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(54) Title: COMMON LYMPHATIC ENDOTHELIAL AND VASCULAR ENDOTHELIAL RECEPTOR-1 (CLEVER-1) AND USES THEREOF

(57) Abstract: A novel protein Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 (CLEVER-1) is described. CLEVER-1 mediates leukocyte and malignant cell binding to vascular and lymphoid endothelial cells. CLEVER-1 is the first protein that has been reported to mediate both influx into and efflux from the lymph nodes. Also provided are methods of treating inflammation and preventing metastasis of malignant cells by providing an inhibitor of CLEVER-1 binding.

COMMON LYMPHATIC ENDOTHELIAL AND VASCULAR EN- DOTHELIAL RECEPTOR-1 (CLEVER-1) AND USES THEREOF

BACKGROUND OF THE INVENTION

Field of the Invention

5 The invention is in the field of cell adhesion proteins. Specifically, the invention is in the field of CLEVER-1, a novel protein that facilitates the influx of leukocytes and malignant cells into the lymphatic system, and also the efflux of the same out of the lymph nodes.

Background Art

10 Leukocytes are the major cellular components of inflammatory and immune responses. Leukocytes include lymphocytes, natural killer (NK) cells, monocytes, dendritic cells and granulocytes (neutrophils, eosinophils and basophils). See, HARRISON'S PRINCIPLES OF INTERNAL MEDICINE, Fauci, A.S. *et al.* eds. (14th ed. 1998). Lymphocytes are composed of B cells and T
15 cells. B cells provide humoral immunity and are the precursors of plasma cells. T cells provide cell mediated immunity. In tissues monocytes differentiate further into macrophages. At sites of inflammation, blood monocytes can attach to inflamed endothelia. Macrophages recognize and take up a
20 wide range of exogenous materials such as bacteria. Granulocytes also have critical roles in inflammation. They are needed to clear infections with extracellular bacteria. The immune response has an important role in the growth, differentiation, and mobilization of granulocytes.

 Continuous lymphocyte recirculation between blood and lymphoid tissues forms a basis for the function of the immune system. However such
25 lymphocyte recirculation inadvertently also facilitates at least two medical conditions: inflammation and metastasis.

 Lymphocytes enter the lymphoid tissues by binding to vascular endothelial cells. Lymphocyte adherence to endothelial cells is mediated by complementary, surface expressed molecules on both cell types. The adhe-
30 sion molecules and mechanisms of lymphocyte entrance into the tissues from the blood have been thoroughly characterized, but mechanisms controlling lymphocyte exit from the non-lymphoid and lymphoid tissues via lymphat-
 ics have remained unknown.

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The majority of lymphocytes extravasate into the lymph nodes via specialized vessels called high endothelial venules, or HEV. The rest of the incoming lymphocytes enter the nodes via afferent lymphatics together with antigens and other types of hematopoietic cells such as dendritic cells, macrophages and granulocytes. However, only lymphocytes are able to leave the nodes via the efferent lymphatic system by first traversing the sinusoidal endothelium and then entering the efferent lymphatic vessel. To maintain the homeostasis in the lymph node the numbers of entering and exiting lymphocytes need to be well balanced. The molecular mechanisms involved in lymphocyte exit are unknown.

In addition to being of fundamental importance in normal lymphocyte recirculation, the lymphatics also regulate seeding of metastasizing cells in approximately 50% of cancers that use this type of vessel for spreading. Lymph nodes are often the first organ to develop metastases, especially in the case of carcinomas. The design of the lymphatic system makes it relatively easy for malignant tumor cells to enter (Sleeman, J.P., *Recent Results Cancer Res.* 157:55-81 (2000)), and thus compounds that prevent the entry and exit of malignant tumor cells from the lymphatics have tremendous therapeutic potential.

BRIEF SUMMARY OF THE INVENTION

Recognizing the need to control lymphocyte recirculation, the inventors initiated a study of the proteins of the efferent lymphatic vessels. These studies have culminated in the discovery of a novel protein, Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 (CLEVER-1), a binding protein that mediates adhesion of lymphocytes (and malignant tumor cells) to endothelium in both the systemic vasculature and in the lymphatics. The inventors have discovered that by blocking the interaction of CLEVER-1 and its lymphocyte substrate, the artisan can, for the first time simultaneously, control lymphocyte recirculation and lymphocyte migration, and related conditions such as inflammation, at the site of lymphocyte influx into, and efflux from, the tissues. The inventors have also discovered that CLEVER-1 also mediates binding of other types of leukocytes such as monocytes and granulocytes to HEV-like vessels. Further, by blocking the interaction of CLEVER-1 and malignant tumor cells, the artisan can also, for the first time,

control metastasis by preventing malignant cells that bind to CLEVER-1 from being taken up by the lymphatic vessels, and thus preventing spread of the malignancy into the lymph nodes.

Accordingly, in a first embodiment, the invention is directed to cellular and subcellular extracts that contain CLEVER-1.

In a further embodiment, the invention is directed to purified or isolated CLEVER-1.

The invention further provides a method for inhibiting CLEVER-1 mediated leukocyte (such as lymphocyte, monocyte, and granulocyte) adhesion in a subject in need of the same, such method comprising administering a CLEVER-1 binding agent, or soluble CLEVER-1, to such subject.

The invention further provides a method for preventing or reducing CLEVER-1 mediated inflammation in a subject in need of the same, such method comprising administering a CLEVER-1 binding agent, or soluble CLEVER-1, to such subject.

The invention further provides a method for inhibiting CLEVER-1 mediated metastasis of malignant cells in a subject in need of the same, especially metastasis into the lymph nodes, such method comprising blocking the interaction of CLEVER-1 and the metastasizing cell in such subject with a CLEVER-1 binding agent, or by administering soluble CLEVER-1, to such subject.

The invention further provides a method for extracting cells that bind to CLEVER-1 from a population of cells, such method comprising mixing a population of cells with a preparation that contains CLEVER-1 and purifying the CLEVER-1-cell complex away from the rest of the non-binding cells in the population. Such method can be used to enrich for a subpopulation of CLEVER-1 binding cells.

The invention further provides a method for identifying an agent that binds to CLEVER-1, such method comprising determining whether a substance will successfully or unsuccessfully compete with leukocytes, malignant cells or with CLEVER-1 antibodies for binding to CLEVER-1.

The invention further provides a method for identifying an agent that is capable of inhibiting CLEVER-1 mediated cell migration, such method comprising assaying CLEVER-1 trafficking of lymphocytes into the afferent lymphatics or HEV, and/or out of the lymph nodes, in the presence of such

agent, and identifying such agent on the basis of its ability to inhibit such trafficking. In another embodiment, CLEVER-1 trafficking of malignant cells is assayed.

The present invention also provides a method of stimulating
5 CLEVER-1 binding, for example, in immunocompromised hosts to facilitate leukocyte (such as lymphocyte, monocyte and granulocyte) trafficking and the function of immune defense systems.

BRIEF DESCRIPTION OF THE DRAWINGS

10 FIGS. 1a-1i. Indirect immunoperoxidase staining showing that monoclonal antibodies 3-266 and 3-372 recognize endothelium both in afferent and efferent lymphatic systems and on HEV. FIGS. 1a-1c are from the skin, FIGS. 1d-1i are from a lymph node. FIGS. 1a, 1d and 1g show the staining with monoclonal antibody 3-266, FIGS. 1b, 1e and 1h show the staining with monoclonal antibody 3-372 and FIGS. 1c, 1f and 1i show the staining
15 with a negative control antibody, 3G6. In FIGS. 1a, 1b and 1c, the arrows point to the epithelium and arrowheads to afferent lymphatics. In FIGS. 1d and 1e, the arrows point to the lymphatic vessels (lymphatic sinusoids that belong to the efferent lymphatic system) within the lymph node. In FIGS. 1g and 1h, the arrows point to HEV.

20 FIG. 2. Monoclonal antibodies 3-266 and 3-372 recognize an about 270-300 kDa molecule. Molecules in lymph node lysates were separated by SDS-PAGE, blotted to nitrocellulose sheets and probed with monoclonal antibody 3-266 and 3-372 or with a negative control antibody (3G6).

FIGS. 3A and 3B. CLEVER-1 is involved in lymphocyte binding to
25 endothelial cells both in HEV and lymphatics. An adhesion assay was performed to measure lymphocyte binding to HEV (FIG. 3A) and to lymphatic endothelium (FIG. 3B). The sections were pre-incubated with monoclonal antibody 3-266 or 3-372, or negative control antibody anti-HLA ABC or 3G6 ("neg co") after which the sections were overlaid with normal lymphocytes.
30 The results of three to four independent inhibition experiments are shown as mean percentage of maximal binding \pm SEM.

FIGS. 4A and 4B. CLEVER-1 is involved in binding of tumor cells to endothelial cells both in HEV and lymphatics. An adhesion assay was performed to measure binding of different tumor cell lines to HEV (FIG. 4A) and

to lymphatic endothelium (FIG. 4B). The sections were pre-incubated with 3-372 or negative control antibody (anti-HLA ABC) after which the sections were overlaid with different tumor cells: Nu, NA, IBW4, KCA and CRL 1648. The results of three to four independent inhibition experiments are shown as
5 mean percentage of maximal binding \pm SEM.

FIG. 5. CLEVER-1 is induced on HEV-like vessels at sites of inflammation in connection to infiltrations of inflammatory cells (FIGS. 5a-5c, synovium; FIGS. 5d-5f, skin). Fig. 5a. Fibrotic type of inflamed synovium without any marked infiltrations of inflammatory cells. Only the afferent lymphatics expressed CLEVER-1 (arrows). FIG. 5b. CLEVER-1 was upregulated
10 on a HEV-like vessel (marked by a dashed line) within a heavy lymphocytic infiltration. FIG. 5c. Staining with a negative control antibody (3G6). FIG. 5d. In normal skin afferent lymphatics expressed CLEVER-1 (arrows), but in inflamed skin HEV-like vessels (dashed line) also expressed CLEVER-1 (FIG.
15 5e). Negative control staining (FIG. 5f). Arrowheads point to epidermis (FIGS. 5d-5f).

FIG. 6. CLEVER-1 mediates binding of monocytes and granulocytes to HEV-like vessels at sites of inflammation. Contribution of CLEVER-1 in binding of monocytes and granulocytes to inflamed synovial vessels and
20 binding of granulocytes to tonsil was tested using Stamper-Woodruff type of binding assay. 3-372 and 3-266 (pooled) but not the class-matched control antibody (3G6) significantly inhibited binding of granulocytes and monocytes to HEV-like vessels in the organs tested. The results of four independent
25 assays are shown as mean percentage of maximal binding ($=100\%$ in the presence of the control antibody) \pm SEM.

FIG. 7. Molecular characterization of CLEVER-1. (a) Antibodies 3-266 and 3-372 recognize a 270 – 300 kDa molecule in immunoblotting. 3G6 is a negative control antibody. (b) In gels run 48 hr for better resolution, at least three different isoforms of CLEVER-1 are seen, and enzymatic digestions with neuraminidase and O-glycans reveal the sialoglycoprotein nature
30 of CLEVER-1. (c) Relative contribution of different isoforms of CLEVER-1 is different in tonsil, lymph nodes and synovium. (d) An alternative spliced form missing exon 27 is present in 1. lung, 2. brain, 3. placenta, 4. heart, 5. liver, 6. skeletal muscle, 7. kidney, 8. pancreas, 9. spleen, 10. thymus, 11. prostate, 12. testis, 13. ovary, 14. small intestine, 15. colon, 16. lymph nodes.
35

Water control negative (lane 17). The upper band represents the standard form and the lower one is the splice variant of CLEVER-1.

Fig. 8. CLEVER-1 is expressed on the surface of endothelium *in vivo* and inhibition of its function blocks lymphocyte trafficking. Intravenously given 3-372 antibody (a) but not a negative class-matched control antibody (b) localized on the surface of HEV in lymph nodes after a 5 min circulation. HEV is pointed out by arrows in a. (c) Anti-CLEVER antibody treatment significantly inhibits the increase of the size of the lymph nodes draining the footpads. (One lymph node of a 3-372 treated rabbit was not found). (d) Lymphatic sinusoids of 3-372 treated animals contained less lymphocytes than those of control treated rabbits (e).

Fig. 9. The nucleotide sequence (7879 nt) of CLEVER-1. Boxed in grey are the translation initiation codon, translation stop codon, the two RGDs, the potential polyadenylation signal and the four nucleotide differences compared to Genbank entry AJ 2752213 (stabilin-1), i.e., nucleotides 1131, 2767, 6629 and 6969. Underlined are the nucleotides corresponding to the alternatively spliced exons.

DETAILED DESCRIPTION OF THE INVENTION

The term "ameliorate" denotes a lessening of an effect. To ameliorate a condition or disease refers to a lessening of the symptoms of the condition or disease.

The term "modulate" means to control in a predictable fashion, either by increasing or by decreasing the targeted parameter, as indicated from the context.

The term "effective amount" refers to that amount of the indicated agent that is sufficient to achieve the desired effect.

The term "inflammatory condition" refers to a physiological or pathological condition that is accompanied by an inflammatory response in a subject, which includes, *inter alia*, an undesired accumulation of leukocytes at one or more sites in such subject. The inflammatory condition can be hyperacute, acute, subacute or chronic. The inflammatory condition can be localized at the site of the inflammatory lesion or diffuse throughout the subject.

The term "drug" denotes any pharmaceutical or physiological agent, composition, bioactive compound, or combinations thereof, useful in the diagnosis, cure, mitigation, treatment, or prevention of a disease, or for any other medical purpose. The term "drug" is intended to be interpreted broadly and is not limited in terms of chemical composition or biological activity.

The term "essentially free of contaminants" refers to a substance that is of, undesired or unnecessary substances that had been present during the in vitro or in vivo synthesis of the desired substance.

The term "treatment" or "treating" refers to the administration of an agent to a subject for purposes which can include prophylaxis, amelioration, prevention or cure of an undesired disorder. Such treatment need not necessarily completely ameliorate the disorder, for example, inflammation; it is sufficient that such treatment ameliorates the disorder to a degree that is beneficial to the subject to which it is administered. Further, such treatment can be used in conjunction with other traditional treatments, for example, alternative treatments for reducing the inflammatory condition, known to those of skill in the art and as desired by the practitioner.

By "systemic vasculature" is meant the vascular network of blood vessels throughout the body of an animal or human.

By "lymphatic system" is meant the specialized part of the circulatory system that consists of lymph, the lymphatics, and the lymph nodes. The lymph nodes are located along the paths of the lymph collecting vessels and in isolated nodules of lymphatic patches in the intestinal wall. Additionally, there are specialized lymphatic organs such as the tonsils, thymus and spleen. B lymphocytes begin their final stages of maturation within the germinal centers of the lymph nodes' cortical nodules. Maturing lymphocytes are then pushed to the more densely packed outer layers as they mature, before being released into the efferent lymphatics. The lymph nodes that are located in the floor of the mouth are called the submental and submaxillary lymph nodes. The superficial cervical lymph nodes are located in the neck. The superficial cubital or supratroclear lymph nodes are located just above the bend in the elbow. The axillary lymph nodes are clustered deep within the underarm and upper chest region. Inguinal lymph nodes are located in the groin. By "lymphatics" is meant the vessels that return lymph to the

blood. Lymph is the clear fluid that flow in the lymphatics. Lymph arises from plasma that filters into the interstitial spaces from blood flowing through the capillaries. Although most of this plasma is taken up and absorbed by cells or the blood, a small amount is not absorbed. The lymphatics act as drains to
5 collect this excess fluid and return it to the venous blood just before it reaches the heart. The lymph nodes act as filters that collect the lymph from several different lymphatics and "percolate" the lymph through spaces termed sinuses before draining into a single efferent draining vessel.

By "afferent lymphatics" is meant the vessels through which anti-
10 gens enter the lymph nodes. Lymphocytes can enter the lymph nodes via the afferent lymphatics or via the high endothelial venules (HEV).

By "high endothelial venules" (HEV) is meant a specialized cortical postcapillary venules whose endothelium is simple cuboidal to columnar instead of simple squamous. HEVs are located mainly in the paracortex of the
15 lymph nodes. Lymphocytes cross the HEV, and thus "traffic" into the lymph nodes by diapedesis, that is, the lymphocytes stick to the luminal surface of the HEV, and then squeeze into the space between two or more HEV cells.

By "efferent lymphatics" meant the vessels that drain the lymph nodules (nodes).

By "lymphocyte recirculation" is meant the continuous movement
20 of lymphocytes throughout the circulatory and lymph system. Lymphocytes leave the lymph node and are first delivered via the lymph to venous system draining into the heart.

The lymphocytes then circulate throughout the body in the blood-
25 stream. Most of the lymphocytes are redelivered to the spleen or to another lymph node. About 10% go to non-lymphoid organs. Lymphocytes that have never been activated cannot enter non-lymphoid organs.

Lymphocyte "trafficking" refers to lymphocyte cell movement to specific locations. Outside of the lymph nodes, the trafficking of circulating
30 lymphocytes allows the lymphocyte to accumulate at sites of inflammation. Activated effector lymphocytes tend to home to areas of inflammation, resulting in a large influx of lymphocytes in areas of inflammation. At the inflamed site, lymphocytes attach to the endothelial cells that line the blood vessels. This attachment localizes the lymphocyte at the site of inflammation and al-

lows for subsequent emigration of the cells into the surrounding tissues (extravasation).

The Identification and Purification of CLEVER-1

The basis of the invention is the discovery of a new molecule, a
5 novel protein herein designated "Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 in the systemic vasculature, and in the afferent and efferent lymphatics. It has been found that leukocytes such as lymphocytes, monocytes, and granulocytes, and malignant cells specifically bind to this protein. It has also been found that this protein acts as a receptor that
10 facilitates entry of bound leukocytes and malignant cells through the walls of the systemic vasculature, into the lymph nodes and out of the lymph nodes.

To search for a protein that played a role in lymphocyte lymphatic efflux, the inventors first identified cell migration-associated lymphatic structures from isolated efferent lymphatic vessels of human lymph nodes. These
15 structures were used to produce monoclonal antibodies. Hybridomas were screened on frozen sections of human lymph nodes using immunoperoxidase staining.

Two of the hybridomas produced antibodies (designated 3-266 and 3-372) that clearly stained lymphatic endothelium both in afferent and efferent lymphatic systems and vascular endothelium on HEV, while other
20 structures remained unstained. This is consistent with the expected pattern for an antibody that recognizes a lymphocyte migration-associated structure. Cell culture of 3-266 (DSM ACC2519) and cell culture of 3-372 (DSM ACC2590) were both deposited under the terms of the Budapest Treaty on
25 the International Recognition of the Deposit of Micro-organisms for the Purposes of Patent Procedure on August 21, 2001, with DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroder Weg 1b, D-38124 Braunschweig.

In molecular weight determinations by immunoblotting, both anti-
30 bodies recognized a molecule of the same size, about 270-300 kDa. Due to this and an identical staining pattern, these antibodies were considered to recognize the same antigen; that antigen was named Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 (CLEVER-1).

CLEVER-1 was purified from CLEVER-1 containing lymph node preparations using affinity chromatography with the 3-372 antibody. The eluted material was subjected to SDS-PAGE analysis and silver staining. The specific band was excised, reduced, alkylated and digested with trypsin.

5 After cleavage with trypsin, mass spectrometric analyses yielded 27 peptides. Twenty-one (77%) of those had identical sequences with stabilin. These sequences covered altogether 268 amino acids (10% of the 2570 amino acids of stabilin-1) and spanned the amino acids between 53 and 2301 (Table 1). The peptide data suggest that CLEVER-1 has some homol-
10 ogy with stabilin-1 at the structural level. No functional information regarding stabilin-1 can be found in the literature.

 Peptide analysis of CLEVER-1 indicates no significant homology with any of the known endothelial homing-associated molecules, such as ICAM-1 (Intercellular Adhesion Molecule), ICAM-2, or VAP-1 (Vascular Ad-
15 hesion Protein).

 CLEVER-1 has several structural motifs that are associated with adhesive functions in other molecules. They include a proteoglycan link homology region important in CD44 for hyaluronan binding and two RGD motifs known to serve as integrin ligands in certain molecules, such as in fi-
20 bornectin. In addition, CLEVER-1 has seven fascicling domains also present in several molecules such as priostin, fasciclin and transforming growth factor- β -induced gene, big-h3 and in all of these cases it is essential for adhesive function of these molecules. Interestingly, twenty-two epidermal growth factor (EGF) repeats are also found in CLEVER-1. This structural domain is
25 also present in all members of the selectin family. Although the lectin domains of selectins are of utmost importance for the transient interaction with their sialomucin ligands, EGF-repeats have been reported to functionally contribute with lectin domains to binding between leukocytes and endothelium. Based on the structural complexity of CLEVER-1 it may turn out have several
30 ligand molecules and be multifunctional in its nature.

 It has been discovered that CLEVER-1 is involved in the process of lymphocyte recirculation. CLEVER-1 is present on the endothelium of the systemic vasculature, especially on the HEV and also on the endothelium of both afferent and efferent lymphatic systems. CLEVER-1 is a protein adhe-
35 sion molecule, and especially, a cell adhesion molecule (CAM), that mediates

adhesion of lymphocytes and of malignant tumor cells to CLEVER-1 in the systemic vasculature and the lymphatic system. These sites are of utmost importance as control points in lymphocyte trafficking.

CLEVER-1 is the first molecule that has been identified to facilitate lymphocyte and malignant cell exit from the lymph nodes. Additionally, CLEVER-1 is the first molecule that has been identified that regulates both entrance of lymphocytes and tumor cells into the lymph nodes and exit of lymphocytes and tumor cells from the lymph nodes. CLEVER-1 has been found to also mediate binding of other leukocytes such as monocytes and granulocytes to HEV-like vessels.

By "CLEVER-1 mediated cell binding" is meant the specific association of CLEVER-1 with either a leukocyte, such as a lymphocyte, monocyte, or granulocyte, or a CLEVER-1-binding malignant cell. CLEVER-1 mediated cell binding can occur with CLEVER-1 in a soluble form or in a particulate form (for example, when CLEVER-1 is present in a form that is membrane associated).

CLEVER-1 Mediated Binding of Leukocytes

According to the invention, adhesion of leukocytes, such as lymphocytes, monocytes, and granulocytes, to the endothelium (that is, to an endothelial cell(s)) in the systemic vasculature, especially the HEV, and to the endothelium in the afferent and efferent lymphatics can be blocked by blocking the binding between such endothelial cell's CLEVER-1 and leukocyte.

In the systemic vasculature, inhibiting or preventing endothelial cell CLEVER-1 mediated lymphocyte binding will inhibit or prevent lymphocytes, especially activated lymphocytes, from accumulating at such sites, and thus prevent or lessen inflammation at such sites. Thus, the invention provides a method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to lymphocytes.

In the afferent lymphatics, inhibiting or preventing lymphocytes from binding to endothelial cell CLEVER-1 will inhibit or prevent such lymphocytes from entering the afferent lymphatics and thus the lymph nodes. Thus, the invention provides a method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell

binding to lymphocytes in the afferent lymphatics and especially at HEV in lymph nodes or HEV-like venules at sites of inflammation. The invention also provides a method of inhibiting lymphocyte trafficking into the lymph nodes, by administering an agent that inhibits or prevents afferent lymphatic
5 CLEVER-1 mediated endothelial cell binding, and especially HEV binding, to lymphocytes and other leukocytes.

In the efferent lymphatics, inhibiting or preventing lymphocytes from binding to endothelial cell CLEVER-1 will prevent the lymphocytes from exiting the lymph node and entering the blood. Thus, the invention provides a
10 method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to lymphocytes in the efferent lymphatics. The invention also provides a method of inhibiting lymphocyte trafficking out of the lymph nodes, by administering an agent that inhibits or prevents efferent lymphatic CLEVER-1 mediated endothelial cell
15 binding to lymphocytes.

Therefore, CLEVER-1 binding with lymphocytes presents a unique, three-prong approach to treat diseases or conditions characterized by an undesired lymphocyte accumulation or trafficking in which the artisan can target lymphocyte entry into the lymph nodes, lymphocyte exit from the
20 lymph nodes, and lymphocyte binding to the systemic vasculature, with the same agent.

The discovery of CLEVER-1 and its role has thus resulted in a new method to control lymphocyte migration by inhibiting CLEVER-1 mediated cell binding to such cells. Thus, the present invention provides a method
25 of inhibiting undesired CLEVER-1 mediated lymphocyte trafficking, and thus blocking harmful or otherwise undesired lymphocyte migration, by preventing the association of CLEVER-1 with lymphocytes. Similarly, the invention provides a method of inhibiting undesired CLEVER-1 mediated binding of other leukocytes by preventing association of CLEVER-1 with the leukocytes.

30 The present invention also provides a method of stimulating CLEVER-1 binding, for example, in immunocompromised hosts to facilitate lymphocyte trafficking and other leukocyte binding and the function of immune defense systems.

CLEVER-1 Mediated Cell Binding to Malignant Cells

Because cancer cells often break away from a malignant tumor and enter the lymphatics, cancer cells travel to and establish themselves in the lymph nodes. According to the invention, the ability of a malignant tumor cell to establish itself in a lymph node can be inhibited or prevented by inhibiting or preventing CLEVER-1 binding to such malignant tumor cell.

The term "tumor" refers to a neoplasm, a tissue mass that is characteristic of a neoplasia. Neoplasia is distinguished from other forms of tissue growth, first, by the formation of a tissue mass, a neoplasm, or tumor. Second, neoplasia is considered to be an irreversible process. Third, neoplastic tissue tends to morphologically resemble its tissue of origin. Fourth, neoplastic tissue tends to functionally resemble its tissue of origin. Fifth, neoplasms grow and function somewhat independently of the homeostatic mechanisms that control normal tissue growth and function.

A neoplasm can be benign or malignant. A benign neoplasm consists of a discrete tissue mass that continues to grow. A benign neoplasm will simply push adjacent tissues out of its way as it grows.

The definitive features of a malignant neoplasm, a malignancy, are invasion and metastasis, that is, the spread of the neoplasm to a distant site. A malignant neoplasm will grow into the adjacent tissue, rather than pushing it away. The terms "malignant neoplasm," "malignant tumor," and "cancer" are synonymous.

Cancer cells typically invade thin-walled vessels such as small veins, venules, capillaries and lymphatics. The passage of cancer cells via lymphatics to lymph nodes, and via blood vessels to other organs and structures, and the subsequent implantation and growth of the cancer cells in those sites is referred to as "metastasis." The lymph nodes are common sites for metastasis.

Cancer cells can also spread by seeding - shedding into, for example, the peritoneal fluid. The cells can be carried by the fluid to a distant site on the peritoneal surface where they can implant and form new foci of cancer growth.

Most neoplasms are one of four types: epithelial, non-epithelial, blastomas and teratomas. Malignant epithelial neoplasms are termed carcinomas. An adenocarcinoma is a carcinoma in which gland-like structures

are present. Carcinomas can be papillary or cystic. Benign epithelial neoplasms are generally adenomas, polyps or papillomas.

Non-epithelial tumors can also be benign or malignant. They are generally named by a prefix that indicates the histologic type and a suffix.

5 The suffix -oma generally means benign while the suffix -sarcoma means malignant. However, several malignant neoplasms have traditional names ending in -oma: for example, melanoma, hepatoma, and lymphoma.

10 Lymphomas are malignant neoplasms arising from cells of the lymphoid series. Blastomas and teratomas contain more than one type of tissue. Malignant teratomas are often termed teratocarcinomas.

A "leukemia" is a tumor of white blood cells that is present in the bone marrow and blood. A "lymphoma" is a tumor of white blood cells that is present in the lymph nodes and tissues.

15 According to the invention, the binding of CLEVER-1-binding malignant cells to the endothelium in the systemic vasculature, especially the HEV, and to the endothelium in the afferent and efferent lymphatics can be inhibited or prevented by inhibiting or preventing the binding between such endothelial cell's CLEVER-1 and such malignant cell. Thus, the invention provides a method of treating cancer, and especially, a method of preventing
20 metastasis, by administration of an agent that inhibits or prevents CLEVER-1 mediated malignant cell binding to the endothelium.

In the systemic vasculature, inhibiting or preventing CLEVER-1 mediated cell binding will inhibit or prevent the establishment of CLEVER-1 binding malignant cells at such sites, and thus lessen or prevent metastasis
25 of such malignant cells. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding tumor cells in the systemic vasculature. The invention also provides a method of
30 inhibiting metastasis, by administering an agent that inhibits or prevents systemic vasculature CLEVER-1 mediated endothelial cell binding to such malignant cells.

In the afferent lymphatics, inhibiting or preventing CLEVER-1 binding malignant cells from binding to endothelial cell CLEVER-1 will inhibit or
35 prevent such CLEVER-1 binding malignant cell from entering and establish-

ing in the lymph node, and thus lessen or prevent metastasis of such cell to the lymph node or thus to other sites in the body. In this context it is worth to note that a metastasizing malignant cell cannot survive long times without matrix support - a condition present for example in blood. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding malignant cells in the afferent lymphatics and at HEV in systemic vasculature. The invention also provides a method of inhibiting metastasis, by administering an agent that inhibits or prevents afferent lymphatic CLEVER-1 mediated endothelial cell binding, and especially HEV binding, to such malignant cells.

In the efferent lymphatics, inhibiting or preventing CLEVER-1 binding malignant cells from binding to endothelial cell CLEVER-1 will inhibit or prevent such CLEVER-1 binding malignant cell from leaving the lymph node, and thus lessen or prevent metastasis of such cell from the lymph node to other sites in the body. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding malignant cells in the efferent lymphatics. The invention also provides a method of inhibiting metastasis, by administering an agent that inhibits or prevents efferent lymphatic CLEVER-1 mediated endothelial cell binding to such malignant cells.

CLEVER-1 interaction with CLEVER-1 binding malignant cells thus presents a unique, three-prong approach to inhibit or prevent metastasis in which not only can the artisan block such malignant cells from entering into and exiting from the lymph system, but also, the artisan can block association of such malignant cell with CLEVER-1 in the vascular endothelium.

30 **Agents that Block or Inhibit CLEVER-1 Mediated Cell Binding**

Soluble CLEVER-1 and antibodies to CLEVER-1 can be provided to the host cell to block or inhibit CLEVER-1 mediated cell binding. Soluble CLEVER-1 can be used to "coat" the CLEVER-1 binding sites on the leukocyte, such as lymphocyte, monocyte, or granulocyte, or tumor cell and thus

prevent the coated cell from association with the native CLEVER-1 on the HEV or afferent or efferent lymphatics.

CLEVER-1 antibodies can be administered to a patient in need of the same to coat CLEVER-1 that is present on the vascular endothelium or lymphatics, of such patient, especially the afferent lymphatics so as to prevent leukocyte or malignant cell binding to such CLEVER-1 in the patient. CLEVER-1 antibody producing cells can be administered directly to the patient so as to provide a source of the same.

Moreover, the present invention provides a method of identifying an agent that inhibits the binding of CLEVER-1 to cells by providing an agent to cells in the presence of CLEVER-1 and comparing the binding of CLEVER-1 to cells provided with the agent to binding of CLEVER-1 in the absence of the agent. Similarly, the invention provides a method of identifying an agent that stimulates the binding of CLEVER-1 to cells by providing an agent to cells in the presence of CLEVER-1 and comparing the binding of CLEVER-1 to cells provided with the agent to binding of CLEVER-1 in the absence of said agent.

The term "antibody" is used in the broadest sense and specifically covers single monoclonal antibodies (including agonist and antagonist antibodies), polyclonal antibodies, as well as antibody fragments and single chain antibodies (e.g., Fab, F(ab')₂, Fv), so long as they exhibit the desired biological activity.

Papain digestion of antibodies produces two identical antigen binding fragments, called Fab fragments, each with a single antigen binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')₂ fragment that has two antigen combining sites and is still capable of cross-linking antigen.

Single chain "Fv" is the minimum antibody fragment which contains a complete antigen recognition and binding site. This region consists of a dimer of one heavy and one light chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen binding site on the surface of the V_H-V_L dimer. Collectively, the six CDRs confer antigen binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recog-

nize and bind antigen, although at a lower affinity than the entire binding site. See, Ladner *et al.*, U.S. Patent No. 4,946,778, and Bird, R.E. *et al.*, *Science*, 242:423-426 (1988).

5 The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal)
10 antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by a hybridoma culture, uncontaminated by other immunoglobulins.
15 The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler and Milstein, *Nature* 256:495 (1975), or may be made by recombinant DNA methods (e.g., Cabilly *et al.*, U.S. Patent No. 4,816,567).
20

Preparation of immunizing antigen, and polyclonal and monoclonal antibody production can be performed as described herein, or using other suitable techniques. A variety of methods have been described (see e.g.,
25 Kohler *et al.*, *Nature* 256:495-497 (1975), and *Eur. J. Immunol.* 6:511-519 (1976); Milstein *et al.*, *Nature* 266:550-552 (1977); Koprowski *et al.*, U.S. Patent No. 4,172,124; Harlow, E. and D. Lane, *Antibodies: A Laboratory Manual* (Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y., 1988); *CURRENT PROTOCOLS IN MOLECULAR BIOLOGY*, Vol. 2 (Supplement 27, 1994),
30 Ausubel, F. M. *et al.*, John Wiley & Sons, eds., New York, N.Y.), Chapter 11, (1991)). Generally, a hybridoma can be produced by fusing a suitable immortal cell line (e.g., a myeloma cell line such as SP2/0) with antibody producing cells. The antibody producing cell, preferably those of the spleen or lymph nodes, are obtained from animals immunized with the antigen of interest.
35 The fused cells (hybridomas) can be isolated using selective culture

conditions, and cloned by limiting dilution. Cells which produce antibodies with the desired binding properties can be selected by a suitable assay (e.g., ELISA).

The term "antibody" also includes chimeric, humanized or primatized (CDR-grafted) antibodies, as well as chimeric or CDR-grafted single chain antibodies, and the like, comprising portions derived from different species. "Chimeric" antibodies (immunoglobulins) have a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (Cabilly *et al.*, U.S. Patent No. 4,816,567; Morrison *et al.*, *Proc. Natl. Acad. Sci. USA* 81:6851-6855 (1984)). The various portions of these antibodies can be joined together chemically by conventional techniques, or can be prepared as a contiguous protein using genetic engineering techniques. For example, nucleic acids encoding a chimeric or humanized chain can be expressed to produce a contiguous protein. See, e.g., Cabilly *et al.*, U.S. Patent No. 4,816,567; Cabilly *et al.*, EP 0 125 023 B1; Boss *et al.*, U.S. Patent No. 4,816,397; Boss *et al.*, EP 0120694 B1; Neuberger, M.S. *et al.*, WO 86/01533; Neuberger, M.S. *et al.*, EP 0194276 B1; Winter, U.S. Patent No. 5,225,539; Winter, EP 0239400 B1; and Queen *et al.*, U.S. Patent Nos. 5,585,089, 5,698,761 and 5,698,762. See also, Newman, R. *et al.*, *Bio-Technology* 10: 1455-1460 (1992), regarding primatized antibody.

By "agonist antibody" is meant an antibody which is able to bind to CLEVER-1 and facilitate adhesion of lymphocytes (and malignant tumor cells) to endothelium. By "antagonist antibody" is meant an antibody that is able to bind to CLEVER-1 and inhibit adhesion of lymphocytes (and malignant tumor cells) to endothelium.

Anti-idiotypic antibodies are also provided. Anti-idiotypic antibodies recognize antigenic determinants associated with the antigen-binding site of another antibody. Anti-idiotypic antibodies can be prepared against second antibody by immunizing an animal of the same species, and preferably of the

same strain, as the animal used to produce the second antibody. See *e.g.*, U.S. Patent No. 4,699,880.

In Vitro Adhesion Assay and Diagnostic Uses Thereof

5 In a further embodiment, the present invention is directed to an adhesion assay in which CLEVER-1 binding is used to assay for the presence of leukocytes or malignant cells that bind to HEV and lymphatic endothelium. Both static and non-static assays are possible. The adhesion assay is exemplified in Example 4. Both static and non-static assays can be used to study leukocyte binding to systemic vasculature.

10 In the static assay, a tissue section is exposed to leukocytes or malignant cells for a desired period of time, without continuous agitation or rotation of the preparation during the exposure. Static assays are preferred for examining the ability of leukocytes and malignant cells to bind to lymphatic endothelium, especially the efferent lymph vessels.

15 In the non-static assay, the CLEVER-1 containing tissue sample and leukocytes are constantly rotated during the time period in which the leukocytes are given to adhere to the CLEVER-1. Non-static assays are preferred for studying leukocyte and malignant cell binding to CLEVER-1 in the HEV. The non-static assay mimics adhesion to the systemic vasculature.

20 In another embodiment, the present invention relates to a method for detection of malignant tumor cells. As explained with detail in Example 1, CLEVER-1 antibodies reduce the binding of malignant tumor cells to the vascular and lymphoid endothelium, demonstrating CLEVER-1 is a receptor for such malignant tumor cells. CLEVER-1 protein, or fragments thereof, or
25 CLEVER-1 binding compounds, including but not restricted to, antibodies against CLEVER-1, both monoclonal and polyclonal, antibodies against antigenic fragments of CLEVER-1, both monoclonal and polyclonal, antigenic polypeptides, small molecule inhibitors or drugs, can be used in both quantitative and qualitative assays to detect the presence of malignant tumor cells
30 in a sample, said sample being tissue or blood from a human or animal.

CLEVER-1 protein, or fragments thereof, or the above-mentioned CLEVER-1 binding compounds can be attached to a solid support matrix, including but not limited to microtiter plates, agarose columns, or magnetic beads. The above-mentioned sample can then be exposed to said solid

support matrix, and the percentage of cells retained by said solid support matrix determined. For example, a sample from a normal, healthy individual, said individual being either a human or an animal, would have a statistically predicted number of leukocytes that bind to CLEVER-1. A sample that contains both leukocytes and malignant tumor cells would have a detectably higher number of cells that bind CLEVER-1.

In a preferred embodiment, a blood or tissue sample that has been taken from a patient who is in need of treatment, especially treatment for cancer, is used as the source of the CLEVER-1 binding cells in the *in vitro* adhesion assay. Such patient can be a patient being treated for a previously diagnosed malignancy, or a patient suspected of having a malignant tumor, or a patient who appears to be cured of such malignant tumor but is in need of monitoring for the reoccurrence of the same. Preferably, such blood or tissue sample is from a patient who is to be tested for the presence of malignant cells that bind to CLEVER-1 in such sample.

The blood or tissue sample that is to be examined in the *in vitro* assay of the invention can be processed, if desired, by methods known in the art so as to further extract or concentrate any CLEVER-1 binding cells that might be present in the sample, prior to the sample's being used in the *in vitro* adhesion assay of the invention.

Additionally, once the *in vitro* adhesion assay is complete, the adherent cells can be studied using other methods. For example, in the static assay, where the bound cells have been fixed, ability of the adherent cells to be recognized by a monoclonal antibody that is diagnostic for the type of tumor can be performed.

Detecting the binding of malignant cells to the CLEVER-1 containing lymphatic endothelium indicates that the patient is in need of treatment for such malignant cells, and especially to prevent the metastasis of such malignant cells

The present invention provides in this aspect a novel, efficient, and convenient assay for identifying antagonists, including but not limited to, monoclonal and polyclonal antibodies, peptides, protein fragments, small molecular inhibitors, drugs, and other agents, which can inhibit the adhesion of leukocytes and malignant tumor cells to the vascular and lymphatic endothelium.

For example, CLEVER-1 containing samples of lymph node sections can be incubated with and without the agent, and the number of bound lymphocytes and/or malignant cells determined. The antagonists can be pre-incubated with lymph node sections (a non-competitive assay) or simultaneously added with lymphocytes to the lymph node sections (a competitive assay).

Such a screen can also be used to customize an anti-metastasis treatment to an individual patient, and allows the practitioner to identify and select those agents or combinations thereof that have the best ability to inhibit CLEVER-1 malignant cell binding to vascular and/or lymphatic endothelium in such patient, and thus maximize the benefit of the treatment with such agents for such patient.

Additionally, such *in vitro* assay allows the practitioner to select for agents that provide a beneficial effect on disrupting malignant cell: CLEVER-1 containing endothelium interactions, nevertheless, minimize, if possible, the effect of such treatment on CLEVER-1 mediated leukocyte binding,

An antagonist can inhibit malignant cell or lymphocyte cell migration into or out of the lymph nodes. In a preferred embodiment, antagonists would inhibit both entrance and exit of an undesired cell into and out of the lymph nodes, respectively. As such, malignant tumor cells would preferably be prevented from entering a lymph node, and establishing there, and any that did enter the lymph nodes via an afferent lymph vessel independent mechanisms would be contained, thus slowing metastasis.

This assay can be used further to monitor the efficacy of chemotherapy treatments administered to an individual, said individual being a human or an animal, in need thereof. Samples can be analyzed before, during, and after chemotherapy for the presence of malignant tumor cells that bind to CLEVER-1 or antigenic fragments thereof, or CLEVER-1 binding compounds.

In a further embodiment, purified CLEVER-1 protein, or fragments thereof can be used for high volume screening of antagonists that are capable of preventing or lowering the ability of a leukocyte or malignant cell to adhere to endothelial cell CLEVER-1. CLEVER-1 protein, or fragments thereof can be attached to a solid support matrix, including but not limited to a microtiter plate, an agarose column, or magnetic beads, using standard methods

well known in the art. Antagonists can be screened for interaction with CLEVER-1 or fragments thereof, either in the absence or presence of leukocytes. Leukocytes or malignant cells can be labeled with fluorescent dyes such as, for example, bis-carboxyethyl carboxyfluorescein or fluorescein isothiocyanate and the number of bound cells in presence or absence of the
5 antagonists can be analyzed by a fluoroimager.

The high volume screen assay of this aspect of the invention can be used to screen combinatorial libraries for molecules that inhibit the binding of leukocytes and malignant tumor cells to CLEVER-1 or a fragments thereof.
10 Antagonists that show strong affinity for purified CLEVER-1 protein or fragments thereof can be screened further using the *in vitro* adhesion assay described above.

Antibodies used in the methods of the invention as CLEVER-1 binding compounds are preferably antibodies with a specificity against
15 CLEVER-1, or an antigenic fragment thereof. Such antibodies can be polyclonal or monoclonal.

Another potential CLEVER-1 antagonist is a peptide derivative of the CLEVER-1 polypeptide that are naturally or synthetically modified analogs of the polypeptides that have lost biological function yet still recognize
20 and bind to the ligand of the polypeptides to thereby effectively block the interaction of said ligand with CLEVER-1. Examples of peptide derivatives include, but are not limited to, small peptides or peptide-like molecules.

Another potential human CLEVER-1 antagonist is a peptide derivative of the ligand polypeptides which are naturally or synthetically modified
25 analogs of the polypeptides that have lost biological function yet still recognize and bind to CLEVER-1 to thereby effectively block CLEVER-1. Examples of peptide derivatives include, but are not limited to, small peptides or peptide-like molecules.

The present invention relates to a diagnostic method for the detection
30 of cells that contain CLEVER-1, that is, CLEVER-1 positive cells, in samples taken from the human or animal body. Such a method would involve the use of CLEVER-1 binding compounds, including but not limited to, monoclonal and polyclonal antibodies, with specificity for CLEVER-1. Such compounds can be labeled with a substance, such as a colorimetric dye or

radioactive molecule, to permit rapid and easy detection of binding of the compound to cells that express CLEVER-1.

Therapeutic Uses of CLEVER-1 Antagonists

In another embodiment, the present invention relates to a method
5 of treating malignant carcinomas. It is common for carcinomas to metasta-
size first to the regional lymph nodes (Sleeman, J.P., *Recent Results Cancer
Res. 157:55-81 (2000)*). As described herein, CLEVER-1 is involved in the
entrance and exit of malignant tumor cells to and from the lymph nodes. As
such, antagonists that inhibit malignant tumor cell binding to CLEVER-1, in-
10 cluding but not limited to, monoclonal and polyclonal antibodies, peptides,
small molecule inhibitors, drugs, and other such agents can reduce metasta-
sis and serve as effective chemotherapeutic agents.

In another aspect, the present invention relates to a method of
treating disorders where the leukocyte-endothelial cell adhesion reaction is
15 associated with acute or chronic inflammatory diseases such as skin inflam-
mations, diabetes, connective tissue diseases (such as lupus, rheumatoid
arthritis, osteoarthritis), obstructive and restrictive lung diseases (such as
asthma, ARDS, sarcoidosis, idiopathic pulmonary fibrosis), inflammatory
bowel diseases (such as ulcerative colitis and Crohn's disease), various ne-
20 phritides, non-viral hepatitis, cirrhosis, cholangitis, atherosclerosis, vasculitis,
thyroiditis, multiple sclerosis, myositis, ischemia reperfusion injury, transplan-
tation rejection.

The antagonists can also be employed to treat histamine-mediated
allergic reactions and immunological disorders including late phase allergic
25 reactions, chronic urticaria, and atopic dermatitis. IgE-mediated allergic reac-
tions such as allergic asthma, rhinitis, and eczema can also be treated.

The antagonists can also be employed to treat chronic and acute
inflammation by preventing the extravasation of leukocytes to a wound area.
They can also be employed to regulate normal pulmonary macrophage popu-
30 lations, since chronic and acute inflammatory pulmonary diseases are asso-
ciated with sequestration of mononuclear phagocytes in the lung.

Antagonists can also be employed to treat rheumatoid arthritis by
preventing the extravasation of leukocytes into synovial fluid in the joints of

patients. Monocyte influx and activation plays a significant role in the pathogenesis of both degenerative and inflammatory arthropathies.

The antagonists can also be employed to treat asthma and allergy by preventing eosinophil accumulation in the lung. The antagonists can also
5 be employed to treat subepithelial basement membrane fibrosis which is a prominent feature of the asthmatic lung.

The antagonists can also be employed for treating atherosclerosis, by preventing monocyte infiltration in the artery wall.

The antagonists can be employed in a composition with a pharmaceutically acceptable carrier, e.g., as hereinafter described.
10

Formulations of Compounds

The antagonists of CLEVER-1 can be used as therapeutic compositions. The antagonists of CLEVER-1 can be administered as a single dose or in multiple doses. The antagonists of the present invention can be administered either as an independent therapeutic regime or in combination with
15 other therapeutic agents. The antagonists can be combined with conventional therapies, which can be administered simultaneously or sequentially.

Such therapeutic compositions can consist solely of the antagonist of CLEVER-1 although, preferably, the compositions will contain the antagonist of CLEVER-1 combined in admixture with a pharmaceutically acceptable
20 carrier vehicle. Suitable vehicles and their formulation, inclusive of other human proteins, e.g., human serum albumin, are described for example in Remington: The Science and Practice of Pharmacy, Gennaro, Alfonso, 20th ed. (2000). In order to form a pharmaceutically acceptable composition that is suitable for effective administration to a patient in need of such composition, such compositions will contain an effective amount of the antagonist of
25 CLEVER-1 together with a suitable amount of carrier vehicle.

Compositions containing antagonists of CLEVER-1 can be administered perorally, intravenously, intramuscularly, or sub-cutaneously at the
30 appropriate dosages, which will depend upon the severity of the condition of the patient and upon such criteria as the patient's height, weight, sex, age, and medical history. The dose will also depend upon whether the compound of the invention is being administered to a human patient or in a veterinary setting to an animal, in need thereof.

For the purpose of parenteral administration, compositions containing the antagonists of CLEVER-1 are preferably dissolved in distilled water and the pH-value is preferably adjusted to about 6 to 8. In order to facilitate the lyophilization process resulting in a suitable product, lactose can be added to the solution. Preferably, the solution is then filtered sterilized, introduced into vials, and lyophilized. In a preferred embodiment, the compound of the invention is administered orally to a patient, at the time of eating or shortly thereafter. The concentration of the antagonists of CLEVER-1 in these composition, whether oral or parenteral, can vary, e.g., from 10^{-12} M to 10^{-3} M.

Additional pharmaceutical methods can be employed to control the duration of action. Controlled release preparations can be achieved by the use of polymers to complex or adsorb the antagonists of CLEVER-1. The controlled delivery can be exercised by selecting appropriate macromolecules (for example, polyesters, polyamino acids, polyvinyl pyrrolidone, ethylenevinylacetate, methylcellulose, carboxymethylcellulose, and protamine sulfate) and the concentration of macromolecules as well as the methods of incorporation in order to control release. Another possible method to control the duration of action by controlled release preparations is to incorporate the antagonists of CLEVER-1 into particles of a polymeric material such as polyesters, polyamino acids, hydrogels, poly (lactic acid) or ethylene vinylacetate copolymers. Alternatively, instead of incorporating the CLEVER-1 antagonists into these polymeric particles, it is possible to entrap these derivatives in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly (methylmethacrylate) microcapsules, respectively, or in colloidal drug delivery systems, for example, liposomes, albumin microspheres, microemulsions, nanoparticles, and nanocapsules or in macroemulsions. Such teachings are disclosed in Remington: The Science and Practice of Pharmacy, Gennaro, Alfonso, 20th ed. (2000).

The following Example serves only to illustrate the invention, and is not to be construed as in any way limiting the invention.

EXAMPLE 1

Production of Monoclonal Antibodies

Balb/c mice were immunized to footpads four times at one week intervals, with incomplete Freund's adjuvant containing suspension made
5 from lymphatic vessels excised from human lymph nodes under stereo microscope. The suspension was made by cutting the vessels to small pieces by scissors and the pieces in phosphate buffered saline were then drawn back and forth into a syringe connected to a 21 g needle. The popliteal lymph node lymphocytes from the immunized mice were isolated by a glass
10 homogenizer. The popliteal lymph node lymphocytes of the immunized mice were fused with Sp2/0 myeloma cells. Hybridoma supernatants were primarily tested on frozen sections of human lymph nodes using immunoperoxidase staining. The testing conditions were the same for antibodies 3-266 and 3-372 generated by two of the hybridomas.

15 Immunoperoxidase stainings were performed as described (Salmi, *Science* 257:1407-1409 (1992)). Briefly, acetone fixed 6 μ m frozen sections from different human tissues (lymph nodes, appendix, bronchus, cerebellum, epididymis, esophagus, heart, small and large intestine, kidney, liver, lung, normal and psoriatic skin, synovium, testis and tonsil) were stained with anti-
20 body 3-266, 3-372 or 3G6, a negative class matched control antibody for 3-266 and 3-372 (mouse IgG1) and 3,3-diaminobenzidine was used as a substrate. Procedures for tissue collection were approved by the Local and National Boards of Medicolegal Affairs in Finland.

Two of the hybridomas produced antibodies (3-266 (DSM
25 ACC2519) and 3-372 (DSM ACC2590)) that clearly stained lymphatic endothelium both in afferent and efferent lymphatic systems and vascular endothelium on HEV, while the other structures remained unstained. The staining of the lymphatic endothelium is shown in Figure 1. Figure 1 is an indirect immunoperoxidase staining that shows that monoclonal antibodies 3-266 and
30 3-372 recognize endothelium both in afferent and efferent lymphatic systems and on HEV. Figures 1a-1c are from the skin. Figures 1d-1i are from a lymph node. Figures 1a, 1d and 1g show the staining with monoclonal antibody 3-266, Figures 1b, 1e and 1h show the staining with monoclonal antibody 3-372 and Figures 1c, 1f and 1i show the staining with a negative con-

-27-

5 trol antibody, 3G6. In Figures 1a, 1b and 1c, the arrows point to the epithelium and arrowheads to afferent lymphatics. In Figures 1d and 1e, the arrows point to the lymphatic vessels (lymphatic sinusoids that belong to the efferent lymphatic system) within the lymph node. In Figures 1g and 1h, the arrows point to HEV.

EXAMPLE 2

Determination of Molecular Weight of CLEVER-1

Molecular weight determination was performed by immunoblotting. One percent NP-40 lysates containing of human lymph nodes was analyzed using 5-12.5% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE). SDS-PAGE was run in non-reducing conditions. The molecules in the gel were blotted overnight to nitrocellulose sheets and probed with 3-266, 3-372 or a negative control antibody (3G6) (Salmi, M. *et al.*, *J. Exp. Med.* 183:569-579 (1996). Peroxidase conjugated rabbit anti-mouse Ig was used as the second stage reagent. Detection was performed using enhanced chemiluminescence system according to the instructions of the manufacturer (Amersham).

Both antibodies recognized a molecule of the same size (about 270-300 kDa; Figure 2). Due to this and an identical staining pattern, these antibodies were assumed to recognize the same antigen and this antigen was named CLEVER-1.

EXAMPLE 3

Purification and Molecular Characterization of CLEVER-1

The molecule recognized by 3-372 antibody was purified from human lymph node lysate overnight (lysis buffer: 150 mM NaCl, 10 mM Tris-base, pH 7.2, 1.5 mM MgCl₂, 1% NP-40, 1% aprotinin, and 1 mM PMSF) as described in Smith, D.J. *et al.*, *J. Exp. Med.* 188:17-27 (1998). After centrifugation, the lysate supernatant was applied sequentially to immunoaffinity columns containing CnBr-activated Sepharose beads armed with irrelevant mAbs and 3-372 (3 mg/ml beads). After washing with lysis buffer, the antigens recognized by 3-372 were eluted with 50 mM triethylamine, frozen and subsequently lyophilized. The eluted material was then subjected to SDS-

PAGE analysis and silver staining (O'Connell, K.L. and Stults, J.T., *Electrophoresis* 18:349-359 (1997)). The specific band was excised, reduced, alkylated and digested with trypsin (Promega) overnight at +37°C as described (Shevchenko, A. *et al.*, *Anal. Chem.* 68:850-855 (1996); O'Connell, K.L. and Stults, J.T., *Electrophoresis* 18:349-359 (1997)). The peptides were analyzed using PerSeptive BioSystems Voyager DE-PRO mass spectrometer operated in the reflectron delayed-extraction mode. Calibration of the spectrum was performed internally by using autolysis products of trypsin or with added calibration mixture 2 (PerSeptive BioSystems). Database search was performed by MS-Fit algorithm (http://prospector.ucsf.edu/_ucsfhtml3.2/msfit.htm) of the University of California, San Francisco mass spectrometry facility.

After cleavage with trypsin, mass spectrometric analyses yielded 27 peptides. 21 (77%) of those had identical nucleotide sequences with two Genebank entries: AJ 275213, a submission for a cDNA clone called stabilin-1, and D87433, a cDAA clone KIAA0246 isolated from the cell line KG-1. The peptide sequences covered altogether 268 amino acids (10% of the 2570 amino acids of stabilin-1) and spanned the amino acids between 53 and 2301.

Next we designed primers based on the peptide sequences, the 5' end of the cDNA for stabilin-1 and the 3' end of the cDNA of KIAA0246 and used them to make several RT-PCR fragments that were then ligated together to clone the full-length 7879 bp cDNA (SEQ ID NO:1). Sequencing of the whole construct revealed a high homology with the existing 3' end KIAA0246 sequence available in the data bank. However, it contained 4 nucleotide differences, when compared to the Stabilin sequence. They all cause a change at the amino acid level. Two of these changes are identical with the genomic sequence data available from the HUGO project (AC 006208), but since the genomic clone only covers about half of the gene for this cDNA, the nature of the two other changes remains to be determined.

Sequencing of several different CLEVER-1 cDNA-clones also revealed the existence of at least two alternatively spliced isoforms of the molecule: the regions covered by exons 23 (nucleotides 2377-2562) and 27 (nucleotides 2914-3009) can be spliced out. We could confirm the existence of one of those splice variants (lacking exon 27) also at the mRNA level (Figure 7) but the second one (lacking exon 23) that we cloned from a human

peripheral lymph node library was not visible in the system suggesting a low abundance/ turnover of the mRNA encoding this form.

The sequence comparisons revealed significant homologies to proteoglycan link protein-like sequence, epidermal growth factor-like repeats
5 and two RGD motifs being well in line with the adhesive properties of CLEVER-1.

EXAMPLE 4

In Vitro Adhesion Assay

Lymph node sections were first incubated with 3-266, 3-372 or
10 control antibodies against human HLA ABC (HB-95, ATCC and 3G6 (against chicken T cells) and then overlaid with Ficoll gradient (Pharmacia) purified peripheral blood mononuclear cells or different human tumor cell lines (lymphoblastoid cell lines, KCA and IBW4; a Burkitt lymphoma CRL-1648; squamocellular carcinoma lines NA and NU). Thereafter, the sections were
15 subjected to two different types of assays: 1. A non-static assay, which optimally measures binding of cells to HEV and is performed under rotatory conditions (60 rpm on orbital shaker for 30 min at +7°C). 2. A static assay, in which the sections overlaid with cells are let to stay in static conditions for 15 min, followed by 5 min of rotation at 60 rpm and then again 15 min without
20 rotation at 7°C. (Static conditions were needed to optimal binding to lymphatic endothelium). The adherent cells were fixed in 1% glutaraldehyde. The number of lymphocytes bound to HEV and to sinusoidal (lymphatic) endothelium was counted single blind under dark-field illumination in which setting the sinusoidal vessels are easily recognizable. The results of the inhibition
25 assays are presented as percentage of control binding (the number of adherent cells/vessel in the presence of control mAb defines 100% adherence).

When the assay was performed in non-static conditions, mimicking the blood flow, lymphocyte binding to HEV was reduced 43.6% and 45.2% by
30 3-266 and 3-372, respectively (Figure 3A). To mimic the conditions at sites of lymphocyte exit, the assay was performed in static conditions. In these assays lymphocyte binding to lymphatic endothelium was decreased 46.4% and 64% by 3-266 and 3-372, respectively (Figure 3B). These data indicate

that the molecule recognized by 3-266 and 3-372 indeed mediate lymphocyte binding both to HEV and to lymphatic endothelium.

To study the role of this molecule in migration of malignant cells, the assays were performed using three lymphoma cell lines (CRL 1648, KCA and IBW4) and two squamocellular carcinoma cell lines (NA and NU). For these assays 3-372 antibody was chosen because of its higher inhibitory capacity (Figure 3). The results of these experiments clearly demonstrated that CLEVER-1 is also involved in binding of malignant cells to endothelium both at entrance and exit sites within the lymph nodes (Figures 4A and 3).

10 EXAMPLE 5

CLEVER-1 is Upregulated at Sites of Inflammation on HEV-like Vessels

Synovial samples from 18 patients suffering from chronic arthritis and undergoing synovectomies, skin samples from diseased skin of patients suffering from psoriasis (n=5), lichen (n=1), mycosis fungoides (n=1), erythrodermia (n=2), exanthema (n=1), folliculitis (n=3) and normal skin samples from 15 individuals were studied for expression of CLEVER-1 using immunoperoxidase method as described above. Like in normal non-lymphoid tissues CLEVER-1 was present in afferent lymphatic vessels in inflamed synovial and both in normal and diseased skin samples. In addition, CLEVER-1 expression was induced on HEV-like vessels that appear at sites of inflammation and are surrounded by heavy infiltrations of inflammatory cells (Figure 5). Table 2 illustrates complete correlation between the extent of inflammatory infiltration and upregulation of CLEVER-1 expression in synovial samples. The same phenomenon was observed in skin samples: all diseased skin samples had inflammatory infiltrations that contained CLEVER-1 positive HEV-like vessels. Those vessels were absent in normal skin samples.

EXAMPLE 6

Clever-1 Also Mediates Binding of Monocytes and Granulocytes to HEV-like Vessels

Human monocytes from peripheral blood were purified from Ficoll-gradient (Pharmacia) isolated mononuclear cells by letting them to adhere to

plastic surfaces for an hour at +37°C. Granulocytes were purified from leukocyte rich buffy coats from human blood using Percoll-gradient (Pharmacia) centrifugation. Their binding was tested to HEV-like vessels in inflamed synovium. In addition, granulocyte binding was tested to tonsil HEV that
5 brightly express CLEVER-1. (When tonsils are removed they always have variable extent of inflammation, although they as a lymphoid organ have HEV without any inflammation). Both granulocytes and monocytes bound efficiently to HEV-like vessels in inflamed synovium and granulocytes adhered avidly to HEV in tonsils. Their binding to these organs was significantly inhibited by the antibody pool containing 3-372 and 3-266 but not with the control
10 antibody (Figure 6).

EXAMPLE 7

CLEVER-1 controls lymphocyte trafficking *in vivo*

In order to verify that CLEVER-1 has a functional role *in vivo*, it
15 was at first confirmed by intravenous injection of 3-372 antibody that rabbits express CLEVER-1 on the surface of endothelium *in vivo*. The presence of CLEVER-1 on HEV was detected after the 3-372 antibody had circulated 5 minutes *in vivo* using frozen sections and FITC labelled anti-mouse IgG second stage antibody after sacrifice (Fig. 8a). In this time frame the intrave-
20 nously given 180 kDa immunoglobulin molecule does not have a possibility to leak and diffuse into the tissue. Based on these results antibody 3-372 (and a class-matched negative control antibody) was given to the rabbits immunized with keyhole limpet hemocyanin to footpads and the effects of the antibody treatment on the size and cellularity of the lymph nodes draining the footpads
25 was analyzed.

Antibody treatment against CLEVER-1 significantly prevented increase of the size of the popliteal lymph nodes (Fig. 8c) indicating that CLEVER-1 has a functional role in lymphocyte traffic *in vivo*. Most likely it exerts its effects both at lymphocyte entrance in HEV and their exit in lymphatic sinusoids, because the rabbits treated with 3-372 antibody had only
30 few lymphocytes in their lymphatic sinusoids when analyzed using histological section (Fig. 8d) Intravenously given 3-372 antibody was also detected to bind CLEVER-1 on lymphatic sinuses when tested at sacrifice 3 days af-

ter the final 3-372 dose. No signal was detected in rabbits which received a control antibody (data not shown).

5 All documents, e.g., scientific publications, patents and patent publications recited herein are hereby incorporated by reference in their entirety to the same extent as if each individual document was specifically and individually indicated to be incorporated by reference in its entirety. Where the document cited only provides the first page of the document, the entire document is intended, including the remaining pages of the document.

Table 1. Matches with CLEVER-1 and stabilin-1

1. 21/27 matches (77%). 275350.0 Da, pI = 6.04. Acc. #6469374. HOMO SAPIENS. (AJ275213) stabilin-1.

m/z submitted	MH ⁺ matched	Delta ppm	start	end	Peptide Sequence (Click for Fragment Ions)	Modifications
775.488	775.483	6.4034	372	377	(R) <u>VFLQLR</u> (V)	
787.36	787.3739	-17.6253	1299	1305	(R) <u>SGFSFSR</u> (G)	
799.495	799.5042	-11.4617	1585	1591	(R) <u>VGLELLR</u> (D)	
812.495	812.4994	-5.4309	1047	1053	(R) <u>TLPNLVR</u> (A)	
917.502	917.4997	2.4556	1040	1046	(R) <u>AFWLQPR</u> (T)	
1017.44	1017.425	14.4862	2295	2301	(R) <u>WDAYCFR</u> (V)	
1104.54	1104.526	12.6406	53	61	(K) <u>QTCPSGWL</u> R(E)	
1212.7	1212.695	3.9482	1021	1032	(R) <u>VTALVPSEAAVR</u> (Q)	
1284.65	1284.622	21.4530	1678	1688	(R) <u>EGSIYLNDFAR</u> (V)	
1291.79	1291.774	12.5434	613	624	(R) <u>ILGPEGVPLQR</u> (V)	
1330.63	1330.575	41.7401	953	965	(R) <u>AGNGGCHGLATCR</u> (A)	
1330.63	1330.633	-1.9754	1873	1882	(R) <u>CDHFETRPLR</u> (L)	
1374.66	1374.632	20.1117	62	72	(R) <u>ELPDQITQDCR</u> (Y)	
1456.79	1456.776	9.6229	1069	1082	(R) <u>LGGQEVATLNPTTR</u> (W)	
1493.8	1493.796	2.4215	508	521	(R) <u>TIGQILASTEAFSR</u> (F)	
1555.7	1555.663	23.5678	219	231	(R) <u>CLPGYTQQGSECR</u> (A)	
1678.94	1678.913	16.1953	1802	1817	(R) <u>NVEALASDLPLNGPLR</u> (T)	
1730.89	1730.887	1.9674	1054	1068	(R) <u>AHFLOGALFEEELAR</u> (L)	
1912.82	1912.832	-6.3742	936	952	(K) <u>LGFAGDGYQCSPIDPCR</u> (A)	
2057.05	2057.03	9.5484	1655	1673	(R) <u>SEDLLEQGYATALS GHPLR</u> (F)	
2165.11	2165.14	-13.6320	1707	1725	(R) <u>VLLPPEALHWEPDDAPIPR</u> (R)	
2295.22	2295.224	-1.5853	389	410	(R) <u>EILTTAGPFTVLVPSVSSFSR</u> (T)	

6 unmatched masses: 871.5410 949.4800 1360.6500 1538.6900 1787.9200 2008.1400

The matched peptides cover 10% (268/2570 AA's) of the protein.
Coverage Map for This Hit (MS-Digest index #): [427477](#)

Table 2. CLEVER-1 Expression Is Induced Mainly on Vessels Surrounded by Lymphocytic Infiltrations in Inflamed Synovia

Expression of CLEVER-1 on HEV-like vessels ¹	The degree of inflammatory infiltration ²	
	0/1 (n=9)	2/3 (n=9)
-/+	100%	0
++/+++	0	100%

¹ Intensity was scored as -, ±, + negative or weak, ++, +++ moderate or strong.

² Degree of the inflammatory cell infiltration in 18 synovial samples was scored as: 0/1, none or few lymphocytes around the vessels, 2/3 marked or massive lymphocytic infiltrations.

WHAT IS CLAIMED IS:

1. A purified lymphatic endothelial glycoprotein, CLEVER-1, having a molecular weight of 270-300 kD in SDS-PAGE under non-reducing conditions, recognisable by a monoclonal antibody selected from the group consisting of
 - (a) monoclonal antibody, DSM ACC 2519; and
 - (b) monoclonal antibody, DSM ACC 2590.
2. The glycoprotein according to claim 1, encoded by a nucleic acid selected from the group of
 - (a) sequence of SEQ ID NO: 1;
 - (b) sequence of SEQ ID NO: 1 lacking nucleotides 2377 – 2562; and
 - (c) sequence of SEQ ID NO: 1 lacking nucleotides 2914 – 3009.
3. A CLEVER-1 antibody.
4. The CLEVER-1 antibody of claim 3, which is monoclonal antibody 3-266 (DSM ACC2519).
5. The CLEVER-1 antibody of claim 3, which is monoclonal antibody 3-372 (DSM ACC2590).
6. A cell-free preparation comprising CLEVER-1.
7. A method of diagnosing inflammatory diseases in a patient, said method comprising:
 - (a) exposing a blood or tissue sample from said patient to CLEVER-1 *in vitro* for a period of time and under conditions sufficient to allow binding of leukocytes if present in said sample; and
 - (b) detecting leukocytes bound to said CLEVER-1 in said blood or tissue sample.
8. A method of detecting malignant cells in a patient, said method comprising:
 - (a) exposing a blood or tissue sample from a patient to CLEVER-1 *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample; and

(b) detecting whether any malignant cells bound to said CLEVER-1 in said blood or tissue sample.

9. The method of claim 8, wherein said CLEVER-1 is on a solid support.

10. The method of claim 8, wherein said CLEVER-1 is provided on lymphoid tissue.

11. The method of claim 8, wherein said CLEVER-1 is present in the membrane of endothelial cells.

12. The method of claim 8, wherein said CLEVER-1 is in a soluble form.

13. The method of claim 8, wherein said detection step is performed by imaging.

14. A method of identifying an agent that inhibits the binding of CLEVER-1 to cells, said method comprising:

- (a) providing an agent to cells in the presence of CLEVER-1; and
- (b) comparing the binding of CLEVER-1 to cells in (a) to binding of CLEVER-1 in the absence of said agent.

15. A method of identifying an agent that stimulates the binding of CLEVER-1 to cells, said method comprising:

- (a) providing an agent to cells in the presence of CLEVER-1; and
- (b) comparing the binding of CLEVER-1 to cells in (A) to binding of CLEVER-1 in the absence of said agent.

16. A method of removing malignant cells from a sample, said method comprising:

- (a) exposing said malignant cells to CLEVER-1 *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample, and
- (b) separating said CLEVER-1 and the malignant cells that adhere thereto from said sample.

17. A method of treating inflammation in a patient in need of the same, said method comprising administering an agent that inhibits CLEVER-1 mediated leukocyte binding to said patient, wherein said inhibiting agent is selected from the group comprising:
- (a) CLEVER-1 antibodies or fragments thereof; and
 - (b) soluble CLEVER-1 or fragments thereof.
18. A method of preventing metastasis in a patient in need of the same, said method comprising administering an agent that inhibits CLEVER-1 mediated malignant cell binding to said patient, wherein said inhibiting agent is selected from the group comprising:
- (a) CLEVER-1 antibodies or fragments thereof; and
 - (b) soluble CLEVER-1 or fragments thereof.
19. The method of claim 18, wherein said CLEVER-1 mediated cell binding inhibits leukocyte binding.
20. The method of claim 18, wherein said CLEVER-1 mediated cell binding inhibits lymphocyte binding.
21. The method of claim 18, wherein said CLEVER-1 mediated cell binding inhibits monocyte binding.
22. The method of claim 18, wherein said CLEVER-1 mediated cell binding inhibits granulocyte binding.
23. The method of claim 18, wherein said CLEVER-1 mediated cell binding inhibits malignant cell binding.
24. The method of claim 18, wherein said CLEVER-1 binding agent is administered to a patient in need of an inhibition of CLEVER-1 mediated cell binding.
25. The method of claim 24, wherein said patient is in need of treatment of inflammation.
26. The method of claim 24, wherein said patient is in need of treatment for a malignancy or possible malignancy.

27. A method of stimulating CLEVER-1 binding in a patient in need of the same, said method comprising: administering an agent that stimulates CLEVER-1 mediated leukocyte binding to said patient.

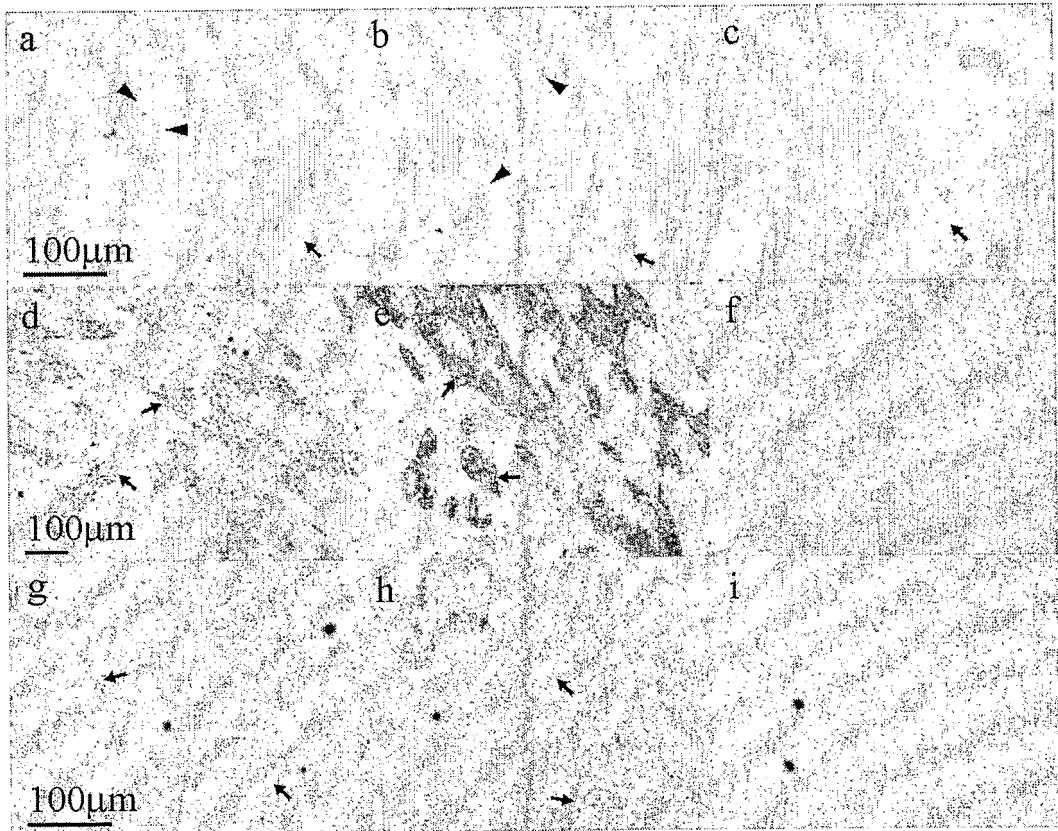


Fig. 1

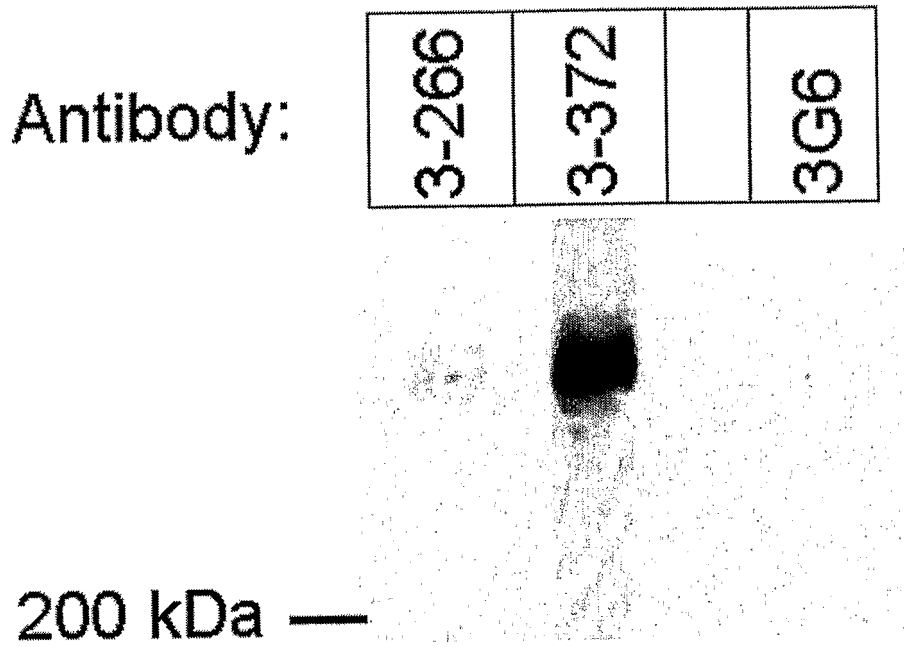
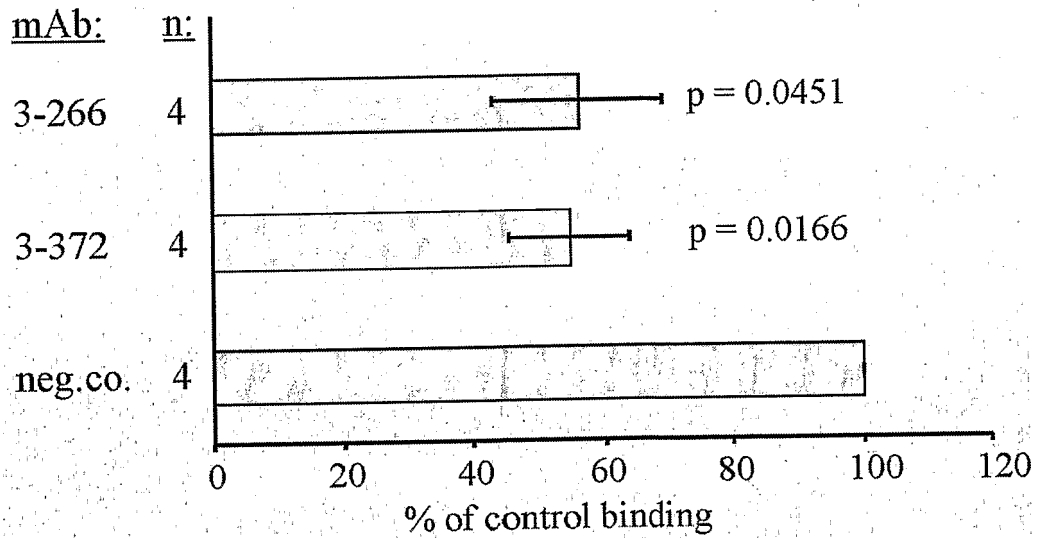


Fig. 2

A Lymphocyte binding to HEVs:



B Lymphocyte binding to lymph endothelium:

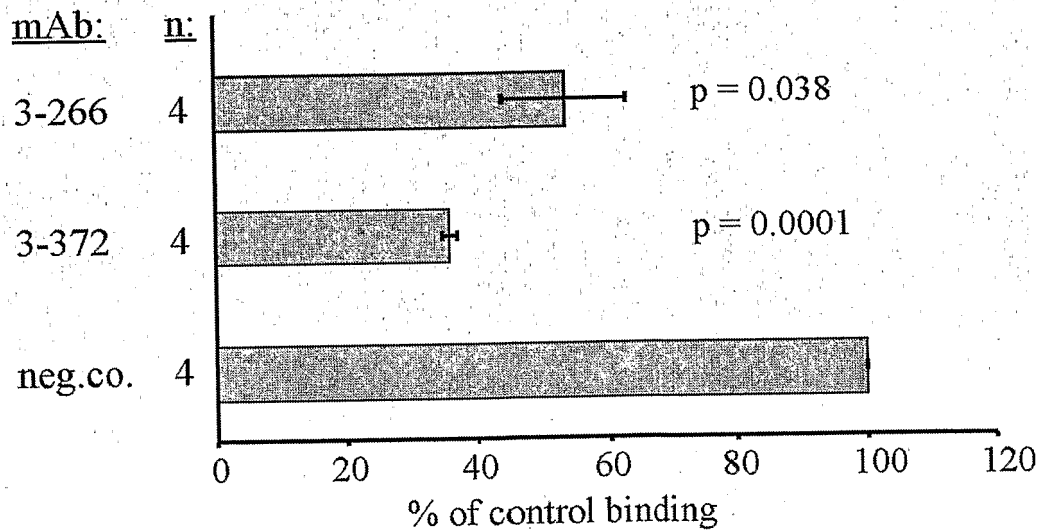
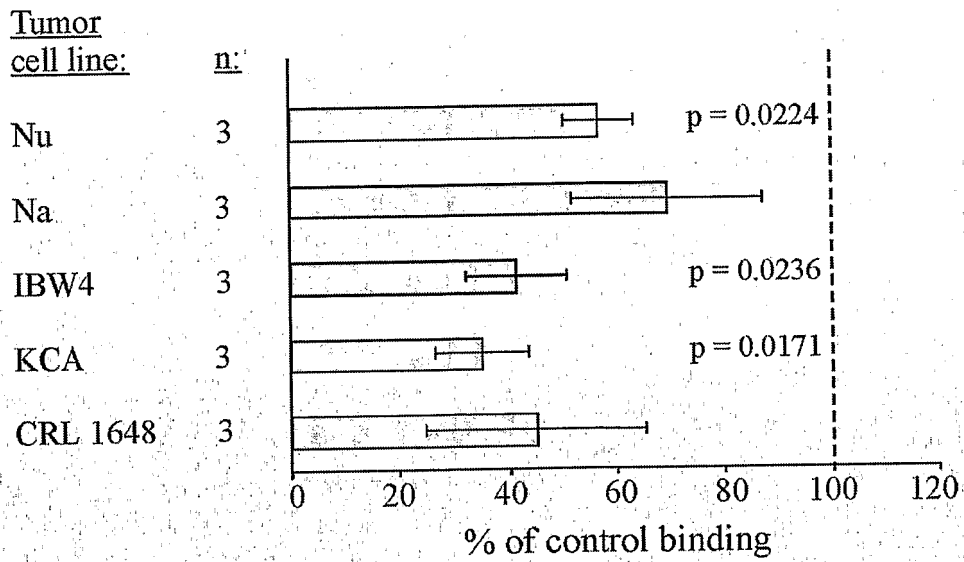


Fig. 3

A Tumor cell binding to HEVs:



B Tumor cell binding to lymphatic endothelium:

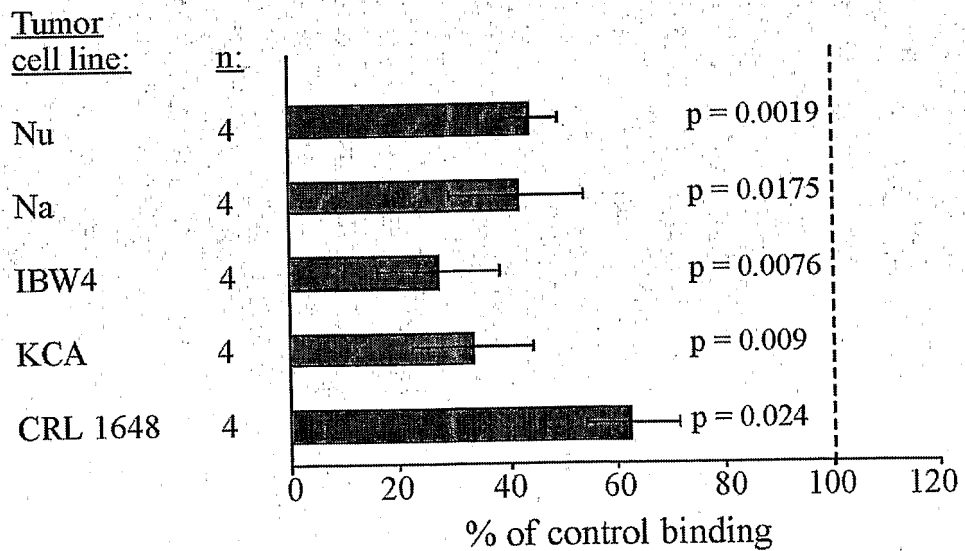


Fig. 4

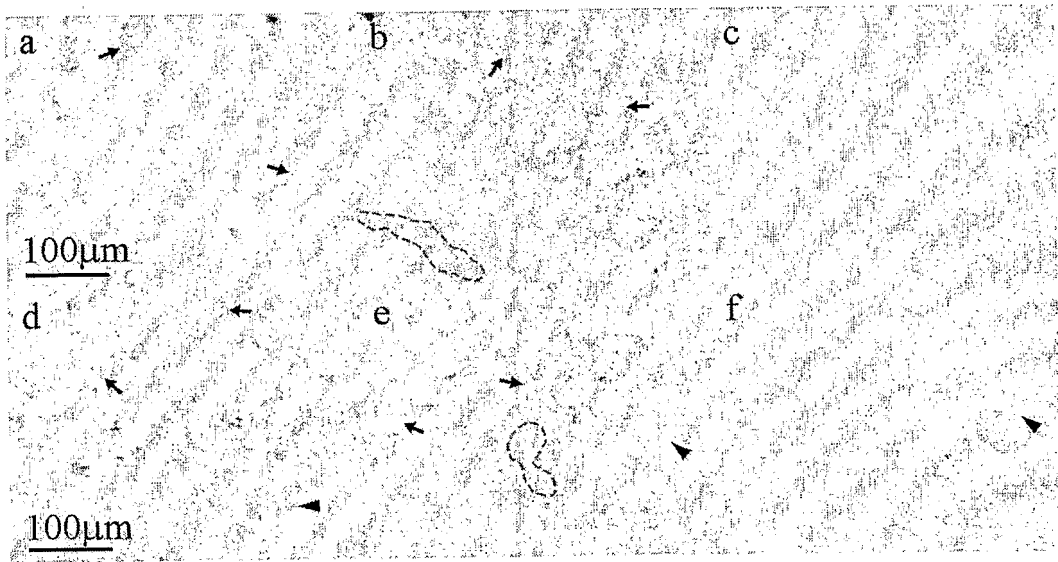


Fig. 5

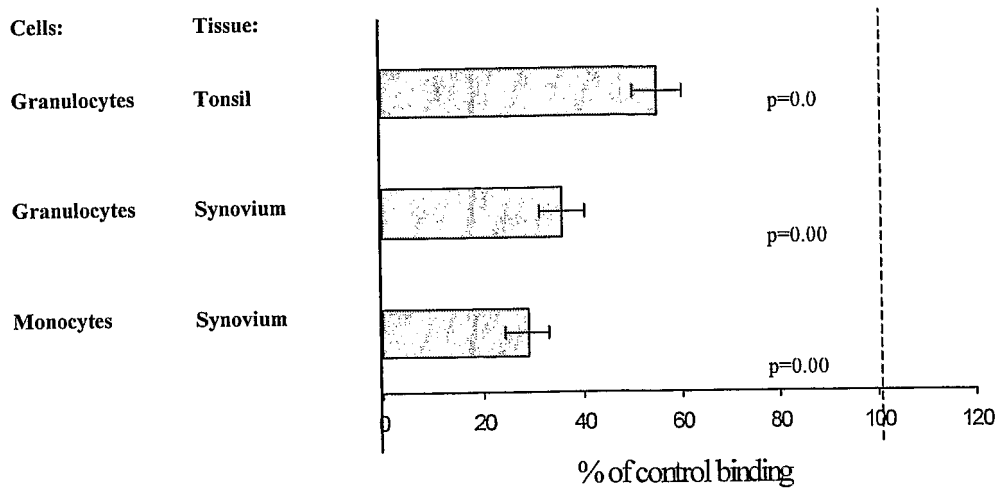


Fig. 6

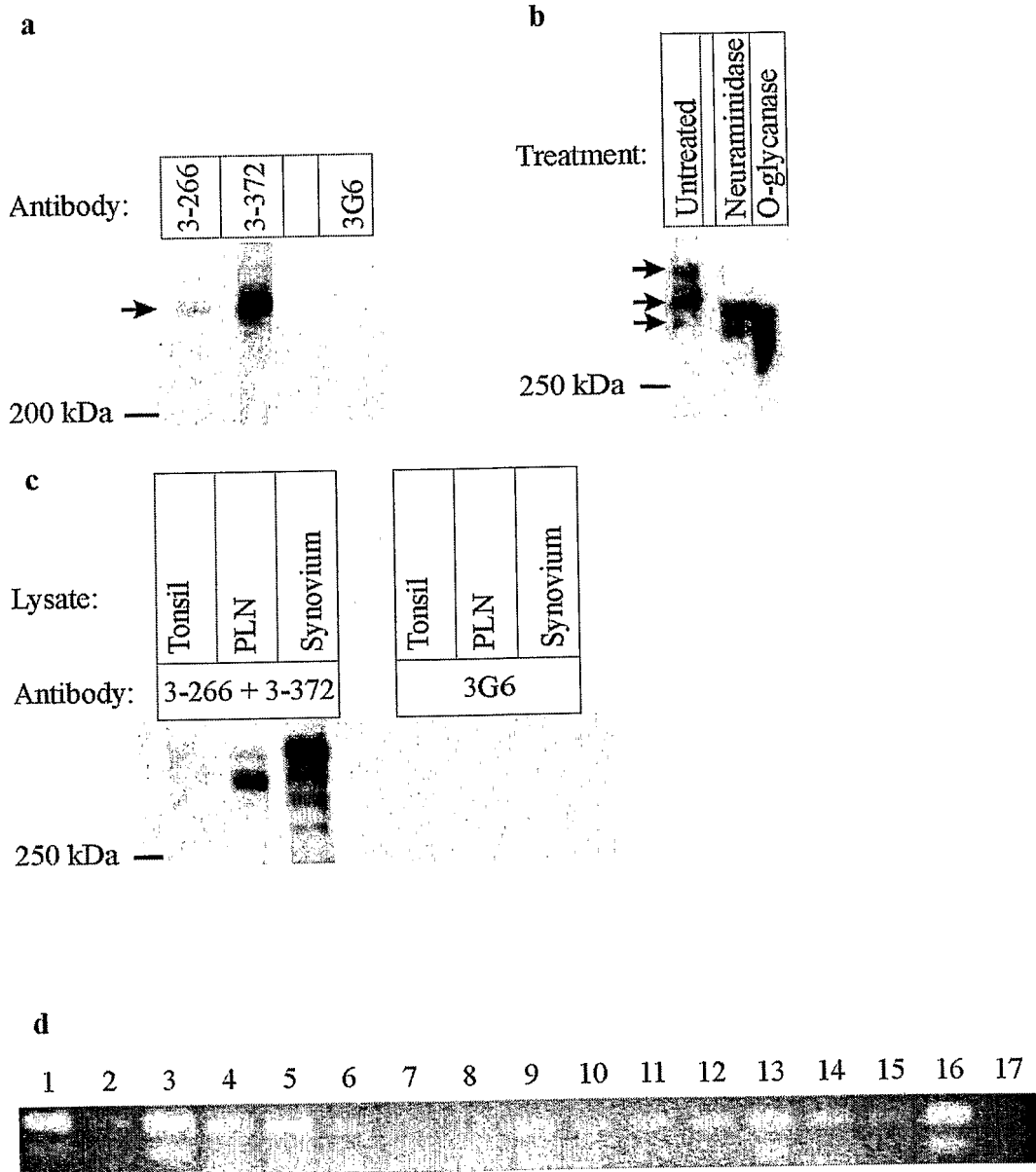


Fig. 7

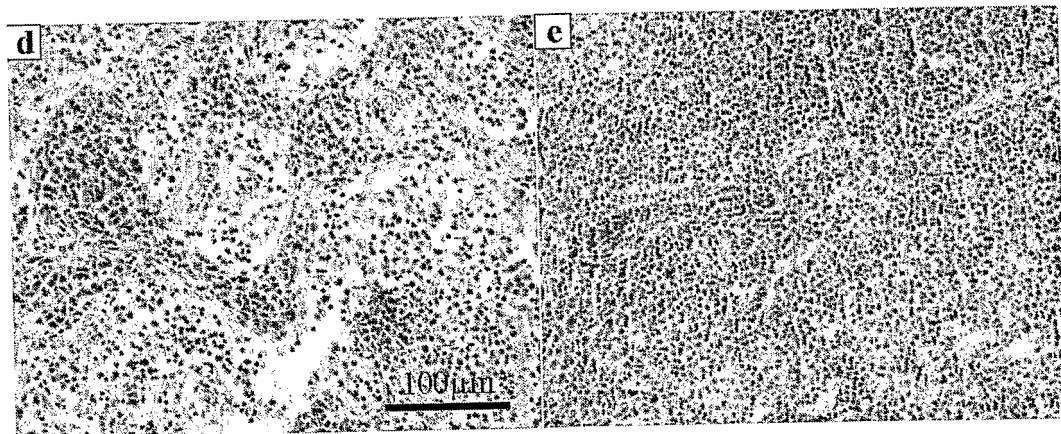
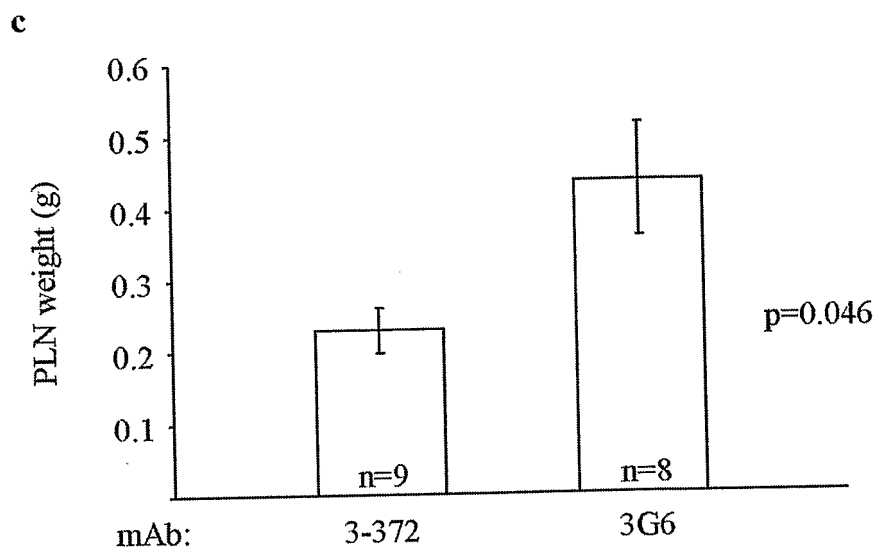
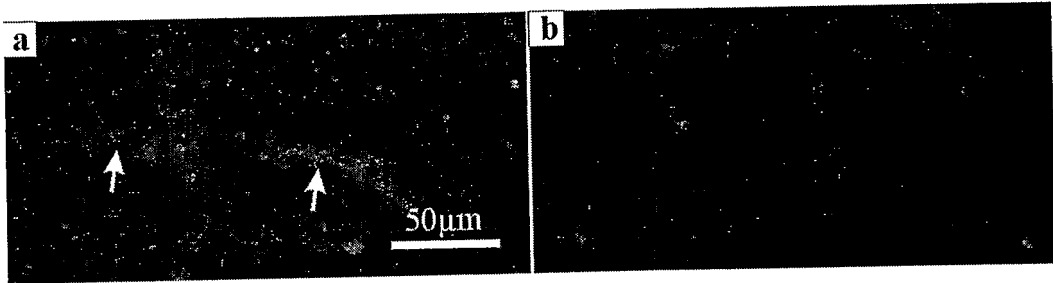


Fig. 8

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Fig. 9

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 ctg gac ccc tgc tct aag aac aat gga gga tgc agc cca tat gcc 4695

Fig. 9 (cont.)

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aca gcc cac acc gtg	ggg gac ggc ctc acc	tgc cgt gcc cga gtc	4785
ggc ctg gag ctc ctg	agg gat aag cat gcc	tca ttc ttc agc ctc	4830
cgc ctc ctg gaa tat	aag gag ctc aag ggc	gat ggg cct ttc acc	4875
atc ttc gtg ccg cac	gca gat cta atg agc	aac ctg tcg cag gat	4920
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Fig. 9 (cont.)

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Fig. 9 (cont.)

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Gln	Thr	Ile	Met	Glu	Gln	Gly	Cys	Cys	Lys	Gly	Phe	Phe	Gly	Pro	Asp	
		715					720					725				
tgc	acg	cag	tgt	cct	ggg	ggc	ttc	tcc	aac	ccc	tgc	tat	ggc	aaa	ggc	2262
Cys	Thr	Gln	Cys	Pro	Gly	Gly	Phe	Ser	Asn	Pro	Cys	Tyr	Gly	Lys	Gly	
	730				735						740					
aat	tgc	agt	gat	ggg	atc	cag	ggc	aat	ggg	gcc	tgc	ctc	tgc	ttc	cca	2310
Asn	Cys	Ser	Asp	Gly	Ile	Gln	Gly	Asn	Gly	Ala	Cys	Leu	Cys	Phe	Pro	
745				750					755						760	
gac	tac	aag	ggc	atc	gcc	tgc	cac	atc	tgc	tcg	aac	cca	aac	aag	cat	2358
Asp	Tyr	Lys	Gly	Ile	Ala	Cys	His	Ile	Cys	Ser	Asn	Pro	Asn	Lys	His	
			765						770					775		
gga	gag	caa	tgc	cag	gaa	gac	tgc	ggc	tgt	gtc	cat	ggg	ctc	tgc	gac	2406
Gly	Glu	Gln	Cys	Gln	Glu	Asp	Cys	Gly	Cys	Val	His	Gly	Leu	Cys	Asp	
			780					785					790			
aac	cgc	cca	ggc	agt	ggg	ggg	gtg	tgc	cag	cag	ggc	acg	tgt	gcc	cct	2454
Asn	Arg	Pro	Gly	Ser	Gly	Gly	Val	Cys	Gln	Gln	Gly	Thr	Cys	Ala	Pro	
		795					800					805				
ggc	ttc	agt	ggc	cgg	ttc	tgc	aac	gag	tcc	atg	ggg	gac	tgt	ggg	ccc	2502
Gly	Phe	Ser	Gly	Arg	Phe	Cys	Asn	Glu	Ser	Met	Gly	Asp	Cys	Gly	Pro	
	810					815					820					
aca	ggg	ctg	gcc	cag	cac	tgc	cac	ctg	cat	gcc	cgc	tgt	gtt	agc	cag	2550
Thr	Gly	Leu	Ala	Gln	His	Cys	His	Leu	His	Ala	Arg	Cys	Val	Ser	Gln	
825				830						835					840	
gag	ggg	gtt	gcc	aga	tgt	cgc	tgt	ctt	gat	ggc	ttt	gag	ggg	gat	ggc	2598
Glu	Gly	Val	Ala	Arg	Cys	Arg	Cys	Leu	Asp	Gly	Phe	Glu	Gly	Asp	Gly	
				845					850					855		
ttc	tcc	tgc	aca	cct	agc	aac	ccc	tgc	tcc	cac	ccg	gac	cgt	gga	ggc	2646
Phe	Ser	Cys	Thr	Pro	Ser	Asn	Pro	Cys	Ser	His	Pro	Asp	Arg	Gly	Gly	
			860					865					870			
tgc	tca	gag	aat	gct	gag	tgt	gtc	cct	ggg	tcc	ctg	ggc	acc	cac	cac	2694
Cys	Ser	Glu	Asn	Ala	Glu	Cys	Val	Pro	Gly	Ser	Leu	Gly	Thr	His	His	
		875					880					885				
tgc	aca	tgc	cac	aaa	ggc	tgg	agt	ggg	gat	ggc	cgc	gtc	tgt	gtg	gct	2742
Cys	Thr	Cys	His	Lys	Gly	Trp	Ser	Gly	Asp	Gly	Arg	Val	Cys	Val	Ala	
	890					895					900					
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Ile	Asp	Glu	Cys	Glu	Leu	Asp	Val	Arg	Gly	Gly	Cys	His	Thr	Asp	Ala	
905				910						915					920	
ctc	tgc	agc	tat	gtg	ggc	ccc	ggg	cag	agc	cga	tgc	acc	tgc	aag	ctg	2838
Leu	Cys	Ser	Tyr	Val	Gly	Pro	Gly	Gln	Ser	Arg	Cys	Thr	Cys	Lys	Leu	
				925					930					935		
ggc	ttt	gcc	ggg	gat	ggc	tac	cag	tgc	agc	ccc	atc	gac	ccc	tgc	cgg	2886
Gly	Phe	Ala	Gly	Asp	Gly	Tyr	Gln	Cys	Ser	Pro	Ile	Asp	Pro	Cys	Arg	
			940					945					950			
gca	ggc	aat	ggc	ggc	tgc	cac	ggc	ctg	gcc	acc	tgc	cgg	gca	gtg	ggg	2934
Ala	Gly	Asn	Gly	Gly	Cys	His	Gly	Leu	Ala	Thr	Cys	Arg	Ala	Val	Gly	
		955					960					965				
gga	ggg	cag	cgg	gtc	tgc	acg	tgc	ccc	cct	ggc	ttt	ggg	ggg	gat	ggc	2982

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Gly 970	Gly	Gln	Arg	Val	Cys	Thr 975	Cys	Pro	Pro	Gly	Phe 980	Gly	Gly	Asp	Gly		
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Phe 985	Ser	Cys	Tyr	Gly	Asp 990	Ile	Phe	Arg	Glu	Leu 995	Glu	Ala	Asn	Ala	His 1000		
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Phe	Ser	Ile	Phe	Tyr 1005	Gln	Trp	Leu	Lys	Ser 1010	Ala	Gly	Ile	Thr	Leu 1015			
cct	gcc	gac	cgc	cga	gtc	aca	gcc	ctg	gtg	ccc	tcc	gag	gct	gca			3120
Pro	Ala	Asp	Arg	Arg 1020	Val	Thr	Ala	Leu	Val 1025	Pro	Ser	Glu	Ala	Ala 1030			
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Val	Arg	Gln	Leu	Ser 1035	Pro	Glu	Asp	Arg	Ala 1040	Phe	Trp	Leu	Gln	Pro 1045			
agg	acg	ctg	ccg	aac	ctg	gtc	agg	gcc	cat	ttt	ctc	cag	ggt	gcc			3210
Arg	Thr	Leu	Pro	Asn 1050	Leu	Val	Arg	Ala	His 1055	Phe	Leu	Gln	Gly	Ala 1060			
ctc	ttc	gag	gag	gag	ctg	gcc	cgg	ctg	ggt	ggg	cag	gaa	gtg	gcc			3255
Leu	Phe	Glu	Glu	Glu 1065	Leu	Ala	Arg	Leu	Gly 1070	Gly	Gln	Glu	Val	Ala 1075			
acc	ctg	aac	ccc	acc	aca	cgc	tgg	gag	att	cgc	aac	att	agt	ggg			3300
Thr	Leu	Asn	Pro	Thr 1080	Thr	Arg	Trp	Glu	Ile 1085	Arg	Asn	Ile	Ser	Gly 1090			
agg	gtc	tgg	gtg	cag	aat	gcc	agc	gtg	gat	gtg	gct	gac	ctc	ctt			3345
Arg	Val	Trp	Val	Gln 1095	Asn	Ala	Ser	Val	Asp 1100	Val	Ala	Asp	Leu	Leu 1105			
gcc	acc	aac	ggt	gtc	cta	cac	atc	ctc	agc	cag	gtc	tta	ctg	ccc			3390
Ala	Thr	Asn	Gly	Val 1110	Leu	His	Ile	Leu	Ser 1115	Gln	Val	Leu	Leu	Pro 1120			
ccc	cga	ggg	gat	gtg	ccc	ggt	ggg	cag	ggg	ttg	ctg	cag	cag	ctg			3435
Pro	Arg	Gly	Asp	Val 1125	Pro	Gly	Gly	Gln	Gly 1130	Leu	Leu	Gln	Gln	Leu 1135			
gac	ttg	gtg	cct	gcc	ttc	agc	ctc	ttc	cgg	gaa	ttg	ctg	cag	cac			3480
Asp	Leu	Val	Pro	Ala 1140	Phe	Ser	Leu	Phe	Arg 1145	Glu	Leu	Leu	Gln	His 1150			
cat	ggg	ttg	gtg	ccc	cag	att	gag	gct	gcc	act	gcc	tac	acc	atc			3525
His	Gly	Leu	Val	Pro 1155	Gln	Ile	Glu	Ala	Ala 1160	Thr	Ala	Tyr	Thr	Ile 1165			
ttt	gtg	ccc	acc	aac	cgc	tcc	ctg	gag	gcc	cag	ggc	aac	agc	agt			3570
Phe	Val	Pro	Thr	Asn 1170	Arg	Ser	Leu	Glu	Ala 1175	Gln	Gly	Asn	Ser	Ser 1180			
cac	ctg	gac	gca	gac	aca	gtg	cgg	cac	cat	gtg	gtc	ctg	ggg	gag			3615
His	Leu	Asp	Ala	Asp 1185	Thr	Val	Arg	His	His 1190	Val	Val	Leu	Gly	Glu 1195			
gcc	ctc	tcc	atg	gaa	acc	ctg	cgg	aag	ggt	gga	cac	cgc	aac	tcc			3660
Ala	Leu	Ser	Met	Glu 1200	Thr	Leu	Arg	Lys	Gly 1205	Gly	His	Arg	Asn	Ser 1210			
ctc	ctg	ggc	cct	gcc	cac	tgg	atc	gtc	ttc	tac	aac	cac	agt	ggc			3705
Leu	Leu	Gly	Pro	Ala 1215	His	Trp	Ile	Val	Phe 1220	Tyr	Asn	His	Ser	Gly 1225			
cag	cct	gag	gtg	aac	cat	gtg	cca	ctg	gaa	ggc	ccc	atg	ctg	gag			3750

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Gln	Pro	Glu	Val	Asn	His	Val	Pro	Leu	Glu	Gly	Pro	Met	Leu	Glu	
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gcc	cct	ggc	cgc	tcg	ctg	att	ggt	ctg	tcg	ggg	gtc	ctg	acg	gtg	3795
Ala	Pro	Gly	Arg	Ser	Leu	Ile	Gly	Leu	Ser	Gly	Val	Leu	Thr	Val	1255
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ggc	tca	agt	cgc	tgc	ctg	cat	agc	cac	gct	gag	gcc	ctg	cgg	gag	3840
Gly	Ser	Ser	Arg	Cys	Leu	His	Ser	His	Ala	Glu	Ala	Leu	Arg	Glu	1270
				1260					1265						
aaa	tgt	gta	aac	tgc	acc	agg	aga	ttc	cgc	tgc	act	cag	ggc	ttc	3885
Lys	Cys	Val	Asn	Cys	Thr	Arg	Arg	Phe	Arg	Cys	Thr	Gln	Gly	Phe	1285
				1275					1280						
cag	ctg	cag	gac	aca	ccc	agg	aag	agc	tgt	gtc	tac	cga	tct	ggc	3930
Gln	Leu	Gln	Asp	Thr	Pro	Arg	Lys	Ser	Cys	Val	Tyr	Arg	Ser	Gly	1300
				1290					1295						
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Phe	Ser	Phe	Ser	Arg	Gly	Cys	Ser	Tyr	Thr	Cys	Ala	Lys	Lys	Ile	1315
				1305					1310						
cag	gtg	ccg	gac	tgc	tgc	cct	ggt	ttc	ttt	ggc	acg	ctg	tgt	gag	4020
Gln	Val	Pro	Asp	Cys	Cys	Pro	Gly	Phe	Phe	Gly	Thr	Leu	Cys	Glu	1330
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cca	tgc	cca	ggg	ggt	cta	ggg	ggg	gtg	tgc	tca	ggc	cat	ggg	cag	4065
Pro	Cys	Pro	Gly	Gly	Leu	Gly	Gly	Val	Cys	Ser	Gly	His	Gly	Gln	1345
				1335					1340						
tgc	cag	gac	agg	ttc	ctg	ggc	agc	ggg	gag	tgc	cac	tgc	cac	gag	4110
Cys	Gln	Asp	Arg	Phe	Leu	Gly	Ser	Gly	Glu	Cys	His	Cys	His	Glu	1360
				1350					1355						
ggc	ttc	cat	gga	acg	gcc	tgt	gag	gtg	tgt	gag	ctg	ggc	cgc	tac	4155
Gly	Phe	His	Gly	Thr	Ala	Cys	Glu	Val	Cys	Glu	Leu	Gly	Arg	Tyr	1375
				1365					1370						
ggg	ccc	aac	tgc	acc	gga	gtg	tgt	gac	tgt	gcc	cat	ggg	ctg	tgc	4200
Gly	Pro	Asn	Cys	Thr	Gly	Val	Cys	Asp	Cys	Ala	His	Gly	Leu	Cys	1390
				1380					1385						
cag	gag	ggg	ctg	caa	ggg	gac	gga	agc	tgt	gtc	tgt	aac	gtg	ggc	4245
Gln	Glu	Gly	Leu	Gln	Gly	Asp	Gly	Ser	Cys	Val	Cys	Asn	Val	Gly	1405
				1395					1400						
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Trp	Gln	Gly	Leu	Arg	Cys	Asp	Gln	Lys	Ile	Thr	Ser	Pro	Gln	Cys	1420
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Pro	Arg	Lys	Cys	Asp	Pro	Asn	Ala	Asn	Cys	Val	Gln	Asp	Ser	Ala	1435
				1425					1430						
gga	gcc	tcc	acc	tgc	gcc	tgt	gct	gcg	gga	tac	tcc	ggc	aat	ggc	4380
Gly	Ala	Ser	Thr	Cys	Ala	Cys	Ala	Ala	Gly	Tyr	Ser	Gly	Asn	Gly	1450
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Ile	Phe	Cys	Ser	Glu	Val	Asp	Pro	Cys	Ala	His	Gly	His	Gly	Gly	1465
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tgc	tcc	cct	cat	gcc	aac	tgt	acc	aag	gtg	gca	cct	ggg	cag	cgg	4470
Cys	Ser	Pro	His	Ala	Asn	Cys	Thr	Lys	Val	Ala	Pro	Gly	Gln	Arg	1480
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Thr	Cys	Thr	Cys	Gln	Asp	Gly	Tyr	Met	Gly	Asp	Gly	Glu	Leu	Cys		
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Gln	Glu	Ile	Asn	Ser	Cys	Leu	Ile	His	His	Gly	Gly	Cys	His	Ile	1510	
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cac	gcc	gag	tgc	atc	ccc	act	ggc	ccc	cag	cag	gtc	tcc	tgc	agc	4605	
His	Ala	Glu	Cys	Ile	Pro	Thr	Gly	Pro	Gln	Gln	Val	Ser	Cys	Ser	1525	
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ctg	gac	ccc	tgc	tct	aag	aac	aat	gga	gga	tgc	agc	cca	tat	gcc	4695	
Leu	Asp	Pro	Cys	Ser	Lys	Asn	Asn	Gly	Gly	Cys	Ser	Pro	Tyr	Ala	1555	
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acc	tgc	aaa	agc	aca	ggg	gat	ggc	cag	agg	aca	tgt	acc	tgc	gac	4740	
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aca	gcc	cac	acc	gtg	ggg	gac	ggc	ctc	acc	tgc	cgt	gcc	cga	gtc	4785	
Thr	Ala	His	Thr	Val	Gly	Asp	Gly	Leu	Thr	Cys	Arg	Ala	Arg	Val	1585	
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Gly	Leu	Glu	Leu	Leu	Arg	Asp	Lys	His	Ala	Ser	Phe	Phe	Ser	Leu	1600	
				1590					1595					1600		
cgc	ctc	ctg	gaa	tat	aag	gag	ctc	aag	ggc	gat	ggg	cct	ttc	acc	4875	
Arg	Leu	Leu	Glu	Tyr	Lys	Glu	Leu	Lys	Gly	Asp	Gly	Pro	Phe	Thr	1615	
				1605					1610					1615		
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Ile	Phe	Val	Pro	His	Ala	Asp	Leu	Met	Ser	Asn	Leu	Ser	Gln	Asp	1630	
				1620					1625					1630		
gag	ctg	gcc	cgg	att	cgt	gcg	cat	cgc	cag	ctg	gtg	ttt	cgc	tac	4965	
Glu	Leu	Ala	Arg	Ile	Arg	Ala	His	Arg	Gln	Leu	Val	Phe	Arg	Tyr	1645	
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Gln	Gly	Tyr	Ala	Thr	Ala	Leu	Ser	Gly	His	Pro	Leu	Arg	Phe	Ser	1675	
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Glu	Arg	Glu	Gly	Ser	Ile	Tyr	Leu	Asn	Asp	Phe	Ala	Arg	Val	Val	1690	
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Ser	Ser	Asp	His	Glu	Ala	Val	Asn	Gly	Ile	Leu	His	Phe	Ile	Asp	1705	
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Arg	Val	Leu	Leu	Pro	Pro	Glu	Ala	Leu	His	Trp	Glu	Pro	Asp	Asp	1720	
				1710					1715					1720		
gct	ccc	atc	ccg	agg	aga	aat	gtc	acc	gcc	gcc	gcc	cag	ggc	ttc	5235	
Ala	Pro	Ile	Pro	Arg	Arg	Asn	Val	Thr	Ala	Ala	Ala	Gln	Gly	Phe	1735	
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Pro	Leu	Leu	Arg	Glu 1755	Ala	Ser	His	Arg	Pro 1760	Phe	Thr	Met	Leu	Trp 1765		
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Pro	Thr	Asp	Ala	Ala 1770	Phe	Arg	Ala	Leu	Pro 1775	Pro	Asp	Arg	Gln	Ala 1780		
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Trp	Leu	Tyr	His	Glu 1785	Asp	His	Arg	Asp	Lys 1790	Leu	Ala	Ala	Ile	Leu 1795		
cgg	ggc	cac	atg	att	cgc	aat	gtc	gag	gcc	ttg	gca	tct	gac	ctg	5460	
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ccc	aac	ctg	ggc	cca	ctt	cga	acc	atg	cat	ggg	acc	ccc	atc	tct	5505	
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Phe	Ser	Cys	Ser	Arg 1830	Thr	Arg	Pro	Gly	Glu 1835	Leu	Met	Val	Gly	Glu 1840		
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gct	cgc	tgt	gac	cac	ttt	gag	acc	cgg	ccc	ctg	cga	ctg	aac	acc	5685	
Ala	Arg	Cys	Asp	His 1875	Phe	Glu	Thr	Arg	Pro 1880	Leu	Arg	Leu	Asn	Thr 1885		
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Cys	Ser	Ile	Cys	Gly 1890	Leu	Glu	Pro	Pro	Cys 1895	Pro	Glu	Gly	Ser	Gln 1900		
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Glu	Gln	Gly	Ser	Pro 1905	Glu	Ala	Cys	Trp	Arg 1910	Phe	Tyr	Pro	Lys	Phe 1915		
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Val	His	Pro	Ser	Leu 1935	Trp	Gly	Arg	Pro	Gln 1940	Gly	Leu	Gly	Arg	Gly 1945		
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Cys	His	Arg	Asn	Cys 1950	Val	Thr	Thr	Thr	Trp 1955	Lys	Pro	Ser	Cys	Cys 1960		
cct	ggt	cac	tat	ggc	agt	gag	tgc	caa	gct	tgc	cct	ggc	ggc	ccc	5955	
Pro	Gly	His	Tyr	Gly 1965	Ser	Glu	Cys	Gln	Ala 1970	Cys	Pro	Gly	Gly	Pro 1975		
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Ser	Ser	Pro	Cys	Ser 1980	Asp	Arg	Gly	Val	Cys 1985	Met	Asp	Gly	Met	Ser 1990		
ggc	agt	ggg	cag	tgt	ctg	tgc	cgt	tca	ggt	ttt	gct	ggg	aca	gcc	6045	

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Gly	Ser	Gly	Gln	Cys	Leu	Cys	Arg	Ser	Gly	Phe	Ala	Gly	Thr	Ala	
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tgt	gaa	ctc	tgt	gct	cct	ggc	gcc	ttt	ggg	ccc	cat	tgt	caa	gcc	6090
Cys	Glu	Leu	Cys	Ala	Pro	Gly	Ala	Phe	Gly	Pro	His	Cys	Gln	Ala	2020
				2010					2015						
tgc	cgc	tgc	act	gtg	cat	ggc	cgc	tgt	gat	gag	ggc	ctt	ggg	ggc	6135
Cys	Arg	Cys	Thr	Val	His	Gly	Arg	Cys	Asp	Glu	Gly	Leu	Gly	Gly	2035
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Ser	Gly	Ser	Cys	Phe	Cys	Asp	Glu	Gly	Trp	Thr	Gly	Pro	Arg	Cys	2050
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gag	gtg	caa	ctg	gag	ctg	cag	cct	gtg	tgt	acc	cca	ccc	tgt	gca	6225
Glu	Val	Gln	Leu	Glu	Leu	Gln	Pro	Val	Cys	Thr	Pro	Pro	Cys	Ala	2065
				2055					2060						
ccc	gag	gct	gtg	tgc	cg	gca	ggc	aac	agc	tgt	gag	tgc	agc	ctg	6270
Pro	Glu	Ala	Val	Cys	Arg	Ala	Gly	Asn	Ser	Cys	Glu	Cys	Ser	Leu	2080
				2070					2075						
ggc	tat	gaa	ggg	gat	ggc	cg	gtg	tgt	aca	gtg	gca	gac	ctg	tgc	6315
Gly	Tyr	Glu	Gly	Asp	Gly	Arg	Val	Cys	Thr	Val	Ala	Asp	Leu	Cys	2095
				2085					2090						
cag	gac	ggg	cat	ggc	ggc	tgc	agt	gag	cac	gcc	aac	tgt	agc	cag	6360
Gln	Asp	Gly	His	Gly	Gly	Cys	Ser	Glu	His	Ala	Asn	Cys	Ser	Gln	2110
				2100					2105						
gta	gga	aca	atg	gtc	act	tgt	acc	tgc	ctg	ccc	gac	tac	gag	ggc	6405
Val	Gly	Thr	Met	Val	Thr	Cys	Thr	Cys	Leu	Pro	Asp	Tyr	Glu	Gly	2125
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gat	ggc	tgg	agc	tgc	cgg	gcc	cg	aac	ccc	tgc	aca	gat	ggc	cac	6450
Asp	Gly	Trp	Ser	Cys	Arg	Ala	Arg	Asn	Pro	Cys	Thr	Asp	Gly	His	2140
				2130					2135						
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Arg	Gly	Gly	Cys	Ser	Glu	His	Ala	Asn	Cys	Leu	Ser	Thr	Gly	Leu	2155
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Gly	Gln	Pro	Pro	Pro	Cys	His	Ser	Asp	Ala	Met	Cys	Thr	Asp	Leu	2200
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Glu	Ala	Gln	Gly	Ala	Val	Leu	Ala	Ser	Phe	Pro	Gln	Leu	Ser	Ala	2245
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Asn	Gly	Ser	Thr	Ala	His	Pro	Val	Val	Phe	Pro	Val	Ala	Asp	Cys		2275
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Gly	Asn	Gly	Arg	Val	Gly	Val	Val	Ser	Leu	Gly	Ala	Arg	Lys	Asn		2280
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Ala	Cys	Arg	Cys	Arg	Asn	Gly	Phe	Val	Gly	Asp	Gly	Ile	Ser	Thr		2310
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Cys	Asn	Gly	Lys	Leu	Leu	Asp	Val	Leu	Ala	Ala	Thr	Ala	Asn	Phe		2325
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Ala	Ser	Pro	Leu	Leu	Ala	Pro	Pro	Gln	Pro	Gln	Ala	Val	Leu	Ala		2460
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cct	gaa	ggc	cca	cct	gtg	gcg	gca	ggc	gtg	ggg	gct	gtg	ctt	gcc		7485
Pro	Glu	Ala	Pro	Pro	Val	Ala	Ala	Gly	Val	Gly	Ala	Val	Leu	Ala		2475
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Ala	Gly	Ala	Leu	Leu	Gly	Leu	Val	Ala	Gly	Ala	Leu	Tyr	Leu	Arg		2490
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Ala	Arg	Gly	Lys	Pro	Thr	Gly	Phe	Gly	Phe	Ser	Ala	Phe	Gln	Ala	
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Glu	Asp	Asp	Ala	Asp	Asp	Asp	Phe	Ser	Pro	Trp	Gln	Glu	Gly	Thr	
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aac	ccc	acc	ctg	gtc	tct	gtc	ccc	aac	cct	gtc	ttt	ggc	agc	gac	7665
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Thr	Phe	Cys	Glu	Pro	Phe	Asp	Asp	Ser	Leu	Leu	Glu	Glu	Asp	Phe	
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cct	gac	acc	cag	agg	atc	ctc	aca	gtc	aag	tgacgaggct	ggggctgaaa				7760
Pro	Asp	Thr	Gln	Arg	Ile	Leu	Thr	Val	Lys						
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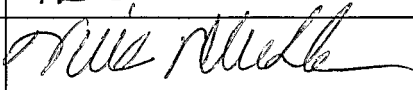
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0-1	Form - PCT/RO/134 (EASY) Indications Relating to Deposited Microorganism(s) or Other Biological Material (PCT Rule 13bis)	
0-1-1	Prepared using	PCT-EASY Version 2.92 (updated 01.10.2002)
0-2	International Application No.	PCT/FI 0 3 / 0 0 0 1 0
0-3	Applicant's or agent's file reference	2020166PC/or
1	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
1-1	page	9
1-2	line	23 - 28
1-3	Identification of Deposit	
1-3-1	Name of depositary institution	DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH
1-3-2	Address of depositary institution	Mascheroder Weg 1b, D-38124 Braunschweig, Germany
1-3-3	Date of deposit	21 August 2001 (21.08.2001)
1-3-4	Accession Number	DSMZ ACC2519
1-4	Additional Indications	According to Rule 13bis.6 PCT the Applicant wishes to make use of the expert provisions in those countries which provide for such
1-5	Designated States for Which Indications are Made	all designated States
1-6	Separate Furnishing of Indications These indications will be submitted to the International Bureau later	NONE
2	The indications made below relate to the deposited microorganism(s) or other biological material referred to in the description on:	
2-1	page	9
2-2	line	23 - 28
2-3	Identification of Deposit	
2-3-1	Name of depositary institution	DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH
2-3-2	Address of depositary institution	Mascheroder Weg 1b, D-38124 Braunschweig, Germany
2-3-3	Date of deposit	21 August 2001 (21.08.2001)
2-3-4	Accession Number	DSMZ ACC2520.>
2-4	Additional Indications	According to Rule 13bis.6 PCT the Applicant wishes to make use of the expert provisions in those countries which provide for such

Original (for **SUBMISSION**) - printed on 08.01.2003 01:45:16 PM

2-5	Designated States for Which Indications are Made	all designated States
2-6	Separate Furnishing of Indications These indications will be submitted to the International Bureau later	NONE

FOR RECEIVING OFFICE USE ONLY

0-4	This form was received with the international application: (yes or no)	YES
0-4-1	Authorized officer	

FOR INTERNATIONAL BUREAU USE ONLY


0-5	This form was received by the international Bureau on:	
0-5-1	Authorized officer	

BUDAPEST TREATY ON THE INTERNATIONAL
 RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
 FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

MediCity Research Laboratory
 University of Turku
 Tykistökatu 6 A
 FIN-20520 Turku

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
 issued pursuant to Rule 7.1 by the
 INTERNATIONAL DEPOSITARY AUTHORITY
 identified at the bottom of this page

I. IDENTIFICATION OF THE MICROORGANISM	
Identification reference given by the DEPOSITOR: 3-266	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: DSM ACC2519
II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION	
The microorganism identified under I. above was accompanied by: <input checked="" type="checkbox"/> a scientific description <input type="checkbox"/> a proposed taxonomic designation (Mark with a cross where applicable).	
III. RECEIPT AND ACCEPTANCE	
This International Depositary Authority accepts the microorganism identified under I. above, which was received by it on 2001-08-21 (Date of the original deposit) ¹ .	
IV. RECEIPT OF REQUEST FOR CONVERSION	
The microorganism identified under I above was received by this International Depositary Authority on (date of original deposit) and a request to convert the original deposit to a deposit under the Budapest Treaty was received by it on (date of receipt of request for conversion).	
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH Address: Mascheroder Weg 1b D-38124 Braunschweig	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):  Date: 2001-08-29

¹ Where Rule 6.4 (d) applies, such date is the date on which the status of international depositary authority was required.


BUDAPEST TREATY ON THE INTERNATIONAL
 RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
 FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

MediCity Research Laboratory
 University of Turku
 Tykistökatu 6 A
 FIN-20520 Turku

VIABILITY STATEMENT

issued pursuant to Rule 10.2 by the
 INTERNATIONAL DEPOSITARY AUTHORITY
 identified at the bottom of this page

I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
Name: MediCity Research Laboratory University of Turku Address: Tykistökatu 6 A FIN-20520 Turku	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: DSM ACC2519 Date of the deposit or the transfer ¹ : 2001-08-21
III. VIABILITY STATEMENT	
The viability of the microorganism identified under II above was tested on 2001-08-21 ² . On that date, the said microorganism was <input checked="" type="checkbox"/> viable <input type="checkbox"/> no longer viable	
IV. CONDITIONS UNDER WHICH THE VIABILITY TEST HAS BEEN PERFORMED ⁴	
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH Address: Mascheroder Weg 1b D-38124 Braunschweig	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):  Date: 2001-08-29

¹ Indicate the date of original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).

² In the cases referred to in Rule 10.2(a) (ii) and (iii), refer to the most recent viability test.

³ Mark with a cross the applicable box.


⁴ Fill in if the information has been requested and if the results of the test were negative.

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

MediCity Research Laboratory
University of Turku
Tykistökatu 6 A
FIN-20520 Turku

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

I. IDENTIFICATION OF THE MICROORGANISM	
Identification reference given by the DEPOSITOR: 3-372	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: DSM ACC2520
II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION	
The microorganism identified under I. above was accompanied by: <input checked="" type="checkbox"/> a scientific description <input type="checkbox"/> a proposed taxonomic designation (Mark with a cross where applicable).	
III. RECEIPT AND ACCEPTANCE	
This International Depositary Authority accepts the microorganism identified under I. above, which was received by it on 2001-08-21 (Date of the original deposit) ¹ .	
IV. RECEIPT OF REQUEST FOR CONVERSION	
The microorganism identified under I above was received by this International Depositary Authority on (date of original deposit) and a request to convert the original deposit to a deposit under the Budapest Treaty was received by it on (date of receipt of request for conversion).	
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH Address: Mascheroder Weg 1b D-38124 Braunschweig	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):  Date: 2001-08-29

¹ Where Rule 6.4 (d) applies, such date is the date on which the status of international depositary authority was acquired.


BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

MediCity Research Laboratory
University of Turku
Tykistökatu 6 A
FIN-20520 Turku

VIABILITY STATEMENT¹

issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

I. DEPOSITOR	II. IDENTIFICATION OF THE MICROORGANISM
Name: MediCity Research Laboratory University of Turku Address: Tykistökatu 6 A FIN-20520 Turku	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: DSM ACC2520 Date of the deposit or the transfer ¹ : 2001-08-21
III. VIABILITY STATEMENT	
The viability of the microorganism identified under II above was tested on 2001-08-21 ² . On that date, the said microorganism was <input checked="" type="checkbox"/> ³ viable <input type="checkbox"/> ³ no longer viable	
IV. CONDITIONS UNDER WHICH THE VIABILITY TEST HAS BEEN PERFORMED ⁴	
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH Address: Mascheroder Weg 1b D-38124 Braunschweig	Signature(s) of person(s) having the power to represent the International Depository Authority or of authorized official(s):  Date: 2001-08-29

¹ Indicate the date of original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).

² In the cases referred to in Rule 10.2(a) (ii) and (iii), refer to the most recent viability test.

³ Mark with a cross the applicable box.

⁴ Fill in if the information has been requested and if the results of the test were negative.

专利名称(译)	常见的淋巴管内皮和血管内皮受体-1 (聪明-1) 及其用途		
公开(公告)号	EP1463760A2	公开(公告)日	2004-10-06
申请号	EP2003729265	申请日	2003-01-08
[标]申请(专利权)人(译)	JALKANEN SIRPA IRJALA HEIKKI 萨尔米MARKO		
申请(专利权)人(译)	JALKANEN , SIRPA IRJALA , HEIKKI 萨尔米 , MARKO		
当前申请(专利权)人(译)	JALKANEN , SIRPA IRJALA , HEIKKI 萨尔米 , MARKO		
[标]发明人	JALKANEN SIRPA IRJALA HEIKKI SALMI MARKO		
发明人	JALKANEN, SIRPA IRJALA, HEIKKI SALMI, MARKO		
IPC分类号	G01N33/50 A61K39/395 A61K45/00 A61P29/00 A61P35/00 A61P35/04 A61P43/00 C07K14/705 C07K16/28 C12N15/02 C12P21/08 C12Q1/02 G01N33/15 G01N33/53 G01N33/574 A61K38/17		
CPC分类号	A61K2039/505 A61P29/00 A61P35/00 A61P35/04 A61P43/00 C07K14/7056 C07K16/28		
优先权	60/346288 2002-01-09 US		
其他公开文献	EP1463760B1		
外部链接	Espacenet		

摘要(译)

描述了一种新的蛋白质共同淋巴内皮和血管内皮细胞受体-1 (CLEVER-1)。CLEVER-1介导白细胞和恶性细胞与血管和淋巴内皮细胞的结合。CLEVER-1是第一种报道介导淋巴结流入和流出的蛋白质。还提供了通过提供CLEVER-1结合抑制剂来治疗炎症和预防恶性细胞转移的方法。