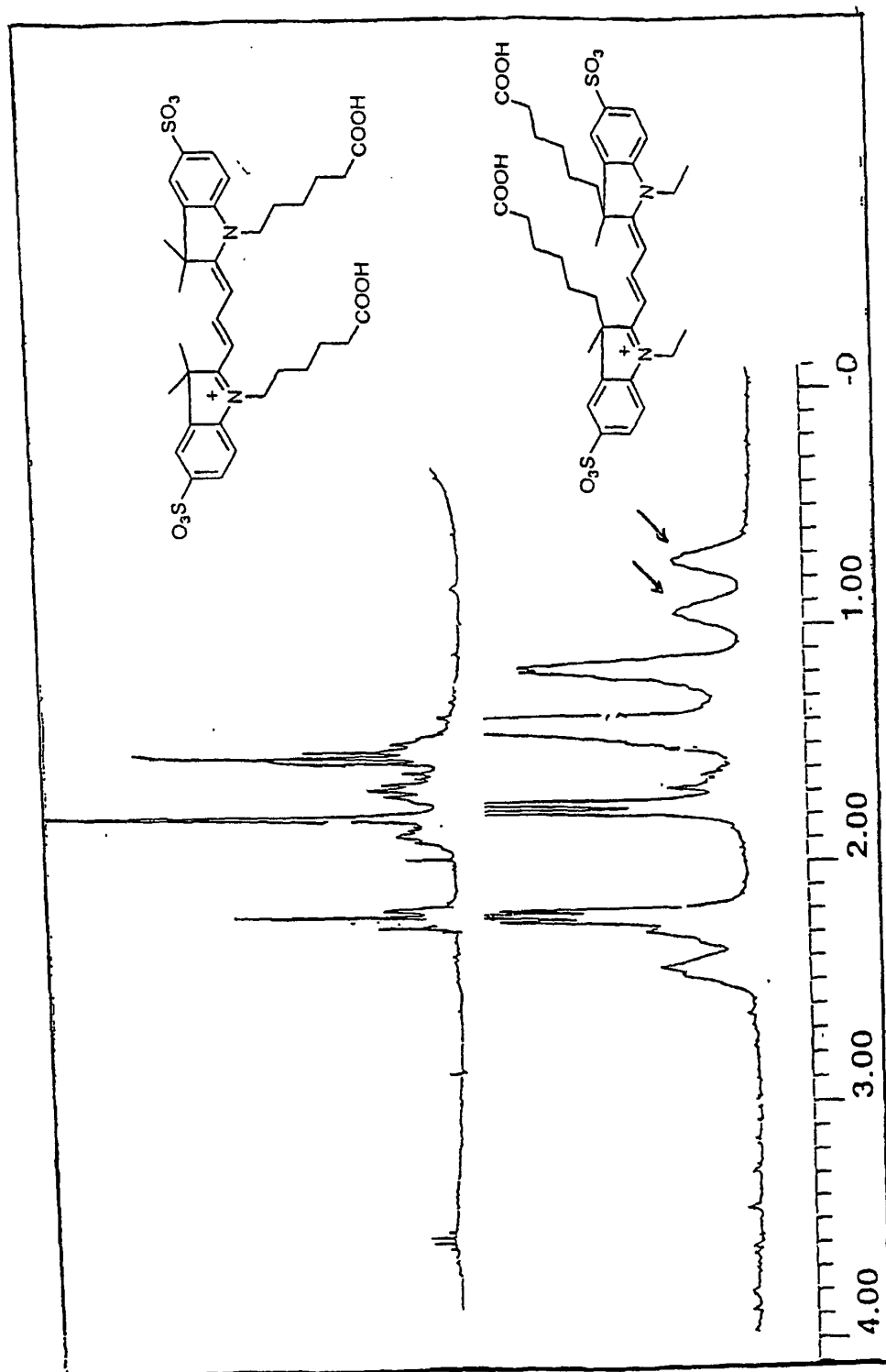
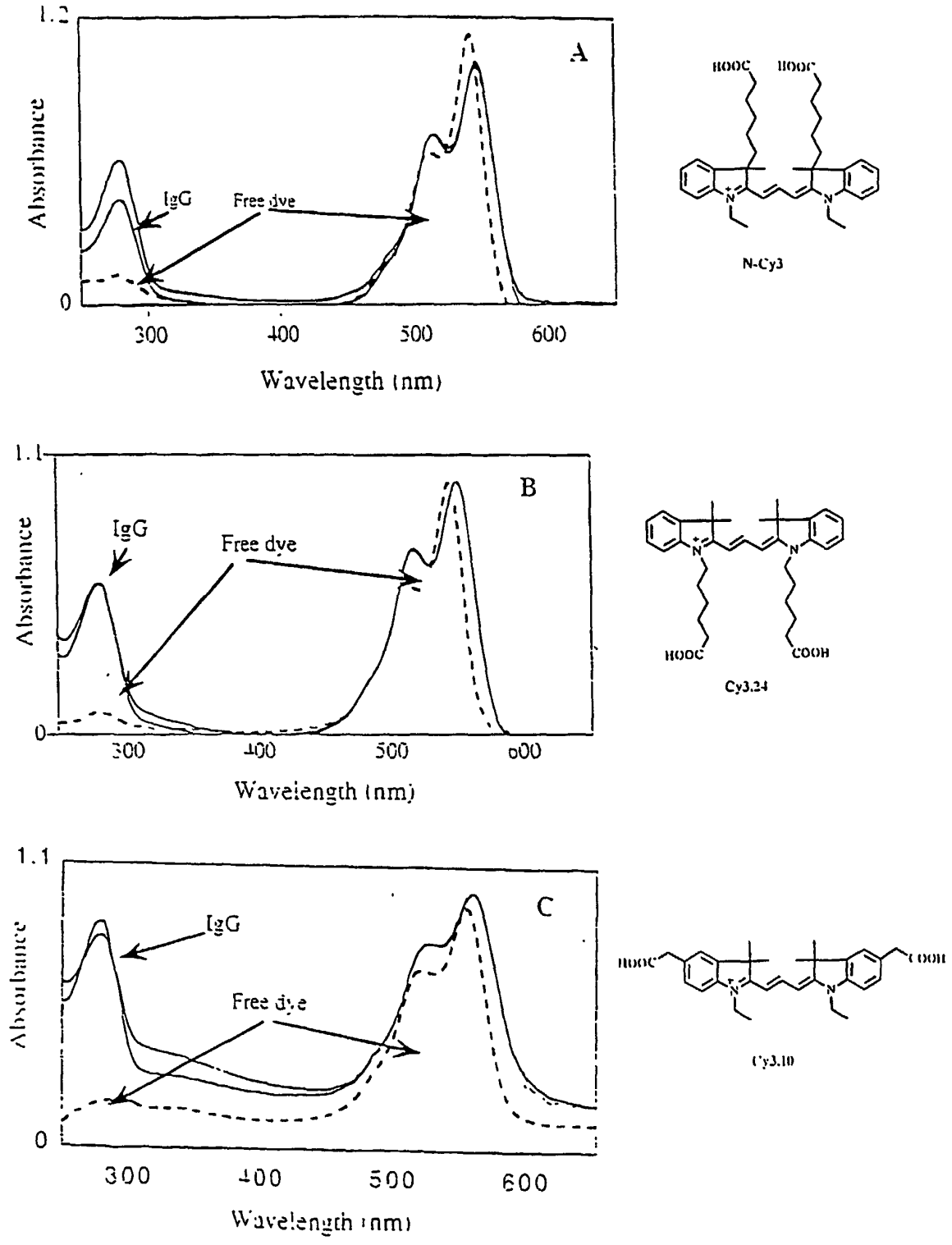


FIGURE 3



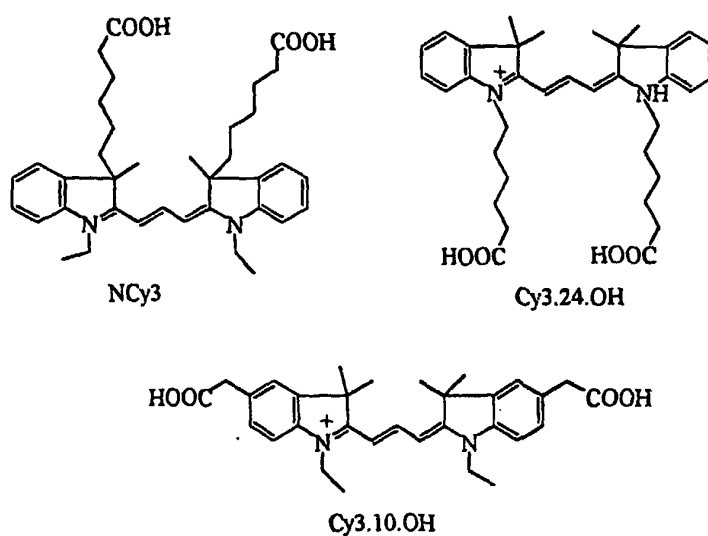
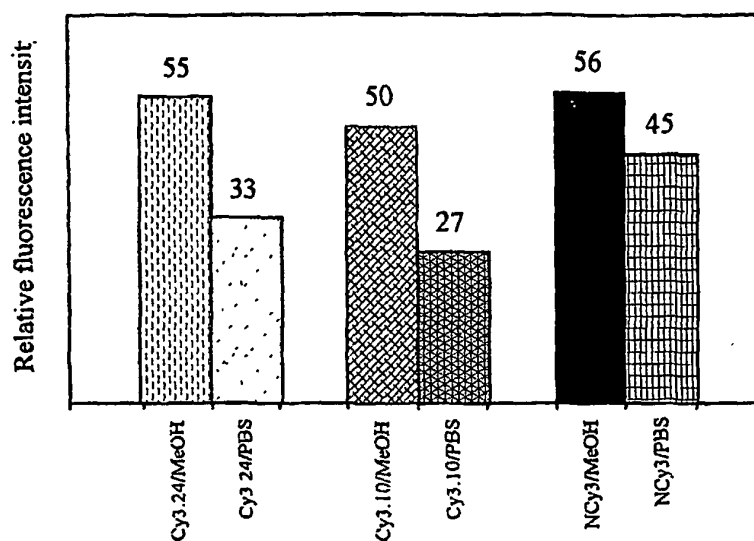
^1H NMR spectra of Cy3.18.OH and NSCy3 in D_2O . Some of the methylene protons of the hexanoic acid chain in NSCy3 are shielded by heterocyclic ring and appear upfield. (shown by arrow).

FIGURE 4



Absorption spectra of non-sulfonated Cy3 isomers and their antibody conjugates.

FIGURE 5



Relative fluorescence intensities of Cy3 dyes. Absorption is 0.05 at 550 nm and excitation wavelength in 514 nm. Ncy3 is brighter in PBS solution suggesting less dye/dye aggregation.

FIGURE 6

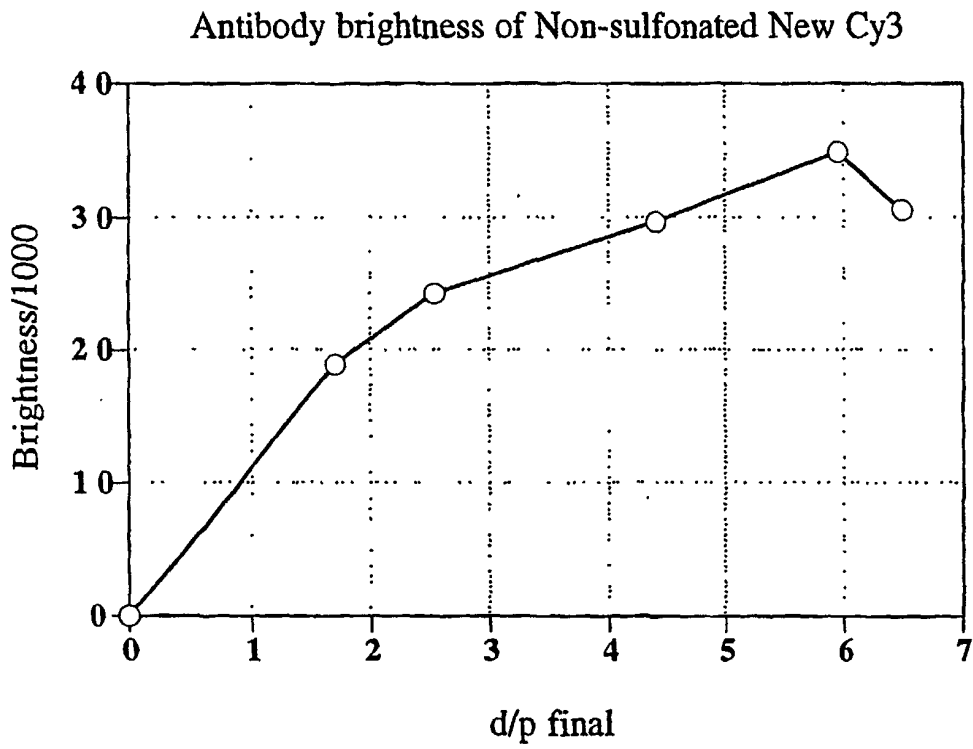
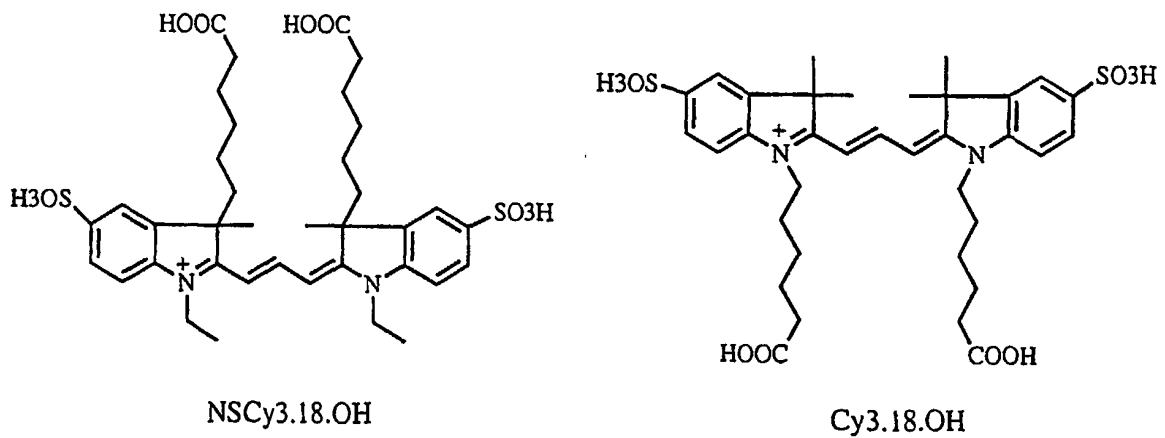
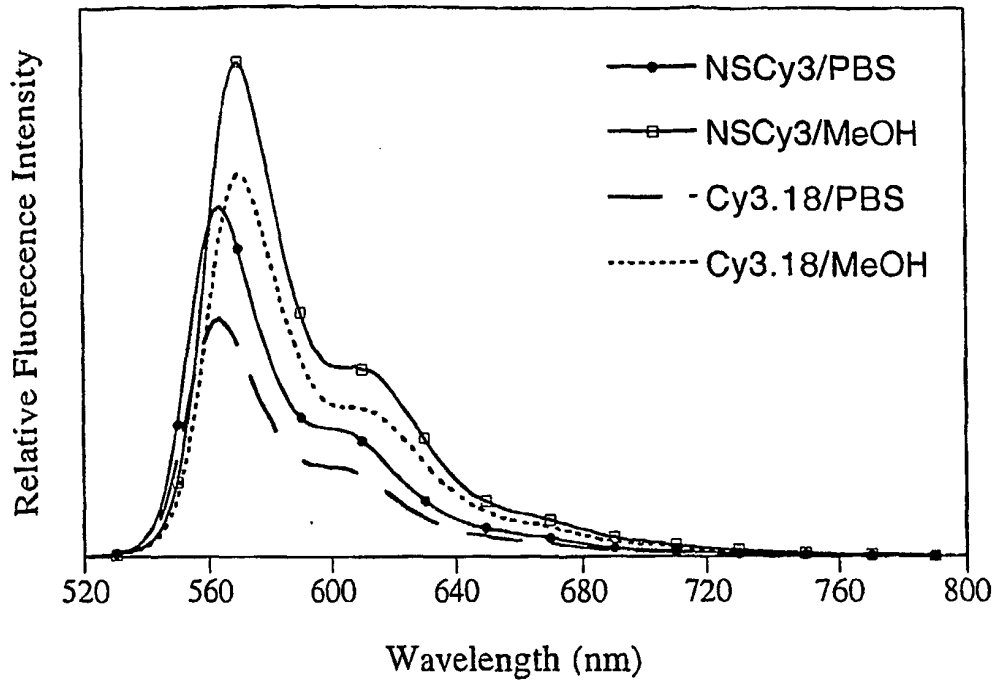


FIGURE 8



Sulfonated Cy3 dicarboxylic acid isomers

FIGURE 9

REFERENCES CITED IN THE DESCRIPTION

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专利名称(译)	手性吲哚中间体及其荧光花青染料含有官能团		
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摘要(译)

本发明涉及功能化的花青染料，更具体地说，涉及作为制备花青染料的中间体的手性3-取代的2, 3'-二甲基-3-H-吲哚及其衍生物的合成，涉及到这些染料的制备方法和制备方法。如此制备的染料。

