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(54) Title: TETHERED LIGANDS AND METHODS OF USE



Control (-)



ELC-Stalkokine (SK)



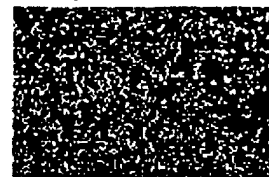
ELC-SK + soluble ELC



ELC-SK + soluble TECK



ELC-SK + soluble SLC



ELC-SK + soluble MCP-3

(57) Abstract: The invention provides methods, reagents and devices for analysis of receptor ligand interactions, determining the profile of receptor expression in cells and cell populations, and diagnostic and drug screening methods. The invention makes use of immobilized tethered ligand fusion proteins having a ligand domain, a stalk domain, and optionally an immobilization domain.



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TETHERED LIGANDS AND METHODS OF USE

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FIELD OF THE INVENTION

The invention relates to reagents and methods for characterizing receptor-ligand interactions, identifying modulators of such interactions, and characterizing the receptor expression profile of cells and cell populations. The invention finds application in the biomedical sciences.

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CROSS-REFERENCES TO RELATED APPLICATIONS

This application claims priority to U.S.S.N. 60/186626, filed March 3, 2000, the contents of which are incorporated by reference herein.

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BACKGROUND OF THE INVENTION

Communication between cells is a fundamental process involved in growth, differentiation, metabolism, and generation of biological responses (e.g., immune responses) in multicellular organisms. Often, cell-to-cell signaling is mediated by extracellular receptors or cell adhesion molecules. These membrane-associated molecules interact with other proteins, such as soluble factors such as peptide hormones, extracellular matrices, and cell surface molecules displayed by other cells. In some cases, signaling involves cell-to-cell contact (e.g., contact between two surface proteins on cells). An example of such an interaction is the binding of the T cell receptor on the surface of a T lymphocyte and the MHC/antigen complex on the surface of an antigen-presenting cell. A different type of cell-to-cell signaling is mediated by soluble polypeptides that, by interacting with a specific receptor on a target cell, lead to changes in target cell activity. An illustrative and important example cell-to-cell signaling mediated by soluble polypeptides is the activity of chemokines in mammalian systems.

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Chemokines are a class of cytokines that play an important role in inflammatory responses, leukocyte trafficking, angiogenesis, and other biological processes related to the migration and activation of cells. As mediators of chemotaxis and inflammation, chemokines play roles in pathological conditions. For

example, the concentration of chemokine MCP-1 is higher in the synovial fluid of patients suffering from rheumatoid arthritis than that of patients suffering from other arthritic diseases.

Known chemokines are typically assigned to one of four subfamilies based on the arrangement of cysteine motifs. In the so-called alpha-chemokines, for example, the first two of four cysteines (starting from the amino terminus) are separated by an intervening amino acid (i.e., having the motif C-X-C). The beta-chemokines are characterized by the absence of an intervening amino acid between first two cysteines (i.e., comprising the motif C-C). The smaller gamma-chemokine family is characterized by a single C residue (gamma). The delta-chemokine family is characterized by a pair of cysteines separated by three residues (i.e., having the motif CX₃C). The sole CX₃C chemokine (fractalkine) is a type 1 membrane protein containing a chemokine domain tethered on a long mucin-like stalk. Fractalkine, also named neurotactin, has contains a chemokine domain at the amino terminus tethered on a long mucin-like stalk. For a recent review on chemokines, see Ward et al., 1998, *Immunity* 9:1-11 and Baggiolini et al., 1998, *Nature* 392:565-568, and the references cited therein.

Some chemokine activities (e.g., promigratory effects) are mediated by binding to an array of cell surface receptors on the surface of target leukocytes. These receptors are of the seven transmembrane spanning, G protein coupled receptor class (alternately referred to as 7TM or GPCR). Several seven-transmembrane-domain G protein-coupled receptors for C-C chemokines have been cloned: a C-C chemokine receptor-1 which recognizes MIP-1 α , RANTES, MCP-2, MCP-3, and MIP-5 (Neote et al., 1993, *Cell*, 72:415-415); CCR2 which is a receptor for MCP1, 2, 3 and 4 or 5; CCR3 which is a receptor for RANTES, MCP-2, 3, 4, MIP-5 and eotaxin; CCR5 which is a receptor for MIP-1 α , MIP-1 β and RANTES; CCR4 which is a receptor for CMDC or TARC; CCR6 which is a receptor for LARC; and CCR7 which is a receptor for SLC and MIP-3 β (reviewed in Sallusto et al., 1998, *Immunol. Today* 19:568 and Ward et al., 1998, *Immunity* 9:1-11).

Due to the importance of interactions between receptors (such as chemokine receptors) and their ligands in biological function, a need exists for rapid and effective methods for the characterization of such interactions.

SUMMARY OF THE INVENTION

In one aspect the invention provides an assay device having a plurality of different immobilized tethered ligand fusion proteins organized in an array, where the ligand-stalk fusion proteins include a ligand domain and a stalk domain ("assay device"). In one embodiment, the ligand-stalk fusion proteins have a ligand domain, an intermediate stalk domain, and an immobilization domain ("assay device"). In an embodiment, the ligand domain and the stalk domain are not associated in a naturally occurring protein. In various embodiments, the immobilized tethered ligand fusion proteins include a mucin-derived stalk sequence, a fractalkine mucin repeat region sequence, and/or a ligand domain encoding a chemokine.

In another aspect, the invention provides a tethered ligand fusion protein, where the ligand domain is other than a chemokine sequence.

In another aspect, the invention provides a method for identifying an interaction between a receptor and a ligand by contacting a cell expressing a receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein and, detecting binding of the cell and the tethered ligand fusion protein, where binding of the cell to the fusion protein is correlated with an interaction between the receptor and the ligand corresponding to the ligand domain of the fusion protein. In an embodiment, the tethered ligand is immobilized on an assay device, supra. In various embodiments, the cell expresses a recombinant receptor (e.g., an orphan receptor). In an embodiment, the cell expresses a chemokine receptor. In one embodiment, an interaction between a receptor and more than one ligand is detected.

In another aspect, the invention provides a method for identifying a modulator of an interaction between a receptor and a ligand by contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the absence of a test compound, and measuring binding of the cells to an immobilized tethered ligand fusion protein, contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the presence of a test compound, and measuring binding of the cells to the immobilized tethered ligand fusion protein; and comparing the levels of binding, where decreased binding of cells in the presence of the test compound indicates that the test compound is an inhibitor of the interaction between the receptor and the ligand corresponding the ligand domain of said fusion protein,

and where increased binding of cells in the presence of the test compound indicates that the test compound is an enhancer of the interaction between the receptor and the receptor and the ligand corresponding the ligand domain of the fusion protein. In an embodiment, the tethered ligand is immobilized on an assay device, supra. In one embodiment, the receptor is a chemokine receptor.

In another aspect, the invention provides a method for detecting a profile of receptor expression in cells in a population of cells by contacting the population of cells with an immobilized tethered ligand fusion protein and detecting binding of cells of the population to the tethered ligand fusion protein, where binding of a cell in the cell population to a tethered ligand fusion protein is correlated with expression of a receptor that binds the ligand corresponding to the ligand domain of the fusion protein. In an embodiment, the tethered ligand is immobilized on an assay device, supra. In an embodiment, the contacting step involves contacting the population with a plurality of different fusion proteins and the detecting step involves detecting binding of cells to zero, one or more than one fusion proteins. In one embodiment, the population is heterogeneous, e.g., the population is obtained from synovial fluid, cerebral-spinal fluid, bronchial alveolar lavage (BAL) fluid, or blood. In an embodiment, the method includes quantitating the level of binding to each tethered ligand fusion protein in the array, or characterizing the cells bound at each sector of the array (e.g., by immunostaining).

In another aspect, the invention provides a method for diagnosis. The method includes the steps of obtaining a population of cells from a patient suspected of suffering from a disease, determining a receptor profile for the population, and, comparing said receptor profile with a profile characteristic of the disease state. In one embodiment the receptor profile is determined by contacting the population of cells with an immobilized tethered ligand fusion protein and identifying a tethered ligand fusion protein bound by cells of said population thereby identifying a receptor profile. In an embodiment, the tethered ligand is immobilized on an assay device, supra. In various embodiments, determining the method also includes quantitating the binding of cells at each sector of the array or characterizing the cells bound at each sector of the array, e.g., by immunostaining. In various embodiments, the disease is an inflammatory or allergic disease, or an autoimmune disease. In various embodiments, the population is obtained from synovial fluid, cerebral-spinal fluid, bronchial alveolar lavage (BAL) fluid, or blood.

In another aspect, the invention provides a method for detecting an effect of a drug or treatment on a patient by determining the receptor profile of a population of cells from the patient for the first time, administering the drug or treatment to the patient, determining the receptor profile of a population of cells from the patient for a second time, and comparing the receptor profiles obtained to determine the effect of the drug or treatment receptor-expressing cells in the patient. In an embodiment, the determining is carried out by contacting said population of cells with an immobilized tethered ligand fusion protein and identifying a subset of arrayed tethered ligand fusion proteins bound by cells of said population thereby identifying a receptor profile. In an embodiment, the tethered ligand is immobilized on an assay device, supra. In various embodiments, determining the method also includes quantitating the binding of cells at each sector of the array or characterizing the cells bound at each sector of the array, e.g., by immunostaining. In various embodiments, the population is obtained from synovial fluid, cerebral-spinal fluid, bronchial alveolar lavage (BAL) fluid, or blood.

In another aspect, the invention provides a method for identifying a modulator of an interaction between a receptor and a ligand by contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the presence of a test compound and determining the level of binding to the fusion protein, contacting cells expressing the receptor or a ligand-binding portion thereof with the immobilized tethered ligand fusion protein in the absence of a test compound; and determining the level of binding to the fusion protein, and comparing the level of binding, where the ligand-stalk fusion proteins include a ligand domain, an intermediate stalk domain, and, an immobilization domain and where decreased binding of cells in the presence of the test compound indicates that the test compound is an inhibitor of the interaction between the receptor and the ligand corresponding the ligand domain of fusion protein, and where increased binding of cells in the presence of the test compound indicates that the test compound is an enhancer of the interaction between the receptor and the ligand corresponding the ligand domain of the fusion protein. In one embodiment, the ligand domain and the stalk domain are not associated in a naturally occurring protein. In an embodiment, the stalk domain is carboxy-terminal to the ligand domain, and the immobilization domain is carboxy-terminal to the stalk domain.

In another aspect, the invention provides the use of tethered ligand fusion proteins in any of the methods described herein.

BRIEF DESCRIPTION OF THE FIGURES

5 Figure 1 shows the structure of a tethered ligand.

Figure 2 shows the nucleotide sequence for a human CCR10 (SEQ ID NO:1) and the predicted amino acid sequence of the human CCR10 polypeptide (SEQ ID NO:2).

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Figure 3 shows the identification of CCR10 ligands by adhesion to stalkokines. Figure 3A shows interrogation of immobilized stalkokine (SK) by HEK293-CCR10 cells, where 'control' shows background adhesion of HEK293-CCR10 cells to wells containing no stalkokine (anchoring antibodies alone are present); "ELC-stalkokine (SK)" shows strong adhesion of HEK293-CCR10 cells to locations containing ELC-stalkokines immobilized via anchoring antibodies; "ELC-SK + soluble ELC", "soluble TECK", or "soluble SLC" shows ablation of adhesion in the presence of excess concentrations of soluble recombinant 'native form' chemokines as shown; "ELC-SK + soluble MCP-3" shows no diminution in adhesion in the presence MCP-3 as representative of many non-competing chemokines. Wild type HEK293 cells showed no adhesion to any of the sites (not shown). Figure 3B shows the quantitation of adhesion of HEK293-CCR10 cells to ELC-stalkokine in the absence and presence of soluble chemokines from a representative experiment. Figure 3C shows the results of homologous competition binding assay using radiolabeled ELC in the presence of increasing concentrations of cold ELC on either HEK293-CCR10 cells (filled squares) or wild type HEK293 cells (open squares).

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Figure 4 shows a schematic diagram of an expression plasmid encoding an ELC-stalkokine. Not all details of the plasmid are shown (e.g., pA site, polylinker, Ori, selection markers are not shown).

Figure 5 shows Western blot analysis of 32 different stalkokines. Each individual stalkokine construct was transfected into 293T and supernatants

containing the stalkokines were subjected to electrophoresis followed by Western blot analysis using the anti-poly histidine antibody. Equivalent volumes of supernatant were analyzed in each lane. Expression levels varied from a very robust accumulation in the case of MCP3 to a low level accumulation (IL-8).

5 Enhanced stalkokine accumulation can be achieved by addition of protease inhibitors. Due to glycosylation, the apparent molecular weight of the soluble stalkokines increased from the typical 35 kDa to around 90 kDa. Partial glycosylation resulted in an intermediate molecular weight, and it demonstrated by the very broad band or smear of signal. MW markers = 97.4, 68, 43, 29, 18.4, 14.3
10 kDa.

Figure 6 shows a displacement assay showing competition by a stalkokine with binding of a radiolabeled tracer chemokine to its cognate receptor.

15 Figure 7 shows a calcium mobilization assay using CEM cells expressing CCR4 receptors contacted with TARC-Stalkokine or MDC-Stalkokine, showing induction of calcium signaling in a receptor dependent manner.

DETAILED DESCRIPTION

20 1. DEFINITIONS

The following definitions are provided to assist the reader in the practice of the invention. As used herein:

The term "receptor" has the ordinary meaning in the art and refers to a protein that specifically binds a second protein. Usually (i.e., in some embodiments)
25 receptors are membrane associated proteins characterized by a transmembrane domain, an extracellular domain and an intracellular domain. Such receptors are called "membrane associated receptors." In other embodiments, the term "receptor" is used to refer to other types of polypeptide binding proteins, e.g., adhesion molecules, soluble binding proteins. It will be apparent that, as used herein, the
30 term "receptor" does not refer to immunoglobulins.

The term "ligand," refers to peptides or polypeptide that bind to a receptor protein. Typically, ligands are involved in cell-to-cell signaling or adhesion of cells to other cells or extracellular matrix.

The term "specific binding" refers to binding between two molecules, for example, a ligand and a receptor, characterized by the ability of a protein (ligand) to associate with another specific molecule (receptor) even in the presence of many other diverse molecules, i.e., to show preferential binding of one molecule for another in a heterogeneous mixture of molecules. Specific binding of a ligand to a receptor is also evidenced by reduced binding of a detectably labeled ligand to the receptor in the presence of excess unlabeled ligand (i.e., a binding competition assay).

The term "orphan receptor" refers to a putative receptor polypeptide for which a naturally occurring cognate ligand is not known. Typically, orphan receptors are identified based on homology (sequence identity) to known receptors based on protein or gene sequence and structure.

The term "fusion protein," as used herein, refers to a composite polypeptide, i.e., a single contiguous amino acid sequence, made up of two or more (e.g., three) distinct polypeptides which are not normally fused together in a single amino acid sequence. Fusion proteins can generally be prepared using either recombinant nucleic acid methods, i.e., as a result of transcription and translation of a recombinant gene fusion product, which fusion comprises a segment encoding a first polypeptide region and a segment encoding a second polypeptide region, or by chemical synthesis methods well known in the art. Thus, for example, a single naturally occurring protein (e.g., Fractalkine) is not a fusion protein.

The term "recombinant" when used with reference to a cell indicates that the cell replicates a heterologous nucleic acid, or expresses a peptide or protein encoded by a heterologous nucleic acid. Recombinant cells can contain genes that are not found within the native (non-recombinant) form of the cell. Recombinant cells can also contain genes found in the native form of the cell wherein the genes are modified and re-introduced into the cell by artificial means (e.g., to increase expression of a receptor). The term also encompasses cells that contain a nucleic acid endogenous to the cell that has been modified without removing the nucleic acid from the cell; such modifications include those obtained by gene replacement, site-specific mutation, and related techniques.

A "recombinant expression cassette" or simply an "expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with nucleic acid elements that are capable of effecting expression of a structural gene in

hosts compatible with such sequences. Expression cassettes include at least promoters and optionally, transcription termination signals. Typically, the recombinant expression cassette includes a nucleic acid to be transcribed (e.g., a nucleic acid encoding a desired polypeptide), and a promoter. Additional factors
5 necessary or helpful in effecting expression may also be used as described herein. Transcription termination signals, enhancers, and other nucleic acid sequences that influence gene expression, can also be included in an expression cassette. An “expression vector” is a vector that contains the necessary elements for the transcription and translation of the inserted coding sequence required for the
10 expression system employed (e.g., a vector containing an expression cassette).

A “recombinant polynucleotide” or a “recombinant polypeptide” is a non-naturally occurring polynucleotide or polypeptide that includes nucleic acid or amino acid sequences, respectively, from more than one source nucleic acid or polypeptide.

15 The term “chemokine” (abbreviated “CK”) refers to a class of cytokines involved in inflammatory responses, leukocyte trafficking, angiogenesis, and other biological processes related to the migration and activation of cells. Known chemokines are typically assigned to one of four subfamilies (α , β , γ , and δ) based on the arrangement of cysteine motifs. A nonexhaustive list of chemokines is
20 described in Table 2.

The term “receptor-bindable portion” when referring to a polypeptide ligand, refers to the portion of the ligand, often less than full-length naturally occurring protein) sufficient for binding to a cognate receptor (i.e., a receptor that binds the naturally occurring ligand).

25

2. INTRODUCTION

The present invention provides reagents, devices, and methods useful for analysis of protein-protein interactions such as receptor-ligand interactions. The invention provides reagents and methods for identification of modulators of such
30 receptor-ligand interactions, and other methods, as described herein. In one embodiment, the invention is used to identify and characterize interactions between chemokines, e.g., human chemokines, and known or orphan chemokine receptors.

In an aspect of the invention, as described in detail *infra*, cells

expressing receptors are contacted with one or more tethered ligands and specific binding, if present, is detected and analyzed. Such contacting, detection and analysis is sometimes referred to as "interrogation." The "tethered ligands" used according to the invention are fusion proteins having at least two, and usually at least three, distinct domains: a ligand domain, a stalk domain, and, optionally, an immobilization domain. The cells may be recombinant cells expressing a predefined receptor protein (including cells expressing an orphan receptor), or may be cells isolated from a biological source (such as a tissue from a patient or non-human animal).

10 Assays in which binding of a cell expressing a receptor and the tethered ligand fusion proteins of the invention is detected may be carried out in a variety of ways.

In one embodiment, the interaction between one or more tethered ligands and one or more receptor expressing cells is detected and characterized. In a particular, the effects of one or more test compound(s) on the interaction can be assayed.

For example, tethered ligand fusion protein of the invention (*i.e.*, a single species) may be immobilized on a surface (*e.g.*, a solid surface such as a slide, plate, bead, dipstick, or the like) and contacted with a test cell or cell population.

20 In another embodiment of the invention, a plurality (*i.e.*, at least 3, typically at least 10, often at least 15) of different tethered ligands (*i.e.*, having different ligand domains) are immobilized in an array. The array can be interrogated by recombinant cells expressing receptors known or suspected of specifically binding the ligand domain of a tethered ligand. The effect of test compounds on the interaction of the cells and tethered ligands of the array can be determined. The array can also be interrogated by populations of naturally occurring cells, and the receptor profile of the population determined (for diagnostic, screening, or other purposes). Notably, the tethered arrays of the invention are useful for determining the ligand binding profile of orphan receptors.

30 The invention finds use in the characterization of a variety of protein-protein interactions. In particular, the invention is useful for the characterization of interactions between 7-transmembrane receptors (*e.g.*, chemokine receptors) and putative ligands (*e.g.*, chemokines). However, as will be apparent *infra*, the

invention is not limited to characterization of 7-transmembrane receptors.

3. TETHERED LIGANDS

As noted *supra*, the methods and devices of the invention make use of
5 “tethered ligands.” Tethered ligands are fusion proteins having a ligand domain, a
stalk domain, and, usually, an immobilization domain. The stalk domain is
positioned in between the ligand domain and the immobilization domain (i.e., it is
“intermediate”). The stalk domain sequence may be contiguous with one or both of
10 the other domains, but typically at least some additional (e.g., linker) sequences will
be interposed. Typically, the ligand domain is located at the amino-terminus of the
fusion protein, and the immobilization domain is at the carboxy-terminus (with the
stalk domain in between). Fig. 1 shows a diagram of an exemplary tethered ligand.

In alternative embodiments, the ligand domain may be located at the
carboxy-terminus of the fusion protein, and the immobilization domain at the amino-
15 terminus. The latter orientation is useful when the ligand domain has the sequence
of an intracellular protein, or intracellular domain of a transmembrane protein.

Techniques used in production of the tethered ligands of the invention
(e.g., cloning and expression of recombinant proteins, including fusion proteins) are
well known in the art and are described, for example, in Sambrook et al., 1989,
20 Molecular Cloning: A Laboratory Manual, 2nd Ed., Vols. 1-3, Cold Spring Harbor
Laboratory and Ausubel et al., Current Protocols In Molecular Biology, Greene
Publishing and Wiley-Interscience, New York, including 1999 and 2000
supplements. For example, it is understood that the various (e.g., at least 2) amino
acid coding segments used to encode the fusion protein will be cloned in frame
25 relative to each other, to encode the fusion protein.

The structure and production of tethered ligands will now be described
in greater detail.

3.1 Ligand Domain

30 The ligand domain of tethered ligands has the sequence of a protein or
peptide ligand of interest, which may be an extracellular ligand or an intracellular
ligand.

Extracellular ligands are those polypeptide ligands that are naturally
bound by extracellular domains of membrane associated receptors; exemplary

extracellular ligands include chemokines (bound by chemokine receptors); other cytokines, interferon (bound by interferon receptors), dopamine (bound by dopamine receptors). Exemplary extracellular receptors are listed in Table 1, although many others will be apparent to those of skill. A particularly useful class of tethered ligands are those having a ligand domain corresponding to a chemokine. A tethered ligand having a chemokine domain is sometimes referred to as a "stalkokine," e.g., "TARC-stalkokine" for a tethered ligand having a TARC sequence in the ligand domain.

Chemokines and cytokines are well known in the art. Exemplary cytokines are listed in Table 2. References describing these and other cytokines are provided in the R&D Systems Catalog (1999) and (2000) R&D Systems Inc., 614 McKinley Place N.E. MN 55413, the R&D online catalog at <http://www.rndsystems.com> (e.g., October 10, 1999), both of which are incorporated by reference for all purposes; the CFB (Cytokine Facts Book, 1994, Academic Press Ltd.), Chemokine Facts Book, 1997, Academic Press Ltd., incorporated by reference for all purposes, and the GenBank protein sequence database (<http://www.ncbi.nlm.nih.gov/entrez/query.fcgi>).

It will be appreciated that a ligand domain need not necessarily include the complete sequence of the intracellular or extracellular ligand to which it corresponds; in various embodiments, the ligand domain will comprise at least a receptor-bindable portion of the naturally occurring ligand.

Table 1

Exemplary Receptors

Receptor Class	Subclasses & Examples	GenBank Accession Number
Chemokine Receptors	1. CCR1 2. CCR2 3. CCR3 4. CCR4 5. CCR5 6. CCR6 7. CCR7 8. CCR8 9. CCR9 10. CCR10 11. CCR11 12. CXCR1 (IL-8R alpha) 13. CXCR2 (IL-8R beta) 14. CXCR3 (GPR9)	NM001295 NM000647 NM001837 AB023892 NM000579 NM004367 NM001838 NM005201 NM006641 AF233281 NM016602 L19591 NM001557 NM001504

	<p>15. CXCR4 16. CXCR5 17. XCR1 18. CX₃CR1 19. CMV US28 20. HHV8ORF74 21. Duffy Ag</p>	<p>NM003467 AJ002211 NM005283 U20350 L20501 AF179931 AF030521</p>
<p>Other Receptor Classes</p>	<p>Peptide Angiotensin Angiotensin type 1 Angiotensin type 2 Bombesin Bradykinin C5a anaphylatoxin Fmet-leu-phe Interleukin-8 Interleukin-8 type A Interleukin-8 type B Endothelin Melanocortin Melanocyte stimulating hormone Adrenocorticotrophic hormone Melanocortin hormone Neuropeptide Y Neurotensin Opioid Opioid type D Opioid type K Opioid type M Opioid type X Somatostatin Somatostatin type 1 Somatostatin type 2 Somatostatin type 3 Somatostatin type 4 Somatostatin type 5 Tachykinin Substance P (NK1) Substance K (NK2) Neuromedin K (NK3) Thrombin Vasopressin-like Vasopressin Oxytocin Galanin Proteinase activated Orexin Chemokine/chemotactic factors like Hormone protein Follicle stimulating hormone Lutropin-choriogonadotropic hormone Thyrotropin Viral Class A Orphan/other Burkitt's lymphoma receptor Mas proto-oncogene RDC1 EDG GPR</p>	

	RBS11 BONZO Chemokine receptor-like 2 G10D GP40 like APJ like
	Growth Factor Receptors Hormone Receptors Class I cytokine receptors (hematopoietin family receptors)
	IL-6 (sharing gp 130) IL-6 IL-11 CNTF LIF Oncostatin M Cardiotrophin 1
	GM-CSF (sharing β chain) GM-CSF IL-3 IL-5
	IL-2 (sharing γ chain) IL-2 IL-4 IL-7 IL-9 IL-15
	IL-13 (sharing α chain) IL-13 IL-14
	IL-12 G-CSF Erythropoietin Growth hormone Prolactin
	IFN- α/β family IFN- γ IL-10
	TNF- α TNF- β (LT- α) LT- α / LT- β heteromes NGF Fas Ligand CD40 ligand TRAIL
	IL-1 α IL-1 β IL-1 receptor antagonist
	TGF- β Bone morphogenetic proteins

Table 2
Exemplary Cytokines

1. TNF-alpha	39. Activins	78. IL-13
2. CD27L	40. TRAIL	79. TWEAK
3. TGF-beta	41. TALL-1	80. GITRL
4. TNF-beta	42. IL-15	81. Lymphotactin
5. CD30L	43. IL-5	82. BRAK
6. IL-2	44. IL-1beta	83. PARC
7. VEGF (VEGF-A)	45. IL-10	84. Eotaxin
8. EGF	46. IGF-II	85. Eotaxin-2
9. HB-EGF	47. MSP	86. MIPF-1
10. IL-6	48. FGF-basic	87. HCC-1
11. SCF	49. BDNF	88. HCC-4
12. BMPs	50. Erythropoietin	89. SCYA26
13. Lymphotoxin-beta	51. PDGF-AB	90. MIP-1alpha
14. CD40L	52. VEGF-D	91. MIP-1beta
15. IL-7	53. Betacellulin	92. MIP-1delta
16. VEGF-B	54. G-CSF	93. I-309
17. TGF-alpha	55. CNTF	94. 6-Ckine
18. SMDF	56. Midkine	95. RANTES
19. LIF	57. Inhibins	96. MCP-1
20. Flt-3 Ligand	58. TRANCE	97. MCP-2
21. GDNFs	59. 4-1BBL	98. MCP-3
22. Fas Ligand	60. IL-4	99. MCP-4
23. April	61. GM-CSF	100. TARC
24. IL-9	62. IL-18	101. MIP-3alpha
25. IL-3	63. IL-17	102. MIP-3beta
26. IL-1alpha	64. IL-16	103. TECK
27. IFN-gamma	65. IL-12	104. MDC
28. IGF-I	66. FGF-3-19	105. IL-8
29. HGF	67. Neurotrophins	106. GCP-2
30. FGF-acidic	68. Angiopoietins (1-4)	107. MIG
31. NGF	69. PDGF-BB	108. SDF-1
32. Thrombopoietin	70. PlGF	109. I-TAC
33. PDGF-AA	71. Heregulins	110. PF4
34. VEGF-C	72. Leptin	111. ENA-78
35. Amphiregulin	73. Oncostatin M	112. IP-10
36. IL-11	74. Pleiotrophin	113. NAP-2
37. Cardiotrophin-1	75. MIS	114. GRO
38. M-CSF	76. LIGHT	115. BLC/BCA-1
	77. OX40L	

In some alternative embodiments, the "receptor" is not the extracellular domain of a membrane associated receptor, but a different protein binding moiety, such as the protein-binding domain of an intracellular protein or protein domain. For

example, the “receptor-ligand pair” used in the invention can be the intracellular domain of a CCR1 (“receptor”) and a G-protein (“ligand”). In this case, the “ligand” is the G-protein domain and is expressed as a fusion protein with a stalk domain (e.g., a fractalkine mucin region sequence). The “receptor” is the intracellular domain of CCR1, that can be expressed as an amino-terminal fusion protein with a single transmembrane receptor (such as EGF receptor with a deleted kinase domain). This allows the CCR1 intracellular domain, for example, to be expressed extracellularly (i.e. on the surface of a cell). Thus, by immobilizing the G-protein tethered ligand onto a solid surface one can interrogate the cell expressing the CCR1 intracellular domain. Another example of an intracellular ligand (a protein that binds an intracellular protein, or intracellular protein domain), is a G protein that binds a G-protein coupled glutamate receptor.

3.2 Stalk Domain

The stalk domain of the tethered ligand fusion protein functions, in part, to display the ligand domain by elevating it a considerable distance above the substrate to which the tethered ligand fusion protein is immobilized. Without intending to be bound by a particular mechanism, it is believed that displaying the ligand a significant distance above the surface increases binding the ligand with the receptor displayed in receptor-expressing cells.

Typically, the length of the stalk domain of the fusion protein is at least about 50 amino acid residues at least about 75 amino acid residues, at least about 100 amino acid residues, often at least about 150 residues, and frequently at least about 200 residues. Typically, the stalk domain is between about 200 residues and 500 residues, and usually between about 200 residues and 300 residues. Generally, the ligand domain of the polypeptide is displayed at least about 20nm (e.g., between about 20 nm and about 60 nm), or at least about 30 nm away from (i.e., above) the surface to which the tethered ligand is immobilized. In embodiments, the ligand domain peptide is displayed at between around 30–40 nm above the surface. The measurement of the extension of the ligand domain is typically done by electron microscopy after heavy metal shadowing (see, e.g., Fong et al, 2000, *J Exp Med* 188:1413-19).

In addition to elevating the ligand domain away from the surface, the flexibility of the stalk region is believed to permit the ligand domain to adopt a variety

of orientations, increasing the likelihood of a strong interaction with the receptor, and having unexpected advantages compared to other methods of immobilizing or displaying a ligand, such as standard ELISA. Typically, the stalk domain is selected to have sufficient rigidity to elevate the ligand but has flexibility to allow the ligand domain to adopt a variety of orientations.

In one embodiment, the stalk domain is derived from the fractalkine polypeptide (Bazan et al., 1997, *Nature* 385:640-644; See also, WO 97/27299). Fractalkine is a naturally occurring type 1 membrane protein containing a chemokine domain tethered on a long mucin-like stalk. Human fractalkine cDNA (Genbank Accession No. U84487) encodes a 397 residue membrane protein with a 24 residue predicted signal peptide, a 76 residue chemokine domain, a 241 residue stalk region containing 17 degenerate mucin-like repeats rich in motifs for O-glycosylated serine and threonine residues, a 19 residue transmembrane segment and a 37 residue cytoplasmic domain.

Thus, in exemplary embodiments, the stalk region has the sequence provided in Table 3A or 3B. In other embodiments, the stalk region includes at least one mucin repeat segment from a fractalkine stalk region, such as the human sequence shown in Table 4. In related embodiments, the stalk region has at least 10, at least 25, or at least 50 contiguous residues of a fractalkine stalk region, such as that shown in Table 3A or 3B, (e.g., subsequences or interactions thereof).

Table 3A

Human Fractalkine Mucin-Repeat Region

IGE VKPRTTPAAGGMDES VVLEPEATGESSSLEPTPSSQEAQRALGTSP ELPTGVT
 25 GSSGTRL PPTPKAQDGGPVGTELFRVPPVSTAATWQSSAPHQPGPSLWAEAKTS
 EAPSTQDPSTQASTASSPAPEENAPSEGQRVWGQGQSPRPENSLEREEMGPVP
 AHTDAFQDWGPGSMAHVSVVPVSSEGTPSREPVASGSWTPKAE EPIHATMDPQR
 LGVLITPVP (SEQ ID NO:3)

Table 3B

Human Fractalkine Mucin-Repeat Sequence Residues 100-336

GGTFEKQIGE VKPRTTPAAGGMDES VVLEPEATGESSSLEPTPSSQEAQRALGT
 30 PELPTGVTGSSGTRL PPTPKAQDGGPVGTELFRVPPVSTAATWQSSAPHQPGPSL
 WAEAKTSEAPSTQDPSTQASTASSPAPEENAPSEGQRVWGQGQSPRPENSLERE
 35 EMGPVPAHTDAFQDWGPGSMAHVSVVPVSSEGTPSREPVASGSWTPKAE EPIHA
 TMDPQRLGVLITPVPDAQA (SEQ ID NO:4)

Table 4
Mucin Repeat Domains

5	1. IGEVKPRTTP	(SEQ ID NO:5)
	2. GGMDSESVLEP	(SEQ ID NO:6)
	3. TGESSSLEPTP	(SEQ ID NO:7)
	4. LGTSPELPTG	(SEQ ID NO:8)
	5. TGSSGTRLPPTP	(SEQ ID NO:9)
	6. VGTELFVRVPPVS	(SEQ ID NO:10)
	7. AATWQSSAPHQ	(SEQ ID NO:11)
10	8. PGPSLWAEAKTS	(SEQ ID NO:12)
	9. EAPSTQDPST	(SEQ ID NO:13)
	10. QASTASSPAP	(SEQ ID NO:14)
	11. VWGQGQSPRP	(SEQ ID NO:15)
	12. SLEREEMGPVP	(SEQ ID NO:16)
15	13. AHTDAFQDWG	(SEQ ID NO:17)
	14. PGSMAHVSVVP	(SEQ ID NO:18)
	15. EGTPSREPVA	(SEQ ID NO:19)
	16. SGSWTPKAEEP	(SEQ ID NO:20)
20	17. QRLGVLITPVP	(SEQ ID NO:21)

In related embodiments, the stalk region of the tethered ligand has a sequence from a fractalkine of a non-human species such as mouse (see, Lloyd et al, 1997, *Nature* 387:611-617; Genbank Accession No. AF010586) and other mammals (e.g., porcine, bovine, ovine, rat, rabbit, and non-human primate mammals).

In a related embodiment, the stalk domain is derived from other mucin family members, such as MUC-type mucins. MUC-type mucins are a family of structurally related molecules that are heavily glycosylated and are expressed in epithelia of the respiratory, gastrointestinal, and reproductive tracts, e.g., MUC1 (GenBank accession number AF125525), MUC2 (L21998), MUC3 (AF113616), MUC4 (AJ000281), MUC5AC (U83139), MUC5B (AJ001402), MUC6 (U97698), MUC7 (L13283), MUC8 (U14383), MUC9 (oviductin) (AW271430). In other embodiments, the stalk domain has sequences from MAdCAM-1, GlyCAM-1, CD34 (See, e.g., Girard & Springer 1995; Van Klinken et al., 1988, *Anal Biochem* 265:103-16), consensus repeats from E- selectin, P- selectin, or L-selectin, or viral glycoprotein spikes (e.g., glycoproteins of viral origin, such as influenza, herpes simplex, human immunodeficiency, or Tobacco mosaic virus) In particular, the influenza virus neuraminidase protein (accession number 091744), and especially the hypervariable stalk region from amino acids 36 through 90 inclusive (i.e., comprising

HFKQYECSSPPNNQVIPCQPTIIERNITEIVYLTNTTIEKEICPKLVEYRNWSKP (SEQ ID NO:22) and concatenations thereof), are useful for a stalk to display immobilized ligands:

5 Additional stalk sequences can be tested (for suitability for use in the methods of the invention) using the assay described in Example 1, *infra*, but substituting the new stalk sequence for the fractalkine domain of Example 1. Briefly, to identify new suitable stalk region sequence, sequences encoding potential display stalks are cloned into an expression vector [e.g., pcDNA3.1/Myc-His(-)B (Invitrogen™ Corp, Carlsbad CA) or other suitable vector (e.g., pcDNA3.1/Myc-His(-)A/C (Invitrogen™ Corp.) or a similar vector], for example as an *EcoRI*-linked PCR
10 fragment to form a fusion protein having an ELC ligand domain at the amino terminus and a 6xHis sequence at the carboxy-terminus. That is, the stalk-encoding sequence is inserted upstream in an analogous position as was used for fractalkine domain of the ELC-stalkokine (i.e., between an ELC binding motif and a poly-
15 histidine immobilization domain. The resulting plasmid is expressed in mammalian cells (e.g., 293 cells) and the tethered ligand protein expressed, e.g., as described in Example 1. The tethered ligand is then assayed for ability to mediate binding of cells expressing CCR10, as described *infra*. Variations of this assay will be apparent to those of skill in the art.

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3.3 Immobilization Domain

In various embodiments, the fusion protein has an immobilization domain to facilitate immobilization of the protein to a solid substrate. Often, immobilization domain is a short (i.e., fewer than 10 residues) epitope tag (i.e., a
25 sequence recognized by an antibody, typically a monoclonal antibody) such as polyhistidine (Bush et al, 1991, *J. Biol Chem* 266:13811-14), SEAP (Berger et al, 1988, *Gene* 66:1-10), or M1 and M2 flag (see, e.g, U.S. Pat. Nos. 5,011,912; 4,851,341; 4,703,004; 4,782,137). In some embodiments of the invention, the tethered ligand does not have a separate immobilization domain. Instead, the stalk
30 domain is directly conjugated to the substrate, the stalk domain is immobilized via an anti-stalk (i.e., mucin) sequence antibody, or via some other immobilization method.

3.4 Production of Tethered Ligands

As noted *supra*, methods for constructing and expressing fusion proteins are well known. Fusion proteins generally are described in Ausubel et al., *supra*, Kroll et al., 1993, *DNA Cell. Biol.* 12:441, and Imai et al., 1997, *Cell* 91:521-530.

The tethered ligand fusion proteins of the invention are typically made by (1) constructing a vector (e.g., plasmid, phage or phagemid) comprising a polynucleotide sequence encoding the desired polypeptide, (2) introducing the vector into a suitable expression system (e.g., a prokaryotic, insect, mammalian, or cell free expression system), (3) expressing the fusion protein and (4) optionally purifying the fusion protein.

(1) In one embodiment, expression of the tethered ligand fusion protein comprises inserting the coding sequence into an appropriate expression vector (i.e., a vector that contains the necessary elements for the transcription and translation of the inserted coding sequence required for the expression system employed, e.g., control elements including enhancers, promoters, transcription terminators, origins of replication, a suitable initiation codon (e.g., methionine), open reading frame, and translational regulatory signals (e.g., a ribosome binding site, a termination codon and a polyadenylation sequence. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, can be used.

The coding sequence of the tethered ligand fusion protein includes ligand, stalk and immobilization domains as described elsewhere herein. Polynucleotides encoding the amino acid sequence for each domain can be obtained in a variety of ways known in the art; typically the polynucleotides are obtained by PCR amplification of cloned plasmids, cDNA libraries, and cDNA generated by reverse transcription of RNA, using primers designed based on sequences determined by the practitioner or, more often, publicly available (e.g., through GenBank). Typically, the primers include linker regions (e.g., sequences including restriction sites) to facilitate cloning and manipulation in production of the fusion construct. The polynucleotides corresponding to the ligand, stalk and immobilization regions are joined in-frame to produce the fusion protein-encoding sequence.

The tethered ligand proteins of the invention may be expressed as

secreted proteins or as nonsecreted proteins. Preferably, the fusion proteins are secreted. When the naturally occurring ligand includes a signal peptide (e.g., chemokine ligands) secretion is easily achieved by including the signal sequence encoding DNA in the fusion gene. Alternatively, and in the case of ligands not naturally secreted, a heterologous or artificial signal peptide is included in the fusion protein (see, e.g., Lui et al, 1993, *PNAS USA*, 90:8957-61).

(2) The tethered ligand fusion protein vectors may be introduced into a cell (e.g., bacterial, yeast, insect, and mammalian cells) by a variety of methods. The nucleic acid expression vectors (typically dsDNA) of the invention can be transferred into the chosen host cell by well-known methods such as calcium chloride transformation (for bacterial systems), electroporation, calcium phosphate treatment, liposome-mediated transformation, injection and microinjection, ballistic methods, virosomes, immunoliposomes, polycation:nucleic acid conjugates, naked DNA, artificial virions, fusion to the herpes virus structural protein VP22 (Elliot and O'Hare, *Cell* 88:223), agent-enhanced uptake of DNA, and other methods known in the art. See Ausubel, *supra*.

(3) A variety of expression systems suitable for expression of tethered ligands are known in the art. Useful bacterial expression systems include *E. coli*, bacilli (such as *Bacillus subtilis*), other enterobacteriaceae (such as *Salmonella*, *Serratia*, and various *Pseudomonas* species) or other bacterial hosts (e.g., *Streptococcus cremoris*, *Streptococcus lactis*, *Streptococcus thermophilus*, *Leuconostoc citrovorum*, *Leuconostoc mesenteroides*, *Lactobacillus acidophilus*, *Lactobacillus lactis*, *Bifidobacterium bifidum*, *Bifidobacterium breve*, and *Bifidobacterium longum*). The tethered ligand fusion protein expression constructs useful in prokaryotes include recombinant bacteriophage, plasmid or cosmid DNA expression vectors, or the like, and typically include promoter sequences. Illustrative promoters include inducible promoters, such as the lac promoter, the hybrid lacZ promoter of the Bluescript7 phagemid (Stratagene, La Jolla CA) or pSport1 (Gibco BRL); phage lambda promoter systems; a tryptophan (*trp*) promoter system; and *ptrp-lac* hybrids and the like. Bacterial expression constructs optionally include a ribosome binding site and transcription termination signal regulatory sequences. Illustrative examples of specific vectors useful for expression include, for example, pTrcHis2, (Invitrogen, San Diego CA),

When tethered ligand fusion protein is expressed in yeast, a number of

suitable vectors are available, including plasmid and yeast artificial chromosomes (YACs) vectors. The vectors typically include expression control sequences, such as constitutive or inducible promoters (e.g., such as alpha factor, alcohol oxidase, PGH, and 3-phosphoglycerate kinase or other glycolytic enzymes), and an origin of replication, termination sequences and the like, as desired. Suitable vectors for use in *Pichia* include pPICZ, His6/pPICZB, pPICZalpha, pPIC3.5K, pPIC9K, pA0815, pGAP2A, B & C, pGAP2alpha A, B, and C (Invitrogen, San Diego, CA) and numerous others known in the art or to be developed. In one embodiment, the vector His6/pPICZB (Invitrogen, San Diego, CA) is used to express a His6-tethered ligand fusion protein fusion protein in the yeast *Pichia pastoris*. An example of a vector useful in *Saccharomyces* is pYES2 (Invitrogen, San Diego, CA).

Another expression system provided by the invention for expression of tethered ligand fusion protein is an insect system. A preferred system uses a baculovirus polyhedrin promoter. In one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia larva*e. The sequence encoding the gene of interest may be cloned into a nonessential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the sequence, e.g., encoding the tethered ligand fusion protein, will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses are then used to infect *S. frugiperda* cells or *Trichoplusia larva*e, in which the tethered ligand fusion protein sequence is then expressed (see, for general methods, Smith et al., 1983, *J. Virol.*, 46:584; Engelhard et al., 1994, *Proc. Natl. Acad. Sci.* 91:3224-7). Useful vectors for baculovirus expression include pBlueBacHis2 A, B & C, pBlueBac4.5, pMelBacB and numerous others known in the art or to be developed. Illustrative examples of tethered ligand fusion protein expression constructs useful in insect cells are provided in Example 6, *infra*.

The present invention also provides expression systems in mammals and mammalian cells. As noted *supra*, tethered ligand fusion protein polynucleotides may be expressed in mammalian cells (e.g., human cells) for production of significant quantities of tethered ligand fusion protein polypeptides are well known in the art and include those described *infra* §5.1.1

Optimum expression times are conveniently determined by analysis of the secreted protein by Western blot analysis (e.g., using a common epitope tag, e.g., poly histidine) for detection.

(4) It is sometimes desirable to purify a secreted tethered ligand fusion protein from cell media, and generally necessary to purify nonsecreted proteins prior to use. Purification can be carried out using a variety of methods, including, for example, metal-chelate affinity chromatography of fusion proteins containing polyhistidine tracts (e.g. His₆), protein A domains or fragments (which allow purification on immobilized immunoglobulin), and the domain utilized in the
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FLAGS extension/affinity purification system (Immunex Corp, Seattle WA).

The fusion proteins may be stored under any suitable conditions (e.g., frozen) and may to preserve integrity, e.g., stored Several stalkokines were seen to be sensitive to long term storage and repeated freeze thaws. To avoid excessive degradation the protein solutions were aliquoted and frozen at -20°C or -80°C for
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long term storage. Addition of protease inhibitors to the supernatants containing the stalkokines reduced the level of degradation and enhanced quality.

3.4.1 Activity Assays

If desired, the fusion protein can be tested to confirm that it retains the
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biological activity of the isolated ligand. For example, in the case of a chemokine ligand, as described in the Examples, standard binding and chemotaxis assays may be carried out using the tethered ligands (e.g., prior to immobilization).

4. IMMOBILIZATION AND PREPARATION OF ARRAYS

4.1 Immobilization on Substrates

In some embodiments, the tethered ligands (e.g., stalkokines) of the invention are immobilized on a solid surface. The substrate to which tethered ligand is bound may be in any of a variety of forms, e.g., a microtiter dish, a test tube, a dipstick, a microcentrifuge tube, a bead, a spinnable disk, and the like. Suitable materials include glass, plastic (e.g., polyethylene, PVC, polypropylene, polystyrene, and the like), protein, paper, carbohydrate, and other solid supports. Other materials that may be employed include ceramics, metals, metalloids, semiconductive materials, cements and the like.

As noted *supra*, in some embodiments, the fusion proteins are organized as an array. The term "array," as used herein, refers to an ordered arrangement of immobilized fusion proteins, in which particular different fusion proteins (i.e., having different ligand domains) are located at different predetermined sites on the substrate. Because the location of particular fusion proteins on the array is known, binding of receptor-bearing cells to that location can be correlated with specific binding of the receptor or receptors displayed by the cell and the ligand (e.g., chemokine) domain of the fusion protein.

Immobilization of fusion proteins on beads (individually or in groups) is another particularly useful approach. In one embodiment, individual tethered ligand fusion proteins are immobilized on beads, allowed to bind to cells expressing cognate receptors, and the resulting complex is separated from unbound cells. In one embodiment, mixtures of distinguishable beads are used. Distinguishable beads are beads that can be separated from each other on the basis of a property such as size, magnetic property, color (e.g., using FACS) or affinity tag (e.g., a bead coated with protein A can be separated from a bead not coated with protein A by using IgG affinity methods). According to one embodiment, each distinguishable bead is associated with a species or particular combination of ligand fusion proteins. Cell binding to particular tethered ligand fusion proteins may be determined; similarly, the effect of test compounds (i.e., agonists and antagonists of binding) may be determined.

4.2 Substrates and Methods for Immobilization

Methods for immobilizing proteins are well known in the art, and

include covalent and non-covalent methods. It will be appreciated that the choice of method will depend in part on the substrate and detection system selected.

4.2.1 Non-covalent immobilization

5 One suitable immobilization method is antibody-mediated immobilization. According to this method, an antibody specific for the sequence of an "immobilization domain" of the tethered ligand is itself immobilized on the substrate (e.g., by adsorption). One advantage of this approach is that a single antibody may be adhered to the substrate and used for immobilization of a number
10 of different tethered ligands (sharing the same immobilization domain). For example, an immobilization domain consisting of poly-histidine (Bush et al, 1991, *J. Biol Chem* 266:13811-14) can be bound by an anti-histidine monoclonal antibody (R&D Systems, Minneapolis, MN); an immobilization domain consisting of secreted alkaline phosphatase ("SEAP") (Berger et al, 1988, *Gene* 66:1-10) can be bound by
15 anti-SEAP (Sigma Chemical Company, St. Louis, MO); an immobilization domain consisting of a FLAG epitope can be bound by anti-FLAG. Other ligand-antiligand immobilization methods are also suitable (e.g., an immobilization domain consisting of protein A sequences (Harlow and Lane, 1988, *ANTIBODIES A LABORATORY MANUAL*, Cold Spring Harbor Laboratory; Sigma Chemical Co., St. Louis, MO) can be bound
20 by IgG; and an immobilization domain consisting of streptavidin can be bound by biotin (Harlow & Lane, *supra*; Sigma Chemical Co., St. Louis, MO).

When antibody-mediated immobilization methods are used, glass is an especially useful substrate (e.g., microscope quality glass slides). When arrays are desired, the glass substrates may be printed with a hydrophobic (e.g., Teflon) mask
25 to form wells. Preprinted glass slides with 3, 10 and 21 wells per 14.5 cm² slide "working area" are available from, e.g., SPI Supplies, West Chester, PA; also see U.S.Pat. No. 4,011,350). In certain applications, a large format (12.4 cm x 8.3 cm) glass slide printed in a 96 well format is used; this format facilitates the use of automated liquid handling equipment and utilization of 96 well format plate readers of
30 various types (fluorescent, colorimetric, scintillation). However, higher densities may be used (e.g., more than 1 tethered ligand per cm²). Often the tethered ligand array will include at least about 3, at least about 5, at least about 10, at least about 15, at least about 20 or more different ligands.

Typically, antibodies are bound to substrates (e.g., glass substrates) by adsorption. Suitable adsorption conditions are well known in the art and include incubation of 0.5-50 ug/ml (e.g., 10 ug/ml) mAb in buffer (e.g., PBS, or 50 to 300 mM Tris, MOPS, HEPES, PIPES, acetate buffers, pHs 6.5 to 8, at 4°C to 37°C and from
5 1hr to more than 24 hours.

Antibody-mediated immobilization may provide certain advantages over other methods for immobilization, due to the flexibility of the antibody hinge region, allowing the ligand domain to adopt additional orientations.

Usually, the fusion protein is immobilized prior to contacting with
10 receptor-expressing cells, however, in some embodiments, the immobilization follows the contacting step.

4.2.2 Covalent immobilization

As noted *supra*, the tethered ligand may be covalently bound or
15 noncovalently attached through nonspecific bonding. If covalent bonding between a the fusion protein and the surface is desired, the surface will usually be polyfunctional or be capable of being polyfunctionalized. Functional groups which may be present on the surface and used for linking can include carboxylic acids, aldehydes, amino groups, cyano groups, ethylenic groups, hydroxyl groups,
20 mercapto groups and the like. The manner of linking a wide variety of compounds to various surfaces is well known and is amply illustrated in the literature.

5. INTERROGATION OF STALKOKINES AND STALKOKINE ARRAYS

According to the methods of the invention, cells expressing receptors
25 (e.g., chemokine receptors) are contacted with one or more tethered ligand fusion proteins (e.g., individual ligands or an array of different tethered ligands) under conditions in which binding between the ligand and cognate receptor (if present) occurs.

Typically, the binding conditions are physiological conditions under
30 which receptor-ligand interactions occur or are expected to occur. Thus, in one embodiment, the cells are contacted with the tethered ligand in a buffered solution (e.g., PBS, TBS, etc).

Binding is typically allowed to proceed for between 0.5 and 10 h (typically between 0.5 and 3 h, e.g., 1.5 h) at 14°C to 37°C (e.g., room temperature),

with or without gentle agitation. Preferred cell concentrations for binding range from 0.5-5x 10⁶/ml (e.g., 1-2 x 10⁶/ml), but concentrations outside this range may be used (e.g., in the case of rare cells from a patient sample).

5 In some embodiments, the cells are pretreated prior to incubation with the array or individual tethered ligands. For example, cells may be decorated with fluorescently labeled antibodies (e.g., anti-CD3 cell marker) or labeled intracellularly with fluorescent dyes such as Calcein (Molecular Probes, Eugene, OR), or labeled using radioisotopes. Such labeling facilitates quantitation of cell binding to tethered ligands (e.g., sites on an array) as well as characterization of the binding cells. For
10 example, in an experiment analyzing chemokine receptors expressed on T lymphocytes, PBMCs may be isolated and T cells stained for with fluorescein labeled anti-CD4 and rhodamine labeled anti-CD8. Typically the cells are stained for 30 minutes, washed 3x in PBS, and resuspended at 1x10⁶ cells per ml in PBS. The cells are then contacted with a tethered ligand array described herein, and allowed
15 to bind at room temperature for 1 hour. The slide is then washed to remove unbound cells, and the adherent cells detected using a fluorescent plate reader. The fluorescein tag and the rhodamine tag are separately detected, with co-localization identifying T cells.

By analysis of the tethered chemokine to which each cell type is
20 bound, and the ratio of the two signals, the cell type being localized at a particular tethered ligand may be determined (i.e., the cell may be characterized and the cells quantitated). It will be apparent that by staining a heterogeneous population of cells (e.g., from a patient with multiple sclerosis or rheumatoid arthritis) subpopulations of cells expressing particular receptors, or combinations of receptors, can be identified,
25 e.g., for diagnostic purposes. Similarly, this protocol is useful in drug development (e.g., screening for compounds that alter the binding pattern of individual T cells to the tethered chemokines).

5.1 Interrogation with recombinant cells expressing receptors

30 In one embodiment, the methods of the invention are practiced using recombinant cells expressing receptors (i.e., cells transiently or stably transfected with expression vectors encoding the receptor protein of interest).

5.1.1 Cells Expression Recombinant Receptor Proteins

The recombinantly expressed receptor proteins used in the assay may be any of a variety of binding proteins, such as seven transmembrane spanning, G protein coupled receptor class (e.g., chemokine receptors), receptor tyrosine kinases (e.g., EGF receptor, insulin receptor, IGF-1 receptor, NGF receptor, PDGF receptor, M-CSF receptor, FGF receptor, VEGF receptor) and receptors for any ligand or ligand class listed in Table 2, *supra*. Exemplary receptors are shown in Table 1. Additional description of 7-transmembrane receptors, including links to sequence databases, is found at <http://swift.embl-heidelberg.de/7tm/phylo/phylo.html>. Numerous additional protein binding receptors are described in the literature and/or in GenBank. Using this information, one of ordinary skill can prepare cells expressing a variety of receptors using routine techniques.

Transformed cell lines expressing receptors of interest and/or expression vectors encoding various receptors are known (see, e.g., American Type Culture Collection, 10801 University Boulevard Manassas, VA 20110-2209; <http://www.atcc.org/>). Alternatively, provided with the DNA sequence of the gene or cDNA encoding a receptor, the receptor may be expressed using routine methods for expression of recombinant proteins. It will be understood that it usually is not necessary to express a full-length or naturally occurring receptor sequence; a portion, variant or fragment capable of binding the ligand is sufficient.

Usually, the receptor-encoding sequence of interest is cloned into a "recombinant expression cassette" expression vector and introduced (e.g., transfected) into an appropriate host cell.

Although receptors may be expressed in a variety of cell types capable of expressing a membrane-bound receptor protein (e.g., insect, and the like), the receptors used in the present invention typically are expressed in mammalian cells.

Host cells useful for receptor expression include, but are not limited to, insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus), mammalian cells expression systems and the like. See, e.g., Ausubel et al., *supra*. In mammalian host cells, a number of expression systems may be utilized, e.g., constitutive expression vectors such as pCMV-myc (Clontech, Palo Alto CA), inducible expression system such as pTRE2 (Clontech, Palo Alto CA) (using tetracycline induction), vectors such as the bicistronic vector pIRES (Clontech, Palo Alto CA) capable of driving the receptor and a drug resistance

gene, and the like. In addition, a host cell strain may be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in a specific fashion desired, e.g., modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such host cells include, but are not limited to, monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293; Graham et al., *J. Gen. Virol.* 36:59 (1977)); baby hamster kidney cells (BHK, ATCC CCL 10); CHO (ATCC CCL 61 and CRL 9618); mouse sertoli cells (TM4, Mather, *Biol. Reprod.* 23:243-251 (1980)); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL 1587); human cervical carcinoma cells (HeLa, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather, et al., *Annals N.Y. Acad. Sci.* 383:44-46 (1982)); MDCK cells (ATCC CCL 34 and CRL 6253); HEK 293 cells (ATCC CRL 1573); and WI-38 cells (ATCC CCL 75; ATCC: American Type Culture Collection, Rockville, MD). The use of mammalian tissue cell culture to express polypeptides is discussed generally in Winnacker, FROM GENES TO CLONES (VCH Publishers, N.Y., N.Y., 1987).

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express receptors may be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with the receptor encoding DNA controlled by appropriate expression control elements (e.g., promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. When appropriate, codon usage may be modified for expression in non-human cells or non-mammalian cells. Following the introduction of foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched medium, and then switched to a selective medium. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines which

express the receptor proteins on the cell surface. A number of selection systems may be used, including but not limited to, the herpes simplex virus thymidine kinase (Wigler *et al.*, 1977, *Cell* 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, 1962, *Proc. Natl. Acad. Sci. USA* 48:2026), and adenine phosphoribosyl transferase (Lowy *et al.*, 1980, *Cell* 22:817) genes which can be employed in tk⁻, hgprt⁻ or aprt⁻ cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for *dhfr*, which confers resistance to methotrexate (Wigler *et al.*, 1980, *Natl. Acad. Sci. USA* 77:3567; O'Hare *et al.*, 1981, *Proc. Natl. Acad. Sci. USA* 78:1527); *gpt*, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981), *Proc. Natl. Acad. Sci. USA* 78:2072); *neo*, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin *et al.*, 1981, *J. Mol. Biol.* 150:1); and *hygro*, which confers resistance to hygromycin (Santerre *et al.*, 1984, *Gene* 30:147). Additional selectable genes have been described, namely *trpB*, which allows cells to utilize indole in place of tryptophan; *hisD*, which allows cells to utilize histinol in place of histidine (Hartman & Mulligan, 1988, *Proc. Natl. Acad. Sci. USA* 85:8047); ODC (ornithine decarboxylase) which confers resistance to the ornithine decarboxylase inhibitor, 2-(difluoromethyl)-DL-ornithine, DFMO (McConlogue L., 1987, In: *Current Communications in Molecular Biology*, Cold Spring Harbor Laboratory ed.) and glutamine synthetase (Bebbington *et al.*, 1992, *Biotech* 10:169). Alternatively, homologous recombination into an advantageous site may be used to introduce DNA into cells (e.g., murine embryonic stem cells).

When interrogating a tethered ligand or tethered ligand array with a recombinant cell, it has been observed that better signal results when the cells are subcloned ("split") 1-2 days before the assay.

5.1.2 Characterization of Orphan Receptors

In one embodiment, the reagents and methods of the invention are useful for identifying ligands that bind to an orphan receptor. There are many putative receptors having various predicted biological functions that have no known ligands. According to the invention, cloned orphan receptors are recombinantly expressed, as described herein, and the binding of the orphan-receptor-expressing cell to a variety of tethered ligands is tested. Based on the profile of binding, a ligand or ligands can be assigned to the receptor.

Usually, one of skill is able to assign an orphan receptor to a specific class of receptors based on sequence homology or other characteristics. It will be recognized that, in such a case, the tethered ligand corresponding to the orphan receptor will usually, or initially, be interrogated with ligands known or believed to bind receptors of this class. For example, an orphan receptor with homology to a CXC chemokine receptor would initially be interrogated with ligands known to interact with CXC chemokine receptors. Similarly, an orphan receptor with homology to one or more classes of chemokine receptors will usually, or initially, be interrogated with chemokine ligands.

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5.2 Interrogation with populations of cells

In various embodiments, the invention provides methods for analyzing the receptor (e.g., chemokine receptor) profile of a population of cells. The terms "receptor profile" or "profile of receptor expression" (used interchangeably) refer to the complement of cell-surface displayed receptors (i.e., membrane bound extracellular receptors) on a cell type (e.g., cell line or purified cell type) or in a heterogeneous population of cells (e.g., a cell sample from a patient), as detected in an assay. Thus, in an assay for a set of chemokine receptors, the term "receptor profile" for a cell line will refer to the set of those chemokine receptors displayed by cells of the cell line (without regard to other receptors, e.g., growth factor receptors, displayed). Similarly, the term "receptor profile" for a heterogeneous mixture of cells refers to the set receptors displayed by the cells of the population, even though not every cell may display the same set of receptors. As is discussed *infra*, the receptor profile of a cell population can also include information about the number and/or type of cells in the population binding each of several receptors.

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Determining the receptor profile for populations of cells is useful in a variety of applications including diagnosis or prognosis of diseases in patients (or nonhuman animals, including medical models); profiling populations of individuals; assessment of the effect of a drug or treatment on a patient, and others.

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5.2.1 Diagnosis and Prognosis

Different types of cells express different extracellular receptors. Moreover, the receptors expressed by any particular cell type may vary with the developmental stage of the cell, the cell environment, and location. For example,

virally-infected cells may express different receptors than similar uninfected cells. Further, the presence or absence of cells expressing certain receptors (i.e., a cell population with a particular receptor profile) at a particular location is indicative of the state of health of an individual. Examples include the presence of leukocytes and other cells at sites of inflammation, and the presence of malignant cells in a sample.

Accordingly, the present invention provides diagnostic methods in which a population of cells is obtained from a patient suspected of suffering from a disease and the receptor profile for the population is determined using the assay methods described herein (i.e., contacting the cell population with one or more immobilized tethered ligand fusion proteins and identifying the ligands bound by the cells in the population). The receptor profile is then compared to a profile characteristic of the disease state and a profile characteristic of a healthy state, and a diagnosis is based on the comparison.

Examples of populations of cells include, without limitation, (1) cells from disease tissues and fluids, (2) cells from patient populations, (3) cells taken from patients at various times during the progression of disease or treatment. Thus, in one embodiment, cells are obtained from a tissue or fluid from an individual suffering from a disease, e.g., synovial fluid from an individual with rheumatoid arthritis; cerebral-spinal fluid from individuals with multiple sclerosis and Alzheimer's disease, bronchial alveolar lavage (BAL) fluid from patients with asthma, sarcoidosis, tuberculosis, adult respiratory distress syndrome and other inflammatory and infectious diseases, fetal cells, tissues, and biopsy (e.g., from tissues such as muscle, bone marrow, lymph node, liver, brain, and others). The reagents and methods of the invention are used to determine the population of receptors (e.g., chemokine receptors or "CKRs") expressed in the population and compared, for example, to the CKRs expressed in normal (e.g., non-disease) tissue. The normal and nondiseased tissue may be from the same subject, or from two or several different subjects. Differentially expressed CKRs define specific molecular disease targets for therapeutic intervention.

Cells may be obtained from patients suffering from any of a variety of diseases including without limitation diseases and conditions associated with inflammation, infection and cancer, e.g., (1) inflammatory or allergic diseases such as systemic anaphylaxis or hypersensitivity responses, drug allergies, insect sting allergies; inflammatory bowel diseases, such as Crohn's disease, ulcerative colitis,

ileitis and enteritis; vaginitis; psoriasis and inflammatory dermatoses such as dermatitis, eczema, atopic dermatitis, allergic contact dermatitis, urticaria; vasculitis; spondyloarthropathies; scleroderma; respiratory allergic diseases such as asthma, allergic rhinitis, hypersensitivity lung diseases, and the like, (2) autoimmune diseases, such as arthritis (rheumatoid and psoriatic), multiple sclerosis, systemic lupus erythematosus, diabetes, glomerulonephritis, and the like, (3) graft rejection (including allograft rejection and graft-vs-host disease), and (4) other diseases in which undesired inflammatory responses are to be inhibited (e.g., atherosclerosis, myositis); cancers, diseases in which angiogenesis or neovascularization play a role (neoplastic diseases, retinopathy and macular degeneration), infectious diseases and immunosuppressive diseases.

Further information about the population is obtained by quantitating and/or characterizing the cells bound to one or more of the ligands (e.g., by counting, sorting, immunostaining, and the like).

5.2.2 Drug Assessment

The reagents and methods of the invention are also useful for monitoring the effect of a drug or treatment on a patient. Cells from an individual or nonhuman animal are obtained from prior to and after treatment for a disease, or before or after onset or progression of a disease, before or after exposure to a pathogen, before and after immunization, and the like, and changes in the profile of receptor expression assayed by comparing the results. Further information about the populations is obtained by quantitating and/or characterizing the cells bound to one or more of the ligands (e.g., by counting, sorting, immunostaining, and the like). The drug assessment can be carried out repeatedly during a course of treatment.

5.2.3 Profiling populations

In another related embodiment, the methods of the invention are used for "profiling" populations of individuals. In an exemplary embodiment, it is useful to determine the receptor profile (e.g., CKR profile) of cells (e.g., blood cells or leukocytes) from specific populations of individuals in order to group the individuals into cohorts of individuals expressing a particular receptor profile, or into cohorts of individuals expressing a particular receptor (e.g., on particular cells). For example, to plan a clinical trial for a CCR3 agonist to inhibit the binding of eotaxin to CCR3

expressed on eosinophils (e.g., for treatment to retard the activity of eosinophils in asthma), it will be useful to profile a large sample of individuals in the general population for the presence of CCR3. A subset of the population deficient in eosinophil binding to eotaxin-stalkokines can be identified and excluded from the clinical trial. Similarly, CCR3 expression can be correlated with other characteristics or markers (e.g., gender, age, or genetic or ethnic variants).

5.3 Interrogation with membrane preparations

In addition to interrogating with recombinant or naturally occurring cells expressing the receptors of interest, a membrane preparation may be obtained from such cells and or cell homogenates may be used in the assays of the invention. Membrane preparations may be prepared using routine methods, e.g., hypotonic lysis and centrifugation (see, e.g., Dairaghi et al, 1999, *J. Biol. Chem.* 274:21569-74).

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6. METHODS OF DETECTION

Cell binding to tethered ligands and arrays can be detected and quantitated in a number of ways. Cells can be observed (and counted) visually (i.e., using a microscopy) with or without prior staining. Alternatively, automated and semiautomated detection systems (e.g., fluorometric imaging plate readers, scintillation counters, "96-well" plate readers) may be used to measure the signal associated with particular tethered ligands (e.g., sites on an array), particularly when the receptor bearing cells are labeled with a detectable label. As used herein, a "detectable label" has the ordinary meaning in the art and refers to an atom (e.g., radionuclide), molecule (e.g., fluorescein), or complex, that is or can be used to detect (e.g., due to a physical or chemical property), indicate the presence of a molecule or to enable binding of another molecule to which it is covalently bound or otherwise associated. Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Labels useful in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads (e.g., DynabeadsTM), fluorescent dyes (e.g., fluorescein, Texas red, rhodamine, green fluorescent protein, enhanced green fluorescent protein, lissamine, phycoerythrin, Cy2, Cy3, Cy3.5, Cy5, Cy5.5, Cy7, FluorX [Amersham],

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SyBR Green I & II [Molecular Probes], and the like), radiolabels (e.g., ^3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P), enzymes (e.g., hydrolases, particularly phosphatases such as alkaline phosphatase, esterases and glycosidases, or oxidoreductases, particularly peroxidases such as horseradish peroxidase, and others commonly used in ELISAs), substrates, cofactors, inhibitors, chemiluminescent groups, chromogenic agents, and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads. Patents teaching the use of such labels include U.S. Patent Nos. 3,817,837; 3,850,752; 3,939,350; 3,996,345; 4,277,437; 4,275,149; and 4,366,241. Means of detecting such labels are well known to those of skill in the art.

In one embodiment, the association of receptor and tethered ligand is detected using the microswitch technology described at <http://www.ece.neu.edu/edsnu/zavracky/mfl/programs/relay/relay.html>.

If desired, cells may be fixed (e.g., using cross-linking agents such as glutaraldehyde) prior to detection.

As noted *supra* it will sometimes be convenient to stain receptor-bearing cells prior to or after contacting the cells with the tethered ligands according to the invention. In one illustrative embodiment, fluorescein isothiocyanate (FITC) labeled anti CD20 antibody is used to mark B cells, and rhodamine labeled anti CD3 is used to mark T cells. Visualization and quantitation under UV illumination and the correct wavelength filter sets would discern between the different cell types.

7. DATA ANALYSIS

Modes of analysis of the binding of receptor bearing cells to the tethered ligands and arrays of the invention will be apparent to one of skill following the guidance of this disclosure. Typically, analysis of the binding of the receptor expression cell line to the tethered array is done after quantitation of cell binding by enumeration, fluorescence, or scintillation. Usually, binding is done in triplicate and the average (mean) of the value is used for analysis.

Initially a baseline of non-specific binding is determined. The measurements for the wells in which a non-binding tethered ligand (e.g. a tethered chemokine of a subclass and does not bind a test chemokine receptor) serves as an appropriate baseline. Additional appropriate baselines can be determined by the binding measured for a non-ligand containing immobilized stalk (i.e., a "tethered

ligand" lacking the ligand domain). Additionally, a mock-transfected control containing supernatant of a cell line that was processed through the transfection protocol but does not contain any plasmid capable of making the tethered ligand (such as the parental vector pcDNA3.1) may be used. A positive control also may
5 be used in some embodiments (e.g., a tethered ligand known to bind to a particular receptor-bearing cell). Suitable positive controls exhibit binding at least about two-fold higher than base line, often about 10-fold higher than base line, and frequently at least about 100-fold higher than base line.

Authentic binding (e.g. binding occurring through the transfected
10 receptor and the tethered ligand) can be verified by competition assays. In one embodiment, a soluble form of the ligand (i.e., corresponding to the ligand motif of the tethered ligand) or a soluble (non-immobilized) form of the tethered ligand is added in excess (e.g. 100 fold molar excess) to the cell line expressing the receptor for a pre-incubation period (e.g., 5 to 20 minutes, typically 15 minutes). The effect
15 on binding is determined as described supra. Authentic binding is inhibited by the addition of soluble ligand. Occasionally, the binding can be mediated through endogenous receptors being expressed on the cell line being used. Use of the parental (i.e, non-recombinant) cell line as a control in a parallel analysis is used to detect this.

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8. BINDING AGONISTS AND ANTAGONISTS

In various embodiments, the invention provides a method for identifying the ability of a test compound to modulate the interaction between a ligand and a receptor, between several ligands and a receptor, or between several
25 ligands and several receptors.

In one embodiment, the binding assays of the invention, using either an array of tethered ligands or one or more individual tethered ligands is carried out in the presence and absence of a test compound. Decreased binding of receptor-bearing cells to the tethered ligand(s) in the presence of the test compound indicates
30 that the test compound is acting as an antagonist of the interaction. Increased binding of receptor-bearing cells to the tethered ligand(s) in the presence of the test compound indicates that the test compound is acting as an agonist of the interaction. Because ligand receptor interactions have physiological effect (e.g., on cell metabolism), modulators of the interaction are expected to modulate that effect, and

such modulators are often therapeutically useful, e.g., for producing pharmaceutical compositions.

The test compounds referred to *supra* may be any of a large variety of compounds, both naturally occurring and synthetic, organic and inorganic, and including polymers (e.g., oligopeptides, polypeptides, oligonucleotides, and polynucleotides), small molecules, antibodies, sugars, fatty acids, nucleotides and nucleotide analogs, analogs of naturally occurring structures (e.g., peptide mimetics, nucleic acid analogs, and the like), and numerous other compounds. Typically, test agents are added at a variety of concentrations, e.g., ranging from about 1 ng to about 1 g/ml, more often between 1 ug/ml and 1 mg/ml. Typically, test agents are added at a variety of molar concentrations, e.g. ranging from 1 picomolar to 1 molar, more often 1 nanomolar to 1 millimolar. Typically, test agents are added at a variety of concentrations, e.g. ranging from about 1 part per billion to about 1 part per hundred, more often from about 1 part per million to about 1 part per thousand.

In one embodiment, the "test compound" is a soluble form of the ligand moiety of the tethered ligand with which the receptor interacts. For example, the chemokines ELC, TECK and SLC have been demonstrated to inhibit (i.e., by competition) the binding of a CCR10-expressing cell to a tethered ligand having an ELC ligand domain (i.e., an ELC-stalkokine). See Example 1. More often, the "test compound" is an "synthetic test compound" usually a non-polypeptide compound, i.e., not having a sequence of a naturally occurring polypeptide.

In assessing the modulatory activity of a test agent, the test agent may be preincubated with the receptor bearing cell(s) prior to contacting of the cell(s) and the tethered ligand(s); added after the contacting of the cell(s) and the tethered ligand(s); added simultaneously with the contacting of the cell(s) and the tethered ligand(s). Test agents may also be administered to animals or in vitro or ex vivo cells or tissues to assay the effects of binding of populations of cells in a particular target tissue, or animal model.

In one embodiment of the invention, the effects of test agent(s) on the interaction between a particular tethered ligand and a specific receptor are tested. This approach is useful, for example, for validating an apparent interaction between a receptor (e.g., an orphan receptor) and a specific ligand and for high-throughput screening of agents that inhibit a specific receptor-ligand interaction (thereby modulating the biological function of the receptor or ligand). Methods for high-

throughput screening are known (see, e.g., Williams, 2000, *Curr. Opin. Biotechnology* 11:42-46, and references cited therein).

In another embodiment, the arrays of the invention are used to assay the effect of test agents on the binding profile of a population of cells to a plurality of
5 ligands.

9. KITS

In one aspect the invention provides kits (e.g., containers) containing (1) a tethered ligand array of the invention, or (2) one or several of the tethered
10 ligand fusion proteins of the invention (e.g., a combination of at least 2, at least 5, or at least 10 different tethered ligands), e.g., in separate vials or (3) one, two or a plurality of the tethered ligand fusion proteins, with at least two of the tethered ligand fusion proteins immobilized different solid substrates (e.g., plates, slides, beads, or the like), e.g., different substrates (such as different beads).

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10. EXAMPLES

The following examples are offered to illustrate, but not to limit, the claimed invention.

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EXAMPLE 1

This example demonstrates the use of the invention to determine the ligand specificity of an orphan receptor.

25 A. Abbreviations

ELC, EBI1 ligand chemokine; SLC, secondary lymphoid-tissue chemokine; TECK, thymus expressed chemokine; HEK293, human embryonic kidney 293 cells; PEI, polyethylenimine; CCR, CC chemokine receptor.

30 B. Materials and Methods

Human, viral and murine recombinant chemokines were obtained from R&D Systems (Minneapolis, MN; http://cytokine.rndsystems.com/cyt_cat/cyt_cat.html). ¹²⁵I-labeled ELC and TECK were obtained from Amersham.

Full length CCR10 expression constructs were made in pIRESpuro expression vector (Clontech, Palo Alto, CA) with a FLAG epitope tag and prolactin signal sequence, and used to generate stable transfectants in HEK293 cells. Transient and stable transfections for CCR10 and stalkokines were done using Superfect reagent (Qiagen, Valencia, CA) following manufacturer's protocol. Stably-transfected cells were generated by selection in 2 ug/mL puromycin for 7 days, and expression was confirmed by FACS analysis of the FLAG epitope using anti-FLAG M1 (Sigma, St. Louis, MO) and 2' anti-mouse PE conjugate (Coulter Immunotech, Miami, FL).

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C. Stable expression of CCR10 protein.

The cloning and characterization of a new human chemokine receptor, designated "CCX CKR" or "CCR10" is described in Gosling *et al.*, 2000, "Cutting edge: identification of a novel chemokine receptor that binds dendritic cell- and T cell-active chemokines including ELC, SLC, and TECK" *J Immunol.* 164:2851-6, incorporated herein by reference in its entirety for all purposes. The coding sequence of CCR10 is shown in Fig. 2

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To assess the functional properties of the protein encoded by the CCR10 cDNA, including its potential chemokine binding profile, expression plasmids encoding CCR10 plus an added N-terminal Flag epitope were constructed. This allowed for detection and selection, using an anti-Flag mAb, of the most highly expressing stable transfectants. Human embryonic kidney 293 (HEK293) cells stably expressing the M1 flag epitope-tagged CCR10 were confirmed by FACS, and were selected for further analysis (HEK293-CCR10 cells).

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D. Receptor interrogation by adhesion to stalkokines.

To determine ligand binding to CCR10, HEK293-CCR10 cells were used to interrogate chemokine "stalkokines" (SK), i.e., molecules in which discrete chemokine domains were engineered to be tethered to the end of an extended stalk structure. Stalkokines were interrogated using 8-well chamber slides coated first with anti-His anchoring antibody (10 ug/ml in PBS overnight at RT), which were washed and 'blocked' (2% FBS/0.5% BSA in PBS); treated with 250 ul of HEK293 cell stalkokine supernatants (1 hr at 37C), and incubated with 500,000 HEK293-CCR10 transfectants (1.5 hrs at RT). Inhibition of adhesion by competition with

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soluble chemokines was done by incubating cells with 5-10 ug/ml of recombinant chemokines. In all cases, nonadherent cells were removed by washing in PBS; remaining adherent cells were fixed with 1% glutaraldehyde, photoimaged and counted. As a primary screen, this adhesion would reveal putative receptor-ligand interactions.

CCR10-expressing cells adhered very well to ELC stalkokines (ELC-SK; Fig 3A). Furthermore ELC-SK mediated adhesion was abolished in the presence of soluble native ELC as a competitor (Fig 3A, top row). A significant reduction in ELC-SK mediated adhesion of HEK293-CCR10 cells in the presence of soluble SLC, as well as soluble TECK, but not soluble MCP-3 (Fig. 3A, bottom row) was also observed. These experiments were performed and quantitated over several independent trials, an example of which is given in Fig. 3B, and were found to be highly reproducible. Moreover, radiolabeled ELC was used in a traditional homologous competition assay in the presence of increasing concentrations of unlabeled ELC. The results revealed significant binding of ELC to HEK293-CCR10 cells, but not to wildtype (wt) HEK293 cells (Fig 3C). Nearly identical results were obtained in homologous competition of radiolabeled TECK with cold TECK (not shown). Taken together, the stalkokine-based adhesion and radiolabeled ligand binding/homologous competition assays indicate that CCR10 is a new chemokine receptor that bound a novel compliment of chemokines.

The spectrum of ligands that bind to CCR10 includes ELC, SLC, and TECK with high affinity, and BLC and vMIPII with lower affinity.

EXAMPLE 2

This example describes the construction and expression of stalkokines, and the adhesion of receptor-expressing cells to immobilized stalkokines.

A. Preparation of Stalkokine expression plasmids

1. Amplification of CK Ligand Sequences

Chemokine ("CK") coding sequences were generated using the polymerase chain reaction (PCR) with oligonucleotides synthesized based on the published literature, as shown in Table 5. The amplified CK coding sequence was cloned into an expression vector encoding a human fractalkine stalk region coding

sequence and a c-terminal epitope tag (poly-histidine). Starting material for the PCR reactions included previously cloned plasmids, cDNA libraries, and cDNA generated by reverse transcription of RNA, as indicated in the table.

5 Amplification was carried out using oligonucleotides containing unique restriction endonuclease sites (EcoR1 in most case, alternatively EcoRV or SmaI) for cloning purposes. The amplified chemokine fragments included the initiation methionine codon and leader peptide sequence, resulting in a secreted product.

10 For example, a polynucleotide encoding the entire MCP2 chemokine was generated by PCR amplification. The amplification oligonucleotides each included an EcoRI site. The PCR fragment contained the regions encoding the whole, unprocessed chemokine (leader sequence plus mature protein), including the codons for the initiating methionine, leader sequence, and entire chemokine coding sequence up to, but not including the stop codon (amino acid 1 (met) to 109 (pro) inclusive, Genbank accession number X99886).

15 Following amplification of the chemokine domain sequence, the DNA fragment was subcloned into the EcoRI (or EcoRV site) of the modified vector based on pcDNA3.1, encoding a human Fractalkine mucin stalk sequence (described below). See Figure 4. The ligand motif is designed to be in frame by the construction of the oligonucleotides for the PCR synthesis of the fragment and the
20 sequence of the construct is determined by DNA sequencing.

Table 5

Chemokine	Forward Primer	Reverse Primer	Tissue Source
MDC GenBank accession number: U83171	5'-GGT GAA TTC ATG GCT CGC CTA CAG ACT GC (SEQ ID NO:23)	3'-GGT GAA TTC TTG GCT CAG CTT ATT GAG AAT CA (SEQ ID NO:24)	RT-PCR from human immature dendritic cell RNA
HCC1 GenBank accession number: Z49270	5'-GGT GAA TTC ATG AAG ATC TCC GTG GCT GCC (SEQ ID NO:25)	3'-GGT GAA TTC GTT CTC CTT CAT GTC CTT GAT ATA G (SEQ ID NO:26)	PCR from human placenta cDNA library
SLC GenBank accession number: AF001979, AB002409	5'-GGT GAA TTC ATG GCT CAG TCA CTG GCT CTG (SEQ ID NO:27)	3'-GGT GAA TTC TGG CCC TTT AGG GGT CTG TG (SEQ ID NO:28)	PCR from human EST clone
MIG GenBank Accession number NM 002416	5' GGT GAA TTC ATG AAG AAAA GT GGT GTT CTT TTC C (SEQ ID NO:29)	3' GGT GAA TTC TGT AGT CTT CTT TTG ACG AGA ACG (SEQ ID NO:30)	reverse transcription of human fetal brain mRNA
IP10 GenBank	5' GGT GAA TTC ATG AAT CAA ACT GCG ATT	3' GGT GAA TTC AGG AGA TCT TTT AGA CAT	human cDNA pool

Accession number NM 001565	CTG A (SEQ ID NO:31)	TTC CTT G (SEQ ID NO:32)	
H174 / I-TAC GenBank accession number AF002985	5' GTG GAA TTC ATG AGT GTG AAG GGC ATG GC (SEQ ID NO:33)	3' GGT GAA TTC AAA ATT CTT TCT TTC AAC TTT TTT GA (SEQ ID NO:34)	human placental cDNA library
Eotaxin1 GenBank accession number U46573	5' GGT GAA TTC ATG AAG GTC TCC GCA GCA CTT C (SEQ ID NO:35)	3' GGT GAA TTC TGG CTT TGG AGT TGG AGA TTT TTG (SEQ ID NO:36)	human EST
TARC GenBank accession number NM 002987	5' GGT GAA TTC ATG GCC CCA CTG AAG ATG CT (SEQ ID NO:37)	3' GGT GAA TTC AGA CCT CTC AAG GCT TTG CAG (SEQ ID NO:38)	reverse transcription of immature dendritic cell mRNA
MIP3alpha GenBank accession number U77035	5' GGT GAA TTC ATG TGC TGT ACC AAG AGT TTG C (SEQ ID NO:39)	3' GGT GAA TTC CAT GTT CTT GAC TTT TTT ACT GAG G (SEQ ID NO:40)	human EST
ELC GenBank accession number U77180, NM 006274	5' GGT GAA TTC ATG GCC CTG CTA CTG GCC CT (SEQ ID NO:41)	3' GGT GAA TTC ACT GCT GCG GCG CTT CAT CT (SEQ ID NO:42)	human EST
TECK GenBank accession number NM 005624	5' GGT GAA TTC ATG AAC CTG TGG CTC CTG GC (SEQ ID NO:43)	3' GGT GAA TTC CAG TCC TGA ATT AGC TGA TAT CAG (SEQ ID NO:44)	human placental cDNA library
PARC GenBank accession number AB012113	5' GGT GAA TTC ATG AAG GGC CTT GCA GCT GC (SEQ ID NO:45)	3' GGT GAA TTC GGC ATT CAG CTT CAG GTC GC (SEQ ID NO:46)	human placental cDNA library
SLC GenBank accession number AF001979	5' GGT GAA TTC ATG GCT CAG TCA CTG GCT CTG (SEQ ID NO:47)	3' GGT GAA TTC TGG CCC TTT AGG GGT CTG TG (SEQ ID NO:48)	human EST
I309 GenBank accession number AA931884	5' GGT GAA TTC ATG CAG ATC ATC ACC ACA GCC C (SEQ ID NO:49)	3' GGT GAA TTC TTT TCT TTT TGA CGG GCA GTG C (SEQ ID NO:50)	human EST
MIP1alpha GenBank accession number AA031820	5' GGT GAA TTC ATG CAG GTC TCC ACT GCT GC (SEQ ID NO:51)	3' GGT GAA TTC GGC ACT CAG CTC TAG GTC GCT (SEQ ID NO:52)	human cDNA pool
MCP-4 GenBank accession number U59808	5' GGT GAA TTC ATG AAA GTC TCT GCA GTG CTT CTG (SEQ ID NO:53)	3' GGT GAA TTC AGT CTT CAG GGT GTG AGC TTT CC (SEQ ID NO:54)	reverse transcription of human brain mRNA
MCP1 GenBank accession number AA024753	5' GGT GAA TTC ATG AAA GTC TCT GCC GCC CTT (SEQ ID NO:55)	3' GGT GAA TTC AGT CTT CGG AGT TTG GGT TTG C (SEQ ID NO:56)	human EST
MCP2 GenBank accession number NM005623	5' GGT GAA TTC ATG AAG GTT TCT GCA GCG CT (SEQ ID NO:57)	3' GGT GAA TTC TGG CTT CAG ATT TTG AAA TAT TTG (SEQ ID NO:58)	reverse transcription of immature dendritic cell mRNA
SDF1a GenBank accession number L36034	5' GGT GAATTC ATG AAC GCC AAG GTC GTG G (SEQ ID NO:59)	3' GGT GAA TTC CTT GTT TAA AGC TTT CTC CAG GT (SEQ ID NO:60)	human EST

MCP5	5' GGT GAT ATC ATG AAG ATT TCC ACA CTT CTA TGC C (SEQ ID NO:61)	3' GGT GAT ATC GCC TAG ACA TGA AGG TTC AAG GAT G (SEQ ID NO:62)	
HCC2	5' GGT GAA TTC ATG AAG GTC TCC GTG GCT GC (SEQ ID NO:63)	3' GGT GAA TTC TAT TGA GTA GGG CTT CAG CTT T (SEQ ID NO:64)	
CLONE 391	5' GGT GAA TTC ATG AAG GTC TTC TCC TTG GTC ATG (SEQ ID NO:65)	3' GGT GAA TTC CGT TGA GGT GTT GCT CAG CTT C (SEQ ID NO:66)	
BCA1	5' GGT GAT ATC ATG AAG TC ATC TCG ACA TCT CTG (SEQ ID NO:67)	3' GGT GAT ATC GGG AAT CTT TCT CTT AAA CAC TGG (SEQ ID NO:68)	
PF4a	5' GGT GAA TTC ATG AGC TCC GCAGCC GGG TTC (SEQ ID NO:69)	3' GGT GAA TTC ACT CTC CAA AAG TTT CTT AAT TAT TTT (SEQ ID NO:70)	
PBP-like1	5' GGT GAA TTC ATG CCA CCC TGC AGC TGT G (SEQ ID NO:71)	3' GGT GAA TTC TAA AGC CAT TGT GAA TAT GAT CTG (SEQ ID NO:72)	
EOTAXIN 2	5' GGT CCC GGG ATG GCA GGC CTG ATG ACC AT (SEQ ID NO:73)	3' GGT CCC GGG GCA GGT GGT TTG GTT GCC AG (SEQ ID NO:74)	
MIP1beta	5' GGT GAA TCC ATG AAG CTC TGC GTG ACT GTC C (SEQ ID NO:75)	3' GGT GAA TTC GTT CAG TTC CAG GTC ATA CAC GTA (SEQ ID NO:76)	
MIP-3	5' GGT GAA TTC ATG AAG GTC TCC GTG GCT GC (SEQ ID NO:77)	3' GGT GAA TTC ATT CTT CCT GGT CTT GAT CCG T (SEQ ID NO:78)	
MRP-1	5' GGT GAA TTC ATG AGA AAC TCC AAG ACT GCC A (SEQ ID NO:79)	3' GGT GAA TTC AGC AAT GAC CTT GTT CCC AGA T (SEQ ID NO:80)	
Lymphotactin (LYNT)	5' GGT GAA TTC ATG ATA CTT CTC ATC CTG GCC C (SEQ ID NO:81)	3' GGT GAA TTC GCC AGT CAG GGT CAC AGC TG (SEQ ID NO:82)	
HCC4	5' GGT GAA TTC ATG AAG GTC TCC GAG GCT GC (SEQ ID NO:83)	3' GGT GAA TTC CTG GGA GTT GAG GAG CTG GG (SEQ ID NO:84)	
MIP-1gamma	5' GGT GAA TTC ATG AAG CCT TTT CAT ACT GCC C (SEQ ID NO:85)	3' GGT GAA TTC TTG TTT GTA GGT CCG TGG TTG T (SEQ ID NO:86)	
IL-8	5' GGT GAT ATC ATG ACT TCC AAG CTG GCC G (SEQ ID NO:87)	3' GGT GAT ATC TGA ATT CTC AGC CCT CTT CAA A (SEQ ID NO:88)	

ENA-78	5' GGT GAA TTC ATG AGC CTC CTG TCC AGC CG (SEQ ID NO:89)	3' GGT GAA TTC GTT TTC CTT GTT TCC ACC GTC C (SEQ ID NO:90)	
GROgamma	5' GGT GAT ATC ATG GCC CAC GCC ACG CTC TC (SEQ ID NO:91)	3' GGT GAT ATC GTT GGT GCT CCC CTT GTT CAG (SEQ ID NO:92)	
GRObeta	5' GGT GAT ATC ATG GCC CGC GCC ACG CTC TC (SEQ ID NO:93)	3' GGT GAT ATC GTT GGA TTT GCC ATT TTT CAG C (SEQ ID NO:94)	
NAP-2	5' GGT GAT ATC ATG AGC CTC AGA CTT GAT ACC ACC (SEQ ID NO:95)	3' GGT GAT ATC ATC AGC AGA TTC ATC ACC TGC C (SEQ ID NO:96)	
GROalpha	5' GGT GAT ATC ATG GCC CGC GCT GCT CTC TC (SEQ ID NO:97)	3' GGT GAT ATC GTT GGA TTT GTC ACT GTT CAG CAT C (SEQ ID NO:98)	
vMIP	5' GGT GAA TTC ATG GCC CCC GTC CAC GTT TT (SEQ ID NO:99)	3' GGT GAA TTC ATG GAC ACC AAG GGC ATC CT (SEQ ID NO:100)	

B. Stalk Expression Cassette

CK ligand coding sequences were cloned into the EcoR1 or EcoRV sites of the polylinker region of the expression vector pcDNA-FRAC. This vector includes a polylinker upstream of a human fractalkine stalk coding region fused to the myc epitope and poly-histidine coding region used for antibody tethering and subsequent purification. The coding region is under control of a CMV promoter.

pcDNA-FRAC was constructed by inserting a fractalkine mucin stalk coding sequence (corresponding to amino acids 100 to 336 (inclusive) of human fractalkine (Genebank Accession number U84487)) into the EcoRI and HindIII restrictions sites of vector pcDNA3.1(-)/Myc-His version B (Invitrogen) [hereinafter, "pcDNA3.1"]. The resulting vector (pcDNA3.1-FRAC) was used for expression of various chemokine ligands. This was accomplished by inserting a PCR-amplified ligand sequence into the vector upstream of, and in frame with, the stalk domain. The resulting vectors (and the encoded proteins) are generally referred to as "chemokine'-FRAC", e.g., "MDC-FRAC", "TARC-FRAC" and the like.

A region of the multiple cloning site polylinker region from pcDNA3.1 is present in between the last amino acid of the chemokine ligand domain and the first amino acid (glycine) of the fractalkine mucin stalk region. Insertion of the PCR fragment encoding the chemokine into the polylinker puts the coding regions in frame, resulting in the proper full length translation of the fusion protein. The

polylinker region encodes various amino acids, depending on the restriction sites used for the cloning of the chemokine domain. For example, if the EcoR1 linkered chemokine domain is cloned into the EcoR1 site of the pcDNA3.1-FRAC, the linker region will encode the additional amino acids glutamate-phenylalanine. As another
5 example, if the EcoRV linkered chemokine domain is cloned into the EcoRV site, the linker region will encode the additional amino acids alanine-glutamate-phenylalanine. As yet another example, if the SmaI linkered chemokine domain is cloned into the EcoRV domain (as a blunt end ligation), the linker region will encode the additional amino acids alanine-isoleucine-proline-alanine-glutamate-phenylalanine.

10 The final plasmid is prepared in bulk using standard protocols and the Qiagen purification system to generated pure plasmid in milligram amounts. Plasmid was stored in 10 mM Tris, 1 mM EDTA at 20°C.

C. Expression of Stalkokine in cell culture

15 The stalkokine synthesis is generated in *in vitro* cell culture using HEK293 (ATCC No. CRL-1573) or 293T (Pear et al, 1993, *PNAS* 80:8392-6) cell lines transiently transfected with the stalkokine expression plasmid. These cell lines were grown in DMEM supplemented with 10% FBS containing antibiotics, at a 60% confluence prior to transfection. On the day of transfection, the cells were
20 transfected by the following protocol:

The cells were washed with PBS. 17 ug of plasmid DNA was resuspended in 825 ul of "Opti-MEM" reduced serum medium (Life Technologies, Rockville, MD)). Superfect reagent (Qiagen) was added to the plasmid, mixed, and allowed to incubate at room temperature for 20 minutes. It is then added to the cells
25 and incubated at 37°C, 5% CO₂ for 3 hours. Next, the cells are washed with PBS, and complete DMEM media containing 10% FBS and antibiotics are added. After overnight culture, the media was aspirated and replaced with DMEM with 2.5% FBS and antibiotics.

The supernatants were harvested from the transfected cells at 24, 48,
30 72. The transfected culture was supplemented with fresh media and incubated as above until the next harvest time. The supernatant was centrifuged to remove any cellular debris, and then transferred to a fresh tube. Several aliquots are made and the samples are stored at -20°C until required. In some experiments, protease inhibitor cocktails were added to the clarified supernatant prior to storage.

Purification of the stalkokines was carried out using poly histidine affinity chromatography. Large scale transfections were done and the supernatants after culture were dialyzed to remove any very small molecule contaminants. The supernatant was passed over the nickel column twice, eluted with imidazole, and the eluate was dialyzed into PBS and analyzed. Usually, significant purification was achieved with reasonable recoveries. This material was compared with the unpurified stalkokine by Western analysis, and total protein was determined. The material purified was typically 50-75% pure.

Purification is useful for a number of reasons: it removes contaminant molecules that could have a detrimental effect on stability of the stalkokine, allows easy comparison of equivalent amounts of material in assays, and facilitates quality control in preparing tethered ligand arrays.

D. Monitoring Expression By Western Blotting

The fusion protein is analyzed by Western blot analysis to confirm the expected length of the fusion protein and the presence of the poly histidine tail (which confirms the carboxy terminal is in the correct reading frame). In many cases, the amino-terminal chemokine domain is also interrogated by Western blot analysis looking with chemokine-specific antibodies which will only react with protein made in the correct frame.

The success of the cell transfection was monitored using Western blotting procedures. Typically, 10 ul of supernatant was mixed with a denaturation solution containing reducing agents (DTT) and heated to 100°C for 5 minutes. The protein sample was then loaded onto a 10-20% acrylamide Tricine gel and electrophoresed for 45 minutes at 100 V. The contents of the gel were transferred to a nitrocellulose membrane using standard transfer protocols. Transfer efficiency was determined by completeness of transfer of the prestained molecular weight markers. The membrane was blocked in a solution of Tris buffered saline, BSA and Tween-20. The membrane was then incubated for 1 hour with a primary antibody such as the anti-HIS antibody, or an antibody recognizing the chemokine domain. The primary antibody was washed off with three consecutive washes in TBS for 5 minutes with gentle agitation. The membrane was then incubated for 1 hour with the secondary antibody recognizing the Fc domain of the primary antibody. This secondary antibody has a covalently linked horseradish peroxidase enzyme

attached for subsequent visualization. After washing off the secondary antibody, the membrane was visualized by incubation with a chromagenic substrate DAB until visible bands were generated, then stopped by washing in water. See Figure 5.

5 E. Stalkokine Activity

The functional activity of the chemokine motif of an expressed stalkokine fusion protein was assessed by two methods: chemotaxis and displacement of labeled chemokines.

10 Fig. 6 shows a displacement assay showing competition by a stalkokine with binding of a radiolabeled tracer chemokine to its cognate receptor. CEM cells expressing CCR4 were used in a binding assay with radiolabeled MDC or TARC, as described in Dairaghi et al, 1999, *J. Biol. Chem.* 274:21569-74.

Briefly, CEM cells were added to radiolabeled chemokine "tracer" (MDC or TARC) in the presence of competitor molecules such as supernatants expressing
15 the soluble MDC-FRAC (a stalkokine with a fractalkine stalk and an MDC ligand domain) or TARC-FRAC (a stalkokine with a fractalkine stalk and an TARC ligand domain). The cells and tracer were incubated and harvested by filtration, capturing the bound tracer. The material was then counted. "Total" is the amount of tracer bound with no competition. "MDC-SK" is the amount of binding in the presence of
20 soluble MDC-FRAC fusion protein, "TARC-SK" is the amount of binding in the presence of soluble TARC-FRAC fusion protein, "ConSup" refers to competition with the control supernatant (conditioned media from CEM cells not expressing any stalkokine), "MDC-CK" is the competition with soluble MDC chemokine, and demonstrated the background binding.

25 As is evident, there is considerable inhibition of binding of either MDC or TARC tracer to the CEM cells in the presence of the experimental supernatants. Part of this inhibition is due to other elements present in the control sup, but there is a significant further decrease specific to the recombinant MDC-FRAC or TARC-FRAC. The MDC-CK inhibition demonstrates the background binding and the
30 maximal level of competition that one can achieve.

Chemotaxis assays were done on cell types known to possess the receptor for the chemokine ligand (Bacon et al, 1988, *Br J Pharmacol* 95:966-74).

Fig. 7 shows an assay in which CEM cells were "loaded" with INDO-1AM (Molecular Probes, Eugene, OR) and subjected to calcium flux analysis as described in Dairaghi et al, 1997, *J. Biol. Chem.* 272:28206-209.

5 Briefly, CEM cells that express CCR4 were placed in a fluorimeter and monitored over time for changes in fluorescence. Two natural ligands for CCR4 are MDC and TARC. TARC-Fractalkine or MDC-Fractalkine (fusion protein supernatants), or control supernatants, were added at the following times: experiment 1, control supernatant added at 50 sec, TARC-FRAC at 80 sec; experiment 2, control supernatant at 50 sec and MDC-FRAC at 80 sec. The control
10 supernatant did not result in any calcium influx, where as the TARC-FRAC or MDC-FRAC resulted in modest influx. This experiment demonstrates that the chemokine ligand fusion proteins are capable of causing signaling.

F. Adhesion Of Stalkokine To Solid Surfaces

15 Stalkokines were immobilized on glass and polystyrene surfaces by binding to anti-histidine mAbs. The glass substrates were in the form of slides, either 8 well chamber slides, or glass slides printed with hydrophobic Teflon mask to generate wells. Standard microscope slides sizes had between 8 and 30 wells per slide.

20 In one set of experiments, five 8 well chamber slides (Lab-Tek II, Nalge Nunc International, Rochester, NY), 250ul of 10ug/ml anti-his antibody (R&D Systems, Minneapolis, MN) in PBS was added to each well and incubated at room temperature overnight (typically 16 hours). Following, the liquid in each well was aspirated and washed once for approximately 5 minutes with 500ul of PBS. The
25 wells were then "blocked" (blocked for any additional protein binding sites) with the addition of 250ul of blocking solution (2% fetal bovine serum, 0.5% BSA in PBS) and incubation for 1 hour at room temp (e.g., 20°C). The liquid in each well was aspirated and 250ul of supernatant containing the chemokine-stalk-immobilization domain fusion proteins (SDF1a-FRAC, finetaxin-FRAC, CCL27-FRAC, I309-FRAC, MIP3a-FRAC, ELC-FRAC, SLC-FRAC, MDC-FRAC, TARC-FRAC, or BRAK-FRAC)
30 or the control supernatant was added to two wells. Each slide contained two wells with control supernatant (from mock-transfected cells) and two wells with the positive control SDF1a (which binds the ubiquitous chemokine receptor CXCR4). The four other wells were for experimental samples. The supernatants were incubated for

one hour at room temperature (typically 20°C), then incubated for an additional hour at 37°C. The supernatants were aspirated off, and each well was washed once for 5 minutes with 500ul of PBS. The slide was then ready for addition of cells for assay.

In a second set of experiments using two 96 well large format glass slides, 50 ul of 10 ug/ml anti-his antibody (R&D Systems, Minneapolis, MN) in PBS was added to each well and incubated at room temperature overnight (typically 16 hours) in a humidified chamber. Following, the liquid in each well was aspirated and washed once for approximately 5 minutes with 50 ul of PBS. The wells were then "blocked" (blocked for any additional protein binding sites) with the addition of 50ul of blocking solution (2% fetal bovine serum, 0.5% BSA in PBS) and incubation for 1 hour at room temp (20°C) in a humidified chamber. The liquid in each well was aspirated and 50ul of supernatant containing one of the 32 chemokine-stalk-immobilization domain fusion proteins shown in Figure 5 was added to 3 wells. Each slide contained 3 wells with the control supernatant (from mock-transfected cells) and 3 wells with the positive control SDF1a (which binds the ubiquitous chemokine receptor CXCR4). The other wells were for experimental samples. The supernatants were incubated for one hour at room temperature (typically 20°C) in a humidified chamber, then incubated for an additional hour at 37C again in a humidified chamber. The supernatants were aspirated off, and each well was washed once for 5 minutes with 50 ul of PBS. The slide was then ready for addition of cells for assay.

G. Expression Of Chemokine Receptors In Cell Culture

Several types of cells were used in binding experiments to stalkokines of the invention, including transfected mouse myeloma NSO cells containing a mammalian expression vector driving expression of the receptor of interest, e.g., CCR1, CCR4, CCR6, CCR7, CCR8, CXCR4, CMV US28 (gift of R&D Systems (Minneapolis, MN)). The results of this experiment are summarized in Table 6.

Table 6

<u>Cell types</u>	<u>Some Known Ligands</u>	<u>Binding to Stalkokine form</u>
CCR1-NSO	Ckb8-1, MCP3	no, yes
CCR4-NSO	MDC, TARC	yes, yes
CCR6-NSO	MIP3a	yes

CCR7-NSO	ELC, SLC	yes, no
CCR8-NSO	I309, vMIP1	yes, yes
CCR10-NSO	ELC	yes
CXCR4-NSO	FK, vMIPII	yes

5

*Negative controls (non-binding stalkokines) were done in every case Ckb8-1 (also MPIF1), ELC (same as MIP3beta), SLC (same as 6Ckine). FK= fractalkine

10

H. Contacting Receptor-Expressing Cells and Stalkokine Arrays

In an experiment, CCR1-NSO cells were grown, harvested, and resuspended in PBS at a 2×10^6 cells/ml. 250 ul of this cell suspension was added to each well of a 8 well chamber slide, with two wells coated with each of CKb8-1
 15 FRAC, MCP3-FRAC, SDF1a-FRAC (positive control), or mock transfected supernatant (negative control) as previously described. The slides were incubated for 1.5 h at room temp, then the supernatant gently aspirated. The chamber portion of the slide was removed (using the apparatus provided by the supplier) and the slide was washed by gentle immersion into a petri dish containing PBS to a depth of
 20 1cm. This was repeated 3 times in total, and the slide was immersed in a 1% glutaraldehyde solution (in PBS) for 5 minutes at room temp. The cells were inspected and enumerated visually by microscopy. The cells bound to MCP3 but not CKb8 tethered ligands.

I. Displacement and competition analysis

In an experiment, CCR4-NSO cells were used to test binding competition. Harvested CCR4 NSO cells were resuspended at 2×10^6 cells per ml in PBS. 500 ul aliquots of cells were transferred into eppendorf tubes, and 5 ug of recombinant human chemokine (MDC or TARC, R&D Systems, Minneapolis, MN)
 30 was added (final concentration was 10 ug/ml), mixed briefly, and incubated at room temperature for 10 minutes. Following, 250 ul of the cells were added to each two wells of an 8 well chamber slide that had been precoated with the MDC-FRAC or TARC-FRAC as previously described. 250 ul of cells that were not pre-treated with soluble chemokine were added to additional wells. The slides were incubated for 1.5
 35 hours at room temp, then the supernatant gently aspirated. The chamber portion of the slide was removed (using the apparatus provided by the supplier) and the slide

was washed by gentle immersion into a petri dish containing PBS to a depth of 1cm. This was repeated 3 times in total, and the slide was immersed in a 1% glutaraldehyde solution (in PBS) for 5 minutes at room temperature. The cells were inspected and enumerated visually by microscopy. The cells bound to both the
5 MDC-FRAC and TARC-FRAC, as expected. Competition with either soluble MDC or TARC reduced binding to background levels in both cases, as expected.

In another experiment, small molecule chemical compounds were tested on the binding of vMIP2-FRAC and US28-NSO. Three small organic compounds previously identified as inhibitors of binding between chemokine
10 receptor US28 and vMIP2, were also observed to compete and inhibit binding between vMIP2-FRAC and US28-expressing NSO cells using the present invention.

EXAMPLE 3

This example describes the binding of a chemokine receptor to
15 an array of stalkokines, to determine the binding specificity of the receptor.

To interrogate the binding of CCR10 to a panel of chemokine elements, an array of tethered ligands is prepared. pc-FRAC-based stalkokines are prepared using at least about 3, at least about 5, at least about 10, at least about 15, at least about 20 or all of the ligand domains listed in Table 5. An array of
20 stalkokines is prepared on a pair of 96 well slides precoated with the anti-polyhistidine antibody overnight and blocked for 1 hour as described *supra*. Each stalkokine is placed on three wells in a 30 ul volume per well and incubated for 1.5 hours in a humidified chamber at 37°C. Also included are negative controls, in triplicate, which include (1) a mock transfected supernatant, (2) an immobilized stalk
25 region (with no chemokine domain), and (3) PBS. A SDF1a stalkokine (which binds CXCR4, a ubiquitously expressed chemokine receptor) is used as a positive control. Each set of positive and negative controls is included on each plate.

The supernatants are aspirated and the wells are washed twice with PBS. The CCR10 cell line is prepared during incubation with the stalkokine. The
30 cell line is trypsinized from the culture flasks, and washed twice with PBS. The cells are resuspended at a concentration of 1 million cells per ml of PBS, 30 ul of the cell suspension are added to each well of plate, and incubated at room temperature without agitation for 1.5 hours at room temperature. The droplets are partially aspirated off (leaving approximately 5 ul, taking care not to disturb the cells resting

on the surface) and placed slowly into a tray with PBS (approximately 150 mls, depth of 1cm). Each of the four edges of the 96 well plate is lifted slowly to the surface of the liquid in each of three consecutive wells. The bound cells are stained with SybrGreen 1 (Molecular Probes, Eugene, OR) for 30 min at a stain diluted 1 to 5000 in PBS. The 96 well plate is removed, drained of excessive liquid, and counted in a Packard Fluorescent plate reader with 480 nm excitation, 530 nm emission filter sets. The results are transferred to an Excel file for further analysis.

10 All references cited herein are incorporated herein by reference in their entirety and for all purposes to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference in its entirety for all purposes.

15 Many modifications and variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described herein are offered by way of example only, and the invention is to be limited only by the terms of the appended claims, along with the full scope of equivalents to which such claims are entitled.

WHAT IS CLAIMED IS:

1. An assay device comprising a plurality of different immobilized tethered ligand fusion proteins organized in an array, wherein said ligand-stalk fusion proteins comprise:
 - 5 (a) a ligand domain, and
 - (b) a stalk domain.

2. An assay device comprising a plurality of different immobilized tethered ligand fusion proteins organized in an array, wherein said ligand-stalk fusion proteins
10 comprise:
 - (a) a ligand domain;
 - (b) an intermediate stalk domain; and,
 - (c) an immobilization domain,wherein said ligand domain and said stalk domain are not associated in a
15 naturally occurring protein.

3. The device of claim 2 wherein the immobilized tethered ligand fusion proteins comprise a mucin-derived stalk sequence.

- 20 4. The device of claim 2 wherein the immobilized tethered ligand fusion proteins comprise a fractalkine mucin repeat region sequence.

5. The device of claim 2 wherein at least some of the of immobilized tethered ligand fusion proteins comprise a ligand domain encoding a chemokine.
25

6. A method for identifying an interaction between a receptor and a ligand, said method comprising:
 - (a) contacting a cell expressing a receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein; and,
 - 30 (b) detecting binding of said cell and the fusion protein,wherein binding of the cell to the fusion protein is correlated with an interaction between the receptor and the ligand corresponding to the ligand domain of the fusion protein.

7. The method of claim 6, wherein the tethered ligand is immobilized on an assay device of claim 1.
8. The method of claim 6 wherein the cell expresses a recombinant receptor.
- 5
9. The method of claim 6 wherein the cell expresses a seven-transmembrane receptor.
10. The method of claim 9 wherein the cell expresses a chemokine receptor or a cytokine receptor.
- 10
11. The method of claim 6 wherein the cell expresses a orphan receptor.
12. The method of claim 6 wherein an interaction between a receptor and more than one ligand is detected.
- 15
13. A method for identifying a modulator of an interaction between a receptor and a ligand comprising
- (a) contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the absence of a test compound, and measuring binding of the cells to the fusion protein;
- 20
- (b) contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the presence of a test compound, and measuring binding of the cells to the fusion protein; and,
- 25
- (c) comparing the level of binding in (a) and (b), wherein decreased binding of cells in the presence of the test compound indicates that the test compound is an inhibitor of the interaction between the receptor and the ligand corresponding the ligand domain of said fusion protein, and wherein increased binding of cells in the presence of the test compound indicates that the test compound is an enhancer of the interaction between the receptor and the receptor and the ligand corresponding the ligand domain of said fusion protein.
- 30

14. The method of claim 13 wherein the tethered ligand is immobilized on an assay device of claim 1.

5 15. The method of claim 13 wherein the receptor is a chemokine receptor.

16. A method for detecting a profile of receptor expression in cells in a population of cells comprising:

- 10 (a) contacting said population of cells with an immobilized tethered ligand fusion protein, and
- (b) detecting binding of cells of said population to the fusion protein, wherein binding of a cell in the cell population to the fusion protein is correlated with expression of a receptor that binds the ligand corresponding to the ligand domain of the fusion protein.

15

17. The method of claim 16 wherein step (a) comprises contacting the population with a plurality of different fusion proteins and step (b) comprises detecting binding of cells to zero, one or more than one fusion protein.

20 18. The method of claim 17 comprising detecting binding of cells of said population to zero, one, or more than one tethered ligand fusion protein.

19. The method of claim 17 wherein the tethered ligand is immobilized on an assay device of claim 1.

25

20. The method of claim 16 wherein the population is heterogeneous.

21. The method of claim 20 wherein the population is obtained from synovial fluid, cerebral-spinal fluid, bronchial alveolar lavage (BAL) fluid, or blood.

30

22. The method of claim 16 further comprising quantitating the level of binding to each tethered ligand fusion protein in the array.

23. The method of claim 16 further comprising characterizing the cells bound at each sector of the array.

24. A method for diagnosis comprising

5 (a) obtaining a population of cells from a patient suspected of suffering from a disease

(b) determining a receptor profile for the population; and,

(c) comparing said receptor profile with a profile characteristic of the disease state.

10

25. A method for diagnosis comprising

(a) obtaining a population of cells from a patient suspected of suffering from a disease

(b) determining a receptor profile for the population by

15

(i) contacting said population of cells with a device of claim 1; and,

(ii) identifying a subset of arrayed tethered ligand fusion proteins bound by cells of said population thereby identifying a receptor profile;

(c) comparing said receptor profile with a profile characteristic of the disease state.

20

26. The method of claim 25, where said determining a receptor profile further comprises characterizing the cells bound at each sector of the array.

27. The method of claim 26 wherein the characterizing is achieved by immunostaining.

25

28. The method of claim 25 further comprising quantitating the binding of cells at each sector of the array.

29. The method of claim 24 wherein the disease is an inflammatory or allergic disease, or an autoimmune disease.

30

30. The method of claim 24 wherein the population is obtained from synovial fluid, cerebral-spinal fluid, bronchial alveolar lavage (BAL) fluid, or blood.

31. A method for detecting an effect of a drug or treatment on a patient comprising:

- 5 (a) determining the receptor profile of a population of cells from the patient for the first time;
- (b) administering the drug or treatment to the patient;
- (c) determining the receptor profile of a population of cells from the patient for a second time,
- 10 (d) comparing the receptor profiles in (a) and (c) to determine the effect of the drug or treatment on receptor-expressing cells in the patient.

32. A method for detecting an effect of a drug or treatment on a patient comprising:

- 15 (a) determining the receptor profile of a population of cells from the patient for the first time, by
- (i) contacting said population of cells with a device of claim 1; and,
- (ii) identifying a subset of arrayed tethered ligand fusion proteins bound by cells of said population thereby identifying a receptor profile;
- (b) administering the drug or treatment to the patient;
- 20 (c) determining the receptor profile of a population of cells from the patient for a second time,
- (d) comparing the receptor profiles observed in (a) and (c) to determine the effect of the drug or treatment receptor-expressing cells in the patient.

25 33. The method of claim 32 further comprising quantitating the level of binding to each tethered ligand fusion protein in the array.

34. The method of claim 32 further comprising characterizing the cells bound at each sector of the array.

30

35. The method of claim 34 wherein the characterizing is achieved by immunostaining.

36. A method for identifying a modulator of an interaction between a receptor and a ligand comprising

(a) contacting cells expressing the receptor or a ligand-binding portion thereof with an immobilized tethered ligand fusion protein in the presence of a test compound and determining the level of binding to the fusion protein,

(b) contacting cells expressing the receptor or a ligand-binding portion thereof with the immobilized tethered ligand fusion protein in the absence of a test compound; and determining the level of binding to the fusion protein, and

(c) comparing the level of binding in (a) and (b) for the immobilized tethered ligand fusion protein,

wherein said ligand-stalk fusion proteins comprise:

(i) a ligand domain;

(ii) an intermediate stalk domain; and,

(iii) an immobilization domain, and

wherein said ligand domain and said stalk domain are not associated in a naturally occurring protein, and,

wherein decreased binding of cells in the presence of the test compound indicates that the test compound is an inhibitor of the interaction between the receptor and the ligand corresponding the ligand domain of said fusion protein, the and wherein increased binding of cells in the presence of the test compound indicates that the test compound is an enhancer of the interaction between the receptor and the receptor and the ligand corresponding the ligand domain of said fusion protein.

37. The method of claim 31 wherein the stalk domain is carboxy-terminal to the ligand domain, and the immobilization domain is carboxy-terminal to the stalk domain.

38. The method of claim 31 wherein the cell expresses a recombinant receptor.

39. The method of claim 31 wherein the cell expresses a chemokine receptor.

40. The method of claim 38 wherein the cell expresses a orphan receptor.

41. The method of claim 36 wherein an interaction between a receptor and more than one ligand is detected.

42. A kit comprising at least two different immobilized ligand fusion proteins,
5 wherein said at least two different immobilized ligand fusion proteins are not immobilized on the same substrate.

43. The kit of claim 42 wherein the fusion proteins are immobilized on beads or
10 slides.

44. A tethered ligand fusion protein, wherein said ligand domain and said stalk
domain are not associated in a naturally occurring protein.

45. The tethered ligand fusion protein of claim 44, wherein the ligand domain is
15 other than a chemokine sequence.

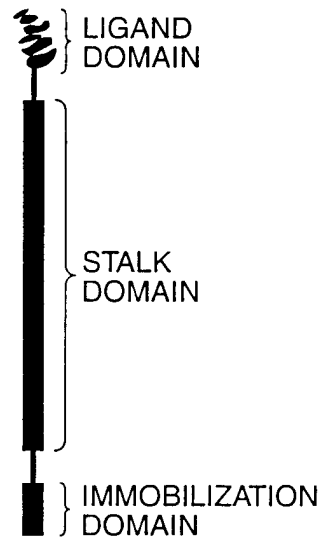


FIG. 1

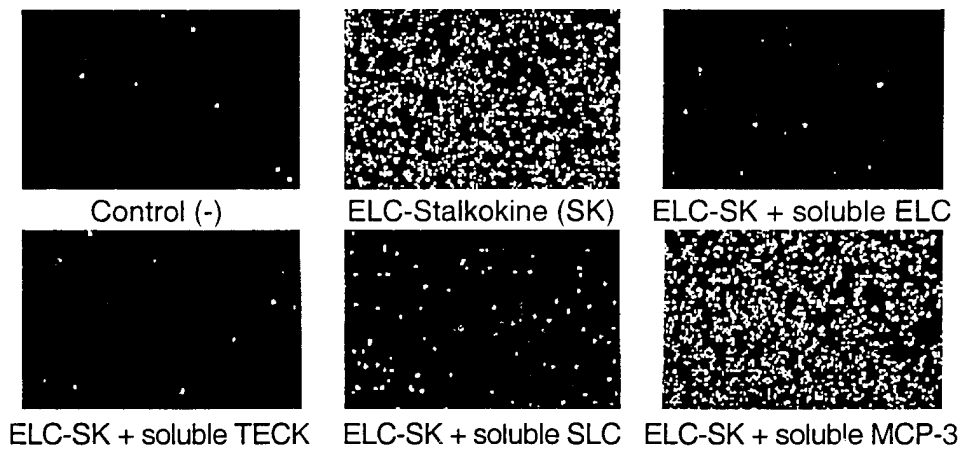


FIG. 3A

FIG. 2

(SEQ. ID NO: 1 & 2)

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M A L E Q N Q S T D Y Y Y B E N E M N G T Y D Y S Q Y E L I

TGTATCAAG AAGATGTCAG AAGATTTCA AAAGTTTCC TCCCTGTTT CCTCACATA GTTTTGGCA TTTCACTGCC AGCCAAATCC 180
C I K B D V R E F A K V F L P V F L T I V P V I G L A G N S

ATGGGATGG CAAATTAATC CTATTTCCAG AAACAGAGA CCAAAACAGA TGTGTACTC CTGATTTGG CTGTAGCACA TTTACTCTCT 270
M V V A I Y A Y Y K K Q R T K T D V Y I L N L A V A D L L L

CTATTCATC TCCCTTTTG CCGTGTAAAT CCAATTCATG GATGGGTTT AGCCAAATA ATGTCCAAA TFACTTCACC CTGTACACA 360
L F T L P F W A V N A V H G W V L G K I M C K I T S A L Y T

CTAACTTG TCTCTGGAT CCGATTTCTG GCTTGTATCA GONTAGCAG APTGTGTGCA GTACTTAAG TCCCCAGCA ATCAGGAGTG 450
L N F V S G M Q F L A C I S I D R Y V A V T K V P S Q S G V

GGAAACACT CCGTGCATC CTGTTCTGTT GTCGTGCG CTCCACTTT CCGACGATA CCCCAGTGG TTTTATATC AGTAAATGAC 540
G K P C W I I C P C V W M A A I L L S I P Q L V F Y T V N D

AATGCPAGT GAAATCCAT TTTCCTGGC TACTTAGGA CATAATGA AGCATTAAT CAATCTGAG AGATCTCCTT TCGATTTGTA 630
N A R C I P I F P R Y L G T S M K A L I Q M L E I C I G F V

GTACCTTC TTTTATGG GGTGTCTPC TTTATACAG CAAGCACT CATGAGTG CCAATGTA AAATATCTGG ACCCCATAA 720
V P F L I M G V C Y F I T A R T L M K M P N I K I S R P L K

GTTCTCTA CAGTGTAT AGTTTTCAT GTACTACAC TCCCTGTAA CATTTCAG TTCTCCGAG TCTCCRA I D I I Y S 810
V L L T V V I V P I V T Q L P Y N I V K P C R A I D I I Y S

CTGATACA GATCAACT GAGCAACC ATGGCAAG CCAATCAGT CACGAAAG ATCCACTT TTACAGTG CTTCAACCA 900
L I T S C N M S K R M D I A I Q V T E S I A L F H S C L N P

ATCCTTAG TTTTATGG AGCATTTT AAATACAG TTTGAAAT GGCACAAA TATGGTCT CCGAGACA GAGCAAGT 990
I L Y V F N G A S F K N Y V M K V A K K Y G S W R R Q R Q S

GTGAGCAT TCCCTTTGA TTCTAGGCT CCTACAGCC CAACTGAC TTTTGCAT TAAAGTAA ACTCCTGCC CTTTGTCTG 1080
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GATCATAG AATGTCCT TCCCTCAA TAAACTCT GCTCTCTT GAAAAAAA AAAA 1147

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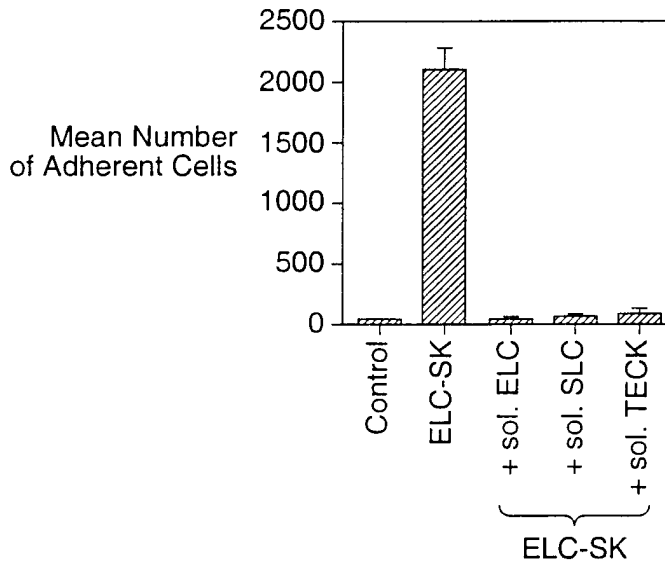


FIG. 3B

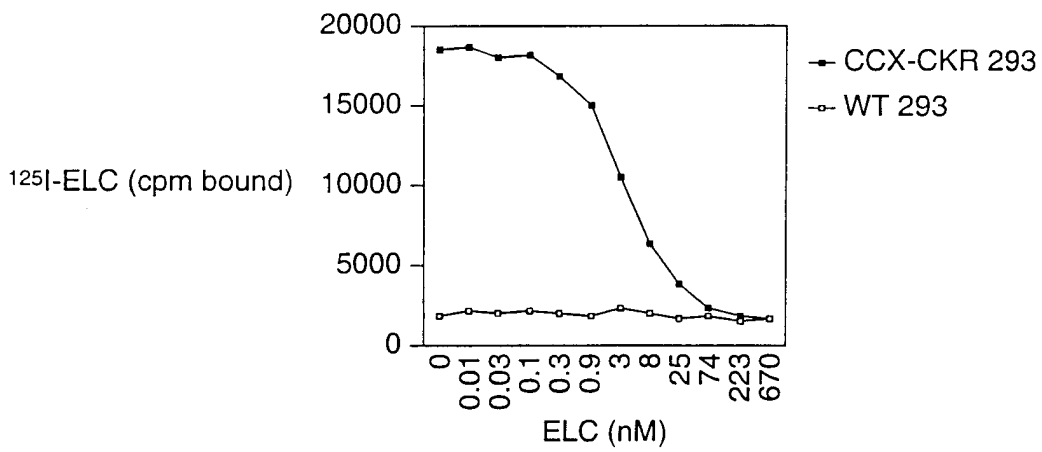


FIG. 3C

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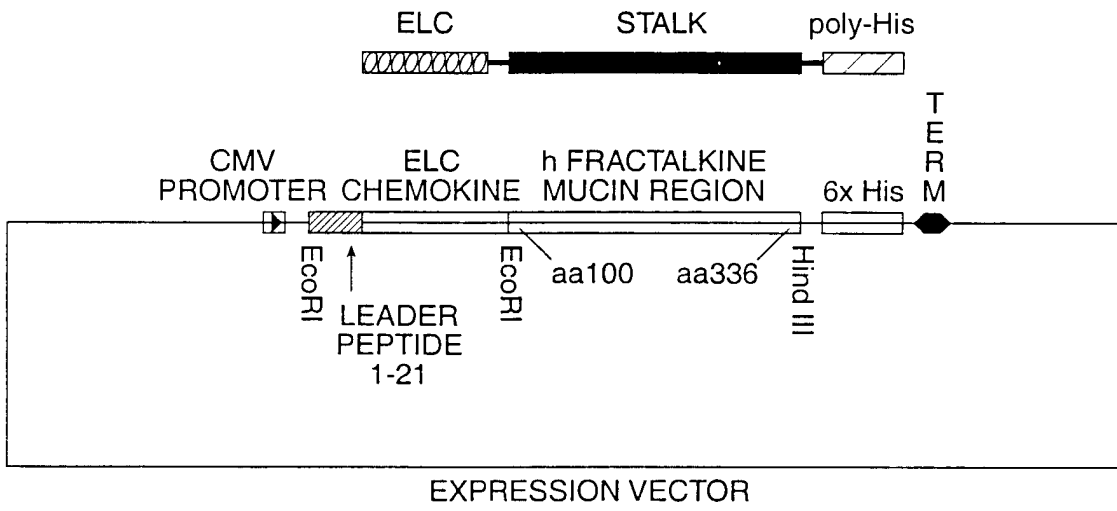


FIG. 4

STALKOKINES EXPRESSION

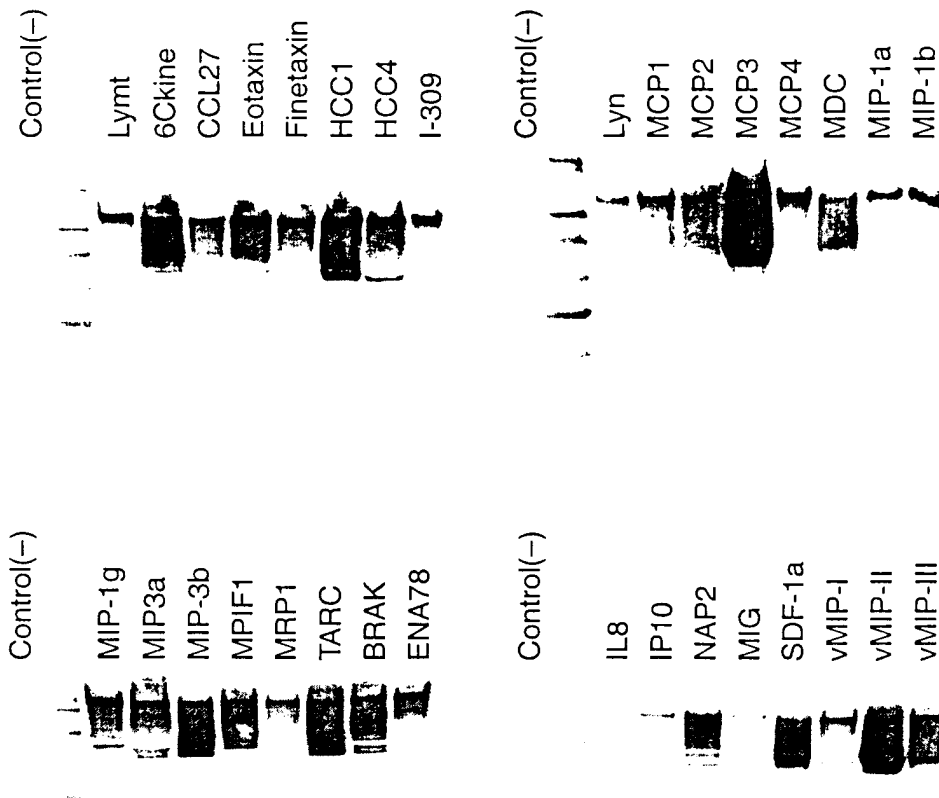
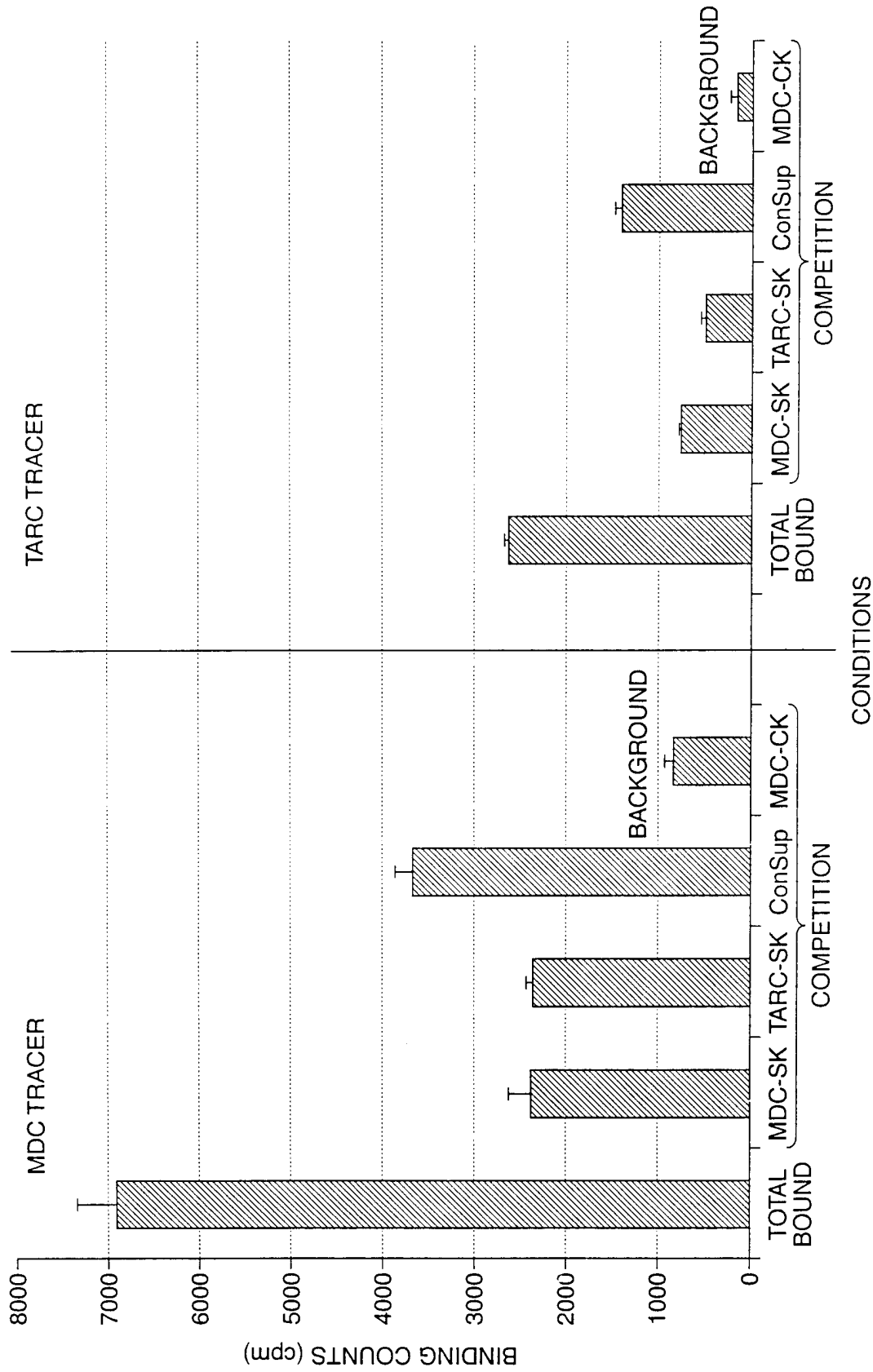


FIG. 5



CONDITIONS

FIG. 6

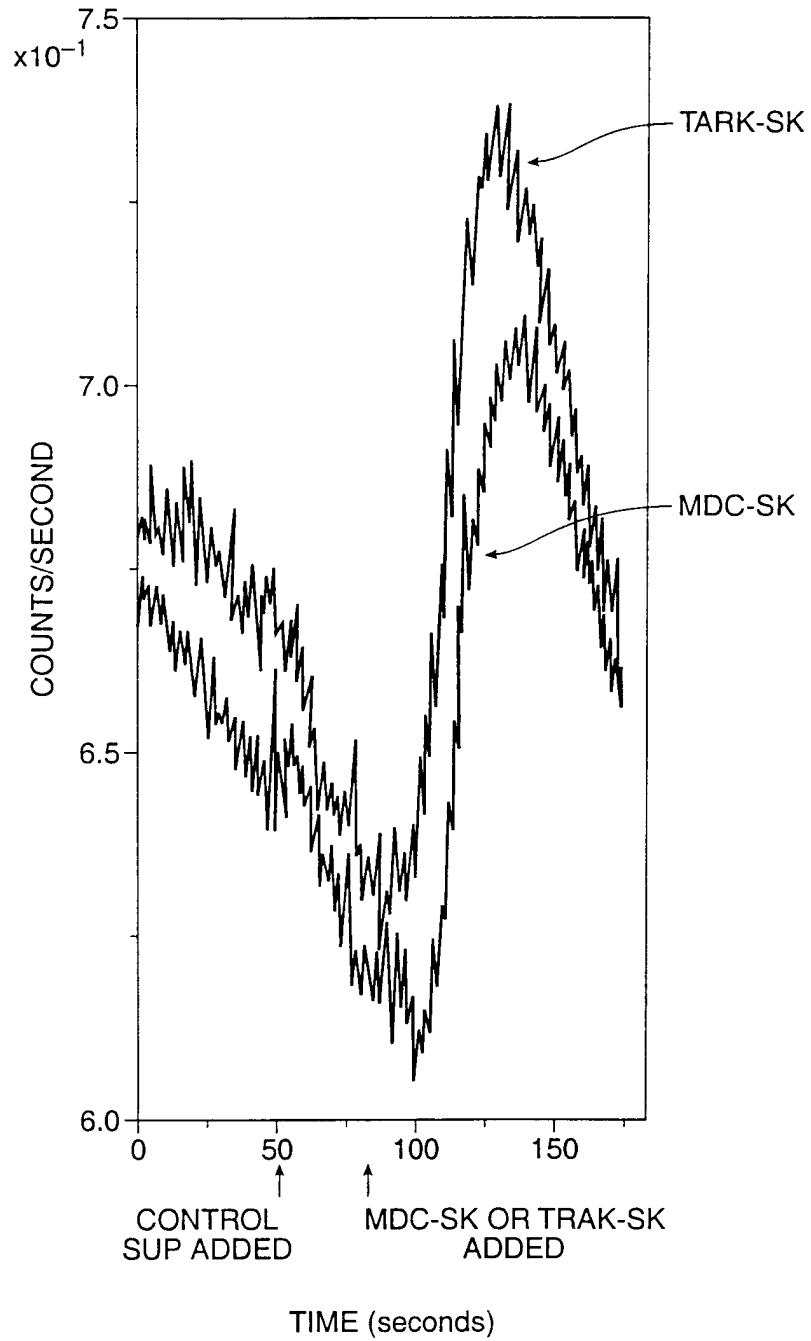


FIG. 7

专利名称(译)	系留配体和使用方法		
公开(公告)号	EP1247103A2	公开(公告)日	2002-10-09
申请号	EP2000988164	申请日	2000-12-18
[标]申请(专利权)人(译)	凯莫森特里克斯股份有限公司		
申请(专利权)人(译)	CHEMOCENTRYX INC.		
当前申请(专利权)人(译)	CHEMOCENTRYX公司		
[标]发明人	SCHALL THOMAS J TALBOT DALE MIAO ZHENHUA WEI ZHENG		
发明人	SCHALL, THOMAS, J. TALBOT, DALE MIAO, ZHENHUA WEI, ZHENG		
IPC分类号	A61K45/00 A61P37/02 A61P37/08 C07K14/47 C07K14/52 C07K14/715 C12N15/09 C12N15/10 C12Q1/02 G01N33/15 G01N33/50 G01N33/53 G01N33/531 G01N33/566 G01N33/68		
CPC分类号	G01N33/6863 C07K14/521 C07K14/7158 C07K2319/00 G01N33/5088 G01N33/531 G01N2500/00		
优先权	60/172979 1999-12-20 US 60/186626 2000-03-03 US 09/721908 2000-11-24 US		
其他公开文献	EP1247103B1		
外部链接	Espacenet		

摘要(译)

本发明提供了用于分析受体配体相互作用，确定细胞和细胞群中受体表达谱以及诊断和药物筛选方法的方法，试剂和装置。本发明利用具有配体结构域，茎结构域和任选的固定化结构域的固定化栓系配体融合蛋白。