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(54) **COMMON LYMPHATIC ENDOTHELIAL AND VASCULAR ENDOTHELIAL RECEPTOR-1 (CLEVER-1) AND USES THEREOF**

GEMEINSAMER LYMPHATISCHER ENDOTHEL- UND GEFÄSSENDOTHEL-REZEPTOR-1 (CLEVER-1) UND SEINE VERWENDUNG

RECEPTEUR-1 ENDOTHELIAL LYMPHATIQUE ET ENDOTHELIAL VASCULAIRE COMMUN (CLEVER-1) ET SES UTILISATIONS

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Remarks:

The file contains technical information submitted after the application was filed and not included in this

specification

Description

BACKGROUND OF THE INVENTION

5 **Field of the Invention**

[0001] The invention is in the field of cell adhesion proteins. Specifically, the invention is in the field of CLEVER-1, a novel protein that facilitates the influx of leukocytes and malignant cells into the lymphatic system, and also the efflux of the same out of the lymph nodes.

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Background Art

[0002] Leukocytes are the major cellular components of inflammatory and immune responses. Leukocytes include lymphocytes, natural killer (NK) cells, monocytes, dendritic cells and granulocytes (neutrophils, eosinophils and basophils). See, HARRISON'S PRINCIPLES OF INTERNAL MEDICINE, Fauci, A.S. et al. eds. (14th ed. 1998). Lymphocytes are composed of B cells and T cells. B cells provide humoral immunity and are the precursors of plasma cells. T cells provide cell mediated immunity. In tissues monocytes differentiate further into macrophages. At sites of inflammation, blood monocytes can attach to inflamed endothelia. Macrophages recognize and take up a wide range of exogenous materials such as bacteria. Granulocytes also have critical roles in inflammation. They are needed to clear infections with extracellular bacteria. The immune response has an important role in the growth, differentiation, and mobilization of granulocytes.

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[0003] Continuous lymphocyte recirculation between blood and lymphoid tissues forms a basis for the function of the immune system. However such lymphocyte recirculation inadvertently also facilitates at least two medical conditions: inflammation and metastasis.

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[0004] Lymphocytes enter the lymphoid tissues by binding to vascular endothelial cells. Lymphocyte adherence to endothelial cells is mediated by complementary, surface expressed molecules on both cell types. The adhesion molecules and mechanisms of lymphocyte entrance into the tissues from the blood have been thoroughly characterized, but mechanisms controlling lymphocyte exit from the non-lymphoid and lymphoid tissues via lymphatics have remained unknown.

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[0005] The majority of lymphocytes extravasate into the lymph nodes via specialized vessels called high endothelial venules, or HEV. The rest of the incoming lymphocytes enter the nodes via afferent lymphatics together with antigens and other types of hematopoietic cells such as dendritic cells, macrophages and granulocytes. However, only lymphocytes are able to leave the nodes via the efferent lymphatic system by first traversing the sinusoidal endothelium and then entering the efferent lymphatic vessel. To maintain the homeostasis in the lymph node the numbers of entering and exiting lymphocytes need to be well balanced. The molecular mechanisms involved in lymphocyte exit are unknown.

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[0006] In addition to being of fundamental importance in normal lymphocyte recirculation, the lymphatics also regulate seeding of metastasizing cells in approximately 50% of cancers that use this type of vessel for spreading. Lymph nodes are often the first organ to develop metastases, especially in the case of carcinomas. The design of the lymphatic system makes it relatively easy for malignant tumor cells to enter (Sleeman, J.P., Recent Results Cancer Res. 157:55-81 (2000)), and thus compounds that prevent the entry and exit of malignant tumor cells from the lymphatics have tremendous therapeutic potential.

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BRIEF SUMMARY OF THE INVENTION

[0007] Recognizing the need to control lymphocyte recirculation, the inventors initiated a study of the proteins of the efferent lymphatic vessels. These studies have culminated in the discovery of a novel protein, Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 (CLEVER-1), a binding protein that mediates adhesion of lymphocytes (and malignant tumor cells) to endothelium in both the systemic vasculature and in the lymphatics. The inventors have discovered that by blocking the interaction of CLEVER-1 and its lymphocyte substrate, the artisan can, for the first time simultaneously, control lymphocyte recirculation and lymphocyte migration, and related conditions such as inflammation, at the site of lymphocyte influx into, and efflux from, the tissues. The inventors have also discovered that CLEVER-1 also mediates binding of other types of leukocytes such as monocytes and granulocytes to HEV-like vessels. Further, by blocking the interaction of CLEVER-1 and malignant tumor cells, the artisan can also, for the first time, control metastasis by preventing malignant cells that bind to CLEVER-1 from being taken up by the lymphatic vessels, and thus preventing spread of the malignancy into the lymph nodes.

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[0008] The publication Hideki Adachi et al., "FEEL-1, a novel scavenger receptor with in vitro bacteria-binding and angio-modulating activities", J Biol. Chem. vol. 277, no.37, September 2002, discloses a polypeptide having a sequence corresponding to CLEVER-1 except for a few mutations.

[0009] Database Genbank, 26 November 1999, Database accession no. HSA275213, discloses the amino acid and

nucleotide sequences of Stabilin-1. The amino acid sequence differs from that of CLEVER-1 with respect to two amino acids. No therapeutic function of the sequences is disclosed.

[0010] According to one aspect, this invention concerns a method of *in vitro* diagnosing inflammatory diseases in a patient, said method comprising:

5 (a) exposing a blood or tissue sample from said patient to a common lymphatic endothelial and vascular endothelial glycoprotein (CLEVER-1) *in vitro*, wherein said glycoprotein has a molecular weight of 270-300 kD in SDS-PAGE under non-reducing conditions, and is recognizable by a monoclonal antibody selected from the group consisting of

- 10 (i) monoclonal antibody 3-266 (DSM ACC 2519); and
(ii) monoclonal antibody 3-372 (DSM ACC 2590),

15 for a period of time and under conditions sufficient to allow binding of leukocytes if present in said sample; and
(b) detecting leukocytes bound to said glycoprotein in said blood or tissue sample.

[0011] According to another aspect, the invention concerns a method of *in vitro* detecting malignant cells in a patient, said method comprising:

- 20 (a) exposing a blood or tissue sample from a patient to the glycoprotein as defined above *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample; and
(b) detecting whether any malignant cells bound to said glycoprotein in said blood or tissue sample.

[0012] According to a third aspect, this invention concerns a method of removing malignant cells from a sample, said method comprising:

- 25 (a) exposing said malignant cells to the glycoprotein as defined above *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample, and
(b) separating said glycoprotein and the malignant cells that adhere thereto from said sample.

30 **[0013]** According to a fourth aspect, the invention concerns the use of an agent that inhibits glycoprotein mediated leukocyte binding, wherein said glycoprotein is defined as above, for the manufacture of a pharmaceutical composition useful in a method of treating inflammation in a patient in need of the same, wherein said inhibiting agent is selected from the group consisting of:

- 35 (a) antibodies or fragments thereof, raised to said glycoprotein; and
(b) said glycoprotein or fragments thereof in soluble form.

40 **[0014]** According to a fifth aspect, the invention concerns the use of an agent that inhibits malignant cell binding mediated by the glycoprotein as defined above, for the manufacture of a pharmaceutical composition useful in a method of preventing metastasis in a patient in need of the same, wherein said inhibiting agent is selected from the group consisting of:

- 45 (a) antibodies or fragments thereof, raised to said glycoprotein; and
(b) said glycoprotein or fragments thereof in soluble form.

[0015] According to a sixth aspect, the invention concerns a method of *in vitro* diagnosing inflammatory diseases in a patient, said method comprising exposing a blood or tissue sample from said patient to an antibody, which is either

- 50 - monoclonal antibody 3-266 (DSM ACC 2519); or
- monoclonal antibody 3-372 (DSM ACC 2590).

BRIEF DESCRIPTION OF THE DRAWINGS

55 **[0016]**

FIGS. 1a-1i. Indirect immunoperoxidase staining showing that monoclonal antibodies 3-266 and 3-372 recognize endothelium both in afferent and efferent lymphatic systems and on HEV. FIGS. 1a-1c are from the skin, FIGS. 1d-

1i are from a lymph node. FIGS. 1a, 1d and 1g show the staining with monoclonal antibody 3-266, FIGS. 1b, 1e and 1h show the staining with monoclonal antibody 3-372 and FIGS. 1c, 1f and 1i show the staining with a negative control antibody, 3G6. In FIGS. 1a, 1b and 1c, the arrows point to the epithelium and arrowheads to afferent lymphatics. In FIGS. 1d and 1e, the arrows point to the lymphatic vessels (lymphatic sinusoids that belong to the efferent lymphatic system) within the lymph node. In FIGS. 1g and 1h, the arrows point to HEV.

FIG. 2. Monoclonal antibodies 3-266 and 3-372 recognize an about 270-300 kDa molecule. Molecules in lymph node lysates were separated by SDS-PAGE, blotted to nitrocellulose sheets and probed with monoclonal antibody 3-266 and 3-372 or with a negative control antibody (3G6).

FIGS. 3A and 3B. CLEVER-1 is involved in lymphocyte binding to endothelial cells both in HEV and lymphatics. An adhesion assay was performed to measure lymphocyte binding to HEV (FIG. 3A) and to lymphatic endothelium (FIG. 3B). The sections were pre-incubated with monoclonal antibody 3-266 or 3-372, or negative control antibody anti-HLA ABC or 3G6 ("neg co") after which the sections were overlaid with normal lymphocytes. The results of three to four independent inhibition experiments are shown as mean percentage of maximal binding \pm SEM.

FIGS. 4A and 4B. CLEVER-1 is involved in binding of tumor cells to endothelial cells both in HEV and lymphatics. An adhesion assay was performed to measure binding of different tumor cell lines to HEV (FIG. 4A) and to lymphatic endothelium (FIG. 4B). The sections were pre-incubated with 3-372 or negative control antibody (anti-HLA ABC) after which the sections were overlaid with different tumor cells: Nu, NA, IBW4, KCA and CRL 1648. The results of three to four independent inhibition experiments are shown as mean percentage of maximal binding \pm SEM.

FIG. 5. CLEVER-1 is induced on HEV-like vessels at sites of inflammation in connection to infiltrations of inflammatory cells (FIGS. 5a-5c, synovium; FIGS. 5d-5f, skin). Fig. 5a. Fibrotic type of inflamed synovium without any marked infiltrations of inflammatory cells. Only the afferent lymphatics expressed CLEVER-1 (arrows). FIG. 5b. CLEVER-1 was upregulated on a HEV-like vessel (marked by a dashed line) within a heavy lymphocytic infiltration. FIG. 5c. Staining with a negative control antibody (3G6). FIG. 5d. In normal skin afferent lymphatics expressed CLEVER-1 (arrows), but in inflamed skin HEV-like vessels (dashed line) also expressed CLEVER-1 (FIG. 5e). Negative control staining (FIG. 5f). Arrowheads point to epidermis (FIGS. 5d-5f).

FIG. 6. CLEVER-1 mediates binding of monocytes and granulocytes to HEV-like vessels at sites of inflammation. Contribution of CLEVER-1 in binding of monocytes and granulocytes to inflamed synovial vessels and binding of granulocytes to tonsil was tested using Stamper-Woodruff type of binding assay. 3-372 and 3-266 (pooled) but not the class-matched control antibody (3G6) significantly inhibited binding of granulocytes and monocytes to HEV-like vessels in the organs tested. The results of four independent assays are shown as mean percentage of maximal binding (=100% in the presence of the control antibody) \pm SEM.

FIG. 7. Molecular characterization of CLEVER-1. (a) Antibodies 3-266 and 3-372 recognize a 270 - 300 kDa molecule in immunoblotting. 3G6 is a negative control antibody. (b) In gels run 48 hr for better resolution, at least three different isoforms of CLEVER-1 are seen, and enzymatic digestions with neuraminidase and O-glycans reveal the sialoglycoprotein nature of CLEVER-1. (c) Relative contribution of different isoforms of CLEVER-1 is different in tonsil, lymph nodes and synovium. (d) An alternative spliced form missing exon 27 is present in 1. lung, 2. brain, 3. placenta, 4. heart, 5. liver, 6. skeletal muscle, 7. kidney, 8. pancreas, 9. spleen, 10. thymus, 11. prostata, 12. testis, 13. ovary, 14. small intestine, 15. colon, 16. lymph nodes. Water control negative (lane 17). The upper band represents the standard form and the lower one is the splice variant of CLEVER-1.

Fig. 8. CLEVER-1 is expressed on the surface of endothelium *in vivo* and inhibition of its function blocks lymphocyte trafficking. Intravenously given 3-372 antibody (a) but not a negative class-matched control antibody (b) localized on the surface of HEV in lymph nodes after a 5 min circulation. HEV is pointed out by arrows in a. (c) Anti-CLEVER antibody treatment significantly inhibits the increase of the size of the lymph nodes draining the footpads. (One lymph node of a 3-372 treated rabbit was not found). (d) Lymphatic sinusoids of 3-372 treated animals contained less lymphocytes than those of control treated rabbits (e).

Fig. 9. The nucleotide sequence (7879 nt) of CLEVER-1. Boxed in grey are the translation initiation codon, translation stop codon, the two RGDs, the potential polyadenylation signal and the four nucleotide differences compared to Genbank entry AJ 2752213 (stabilin-1), i.e., nucleotides 1131, 2767, 6629 and 6969. Underlined are the nucleotides corresponding to the alternatively spliced exons.

DETAILED DESCRIPTION OF THE INVENTION

[0017] The term "ameliorate" denotes a lessening of an effect. To ameliorate a condition or disease refers to a lessening of the symptoms of the condition or disease.

[0018] The term "modulate" means to control in a predictable fashion, either by increasing or by decreasing the targeted parameter, as indicated from the context.

[0019] The term "effective amount" refers to that amount of the indicated agent that is sufficient to achieve the desired effect.

[0020] The term "inflammatory condition" refers to a physiological or pathological condition that is accompanied by an inflammatory response in a subject, which includes, *inter alia*, an undesired accumulation of leukocytes at one or more sites in such subject. The inflammatory condition can be hyperacute, acute, subacute or chronic. The inflammatory condition can be localized at the site of the inflammatory lesion or diffuse throughout the subject.

[0021] The term "drug" denotes any pharmaceutical or physiological agent, composition, bioactive compound, or combinations thereof, useful in the diagnosis, cure, mitigation, treatment, or prevention of a disease, or for any other medical purpose. The term "drug" is intended to be interpreted broadly and is not limited in terms of chemical composition or biological activity.

[0022] The term "essentially free of contaminants" refers to a substance that is of, undesired or unnecessary substances that had been present during the *in vitro* or *in vivo* synthesis of the desired substance.

[0023] The term "treatment" or "treating" refers to the administration of an agent to a subject for purposes which can include prophylaxis, amelioration, prevention or cure of an undesired disorder. Such treatment need not necessarily completely ameliorate the disorder, for example, inflammation; it is sufficient that such treatment ameliorates the disorder to a degree that is beneficial to the subject to which it is administered. Further, such treatment can be used in conjunction with other traditional treatments, for example, alternative treatments for reducing the inflammatory condition, known to those of skill in the art and as desired by the practitioner.

[0024] By "systemic vasculature" is meant the vascular network of blood vessels throughout the body of an animal or human.

[0025] By "lymphatic system" is meant the specialized part of the circulatory system that consists of lymph, the lymphatics, and the lymph nodes. The lymph nodes are located along the paths of the lymph collecting vessels and in isolated nodules of lymphatic patches in the intestinal wall. Additionally, there are specialized lymphatic organs such as the tonsils, thymus and spleen. B lymphocytes begin their final stages of maturation within the germinal centers of the lymph nodes' cortical nodules. Maturing lymphocytes are then pushed to the more densely packed outer layers as they mature, before being released into the efferent lymphatics. The lymph nodes that are located in the floor of the mouth are called the submental and submaxillary lymph nodes. The superficial cervical lymph nodes are located in the neck. The superficial cubital or supratroclear lymph nodes are located just above the bend in the elbow. The axillary lymph nodes are clustered deep within the underarm and upper chest region. Inguinal lymph nodes are located in the groin. By "lymphatics" is meant the vessels that return lymph to the blood. Lymph is the clear fluid that flow in the lymphatics. Lymph arises from plasma that filters into the interstitial spaces from blood flowing through the capillaries. Although most of this plasma is taken up and absorbed by cells or the blood, a small amount is not absorbed. The lymphatics act as drains to collect this excess fluid and return it to the venous blood just before it reaches the heart. The lymph nodes act as filters that collect the lymph from several different lymphatics and "percolate" the lymph through spaces termed sinuses before draining into a single efferent draining vessel.

[0026] By "afferent lymphatics" is meant the vessels through which antigens enter the lymph nodes. Lymphocytes can enter the lymph nodes via the afferent lymphatics or via the high endothelial venules (HEV).

[0027] By "high endothelial venules" (HEV) is meant a specialized cortical postcapillary venules whose endothelium is simple cuboidal to columnar instead of simple squamous. HEVs are located mainly in the paracortex of the lymph nodes. Lymphocytes cross the HEV, and thus "traffic" into the lymph nodes by diapedesis, that is, the lymphocytes stick to the luminal surface of the HEV, and then squeeze into the space between two or more HEV cells.

[0028] By "efferent lymphatics" meant the vessels that drain the lymph nodules (nodes).

[0029] By "lymphocyte recirculation" is meant the continuous movement of lymphocytes throughout the circulatory and lymph system. Lymphocytes leave the lymph node and are first delivered via the lymph to venous system draining into the heart.

[0030] The lymphocytes then circulate throughout the body in the bloodstream. Most of the lymphocytes are redelivered to the spleen or to another lymph node. About 10% go to non-lymphoid organs. Lymphocytes that have never been activated cannot enter non-lymphoid organs.

[0031] Lymphocyte "trafficking" refers to lymphocyte cell movement to specific locations. Outside of the lymph nodes, the trafficking of circulating lymphocytes allows the lymphocyte to accumulate at sites of inflammation. Activated effector lymphocytes tend to home to areas of inflammation, resulting in a large influx of lymphocytes in areas of inflammation. At the inflamed site, lymphocytes attach to the endothelial cells that line the blood vessels. This attachment localizes the lymphocyte at the site of inflammation and allows for subsequent emigration of the cells into the surrounding tissues (extravasation).

The Identification and Purification of CLEVER-1

[0032] The basis of the invention is the discovery of a new molecule, a novel protein herein designated "Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 in the systemic vasculature, and in the afferent and efferent lymphatics. It has been found that leukocytes such as lymphocytes, monocytes, and granulocytes, and malignant cells

specifically bind to this protein. It has also been found that this protein acts as a receptor that facilitates entry of bound leukocytes and malignant cells through the walls of the systemic vasculature, into the lymph nodes and out of the lymph nodes.

[0033] To search for a protein that played a role in lymphocyte lymphatic efflux, the inventors first identified cell migration-associated lymphatic structures from isolated efferent lymphatic vessels of human lymph nodes. These structures were used to produce monoclonal antibodies. Hybridomas were screened on frozen sections of human lymph nodes using immunoperoxidase staining.

[0034] Two of the hybridomas produced antibodies (designated 3-266 and 3-372) that clearly stained lymphatic endothelium both in afferent and efferent lymphatic systems and vascular endothelium on HEV, while other structures remained unstained. This is consistent with the expected pattern for an antibody that recognizes a lymphocyte migration-associated structure. Cell culture of 3-266 (DSM ACC2519) and cell culture of 3-372 (DSM ACC2590) were both deposited under the terms of the Budapest Treaty on the International Recognition of the Deposit of Micro-organisms for the Purposes of Patent Procedure on August 21, 2001, with DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroder Weg 1b, D-38124 Braunschweig.

[0035] In molecular weight determinations by immunoblotting, both antibodies recognized a molecule of the same size, about 270-300 kDa. Due to this and an identical staining pattern, these antibodies were considered to recognize the same antigen; that antigen was named Common Lymphatic Endothelial and Vascular Endothelial Receptor-1 (CLEVER-1).

[0036] CLEVER-1 was purified from CLEVER-1 containing lymph node preparations using affinity chromatography with the 3-372 antibody. The eluted material was subjected to SDS-PAGE analysis and silver staining. The specific band was excised, reduced, alkylated and digested with trypsin.

[0037] After cleavage with trypsin, mass spectrometric analyses yielded 27 peptides. Twenty-one (77%) of those had identical sequences with stabilin. These sequences covered altogether 268 amino acids (10% of the 2570 amino acids of stabilin-1) and spanned the amino acids between 53 and 2301 (Table 1). The peptide data suggest that CLEVER-1 has some homology with stabilin-1 at the structural level. No functional information regarding stabilin-1 can be found in the literature.

[0038] Peptide analysis of CLEVER-1 indicates no significant homology with any of the known endothelial homing-associated molecules, such as ICAM-1 (Intercellular Adhesion Molecule), ICAM-2, or VAP-1 (Vascular Adhesion Protein).

[0039] CLEVER-1 has several structural motifs that are associated with adhesive functions in other molecules. They include a proteoglycan link homology region important in CD44 for hyaluronan binding and two RGD motifs known to serve as integrin ligands in certain molecules, such as in fibronectin. In addition, CLEVER-1 has seven fascicling domains also present in several molecules such as priostin, fasciclin and transforming growth factor- β -induced gene, big-h3 and in all of these cases it is essential for adhesive function of these molecules. Interestingly, twenty-two epidermal growth factor (EGF) repeats are also found in CLEVER-1. This structural domain is also present in all members of the selectin family. Although the lectin domains of selectins are of utmost importance for the transient interaction with their sialomucin ligands, EGF-repeats have been reported to functionally contribute with lectin domains to binding between leukocytes and endothelium. Based on the structural complexity of CLEVER-1 it may turn out have several ligand molecules and be multifunctional in its nature.

[0040] It has been discovered that CLEVER-1 is involved in the process of lymphocyte recirculation. CLEVER-1 is present on the endothelium of the systemic vasculature, especially on the HEV and also on the endothelium of both afferent and efferent lymphatic systems. CLEVER-1 is a protein adhesion molecule, and especially, a cell adhesion molecule (CAM), that mediates adhesion of lymphocytes and of malignant tumor cells to CLEVER-1 in the systemic vasculature and the lymphatic system. These sites are of utmost importance as control points in lymphocyte trafficking.

[0041] CLEVER-1 is the first molecule that has been identified to facilitate lymphocyte and malignant cell exit from the lymph nodes. Additionally, CLEVER-1 is the first molecule that has been identified that regulates both entrance of lymphocytes and tumor cells into the lymph nodes and exit of lymphocytes and tumor cells from the lymph nodes. CLEVER-1 has been found to also mediate binding of other leukocytes such as monocytes and granulocytes to HEV-like Vessels.

[0042] By "CLEVER-1 mediated cell binding" is meant the specific association of CLEVER-1 with either a leukocyte, such as a lymphocyte, monocyte, or granulocyte, or a CLEVER-1-binding malignant cell. CLEVER-1 mediated cell binding can occur with CLEVER-1 in a soluble form or in a particulate form (for example, when CLEVER-1 is present in a form that is membrane associated).

CLEVER-1 Mediated Binding of Leukocytes

[0043] According to the invention, adhesion of leukocytes, such as lymphocytes, monocytes, and granulocytes, to the endothelium (that is, to an endothelial cell(s)) in the systemic vasculature, especially the HEV, and to the endothelium in the afferent and efferent lymphatics can be blocked by blocking the binding between such endothelial cell's CLEVER-

1 and leukocyte.

[0044] In the systemic vasculature, inhibiting or preventing endothelial cell CLEVER-1 mediated lymphocyte binding will inhibit or prevent lymphocytes, especially activated lymphocytes, from accumulating at such sites, and thus prevent or lessen inflammation at such sites. Thus, the invention provides a method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to lymphocytes.

[0045] In the afferent lymphatics, inhibiting or preventing lymphocytes from binding to endothelial cell CLEVER-1 will inhibit or prevent such lymphocytes from entering the afferent lymphatics and thus the lymph nodes. Thus, the invention provides a method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to lymphocytes in the afferent lymphatics and especially at HEV in lymph nodes or HEV-like venules at sites of inflammation. The invention also provides a method of inhibiting lymphocyte trafficking into the lymph nodes, by administering an agent that inhibits or prevents afferent lymphatic CLEVER-1 mediated endothelial cell binding, and especially HEV binding, to lymphocytes and other leukocytes.

[0046] In the efferent lymphatics, inhibiting or preventing lymphocytes from binding to endothelial cell CLEVER-1 will prevent the lymphocytes from exiting the lymph node and entering the blood. Thus, the invention provides a method of treating inflammation, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to lymphocytes in the efferent lymphatics. The invention also provides a method of inhibiting lymphocyte trafficking out of the lymph nodes, by administering an agent that inhibits or prevents efferent lymphatic CLEVER-1 mediated endothelial cell binding to lymphocytes.

[0047] Therefore, CLEVER-1 binding with lymphocytes presents a unique, three-prong approach to treat diseases or conditions characterized by an undesired lymphocyte accumulation or trafficking in which the artisan can target lymphocyte entry into the lymph nodes, lymphocyte exit from the lymph nodes, and lymphocyte binding to the systemic vasculature, with the same agent.

[0048] The discovery of CLEVER-1 and its role has thus resulted in a new method to control lymphocyte migration by inhibiting CLEVER-1 mediated cell binding to such cells. Thus, the present invention provides a method of inhibiting undesired CLEVER-1 mediated lymphocyte trafficking, and thus blocking harmful or otherwise undesired lymphocyte migration, by preventing the association of CLEVER-1 with lymphocytes. Similarly, the invention provides a method of inhibiting undesired CLEVER-1 mediated binding of other leukocytes by preventing association of CLEVER-1 with the leukocytes.

[0049] The present invention also provides a method of stimulating CLEVER-1 binding, for example, in immunocompromised hosts to facilitate lymphocyte trafficking and other leukocyte binding and the function of immune defense systems.

CLEVER-1 Mediated Cell Binding to Malignant Cells

[0050] Because cancer cells often break away from a malignant tumor and enter the lymphatics, cancer cells travel to and establish themselves in the lymph nodes. According to the invention, the ability of a malignant tumor cell to establish itself in a lymph node can be inhibited or prevented by inhibiting or preventing CLEVER-1 binding to such malignant tumor cell:

[0051] The term "tumor" refers to a neoplasm, a tissue mass that is characteristic of a neoplasia. Neoplasia is distinguished from other forms of tissue growth, first, by the formation of a tissue mass, a neoplasm, or tumor. Second, neoplasia is considered to be an irreversible process. Third, neoplastic tissue tends to morphologically resemble its tissue of origin. Fourth, neoplastic tissue tends to functionally resemble its tissue of origin. Fifth, neoplasms grow and function somewhat independently of the homeostatic mechanisms that control normal tissue growth and function.

[0052] A neoplasm can be benign or malignant. A benign neoplasm consists of a discrete tissue mass that continues to grow. A benign neoplasm will simply push adjacent tissues out of its way as it grows.

[0053] The definitive features of a malignant neoplasm, a malignancy, are invasion and metastasis, that is, the spread of the neoplasm to a distant site. A malignant neoplasm will grow into the adjacent tissue, rather than pushing it away. The terms "malignant neoplasm," "malignant tumor," and "cancer" are synonymous.

[0054] Cancer cells typically invade thin-walled vessels such as small veins, venules, capillaries and lymphatics. The passage of cancer cells via lymphatics to lymph nodes, and via blood vessels to other organs and structures, and the subsequent implantation and growth of the cancer cells in those sites is referred to as "metastasis." The lymph nodes are common sites for metastasis.

[0055] Cancer cells can also spread by seeding - shedding into, for example, the peritoneal fluid. The cells can be carried by the fluid to a distant site on the peritoneal surface where they can implant and form new foci of cancer growth.

[0056] Most neoplasms are one of four types: epithelial, non-epithelial, blastomas and teratomas. Malignant epithelial neoplasms are termed carcinomas. An adenocarcinoma is a carcinoma in which gland-like structures are present. Carcinomas can be papillary or cystic. Benign epithelial neoplasms are generally adenomas, polyps or papillomas.

[0057] Non-epithelial tumors can also be benign or malignant. They are generally named by a prefix that indicates the

histologic type and a suffix. The suffix -oma generally means benign while the suffix -sarcoma means malignant. However, several malignant neoplasms have traditional names ending in -oma: for example, melanoma, hepatoma, and lymphoma.

[0058] Lymphomas are malignant neoplasms arising from cells of the lymphoid series. Blastomas and teratomas contain more than one type of tissue. Malignant teratomas are often termed teratocarcinomas.

5 [0059] A "leukemia" is a tumor of white blood cells that is present in the bone marrow and blood. A "lymphoma" is a tumor of white blood cells that is present in the lymph nodes and tissues.

[0060] According to the invention, the binding of CLEVER-1-binding malignant cells to the endothelium in the systemic vasculature, especially the HEV, and to the endothelium in the afferent and efferent lymphatics can be inhibited or prevented by inhibiting or preventing the binding between such endothelial cell's CLEVER-1 and such malignant cell. Thus, the invention provides a method of treating cancer, and especially, a method of preventing metastasis, by administration of an agent that inhibits or prevents CLEVER-1 mediated malignant cell binding to the endothelium.

10 [0061] In the systemic vasculature, inhibiting or preventing CLEVER-1 mediated cell binding will inhibit or prevent the establishment of CLEVER-1 binding malignant cells at such sites, and thus lessen or prevent metastasis of such malignant cells. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding tumor cells in the systemic vasculature. The invention also provides a method of inhibiting metastasis, by administering an agent that inhibits or prevents systemic vasculature CLEVER-1 mediated endothelial cell binding to such malignant cells.

20 [0062] In the afferent lymphatics, inhibiting or preventing CLEVER-1 binding malignant cells from binding to endothelial cell CLEVER-1 will inhibit or prevent such CLEVER-1 binding malignant cell from entering and establishing in the lymph node, and thus lessen or prevent metastasis of such cell to the lymph node or thus to other sites in the body. In this context it is worth to note that a metastasizing malignant cell cannot survive long times without matrix support - a condition present for example in blood. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding malignant cells in the afferent lymphatics and at HEV in systemic vasculature. The invention also provides a method of inhibiting metastasis, by administering an agent that inhibits or prevents afferent lymphatic CLEVER-1 mediated endothelial cell binding, and especially HEV binding, to such malignant cells.

25 [0063] In the efferent lymphatics, inhibiting or preventing CLEVER-1 binding malignant cells from binding to endothelial cell CLEVER-1 will inhibit or prevent such CLEVER-1 binding malignant cell from leaving the lymph node, and thus lessen or prevent metastasis of such cell from the lymph node to other sites in the body. Thus, the invention provides a method of treating cancer, and especially, a method for preventing metastasis of a CLEVER-1 binding malignant cell, by administering an agent that inhibits or prevents CLEVER-1 mediated endothelial cell binding to CLEVER-1-binding malignant cells in the efferent lymphatics. The invention also provides a method of inhibiting metastasis, by administering an agent that inhibits or prevents efferent lymphatic CLEVER-1 mediated endothelial cell binding to such malignant cells.

30 [0064] CLEVER-1 interaction with CLEVER-1 binding malignant cells thus presents a unique, three-prong approach to inhibit or prevent metastasis in which not only can the artisan block such malignant cells from entering into and exiting from the lymph system, but also, the artisan can block association of such malignant cell with CLEVER-1 in the vascular endothelium.

40 **Agents that Block or Inhibit CLEVER-1 Mediated Cell Binding**

[0065] Soluble CLEVER-1 and antibodies to CLEVER-1 can be provided to the host cell to block or inhibit CLEVER-1 mediated cell binding. Soluble CLEVER-1 can be used to "coat" the CLEVER-1 binding sites on the leukocyte, such as lymphocyte, monocyte, or granulocyte, or tumor cell and thus prevent the coated cell from association with the native CLEVER-1 on the HEV or afferent or efferent lymphatics.

45 [0066] CLEVER-1 antibodies can be administered to a patient in need of the same to coat CLEVER-1 that is present on the vascular endothelium or lymphatics, of such patient, especially the afferent lymphatics so as to prevent leukocyte or malignant cell binding to such CLEVER-1 in the patient. CLEVER-1 antibody producing cells can be administered directly to the patient so as to provide a source of the same.

50 [0067] Moreover, the present invention provides a method of identifying an agent that inhibits the binding of CLEVER-1 to cells by providing an agent to cells in the presence of CLEVER-1 and comparing the binding of CLEVER-1 to cells provided with the agent to binding of CLEVER-1 in the absence of the agent. Similarly, the invention provides a method of identifying an agent that stimulates the binding of CLEVER-1 to cells by providing an agent to cells in the presence of CLEVER-1 and comparing the binding of CLEVER-1 to cells provided with the agent to binding of CLEVER-1 in the absence of said agent.

55 [0068] The term "antibody" is used in the broadest sense and specifically covers single monoclonal antibodies (including agonist and antagonist antibodies), polyclonal antibodies, as well as antibody fragments and single chain antibodies

(e.g., Fab, F(ab')₂, Fv), so long as they exhibit the desired biological activity.

[0069] Papain digestion of antibodies produces two identical antigen binding fragments, called Fab fragments, each with a single antigen binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an F(ab')₂ fragment that has two antigen combining sites and is still capable of cross-linking antigen.

[0070] Single chain "Fv" is the minimum antibody fragment which contains a complete antigen recognition and binding site. This region consists of a dimer of one heavy and one light chain variable domain in tight, noncovalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen binding site on the surface of the V_H-V_L dimer. Collectively, the six CDRs confer antigen binding specificity, to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site. See, Ladner et al., U.S. Patent No. 4,946,778, and Bird, R.E. et al., *Science*, 242:423-426 (1988).

[0071] The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by a hybridoma culture, uncontaminated by other immunoglobulins. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler and Milstein, *Nature* 256:495 (1975), or may be made by recombinant DNA methods (e.g., Cabilly et al., U.S. Patent No. 4,816,567).

[0072] Preparation of immunizing antigen, and polyclonal and monoclonal antibody production can be performed as described herein, or using other suitable techniques. A variety of methods have been described (*see e.g.*, Kohler et al., *Nature* 256:495-497 (1975), and *Eur. J. Immunol.* 6:511-519 (1976); Milstein et al., *Nature* 266:550-552 (1977); Koprowski et al., U.S. Patent No. 4,172,124; Harlow, E. and D. Lane, *Antibodies: A Laboratory Manual* (Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y., 1988); *CURRENT PROTOCOLS IN MOLECULAR BIOLOGY*, Vol. 2 (Supplement 27, 1994), Ausubel, F. M. et al., John Wiley & Sons, eds., New York, N.Y.), Chapter 11, (1991)). Generally, a hybridoma can be produced by fusing a suitable immortal cell line (e.g., a myeloma cell line such as SP2/0) with antibody producing cells. The antibody producing cell, preferably those of the spleen or lymph nodes, are obtained from animals immunized with the antigen of interest. The fused cells (hybridomas) can be isolated using selective culture conditions, and cloned by limiting dilution. Cells which produce antibodies with the desired binding properties can be selected by a suitable assay (e.g., ELISA).

[0073] The term "antibody" also includes chimeric, humanized or primatized (CDR-grafted) antibodies, as well as chimeric or CDR-grafted single chain antibodies, and the like, comprising portions derived from different species. "Chimeric" antibodies (immunoglobulins) have a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (Cabilly *et al.*, U.S. Patent No. 4,816,567; Morrison *et al.*, *Proc. Natl. Acad. Sci. USA* 81:6851-6855 (1984)). The various portions of these antibodies can be joined together chemically by conventional techniques, or can be prepared as a contiguous protein using genetic engineering techniques. For example, nucleic acids encoding a chimeric or humanized chain can be expressed to produce a contiguous protein. See, e.g., Cabilly et al., U.S. Patent No. 4,816,567; Cabilly et al., EP 0 125 023 B1; Boss et al., U.S. Patent No. 4,816,397; Boss et al., EP 0120694 B1; Neuberger, M.S. et al., WO 86/01533; Neuberger, M.S. et al., EP 0194276 B1; Winter, U.S. Patent No. 5,225,539; Winter, EP 0239400 B1; and Queen et al., U.S. Patent Nos. 5,585,089, 5,698,761 and 5,698,762. See also, Newman, R. et al., *BioTechnology* 10: 1455-1460 (1992), regarding primatized antibody.

[0074] By "agonist antibody" is meant an antibody which is able to bind to CLEVER-1 and facilitate adhesion of lymphocytes (and malignant tumor cells) to endothelium. By "antagonist antibody" is meant an antibody that is able to bind to CLEVER-1 and inhibit adhesion of lymphocytes (and malignant tumor cells) to endothelium.

[0075] Anti-idiotypic antibodies are also provided. Anti-idiotypic antibodies recognize antigenic determinants associated with the antigen-binding site of another antibody. Anti-idiotypic antibodies can be prepared against second antibody by immunizing an animal of the same species, and preferably of the same strain, as the animal used to produce the second antibody. See *e.g.*, U.S. Patent No. 4,699,880.

In Vitro Adhesion Assay and Diagnostic Uses Thereof

5 [0076] In a further embodiment, the present invention is directed to an adhesion assay in which CLEVER-1 binding is used to assay for the presence of leukocytes or malignant cells that bind to HEV and lymphatic endothelium. Both static and non-static assays are possible. The adhesion assay is exemplified in Example 4. Both static and non-static assays can be used to study leukocyte binding to systemic vasculature.

[0077] In the static assay, a tissue section is exposed to leukocytes or malignant cells for a desired period of time, without continuous agitation or rotation of the preparation during the exposure. Static assays are preferred for examining the ability of leukocytes and malignant cells to bind to lymphatic endothelium, especially the efferent lymph vessels.

10 [0078] In the non-static assay, the CLEVER-1 containing tissue sample and leukocytes are constantly rotated during the time period in which the leukocytes are given to adhere to the CLEVER-1. Non-static assays are preferred for studying leukocyte and malignant cell binding to CLEVER-1 in the HEV. The non-static assay mimics adhesion to the systemic vasculature.

15 [0079] In another embodiment, the present invention relates to a method for detection of malignant tumor cells. As explained with detail in Example 1, CLEVER-1 antibodies reduce the binding of malignant tumor cells to the vascular and lymphoid endothelium, demonstrating CLEVER-1 is a receptor for such malignant tumor cells. CLEVER-1 protein, or fragments thereof, or CLEVER-1 binding compounds, including but not restricted to, antibodies against CLEVER-1, both monoclonal and polyclonal, antibodies against antigenic fragments of CLEVER-1, both monoclonal and polyclonal, antigenic polypeptides, small molecule inhibitors or drugs, can be used in both quantitative and qualitative assays to detect the presence of malignant tumor cells in a sample, said sample being tissue or blood from a human or animal.

20 [0080] CLEVER-1 protein, or fragments thereof, or the above-mentioned CLEVER-1 binding compounds can be attached to a solid support matrix, including but not limited to microtiter plates, agarose columns, or magnetic beads. The above-mentioned sample can then be exposed to said solid support matrix, and the percentage of cells retained by said solid support matrix determined. For example, a sample from a normal, healthy individual, said individual being either a human or an animal, would have a statistically predicted number of leukocytes that bind to CLEVER-1. A sample that contains both leukocytes and malignant tumor cells would have a detectably higher number of cells that bind CLEVER-1.

25 [0081] In a preferred embodiment, a blood or tissue sample that has been taken from a patient who is in need of treatment, especially treatment for cancer, is used as the source of the CLEVER-1 binding cells in the *in vitro* adhesion assay. Such patient can be a patient being treated for a previously diagnosed malignancy, or a patient suspected of having a malignant tumor, or a patient who appears to be cured of such malignant tumor but is in need of monitoring for the reoccurrence of the same. Preferably, such blood or tissue sample is from a patient who is to be tested for the presence of malignant cells that bind to CLEVER-1 in such sample.

30 [0082] The blood or tissue sample that is to be examined in the *in vitro* assay of the invention can be processed, if desired, by methods known in the art so as to further extract or concentrate any CLEVER-1 binding cells that might be present in the sample, prior to the sample's being used in the *in vitro* adhesion assay of the invention.

[0083] Additionally, once the *in vitro* adhesion assay is complete, the adherent cells can be studied using other methods. For example, in the static assay, where the bound cells have been fixed, ability of the adherent cells to be recognized by a monoclonal antibody that is diagnostic for the type of tumor can be performed.

35 [0084] Detecting the binding of malignant cells to the CLEVER-1 containing lymphatic endothelium indicates that the patient is in need of treatment for such malignant cells, and especially to prevent the metastasis of such malignant cells

[0085] The present invention provides in this aspect a novel, efficient, and convenient assay for identifying antagonists, including but not limited to, monoclonal and polyclonal antibodies, peptides, protein fragments, small molecular inhibitors, drugs, and other agents, which can inhibit the adhesion of leukocytes and malignant tumor cells to the vascular and lymphatic endothelium.

40 [0086] For example, CLEVER-1 containing samples of lymph node sections can be incubated with and without the agent, and the number of bound lymphocytes and/or malignant cells determined. The antagonists can be pre-incubated with lymph node sections (a non-competitive assay) or simultaneously added with lymphocytes to the lymph node sections (a competitive assay).

45 [0087] Such a screen can also be used to customize an anti-metastasis treatment to an individual patient, and allows the practitioner to identify and select those agents or combinations thereof that have the best ability to inhibit CLEVER-1 malignant cell binding to vascular and/or lymphatic endothelium in such patient, and thus maximize the benefit of the treatment with such agents for such patient.

50 [0088] Additionally, such *in vitro* assay allows the practitioner to select for agents that provide a beneficial effect on disrupting malignant cell: CLEVER-1 containing endothelium interactions, nevertheless, minimize, if possible, the effect of such treatment on CLEVER-1 mediated leukocyte binding,

55 [0089] An antagonist can inhibit malignant cell or lymphocyte cell migration into or out of the lymph nodes. In a preferred embodiment, antagonists would inhibit both entrance and exit of an undesired cell into and out of the lymph nodes,

respectively. As such, malignant tumor cells would preferably be prevented from entering a lymph node, and establishing there, and any that did enter the lymph nodes via an afferent lymph vessel independent mechanisms would be contained, thus slowing metastasis.

[0090] This assay can be used further to monitor the efficacy of chemotherapy treatments administered to an individual, said individual being a human or an animal, in need thereof. Samples can be analyzed before, during, and after chemotherapy for the presence of malignant tumor cells that bind to CLEVER-1 or antigenic fragments thereof, or CLEVER-1 binding compounds.

[0091] In a further embodiment, purified CLEVER-1 protein, or fragments thereof can be used for high volume screening of antagonists that are capable of preventing or lowering the ability of a leukocyte or malignant cell to adhere to endothelial cell CLEVER-1. CLEVER-1 protein, or fragments thereof can be attached to a solid support matrix, including but not limited to a microtiter plate, an agarose column, or magnetic beads, using standard methods well known in the art. Antagonists can be screened for interaction with CLEVER-1 or fragments thereof, either in the absence or presence of leukocytes. Leukocytes or malignant cells can be labeled with fluorescent dyes such as, for example, bis-carboxyethyl carboxyfluorescein or fluorescein isothiocyanate and the number of bound cells in presence or absence of the antagonists can be analyzed by a fluoroimager.

[0092] The high volume screen assay of this aspect of the invention can be used to screen combinatorial libraries for molecules that inhibit the binding of leukocytes and malignant tumor cells to CLEVER-1 or a fragments thereof. Antagonists that show strong affinity for purified CLEVER-1 protein or fragments thereof can be screened further using the *in vitro* adhesion assay described above.

[0093] Antibodies used in the methods of the invention as CLEVER-1 binding compounds are preferably antibodies with a specificity against CLEVER-1, or an antigenic fragment thereof. Such antibodies can be polyclonal or monoclonal.

[0094] Another potential CLEVER-1 antagonist is a peptide derivative of the CLEVER-1 polypeptide that are naturally or synthetically modified analogs of the polypeptides that have lost biological function yet still recognize and bind to the ligand of the polypeptides to thereby effectively block the interaction of said ligand with CLEVER-1. Examples of peptide derivatives include, but are not limited to, small peptides or peptide-like molecules.

[0095] Another potential human CLEVER-1 antagonist is a peptide derivative of the ligand polypeptides which are naturally or synthetically modified analogs of the polypeptides that have lost biological function yet still recognize and bind to CLEVER-1 to thereby effectively block CLEVER-1. Examples of peptide derivatives include, but are not limited to, small peptides or peptide-like molecules.

[0096] The present invention relates to a diagnostic method for the detection of cells that contain CLEVER-1, that is, CLEVER-1 positive cells, in samples taken from the human or animal body. Such a method would involve the use of CLEVER-1 binding compounds, including but not limited to, monoclonal and polyclonal antibodies, with specificity for CLEVER-1. Such compounds can be labeled with a substance, such as a colorimetric dye or radioactive molecule, to permit rapid and easy detection of binding of the compound to cells that express CLEVER-1.

Therapeutic Uses of CLEVER-1 Antagonists

[0097] In another embodiment, the present invention relates to a method of treating malignant carcinomas. It is common for carcinomas to metastasize first to the regional lymph nodes (Sleeman, J.P., Recent Results Cancer Res. 157:55-81 (2000)). As described herein, CLEVER-1 is involved in the entrance and exit of malignant tumor cells to and from the lymph nodes. As such, antagonists that inhibit malignant tumor cell binding to CLEVER-1 are monoclonal or polyclonal antibodies and peptides, able to reduce metastasis and serve as effective chemotherapeutic agents.

[0098] In another aspect, the present invention relates to a method of treating disorders where the leukocyte-endothelial cell adhesion reaction is associated with acute or chronic inflammatory diseases such as skin inflammations, diabetes, connective tissue diseases (such as lupus, rheumatoid arthritis, osteoarthritis), obstructive and restrictive lung diseases (such as asthma, ARDS, sarcoidosis, idiopathic pulmonary fibrosis), inflammatory bowel diseases (such as ulcerative colitis and Crohn's disease), various nephritides, non-viral hepatitis, cirrhosis, cholangitis, atherosclerosis, vasculitis, thyroiditis, multiple sclerosis, myositis, ischemia reperfusion injury, transplantation rejection.

[0099] The antagonists can also be employed to treat histamine-mediated allergic reactions and immunological disorders including late phase allergic reactions, chronic urticaria, and atopic dermatitis. IgE-mediated allergic reactions such as allergic asthma, rhinitis, and eczema can also be treated.

[0100] The antagonists can also be employed to treat chronic and acute inflammation by preventing the extravasation of leukocytes to a wound area. They can also be employed to regulate normal pulmonary macrophage populations, since chronic and acute inflammatory pulmonary diseases are associated with sequestration of mononuclear phagocytes in the lung.

[0101] Antagonists can also be employed to treat rheumatoid arthritis by preventing the extravasation of leukocytes into synovial fluid in the joints of patients. Monocyte influx and activation plays a significant role in the pathogenesis of both degenerative and inflammatory arthropathies.

[0102] The antagonists can also be employed to treat asthma and allergy by preventing eosinophil accumulation in the lung. The antagonists can also be employed to treat subepithelial basement membrane fibrosis which is a prominent feature of the asthmatic lung.

[0103] The antagonists can also be employed for treating atherosclerosis, by preventing monocyte infiltration in the artery wall.

[0104] The antagonists can be employed in a composition with a pharmaceutically acceptable carrier, e.g., as hereinafter described.

Formulations of Compounds

[0105] The antagonists of CLEVER-1 can be used as therapeutic compositions. The antagonists of CLEVER-1 can be administered as a single dose or in multiple doses. The antagonists of the present invention can be administered either as an independent therapeutic regime or in combination with other therapeutic agents. The antagonists can be combined with conventional therapies, which can be administered simultaneously or sequentially.

[0106] Such therapeutic compositions can consist solely of the antagonist of CLEVER-1 although, preferably, the compositions will contain the antagonist of CLEVER-1 combined in admixture with a pharmaceutically acceptable carrier vehicle. Suitable vehicles and their formulation, inclusive of other human proteins, e.g., human serum albumin, are described for example in Remington: The Science and Practice of Pharmacy, Gennaro, Alfonso, 20th ed. (2000). In order to form a pharmaceutically acceptable composition that is suitable for effective administration to a patient in need of such composition, such compositions will contain an effective amount of the antagonist of CLEVER-1 together with a suitable amount of carrier vehicle.

[0107] Compositions containing antagonists of CLEVER-1 can be administered perorally, intravenously, intramuscularly, or sub-cutaneously at the appropriate dosages, which will depend upon the severity of the condition of the patient and upon such criteria as the patient's height, weight, sex, age, and medical history. The dose will also depend upon whether the compound of the invention is being administered to a human patient or in a veterinary setting to an animal, in need thereof.

[0108] For the purpose of parenteral administration, compositions containing the antagonists of CLEVER-1 are preferably dissolved in distilled water and the pH-value is preferably adjusted to about 6 to 8. In order to facilitate the lyophilization process resulting in a suitable product, lactose can be added to the solution. Preferably, the solution is then filtered sterilized, introduced into vials, and lyophilized. In a preferred embodiment, the compound of the invention is administered orally to a patient, at the time of eating or shortly thereafter. The concentration of the antagonists of CLEVER-1 in these composition, whether oral or parenteral, can vary, e.g., from 10^{-12} M to 10^{-3} M.

[0109] Additional pharmaceutical methods can be employed to control the duration of action. Controlled release preparations can be achieved by the use of polymers to complex or adsorb the antagonists of CLEVER-1. The controlled delivery can be exercised by selecting appropriate macromolecules (for example, polyesters, polyamino acids, polyvinyl pyrrolidone, ethylenevinylacetate, methylcellulose, carboxymethylcellulose, and protamine sulfate) and the concentration of macromolecules as well as the methods of incorporation in order to control release. Another possible method to control the duration of action by controlled release preparations is to incorporate the antagonists of CLEVER-1 into particles of a polymeric material such as polyesters, polyamino acids, hydrogels, poly (lactic acid) or ethylene vinylacetate copolymers. Alternatively, instead of incorporating the CLEVER-1 antagonists into these polymeric particles, it is possible to entrap these derivatives in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, bydroxymethylcellulose or gelatin-microcapsules and poly (methylmethacrylate) microcapsules, respectively, or in colloidal drug delivery systems, for example, liposomes, albumin microspheres, microemulsions, nanoparticles, and nanocapsules or in macroemulsions. Such teachings are disclosed in Remington: The Science and Practice of Pharmacy, Gennaro, Alfonso, 20th ed. (2000).

[0110] The following Example serves only to illustrate the invention, and is not to be construed as in any way limiting the invention.

EXAMPLE 1

Production of Monoclonal Antibodies

[0111] Balb/c mice were immunized to footpads four times at one week intervals, with incomplete Freund's adjuvant containing suspension made from lymphatic vessels excised from human lymph nodes under stereo microscope. The suspension was made by cutting the vessels to small pieces by scissors and the pieces in phosphate buffered saline were then drawn back and forth into a syringe connected to a 21 g needle. The popliteal lymph node lymphocytes from the immunized mice were isolated by a glass homogenizer. The popliteal lymph node lymphocytes of the immunized mice were fused with Sp2/0 myeloma cells. Hybridoma supernatants were primarily tested on frozen sections of human

lymph nodes using immunoperoxidase staining. The testing conditions were the same for antibodies 3-266 and 3-372 generated by two of the hybridomas.

[0112] Immunoperoxidase stainings were performed as described (Salmi, Science 257:1407-1409 (1992)). Briefly, acetone fixed 6 μ m frozen sections from different human tissues (lymph nodes, appendix, bronchus, cerebellum, epididymis, esophagus, heart, small and large intestine, kidney, liver, lung, normal and psoriatic skin, synovium, testis and tonsil) were stained with antibody 3-266, 3-372 or 3G6, a negative class matched control antibody for 3-266 and 3-372 (mouse IgG1) and 3,3-diaminobenzidine was used as a substrate. Procedures for tissue collection were approved by the Local and National Boards of Medicolegal Affairs in Finland.

[0113] Two of the hybridomas produced antibodies (3-266 (DSM ACC2519) and 3-372 (DSM ACC2590)) that clearly stained lymphatic endothelium both in afferent and efferent lymphatic systems and vascular endothelium on HEV, while the other structures remained unstained. The staining of the lymphatic endothelium is shown in Figure 1. Figure 1 is an indirect immunoperoxidase staining that shows that monoclonal antibodies 3-266 and 3-372 recognize endothelium both in afferent and efferent lymphatic systems and on HEV. Figures 1a-1c are from the skin. Figures 1d-1i are from a lymph node. Figures 1a, 1d and 1g show the staining with monoclonal antibody 3-266, Figures 1b, 1e and 1h show the staining with monoclonal antibody 3-372 and Figures 1c, 1f and 1i show the staining with a negative control antibody, 3G6. In Figures 1a, 1b and 1c, the arrows point to the epithelium and arrowheads to afferent lymphatics. In Figures 1d and 1e, the arrows point to the lymphatic vessels (lymphatic sinusoids that belong to the efferent lymphatic system) within the lymph node. In Figures 1g and 1h, the arrows point to HEV.

EXAMPLE 2

Determination of Molecular Weight of CLEVER-1

[0114] Molecular weight determination was performed by immunoblotting. One percent NP-40 lysates containing of human lymph nodes was analyzed using 5-12.5% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE). SDS-PAGE was run in non-reducing conditions. The molecules in the gel were blotted overnight to nitrocellulose sheets and probed with 3-266, 3-372 or a negative control antibody (3G6) (Salmi, M. et al., J. Exp. Med. 183:569-579 (1996)). Peroxidase conjugated rabbit anti-mouse Ig was used as the second stage reagent. Detection was performed using enhanced chemiluminescence system according to the instructions of the manufacturer (Amersham).

[0115] Both antibodies recognized a molecule of the same size (about 270-300 kDa; Figure 2). Due to this and an identical staining pattern, these antibodies were assumed to recognize the same antigen and this antigen was named CLEVER-1.

EXAMPLE 3

Purification and Molecular Characterization of CLEVER-1

[0116] The molecule recognized by 3-372 antibody was purified from human lymph node lysate overnight (lysis buffer: 150 mM NaCl, 10 mM Tris-base, pH 7.2, 1.5 mM MgCl₂, 1% NP-40, 1% aprotinin, and 1 mM PMSF) as described in Smith, D.J. et al., J. Exp. Med. 188:17-27 (1998). After centrifugation, the lysate supernatant was applied sequentially to immunoaffinity columns containing CnBr-activated Sepharose beads armed with irrelevant mAbs and 3-372 (3 mg/ml beads). After washing with lysis buffer, the antigens recognized by 3-372 were eluted with 50 mM triethylamine, frozen and subsequently lyophilized. The eluted material was then subjected to SDS-PAGE analysis and silver staining (O'Connell, K.L. and Stults, J.T., Electrophoresis 18:349-359 (1997)). The specific band was excised, reduced, alkylated and digested with trypsin (Promega) overnight at +37°C as described (Shevchenko, A. et al., Anal. Chem. 68:850-855 (1996); O'Connell, K.L. and Stults, J.T., Electrophoresis 18:349-359 (1997)). The peptides were analyzed using PerSeptive BioSystems Voyager DE-PRO mass spectrometer operated in the reflectron delayed-extraction mode. Calibration of the spectrum was performed internally by using autolysis products of trypsin or with added calibration mixture 2 (PerSeptive BioSystems). Database search was performed by MS-Fit algorithm (<http://prospector.ucsf.edu/ucsfhtml3.2/ms-fit.htm>) of the University of California, San Francisco mass spectrometry facility.

[0117] After cleavage with trypsin, mass spectrometric analyses yielded 27 peptides. 21 (77%) of those had identical nucleotide sequences with two Genebank entries: AJ 275213, a submission for a cDNA clone called stabilin-1, and D87433, a cDNA clone KIAA0246 isolated from the cell line KG-1. The peptide sequences covered altogether 268 amino acids (10% of the 2570 amino acids of stabilin-1) and spanned the amino acids between 53 and 2301.

[0118] Next we designed primers based on the peptide sequences, the 5' end of the cDNA for stabilin-1 and the 3' end of the cDNA of KIAA0246 and used them to make several RT-PCR fragments that were then ligated together to clone the full-length 7879 bp cDNA (SEQ ID NO:1). Sequencing of the whole construct revealed a high homology with the existing 3' end KIAA0246 sequence available in the data bank. However, it contained 4 nucleotide differences, when

compared to the Stabilin sequence. They all cause a change at the amino acid level. Two of these changes are identical with the genomic sequence data available from the HUGO project (AC 006208), but since the genomic clone only covers about half of the gene for this cDNA, the nature of the two other changes remains to be determined.

5 [0119] Sequencing of several different CLEVER-1 cDNA-clones also revealed the existence of at least two alternatively spliced isoforms of the molecule: the regions covered by exons 23 (nucleotides 2377-2562) and 27 (nucleotides 2914-3009) can be spliced out. We could confirm the existence of one of those splice variants (lacking exon 27) also at the mRNA level (Figure 7) but the second one (lacking exon 23) that we cloned from a human peripheral lymph node library was not visible in the system suggesting a low abundance/ turnover of the mRNA encoding this form.

10 [0120] The sequence comparisons revealed significant homologies to proteoglycan link protein-like sequence, epidermal growth factor-like repeats and two RGD motifs being well in line with the adhesive properties of CLEVER-1.

EXAMPLE 4

In Vitro Adhesion Assay

15 [0121] Lymph node sections were first incubated with 3-266, 3-372 or control antibodies against human HLA ABC (HB-95, ATCC and 3G6 (against chicken T cells) and then overlaid with Ficoll gradient (Pharmacia) purified peripheral blood mononuclear cells or different human tumor cell lines (lymphoblastoid cell lines, KCA and IBW4; a Burkitt lymphoma CRL-1648; squamocellular carcinoma lines NA and NU). Thereafter, the sections were subjected to two different types
20 of assays: 1. A non-static assay, which optimally measures binding of cells to HEV and is performed under rotatory conditions (60 rpm on orbital shaker for 30 min at +7°C). 2. A static assay, in which the sections overlaid with cells are let to stay in static conditions for 15 min, followed by 5 min of rotation at 60 rpm and then again 15 min without rotation at 7°C. (Static conditions were needed to optimal binding to lymphatic endothelium). The adherent cells were fixed in 1% glutaraldehyde. The number of lymphocytes bound to HEV and to sinusoidal (lymphatic) endothelium was counted
25 single blind under dark-field illumination in which setting the sinusoidal vessels are easily recognizable. The results of the inhibition assays are presented as percentage of control binding (the number of adherent cells/vessel in the presence of control mAb defines 100% adherence).

30 [0122] When the assay was performed in non-static conditions, mimicking the blood flow, lymphocyte binding to HEV was reduced 43.6% and 45.2% by 3-266 and 3-372, respectively (Figure 3A). To mimic the conditions at sites of lymphocyte exit, the assay was performed in static conditions. In these assays lymphocyte binding to lymphatic endothelium was decreased 46.4% and 64% by 3-266 and 3-372, respectively (Figure 3B). These data indicate that the molecule recognized by 3-266 and 3-372 indeed mediate lymphocyte binding both to HEV and to lymphatic endothelium.

35 [0123] To study the role of this molecule in migration of malignant cells, the assays were performed using three lymphoma cell lines (CRL 1648, KCA and IBW4) and two squamocellular carcinoma cell lines (NA and NU). For these assays 3-372 antibody was chosen because of its higher inhibitory capacity (Figure 3). The results of these experiments clearly demonstrated that CLEVER-1 is also involved in binding of malignant cells to endothelium both at entrance and exit sites within the lymph nodes (Figures 4A and 3).

EXAMPLE 5

CLEVER-1 is Upregulated at Sites of Inflammation on HEV-like Vessels

40 [0124] Synovial samples from 18 patients suffering from chronic arthritis and undergoing synovectomies, skin samples from diseased skin of patients suffering from psoriasis (n=5), lichen (n=1), mycosis fungoides (n=1), erythrodermia (n=2),
45 exanthema (n=1), folliculitis (n=3) and normal skin samples from 15 individuals were studied for expression of CLEVER-1 using immunoperoxidase method as described above. Like in normal non-lymphoid tissues CLEVER-1 was present in afferent lymphatic vessels in inflamed synovial and both in normal and diseased skin samples. In addition, CLEVER-1 expression was induced on HEV-like vessels that appear at sites of inflammation and are surrounded by heavy infiltrations of inflammatory cells (Figure 5). Table 2 illustrates complete correlation between the extent of inflammatory
50 infiltration and upregulation of CLEVER-1 expression in synovial samples. The same phenomenon was observed in skin samples: all diseased skin samples had inflammatory infiltrations that contained CLEVER-1 positive HEV-like vessels. Those vessels were absent in normal skin samples.

EXAMPLE 6

Cleaver-1 Also Mediates Binding of Monocytes and Granulocytes to HEV-like Vessels

55 [0125] Human monocytes from peripheral blood were purified from Ficoll -gradient (Pharmacia) isolated mononuclear

cells by letting them to adhere to plastic surfaces for an hour at +37°C. Granulocytes were purified from leukocyte rich buffy coats from human blood using Percoll-gradient (Pharmacia) centrifugation. Their binding was tested to HEV-like vessels in inflamed synovium. In addition, granulocyte binding was tested to tonsil HEV that brightly express CLEVER-1. (When tonsils are removed they always have variable extent of inflammation, although they as a lymphoid organ have HEV without any inflammation). Both granulocytes and monocytes bound efficiently to HEV-like vessels in inflamed synovium and granulocytes adhered avidly to HEV in tonsils. Their binding to these organs was significantly inhibited by the antibody pool containing 3-372 and 3-266 but not with the control antibody (Figure 6).

EXAMPLE 7

CLEVER-1 controls lymphocyte trafficking *in vivo*

[0126] In order to verify that CLEVER-1 has a functional role *in vivo*, it was at first confirmed by intravenous injection of 3-372 antibody that rabbits express CLEVER-1 on the surface of endothelium *in vivo*. The presence of CLEVER-1 on HEV was detected after the 3-372 antibody had circulated 5 minutes *in vivo* using frozen sections and FITC labelled anti-mouse IgG second stage antibody after sacrifice (Fig. 8a). In this time frame the intravenously given 180 kDa immunoglobulin molecule does not have a possibility to leak and diffuse into the tissue. Based on these results antibody 3-372 (and a class-matched negative control antibody) was given to the rabbits immunized with keyhole limpet hemocyanin to footpads and the effects of the antibody treatment on the size and cellularity of the lymph nodes draining the footpads was analyzed.

[0127] Antibody treatment against CLEVER-1 significantly prevented increase of the size of the popliteal lymph nodes (Fig. 8c) indicating that CLEVER-1 has a functional role in lymphocyte traffic *in vivo*. Most likely it exerts its effects both at lymphocyte entrance in HEV and their exit in lymphatic sinusoids, because the rabbits treated with 3-372 antibody had only few lymphocytes in their lymphatic sinusoids when analyzed using histological section (Fig. 8d) Intravenously given 3-372 antibody was also detected to bind CLEVER-1 on lymphatic sinuses when tested at sacrifice 3 days after the final 3-372 dose. No signal was detected in rabbits which received a control antibody (data not shown).

[0128] All documents, e.g., scientific publications, patents and patent publications recited herein are hereby incorporated by reference in their entirety to the same extent as if each individual document was specifically and individually indicated to be incorporated by reference in its entirety. Where the document cited only provides the first page of the document, the entire document is intended, including the remaining pages of the document.

Table 1. Matches with CLEVER-1 and stabilin-1

1. 21/27 matches (77%). 275350.0 Da, pI = 6.04. Acc. #6469374. HOMO SAPIENS. (AJ275213) stabilin-1.

m/z	MH ⁺	Delta	start	end	Peptide Sequence	Modifications
submitted	matched	ppm			(Click for Fragment Ions)	
775.488	775.483	6.4034	372	377	(R)VFLQLR(V)	
787.36	787.3739	-17.6253	1299	1305	(R)SGFSFSR(G)	
799.495	799.5042	-11.4617	1585	1591	(R)VGLELLR(D)	
812.495	812.4994	-5.4309	1047	1053	(R)TLPNLVR(A)	
917.502	917.4997	2.4556	1040	1046	(R)AFWLQPR(T)	
1017.44	1017.425	14.4862	2295	2301	(R)WDAYCFR(V)	
1104.54	1104.526	12.6406	53	61	(K)QTCPGWLR(E)	
1212.7	1212.695	3.9482	1021	1032	(R)VTALVPSEAAVR(Q)	
1284.65	1284.622	21.4530	1678	1688	(R)EGSIYLNDFAR(V)	
1291.79	1291.774	12.5434	613	624	(R)ILLGPEGVPLQR(V)	
1330.63	1330.575	41.7401	953	965	(R)AGNGGCHGLATCR(A)	
1330.63	1330.633	-1.9754	1873	1882	(R)CDHFETRPLR(L)	
1374.66	1374.632	20.1117	62	72	(R)ELPDQITQDCR(Y)	
1456.79	1456.776	9.6229	1069	1082	(R)LGGQEVATLNPTTR(W)	
1493.8	1493.796	2.4215	508	521	(R)TIGQILASTEAFSR(F)	
1555.7	1555.663	23.5678	219	231	(R)CLPGYTQQGSECR(A)	
1678.94	1678.913	16.1953	1802	1817	(R)NVEALASDLPNLGPLR(T)	
1730.89	1730.887	1.9674	1054	1068	(R)AHFLQGALFEEELAR(L)	
1912.82	1912.832	-6.3742	936	952	(K)LGFAGDGYQCSPIDPCR(A)	
2057.05	2057.03	9.5484	1655	1673	(R)SEDLLEQGYATALSGHPLR(F)	

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(continued)

1. 21/27 matches (77%). 275350.0 Da, pI = 6.04. Acc. #6469374. HOMO SAPIENS. (AJ275213) stabilin-1.

m/z	MH ⁺	Delta	start	end	Peptide Sequence	Modifications
submitted	matched	ppm			(Click for Fragment Ions)	
2165.11	2165.14	-13.6320	1707	1725	(R)VLLPPEALHWEPDDAPIPR(R)	
2295.22	2295.224	-1.5853	389	410	(R)EILTTAGPFTVLVPSVSSFSSR(T)	

6 unmatched masses: 871.5410 949.4800 1360.6500 1538.6900 1787.9200 2008.1400

The matched peptides cover 10% (268/2570 AA's) of the protein.

Coverage Map for This Hit (MS-Digest index #): [427477](#)

Table 2. CLEVER-1 Expression Is Induced Mainly on Vessels Surrounded by Lymphocytic Infiltrations in Inflamed Synovia

Expression of CLEVER-1 on HEV-like vessels ¹	The degree of inflammatory infiltration ²	
	0/1 (n=9)	2/3 (n=9)
-/+	100%	0
++/+++	0	100%

¹ Intensity was scored as -, ±, + negative or weak, ++, +++ moderate or strong.

² Degree of the inflammatory cell infiltration in 18 synovial samples was scored as: 0/1, none or few lymphocytes around the vessels, 2/3 marked or massive lymphocytic infiltrations.

SEQUENCE LISTING

[0129]

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15	gtc Val	cgt Arg	cag Gln	ctg Leu	agc Ser 1035	ccc Pro	gag Glu	gac Asp	cga Arg	gct Ala 1040	ttc Phe	tgg Trp	ctg Leu	cag Gln	cca Pro 1045		3165
	agg Arg	acg Thr	ctg Leu	ccg Pro	aac Asn 1050	ctg Leu	gtc Val	agg Arg	gcc Ala	cat His 1055	ttt Phe	ctc Leu	cag Gln	ggt Gly	gcc Ala 1060		3210
20	ctc Leu	ttc Phe	gag Glu	gag Glu	gag Glu 1065	ctg Leu	gcc Ala	cgg Arg	ctg Leu	ggt Gly 1070	ggg Gly	cag Gln	gaa Glu	gtg Val	gcc Ala 1075		3255
25	acc Thr	ctg Leu	aac Asn	ccc Pro	acc Thr 1080	aca Thr	cgc Arg	tgg Trp	gag Glu	att Ile 1085	cgc Arg	aac Asn	att Ile	agt Ser	ggg Gly 1090		3300
	agg Arg	gtc Val	tgg Trp	gtg Val	cag Gln 1095	aat Asn	gcc Ala	agc Ser	gtg Val	gat Asp 1100	gtg Val	gct Ala	gac Asp	ctc Leu	ctt Leu 1105		3345
30	gcc Ala	acc Thr	aac Asn	ggt Gly	gtc Val 1110	cta Leu	cac His	atc Ile	ctc Leu	agc Ser 1115	cag Gln	gtc Val	tta Leu	ctg Leu	ccc Pro 1120		3390
35	ccc Pro	cga Arg	ggg Gly	gat Asp	gtg Val 1125	ccc Pro	ggt Gly	ggg Gly	cag Gln	ggg Gly 1130	ttg Leu	ctg Leu	cag Gln	cag Gln	ctg Leu 1135		3435
	gac Asp	ttg Leu	gtg Val	cct Pro	gcc Ala 1140	ttc Phe	agc Ser	ctc Leu	ttc Phe	cgg Arg 1145	gaa Glu	ttg Leu	ctg Leu	cag Gln	cac His 1150		3480
40	cat His	ggg Gly	ttg Leu	gtg Val	ccc Pro 1155	cag Gln	att Ile	gag Glu	gct Ala	gcc Ala 1160	act Thr	gcc Ala	tac Tyr	acc Thr	atc Ile 1165		3525
45	ttt Phe	gtg Val	ccc Pro	acc Thr	aac Asn 1170	cgc Arg	tcc Ser	ctg Leu	gag Glu	gcc Ala 1175	cag Gln	ggc Gly	aac Asn	agc Ser	agt Ser 1180		3570
	cac His	ctg Leu	gac Asp	gca Ala	gac Asp 1185	aca Thr	gtg Val	cgg Arg	cac His	cat His 1190	gtg Val	gtc Val	ctg Leu	ggg Gly	gag Glu 1195		3615
50	gcc Ala	ctc Leu	tcc Ser	atg Met	gaa Glu 1200	acc Thr	ctg Leu	cgg Arg	aag Lys	ggt Gly 1205	gga Gly	cac His	cgc Arg	aac Asn	tcc Ser 1210		3660
	ctc Leu	ctg Leu	ggc Gly	cct Pro	gcc Ala 1215	cac His	tgg Trp	atc Ile	gtc Val	ttc Phe 1220	tac Tyr	aac Asn	cac His	agt Ser	ggc Gly 1225		3705
55	cag	cct	gag	gtg	aac	cat	gtg	cca	ctg	gaa	ggc	ccc	atg	ctg	gag		3750

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	Gln	Pro	Glu	Val	Asn 1230	His	Val	Pro	Leu	Glu 1235	Gly	Pro	Met	Leu	Glu 1240	
5	gcc Ala	cct Pro	ggc Gly	cgc Arg	tcg Ser 1245	ctg Leu	att Ile	ggc Gly	ctg Leu	tcg Ser 1250	ggg Gly	gtc Val	ctg Leu	acg Thr	gtg Val 1255	3795
10	ggc Gly	tca Ser	agt Ser	cgc Arg	tgc Cys 1260	ctg Leu	cat His	agc Ser	cac His	gct Ala 1265	gag Glu	gcc Ala	ctg Leu	cgg Arg	gag Glu 1270	3840
	aaa Lys	tgt Cys	gta Val	aac Asn	tgc Cys 1275	acc Thr	agg Arg	aga Arg	ttc Phe	cgc Arg 1280	tgc Cys	act Thr	cag Gln	ggc Gly	ttc Phe 1285	3885
15	cag Gln	ctg Leu	cag Gln	gac Asp	aca Thr 1290	ccc Pro	agg Arg	aag Lys	agc Ser	tgt Cys 1295	gtc Val	tac Tyr	cga Arg	tct Ser	ggc Gly 1300	3930
20	ttc Phe	tcc Ser	ttc Phe	tcc Ser	cgg Arg 1305	ggc Gly	tgc Cys	tct Ser	tac Tyr	aca Thr 1310	tgt Cys	gcc Ala	aag Lys	aag Lys	atc Ile 1315	3975
	cag Gln	gtg Val	ccg Pro	gac Asp	tgc Cys 1320	tgc Cys	cct Pro	ggc Gly	ttc Phe	ttt Phe 1325	ggc Gly	acg Thr	ctg Leu	tgt Cys	gag Glu 1330	4020
25	cca Pro	tgc Cys	cca Pro	ggg Gly	ggc Gly 1335	cta Leu	ggg Gly	ggg Gly	gtg Val	tgc Cys 1340	tca Ser	ggc Gly	cat His	ggg Gly	cag Gln 1345	4065
	tgc Cys	cag Gln	gac Asp	agg Arg	ttc Phe 1350	ctg Leu	ggc Gly	agc Ser	ggg Gly	gag Glu 1355	tgc Cys	cac His	tgc Cys	cac His	gag Glu 1360	4110
30	ggc Gly	ttc Phe	cat His	gga Gly	acg Thr 1365	gcc Ala	tgt Cys	gag Glu	gtg Val	tgt Cys 1370	gag Glu	ctg Leu	ggc Gly	cgc Arg	tac Tyr 1375	4155
35	ggg Gly	ccc Pro	aac Asn	tgc Cys	acc Thr 1380	gga Gly	gtg Val	tgt Cys	gac Asp	tgt Cys 1385	gcc Ala	cat His	ggg Gly	ctg Leu	tgc Cys 1390	4200
	cag Gln	gag Glu	ggg Gly	ctg Leu	caa Gln 1395	ggg Gly	gac Asp	gga Gly	agc Ser	tgt Cys 1400	gtc Val	tgt Cys	aac Asn	gtg Val	ggc Gly 1405	4245
40	tgg Trp	cag Gln	ggc Gly	ctc Leu	cgc Arg 1410	tgt Cys	gac Asp	cag Gln	aaa Lys	atc Ile 1415	acc Thr	agc Ser	cct Pro	cag Gln	tgc Cys 1420	4290
45	cct Pro	agg Arg	aag Lys	tgc Cys	gac Asp 1425	ccc Pro	aat Asn	gcc Ala	aac Asn	tgc Cys 1430	gtg Val	cag Gln	gac Asp	tcg Ser	gcc Ala 1435	4335
	gga Gly	gcc Ala	tcc Ser	acc Thr	tgc Cys 1440	gcc Ala	tgt Cys	gct Ala	gcg Ala	gga Gly 1445	tac Tyr	tcc Ser	ggc Gly	aat Asn	ggc Gly 1450	4380
50	atc Ile	ttc Phe	tgt Cys	tca Ser	gag Glu 1455	gtg Val	gac Asp	ccc Pro	tgc Cys	gcc Ala 1460	cac His	ggc Gly	cat His	ggg Gly	ggc Gly 1465	4425
	tgc Cys	tcc Ser	cct Pro	cat His	gcc Ala 1470	aac Asn	tgt Cys	acc Thr	aag Lys	gtg Val 1475	gca Ala	cct Pro	ggg Gly	cag Gln	cgg Arg 1480	4470
55	aca	tgc	acc	tgc	cag	gat	ggc	tac	atg	ggc	gac	ggg	gag	ctg	tgc	4515

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	Thr	Cys	Thr	Cys	Gln	Asp	Gly	Tyr	Met	Gly	Asp	Gly	Glu	Leu	Cys	
					1485					1490					1495	
5	cag	gaa	att	aac	agc	tgt	ctc	atc	cac	cac	ggg	ggc	tgc	cac	att	4560
	Gln	Glu	Ile	Asn	Ser	Cys	Leu	Ile	His	His	Gly	Gly	Cys	His	Ile	1510
					1500					1505						
10	cac	gcc	gag	tgc	atc	ccc	act	ggc	ccc	cag	cag	gtc	tcc	tgc	agc	4605
	His	Ala	Glu	Cys	Ile	Pro	Thr	Gly	Pro	Gln	Gln	Val	Ser	Cys	Ser	1525
					1515					1520						
15	tgc	cgt	gag	ggc	tac	agc	ggg	gat	ggc	atc	cgg	acc	tgc	gag	ctc	4650
	Cys	Arg	Glu	Gly	Tyr	Ser	Gly	Asp	Gly	Ile	Arg	Thr	Cys	Glu	Leu	1540
					1530					1535						
20	ctg	gac	ccc	tgc	tct	aag	aac	aat	gga	gga	tgc	agc	cca	tat	gcc	4695
	Leu	Asp	Pro	Cys	Ser	Lys	Asn	Asn	Gly	Gly	Cys	Ser	Pro	Tyr	Ala	1555
					1545					1550						
25	acc	tgc	aaa	agc	aca	ggg	gat	ggc	cag	agg	aca	tgt	acc	tgc	gac	4740
	Thr	Cys	Lys	Ser	Thr	Gly	Asp	Gly	Gln	Arg	Thr	Cys	Thr	Cys	Asp	1570
					1560					1565						
30	aca	gcc	cac	acc	gtg	ggg	gac	ggc	ctc	acc	tgc	cgt	gcc	cga	gtc	4785
	Thr	Ala	His	Thr	Val	Gly	Asp	Gly	Leu	Thr	Cys	Arg	Ala	Arg	Val	1585
					1575					1580						
35	ggc	ctg	gag	ctc	ctg	agg	gat	aag	cat	gcc	tca	ttc	ttc	agc	ctc	4830
	Gly	Leu	Glu	Leu	Leu	Arg	Asp	Lys	His	Ala	Ser	Phe	Phe	Ser	Leu	1600
					1590					1595						
40	cgc	ctc	ctg	gaa	tat	aag	gag	ctc	aag	ggc	gat	ggg	cct	ttc	acc	4875
	Arg	Leu	Leu	Glu	Tyr	Lys	Glu	Leu	Lys	Gly	Asp	Gly	Pro	Phe	Thr	1615
					1605					1610						
45	atc	ttc	gtg	ccg	cac	gca	gat	cta	atg	agc	aac	ctg	tgc	cag	gat	4920
	Ile	Phe	Val	Pro	His	Ala	Asp	Leu	Met	Ser	Asn	Leu	Ser	Gln	Asp	1630
					1620					1625						
50	gag	ctg	gcc	cgg	att	cgt	gcg	cat	cgc	cag	ctg	gtg	ttt	cgc	tac	4965
	Glu	Leu	Ala	Arg	Ile	Arg	Ala	His	Arg	Gln	Leu	Val	Phe	Arg	Tyr	1645
					1635					1640						
55	cac	gtg	ggt	ggc	tgt	cgg	cgg	ctg	cgg	agc	gag	gac	ctg	ctg	gag	5010
	His	Val	Val	Gly	Cys	Arg	Arg	Leu	Arg	Ser	Glu	Asp	Leu	Leu	Glu	1660
					1650					1655						
60	cag	ggg	tac	gcc	acg	gcc	ctc	tca	ggg	cac	cca	ctg	cgc	ttc	agc	5055
	Gln	Gly	Tyr	Ala	Thr	Ala	Leu	Ser	Gly	His	Pro	Leu	Arg	Phe	Ser	1675
					1665					1670						
65	gag	agg	gag	ggc	agc	ata	tac	ctc	aat	gac	ttc	gcg	cgc	gtg	gtg	5100
	Glu	Arg	Glu	Gly	Ser	Ile	Tyr	Leu	Asn	Asp	Phe	Ala	Arg	Val	Val	1690
					1680					1685						
70	agc	agc	gac	cat	gag	gcc	gtg	aac	ggc	atc	ctg	cac	ttc	att	gac	5145
	Ser	Ser	Asp	His	Glu	Ala	Val	Asn	Gly	Ile	Leu	His	Phe	Ile	Asp	1705
					1695					1700						
75	cgt	gtc	ctg	ctg	ccc	ccc	gag	gcg	ctg	cac	tgg	gag	cct	gat	gat	5190
	Arg	Val	Leu	Leu	Pro	Pro	Glu	Ala	Leu	His	Trp	Glu	Pro	Asp	Asp	1720
					1710					1715						
80	gct	ccc	atc	ccg	agg	aga	aat	gtc	acc	gcc	gcc	gcc	cag	ggc	ttc	5235
	Ala	Pro	Ile	Pro	Arg	Arg	Asn	Val	Thr	Ala	Ala	Ala	Gln	Gly	Phe	1735
					1725					1730						
85	ggc	tac	aag	atc	ttc	agc	ggc	ctc	ctg	aag	gtg	gcc	ggc	ctc	ctg	5280

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	Gly	Tyr	Lys	Ile	Phe 1740	Ser	Gly	Leu	Leu	Lys 1745	Val	Ala	Gly	Leu	Leu 1750	
5	ccc Pro	ctg Leu	ctt Leu	cga Arg	gag Glu 1755	gca Ala	tcc Ser	cat His	agg Arg	ccc Pro 1760	ttc Phe	aca Thr	atg Met	ctg Leu	tgg Trp 1765	5325
10	ccc Pro	aca Thr	gac Asp	gcc Ala	gcc Ala 1770	ttt Phe	cga Arg	gct Ala	ctg Leu	cct Pro 1775	ccg Pro	gat Asp	cgc Arg	cag Gln	gcc Ala 1780	5370
	tgg Trp	ctg Leu	tac Tyr	cat His	gag Glu 1785	gac Asp	cac His	cgt Arg	gac Asp	aag Lys 1790	cta Leu	gca Ala	gcc Ala	att Ile	ctg Leu 1795	5415
15	cgg Arg	ggc Gly	cac His	atg Met	att Ile 1800	cgc Arg	aat Asn	gtc Val	gag Glu	gcc Ala 1805	ttg Leu	gca Ala	tct Ser	gac Asp	ctg Leu 1810	5460
20	ccc Pro	aac Asn	ctg Leu	ggc Gly	cca Pro 1815	ctt Leu	cga Arg	acc Thr	atg Met	cat His 1820	ggg Gly	acc Thr	ccc Pro	atc Ile	tct Ser 1825	5505
	ttc Phe	tcc Ser	tgc Cys	agc Ser	cga Arg 1830	acg Thr	cgg Arg	ccc Pro	ggt Gly	gag Glu 1835	ctc Leu	atg Met	gtg Val	ggt Gly	gag Glu 1840	5550
25	gat Asp	gat Asp	gct Ala	cgc Arg	att Ile 1845	gtg Val	cag Gln	cgg Arg	cac His	ttg Leu 1850	ccc Pro	ttt Phe	gag Glu	ggt Gly	ggc Gly 1855	5595
	ctg Leu	gcc Ala	tat Tyr	ggc Gly	atc Ile 1860	gac Asp	cag Gln	ctg Leu	ctg Leu	gag Glu 1865	cca Pro	cct Pro	ggc Gly	ctt Leu	ggt Gly 1870	5640
30	gct Ala	cgc Arg	tgt Cys	gac Asp	cac His 1875	ttt Phe	gag Glu	acc Thr	cgg Arg	ccc Pro 1880	ctg Leu	cga Arg	ctg Leu	aac Asn	acc Thr 1885	5685
35	tgc Cys	agc Ser	atc Ile	tgt Cys	ggg Gly 1890	ctg Leu	gag Glu	cca Pro	ccc Pro	tgt Cys 1895	cct Pro	gag Glu	ggg Gly	tca Ser	cag Gln 1900	5730
	gag Glu	cag Gln	ggc Gly	agc Ser	cct Pro 1905	gag Glu	gcc Ala	tgc Cys	tgg Trp	cgc Arg 1910	ttc Phe	tac Tyr	ccg Pro	aag Lys	ttc Phe 1915	5775
40	tgg Trp	acg Thr	tcc Ser	cct Pro	ccg Pro 1920	ctg Leu	cac His	tct Ser	ttg Leu	gga Gly 1925	tta Leu	cgc Arg	agc Ser	gtc Val	tgg Trp 1930	5820
45	gtc Val	cac His	ccc Pro	agc Ser	ctt Leu 1935	tgg Trp	ggt Gly	agg Arg	ccc Pro	caa Gln 1940	ggc Gly	ctg Leu	ggc Gly	agg Arg	ggc Gly 1945	5865
	tgc Cys	cac His	cgc Arg	aat Asn	tgt Cys 1950	gtc Val	acc Thr	acc Thr	acc Thr	tgg Trp 1955	aag Lys	ccc Pro	agc Ser	tgc Cys	tgc Cys 1960	5910
50	cct Pro	ggt Gly	cac His	tat Tyr	ggc Gly 1965	agt Ser	gag Glu	tgc Cys	caa Gln	gct Ala 1970	tgc Cys	cct Pro	ggc Gly	ggc Gly	ccc Pro 1975	5955
	agc Ser	agc Ser	cct Pro	tgt Cys	agt Ser 1980	gac Asp	cgt Arg	ggc Gly	gtg Val	tgc Cys 1985	atg Met	gac Asp	ggc Gly	atg Met	agt Ser 1990	6000
55	ggc	agt	ggg	cag	tgt	ctg	tgc	cgt	tca	ggt	ttt	gct	ggg	aca	gcc	6045

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	Gly	Ser	Gly	Gln	Cys 1995	Leu	Cys	Arg	Ser	Gly 2000	Phe	Ala	Gly	Thr	Ala 2005	
5	tgt	gaa	ctc	tgt	gct	cct	ggt	gcc	ttt	ggg	ccc	cat	tgt	caa	gcc	6090
	Cys	Glu	Leu	Cys	Ala 2010	Pro	Gly	Ala	Phe	Gly 2015	Pro	His	Cys	Gln	Ala 2020	
	tgc	cgc	tgc	act	gtg	cat	ggc	cgc	tgt	gat	gag	ggc	ctt	ggg	ggc	6135
10	Cys	Arg	Cys	Thr	Val 2025	His	Gly	Arg	Cys	Asp 2030	Glu	Gly	Leu	Gly	Gly 2035	
	tct	ggc	tcc	tgc	ttc	tgt	gat	gaa	ggc	tgg	act	ggg	cca	cgc	tgt	6180
	Ser	Gly	Ser	Cys	Phe 2040	Cys	Asp	Glu	Gly	Trp 2045	Thr	Gly	Pro	Arg	Cys 2050	
15	gag	gtg	caa	ctg	gag	ctg	cag	cct	gtg	tgt	acc	cca	ccc	tgt	gca	6225
	Glu	Val	Gln	Leu	Glu 2055	Leu	Gln	Pro	Val	Cys 2060	Thr	Pro	Pro	Cys	Ala 2065	
	ccc	gag	gct	gtg	tgc	cgt	gca	ggc	aac	agc	tgt	gag	tgc	agc	ctg	6270
20	Pro	Glu	Ala	Val	Cys 2070	Arg	Ala	Gly	Asn	Ser 2075	Cys	Glu	Cys	Ser	Leu 2080	
	ggc	tat	gaa	ggg	gat	ggc	cgc	gtg	tgt	aca	gtg	gca	gac	ctg	tgc	6315
	Gly	Tyr	Glu	Gly	Asp 2085	Gly	Arg	Val	Cys	Thr 2090	Val	Ala	Asp	Leu	Cys 2095	
25	cag	gac	ggg	cat	ggt	ggc	tgc	agt	gag	cac	gcc	aac	tgt	agc	cag	6360
	Gln	Asp	Gly	His	Gly 2100	Gly	Cys	Ser	Glu	His 2105	Ala	Asn	Cys	Ser	Gln 2110	
	gta	gga	aca	atg	gtc	act	tgt	acc	tgc	ctg	ccc	gac	tac	gag	ggt	6405
	Val	Gly	Thr	Met	Val 2115	Thr	Cys	Thr	Cys	Leu 2120	Pro	Asp	Tyr	Glu	Gly 2125	
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	Asp	Gly	Trp	Ser	Cys 2130	Arg	Ala	Arg	Asn	Pro 2135	Cys	Thr	Asp	Gly	His 2140	
	cgc	ggg	ggc	tgc	agc	gag	cac	gcc	aac	tgc	ttg	agc	acc	ggc	ctg	6495
35	Arg	Gly	Gly	Cys	Ser 2145	Glu	His	Ala	Asn	Cys 2150	Leu	Ser	Thr	Gly	Leu 2155	
	aac	aca	cgg	cgc	tgt	gag	tgc	cac	gca	ggc	tac	gta	ggc	gat	gga	6540
	Asn	Thr	Arg	Arg	Cys 2160	Glu	Cys	His	Ala	Gly 2165	Tyr	Val	Gly	Asp	Gly 2170	
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	Leu	Gln	Cys	Leu	Glu 2175	Glu	Ser	Glu	Pro	Pro 2180	Val	Asp	Arg	Cys	Leu 2185	
	ggc	cag	cca	ccg	ccc	tgc	cac	tca	gat	gcc	atg	tgc	act	gac	ctg	6630
45	Gly	Gln	Pro	Pro	Pro 2190	Cys	His	Ser	Asp	Ala 2195	Met	Cys	Thr	Asp	Leu 2200	
	cac	ttc	cag	gag	aaa	cgg	gct	ggc	gtt	ttc	cac	ctc	cag	gcc	acc	6675
	His	Phe	Gln	Glu	Lys 2205	Arg	Ala	Gly	Val	Phe 2210	His	Leu	Gln	Ala	Thr 2215	
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	Ser	Gly	Pro	Tyr	Gly 2220	Leu	Asn	Phe	Ser	Glu 2225	Ala	Glu	Ala	Ala	Cys 2230	
	gaa	gca	cag	gga	gcc	gtc	ctt	gct	tca	ttc	cct	cag	ctc	tct	gct	6765
	Glu	Ala	Gln	Gly	Ala 2235	Val	Leu	Ala	Ser	Phe 2240	Pro	Gln	Leu	Ser	Ala 2245	
55	gcc	cag	cag	ctg	ggc	ttc	cac	ctg	tgc	ctc	atg	ggc	tgg	ctg	gcc	6810

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	Ala	Gln	Gln	Leu	Gly 2250	Phe	His	Leu	Cys	Leu 2255	Met	Gly	Trp	Leu	Ala 2260	
5	aat	ggc	tcc	act	gcc Ala 2265	cac	cct	gtg	gtt	ttc Phe 2270	cct	gtg	gcg	gac	tgt Cys 2275	6855
	Asn	Gly	Ser	Thr		His	Pro	Val	Val		Pro	Val	Ala	Asp		
10	ggc	aat	ggt	cgg	gtg Val 2280	ggc	gta	gtc	agc	ctg Leu 2285	ggt	gcc	cgc	aag	aac Asn 2290	6900
	Gly	Asn	Gly	Arg		Gly	Val	Val	Ser		Gly	Ala	Arg	Lys		
15	ctc	tca	gaa	cgc	tgg Trp 2295	gat	gcc	tac	tgc	ttc Phe 2300	cg	gtg	caa	gat	gtg Val 2305	6945
	Leu	Ser	Glu	Arg		Asp	Ala	Tyr	Cys		Arg	Val	Gln	Asp		
20	gcc	tgc	cga	tgc	cga Arg 2310	aat	ggc	ttc	gtg	ggt Gly 2315	gac	ggg	atc	agc	acg Thr 2320	6990
	Ala	Cys	Arg	Cys		Asn	Gly	Phe	Val		Asp	Gly	Ile	Ser		
25	tgc	aat	ggg	aag	ctg Leu 2325	ctg	gat	gtg	ctg	gct Ala 2330	gcc	act	gcc	aac	ttc Phe 2335	7035
	Cys	Asn	Gly	Lys		Leu	Asp	Val	Leu		Ala	Thr	Ala	Asn		
30	tcc	acc	ttc	tat	ggg Gly 2340	atg	cta	ttg	ggc	tat Tyr 2345	gcc	aat	gcc	acc	cag Gln 2350	7080
	Ser	Thr	Phe	Tyr		Met	Leu	Leu	Gly		Ala	Asn	Ala	Thr		
35	cgg	ggt	ctc	gac	ttc Phe 2355	ctg	gac	ttc	ctg	gat Asp 2360	gat	gag	ctc	acg	tat Tyr 2365	7125
	Arg	Gly	Leu	Asp		Leu	Asp	Phe	Leu		Asp	Glu	Leu	Thr		
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	Lys	Thr	Leu	Phe		Pro	Val	Asn	Glu		Phe	Val	Asp	Asn		
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	Thr	Leu	Ser	Gly		Asp	Leu	Glu	Leu		Ala	Ser	Asn	Ala		
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	Leu	Leu	Ser	Ala		Ala	Ser	Gln	Gly		Leu	Leu	Pro	Ala		
55	tca	ggc	ctc	agc	ctc Leu 2415	atc	atc	agt	gac	gca Ala 2420	ggc	cct	gac	aac	agt Ser 2425	7305
	Ser	Gly	Leu	Ser		Ile	Ile	Ser	Asp		Gly	Pro	Asp	Asn		
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	Ser	Trp	Ala	Pro		Ala	Pro	Gly	Thr		Val	Val	Ser	Arg		
65	att	gtg	tgg	gac	atc Ile 2445	atg	gcc	ttc	aat	ggc Gly 2450	atc	atc	cat	gct	ctg Leu 2455	7395
	Ile	Val	Trp	Asp		Met	Ala	Phe	Asn		Ile	Ile	His	Ala		
70	gcc	agc	ccc	ctc	ctg Leu 2460	gca	ccc	cca	cag	ccc Pro 2465	cag	gca	gtg	ctg	gcg Ala 2470	7440
	Ala	Ser	Pro	Leu		Ala	Pro	Pro	Gln		Gln	Ala	Val	Leu		
75	cct	gaa	gcc	cca	cct Pro 2475	gtg	gcg	gca	ggc	gtg Val 2480	ggg	gct	gtg	ctt	gcc Ala 2485	7485
	Pro	Glu	Ala	Pro		Val	Ala	Ala	Gly		Gly	Ala	Val	Leu		
80	gct	gga	gca	ctg	ctt Leu 2490	ggc	ttg	gtg	gcc	gga Gly 2495	gct	ctc	tac	ctc	cg	7530
	Ala	Gly	Ala	Leu		Gly	Leu	Val	Ala		Ala	Leu	Tyr	Leu	Arg 2500	
85	gcc	cga	ggc	aag	ccc	acg	ggc	ttt	ggc	ttc	tct	gcc	ttc	cag	gcg	7575

	Ala	Arg	Gly	Lys	Pro 2505	Thr	Gly	Phe	Gly	Phe 2510	Ser	Ala	Phe	Gln	Ala 2515	
5	gaa	gat	gat	gct	gac	gac	gac	ttc	tca	ccg	tgg	caa	gaa	ggg	acc	7620
	Glu	Asp	Asp	Ala	Asp 2520	Asp	Asp	Phe	Ser	Pro 2525	Trp	Gln	Glu	Gly	Thr 2530	
	aac	ccc	acc	ctg	gtc	tct	gtc	ccc	aac	cct	gtc	ttt	ggc	agc	gac	7665
10	Asn	Pro	Thr	Leu	Val 2535	Ser	Val	Pro	Asn	Pro 2540	Val	Phe	Gly	Ser	Asp 2545	
	acc	ttt	tgt	gaa	ccc	ttc	gat	gac	tca	ctg	ctg	gag	gag	gac	ttc	7710
	Thr	Phe	Cys	Glu	Pro 2550	Phe	Asp	Asp	Ser	Leu 2555	Leu	Glu	Glu	Asp	Phe 2560	
15	cct	gac	acc	cag	agg	atc	ctc	aca	gtc	aag	tgacgaggct	ggggctgaaa			7760	
	Pro	Asp	Thr	Gln	Arg 2565	Ile	Leu	Thr	Val	Lys 2570						
	gcagaagcat	gcacagggag	gagaccactt	ttattgcttg	tctgggtgga	tggggcagga									7820	
20	ggggctgagg	gcctgtccca	gacaataaag	tgccctcagc	ggatgtgggc	catgtcacc									7879	

Claims

- 25
1. A method of *in vitro* diagnosing inflammatory diseases in a patient, said method comprising:
- (a) exposing a blood or tissue sample from said patient to a common lymphatic endothelial and vascular endothelial glycoprotein (CLEVER-1) *in vitro*,
- 30 wherein said glycoprotein has a molecular weight of 270-300 kD in SDS-PAGE under non-reducing conditions, and is recognizable by a monoclonal antibody selected from the group consisting of
- (i) monoclonal antibody 3-266 (DSM ACC 2519); and
- (ii) monoclonal antibody 3-372 (DSM ACC 2590),
- 35 for a period of time and under conditions sufficient to allow binding of leukocytes if present in said sample; and
- (b) detecting leukocytes bound to said glycoprotein in said blood or tissue sample.
- 40
2. A method of *in vitro* detecting malignant cells in a patient, said method comprising:
- (a) exposing a blood or tissue sample from a patient to the glycoprotein as defined in claim 2 *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample; and
- (b) detecting whether any malignant cells bound to said glycoprotein in said blood or tissue sample.
- 45
3. The method of claim 2, wherein said glycoprotein is on a solid support.
4. The method of claim 2, wherein said glycoprotein is provided on lymphoid tissue.
5. The method of claim 2, wherein said glycoprotein is present in the membrane of endothelial cells.
- 50
6. The method of claim 2, wherein said glycoprotein is in a soluble form.
7. The method of claim 2, wherein said detection step is performed by imaging.
- 55
8. A method of removing malignant cells from a sample, said method comprising:
- (a) exposing said malignant cells to the glycoprotein as defined in claim 1 *in vitro* for a period of time and under conditions sufficient to allow binding of said malignant cells if present in said sample, and

(b) separating said glycoprotein and the malignant cells that adhere thereto from said sample.

5 9. Use of an agent that inhibits glycoprotein mediated leukocyte binding, wherein said glycoprotein is defined in claim 1, for the manufacture of a pharmaceutical composition useful in a method of treating inflammation in a patient in need of the same, wherein said inhibiting agent is selected from the group consisting of:

- (a) antibodies or fragments thereof, raised to said glycoprotein; and
- (b) said glycoprotein or fragments thereof in soluble form.

10 10. Use of an agent that inhibits malignant cell binding mediated by the glycoprotein as defined in claim 1, for the manufacture of a pharmaceutical composition useful in a method of preventing metastasis in a patient in need of the same, wherein said inhibiting agent is selected from the group consisting of:

- 15
- (a) antibodies or fragments thereof, raised to said glycoprotein; and
 - (b) said glycoprotein or fragments thereof in soluble form.

11. The use of claim 9, wherein said glycoprotein mediated cell binding inhibits leukocyte binding.

20 12. The use of claim 9 wherein said glycoprotein mediated cell binding inhibits lymphocyte binding.

13. The use of claim 9, wherein said glycoprotein mediated cell binding inhibits monocyte binding.

14. The use of claim 9, wherein said glycoprotein mediated cell binding inhibits granulocyte binding.

25 15. The use of claim 10, wherein said glycoprotein mediated cell binding inhibits malignant cell binding.

16. The use of claim 9 or 10, wherein said glycoprotein binding agent is administered to a patient in need of an inhibition of glycoprotein mediated cell binding.

30 17. The use of claim 16, wherein said patient is in need of treatment of inflammation.

18. The use of claim 17, wherein said patient is in need of treatment for a malignancy or possible malignancy.

35 19. A method of *in vitro* diagnosing inflammatory diseases in a patient, said method comprising exposing a blood or tissue sample from said patient to an antibody, which is either

- monoclonal antibody 3-266 (DSM ACC 2519); or
- monoclonal antibody 3-372 (DSM ACC 2590).

40

Patentansprüche

45 1. Verfahren zum In-vitro-Diagnostizieren von Entzündungserkrankungen bei einem Patienten, wobei das Verfahren umfasst:

(a) Exponieren einer Blut- oder Gewebeprobe von einem Patienten gegenüber einem gemeinsamen Lymphendothel- und Gefäßendothel-Glycoprotein (CLEVER-1) *in vitro*, wobei das Glycoprotein ein Molekulargewicht von 270 bis 300 kD in einer SDS-PAGE unter nicht-reduzierenden Bedingungen hat und durch einen monoklonalen Antikörper, ausgewählt aus der Gruppe, bestehend aus

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- (i) monoklonaler Antikörper 3-266 (DSM ACC 2519); und
- (ii) monoklonaler Antikörper 3-372 (DSM ACC 2590), erkennbar ist,

für eine Zeitdauer und unter Bedingungen, die ausreichend sind, um das Binden von Leukozyten, falls in der Probe vorhanden, zu ermöglichen, und

55

(b) Detektieren von Leukozyten, die an das Glycoprotein in der Blut- oder Gewebeprobe gebunden sind.

2. Verfahren zum In-vitro-Detektieren maligner Zellen bei einem Patienten, wobei das Verfahren umfasst:

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- (a) Exponieren einer Blut- oder Gewebeprobe von einem Patienten gegenüber dem Glycoprotein, wie in Anspruch 2 definiert, *in vitro* für eine Zeitdauer und unter Bedingungen, die ausreichend sind, um das Binden der malignen Zellen, falls in der Probe vorhanden, zu ermöglichen, und
- 5 (b) Detektieren, ob irgendwelche malignen Zellen in der Blut- oder Gewebeprobe an das Glycoprotein gebunden haben.
3. Verfahren nach Anspruch 2, wobei das Glycoprotein auf einem festen Träger ist.
4. Verfahren nach Anspruch 2, wobei das Glycoprotein auf lymphoidem Gewebe bereitgestellt wird.
- 10 5. Verfahren nach Anspruch 2, wobei das Glycoprotein in der Membran von Endothelzellen vorhanden ist.
6. Verfahren nach Anspruch 2, wobei das Glycoprotein in einer löslichen Form vorliegt.
- 15 7. Verfahren nach Anspruch 2, wobei der Detektionsschritt mittels Bildgebung durchgeführt wird.
8. Verfahren zum Entfernen von malignen Zellen aus einer Probe, wobei das Verfahren umfasst:
- 20 (a) Exponieren der malignen Zellen gegenüber dem Glycoprotein, wie in Anspruch 1 definiert, *in vitro* für eine Zeitdauer und unter Bedingungen, die ausreichend sind, um die Bindung maligner Zellen, falls in der Probe vorhanden, zu ermöglichen, und
- (b) Abtrennen des Glycoproteins und der malignen Zellen, die daran haften, aus der Probe.
9. Verwendung eines Mittels, das die Glycoproteinvermittelte Leukozytenbindung inhibiert, wobei das Glycoprotein in Anspruch 1 definiert ist, zur Herstellung einer pharmazeutischen Zusammensetzung, die in einem Verfahren zum Behandeln von Entzündung bei einem Patienten, der dessen bedarf, zweckmäßig ist, wobei das inhibierende Mittel ausgewählt ist aus der Gruppe, bestehend aus:
- 25 (a) Antikörpern oder Fragmenten davon, erzeugt gegen das Glycoprotein; und
- 30 (b) dem Glycoprotein oder Fragmenten davon in löslicher Form.
10. Verwendung eines Mittels, das die Bindung maligner Zellen, vermittelt durch das Glycoprotein, wie in Anspruch 1 definiert, inhibiert, zur Herstellung einer pharmazeutischen Zusammensetzung, die in einem Verfahren zum Verhindern von Metastase bei einem Patienten, der dessen bedarf, zweckmäßig ist, wobei das inhibierende Mittel ausgewählt ist aus der Gruppe, bestehend aus:
- 35 (a) Antikörpern oder Fragmenten davon, erzeugt gegen das Glycoprotein; und
- (b) dem Glycoprotein oder Fragmenten davon in löslicher Form.
- 40 11. Verwendung nach Anspruch 9, wobei die Glycoproteinvermittelte Zellbindung Leukozytenbindung inhibiert.
12. Verwendung nach Anspruch 9, wobei die Glycoproteinvermittelte Zellbindung Lymphozytenbindung inhibiert.
13. Verwendung nach Anspruch 9, wobei die Glycoproteinvermittelte Zellbindung Monozytenbindung inhibiert.
- 45 14. Verwendung nach Anspruch 9, wobei die Glycoproteinvermittelte Zellbindung Granulozytenbindung inhibiert.
15. Verwendung nach Anspruch 10, wobei die Glycoproteinvermittelte Zellbindung die Bindung maligner Zellen inhibiert.
- 50 16. Verwendung nach Anspruch 9 oder 10, wobei das Glycoproteinbindungsmittel an einen Patienten verabreicht wird, der einer Inhibierung von Glycoprotein-vermittelter Zellbindung bedarf.
17. Verwendung nach Anspruch 16, wobei der Patient einer Behandlung von Entzündung bedarf.
- 55 18. Verwendung nach Anspruch 17, wobei der Patient einer Behandlung für eine Malignität oder eine mögliche Malignität bedarf.
19. Verfahren zum *In-vitro*-Diagnostizieren von Entzündungserkrankungen bei einem Patienten, wobei das Verfahren

das Exponieren einer Blut- oder Gewebeprobe von dem Patienten gegenüber einem Antikörper umfasst, der entweder

- monoklonaler Antikörper 3-266 (DSM ACC 2519); oder
- monoklonaler Antikörper 3-372 (DSM ACC 2590)

ist.

Revendications

1. Procédé de diagnostic *in vitro* de maladies inflammatoires chez un patient, ledit procédé comprenant :

(a) l'exposition, *in vitro*, d'un échantillon de sang ou de tissu provenant dudit patient à une glycoprotéine lymphatique endothéliale et vasculaire commune (CLEVER-1), dans lequel ladite glycoprotéine a une masse moléculaire de 270 à 300 kD par SDS-PAGE dans des conditions non réductrices et est reconnaissable par un anticorps monoclonal choisi dans le groupe constitué par :

- (i) un anticorps monoclonal 3-266 (DSM ACC 2519) ; et
- (ii) un anticorps monoclonal 3-372 (DSM ACC 2590), pendant une durée de temps et dans des conditions suffisantes qui permettent la liaison des leucocytes, si présents, dans ledit échantillon;

et

(b) la détection de leucocytes liés à ladite glycoprotéine dans ledit échantillon de sang ou de tissu.

2. Procédé de détection, *in vitro*, de cellules malignes chez un patient, ledit procédé comprenant :

- (a) l'exposition, *in vitro*, d'un échantillon de sang ou de tissu provenant d'un patient à la glycoprotéine telle que définie dans la revendication 1 pendant une durée de temps et dans des conditions suffisantes qui permettent la liaison desdites cellules malignes, si présentes, dans ledit échantillon ; et
- (b) la détection, le cas échéant, de cellules malignes liées à ladite glycoprotéine dans ledit échantillon de sang ou de tissu.

3. Procédé selon la revendication 2, dans lequel ladite glycoprotéine est sur un support solide.

4. Procédé selon la revendication 2, dans lequel ladite glycoprotéine est fournie sur un tissu lymphoïde.

5. Procédé selon la revendication 2, dans lequel ladite glycoprotéine est présente dans la membrane de cellules endothéliales.

6. Procédé selon la revendication 2, dans lequel ladite glycoprotéine est sous une forme soluble.

7. Procédé selon la revendication 2, dans lequel ladite étape de détection est réalisée par imagerie.

8. Procédé d'élimination de cellules malignes d'un échantillon, ledit procédé comprenant :

- (a) l'exposition, *in vitro*, desdites cellules malignes à la glycoprotéine telle que définie dans la revendication 1 pendant une durée de temps et dans des conditions suffisantes qui permettent la liaison desdites cellules malignes, si présentes, dans ledit échantillon, et ;
- (b) la séparation de ladite glycoprotéine et des cellules malignes qui adhèrent à celle-ci dudit échantillon.

9. Utilisation d'un agent qui inhibe la liaison des leucocytes induite par les glycoprotéines, dans laquelle ladite glycoprotéine est définie dans la revendication 1, pour la fabrication d'une composition pharmaceutique utile dans un procédé de traitement d'une inflammation chez un patient nécessitant celui-ci, dans lequel ledit agent inhibiteur est choisi dans le groupe constitué par :

- (a) des anticorps ou des fragments de ceux-ci, dirigés contre ladite glycoprotéine ; et
- (b) ladite glycoprotéine ou des fragments de celle-ci sous une forme soluble.

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10. Utilisation d'un agent qui inhibe la liaison de cellules malignes induite par la glycoprotéine telle que définie dans la revendication 1, pour la fabrication d'une composition pharmaceutique utile dans un procédé de prévention de métastases chez un patient nécessitant celui-ci, dans lequel ledit agent inhibiteur est choisi dans le groupe constitué par :

5

- (a) des anticorps ou des fragments de celui-ci, dirigé contre ladite glycoprotéine ; et
- (b) ladite glycoprotéine ou des fragments de celle-ci sous une forme soluble.

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11. Utilisation selon la revendication 9 dans laquelle ladite liaison de cellules induite par la glycoprotéine inhibe la liaison des leucocytes.

12. Utilisation selon la revendication 9 dans laquelle ladite liaison de cellules induite par la glycoprotéine inhibe la liaison des lymphocytes.

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13. Utilisation selon la revendication 9 dans laquelle ladite liaison de cellules induite par la glycoprotéine inhibe la liaison des monocytes.

14. Utilisation selon la revendication 9 dans laquelle ladite liaison de cellules induite par la glycoprotéine inhibe la liaison des granulocytes.

20

15. Utilisation selon la revendication 10 dans laquelle ladite liaison de cellules induite par la glycoprotéine inhibe la liaison de cellules malignes.

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16. Utilisation selon la revendication 9 ou 10 dans laquelle ledit agent de liaison à la glycoprotéine est administré à un patient nécessitant une inhibition de la liaison des cellules induite par la glycoprotéine.

17. Utilisation selon la revendication 16, dans laquelle ledit patient a besoin d'un traitement d'une inflammation.

30

18. Utilisation selon la revendication 17, dans laquelle ledit patient a besoin d'un traitement d'une affection maligne ou d'une affection maligne éventuelle.

19. Procédé de diagnostic *in vitro* de maladies inflammatoires chez un patient, ledit procédé comprenant l'exposition d'un échantillon de sang ou de tissu provenant dudit patient à un anticorps, qui est soit :

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- l'anticorps monoclonal 3-266 (DSM ACC 2519) ; ou
- l'anticorps monoclonal 3-372 (DSM ACC 2590).

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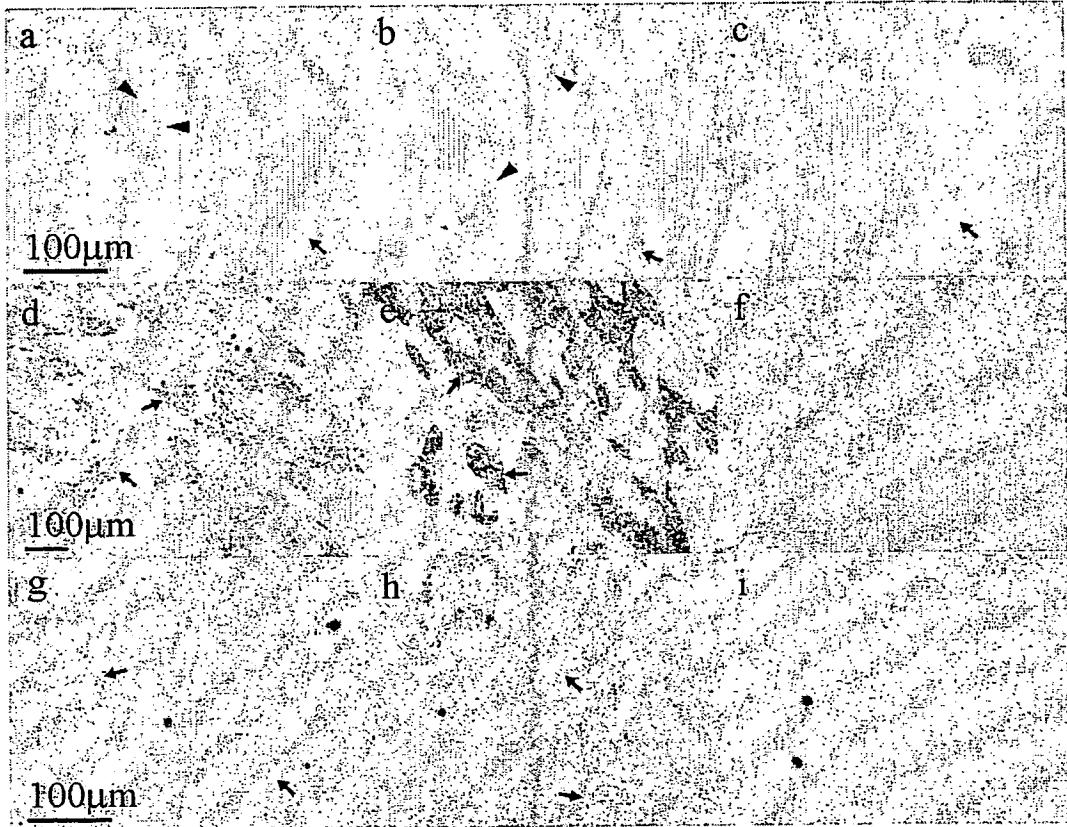


Fig. 1

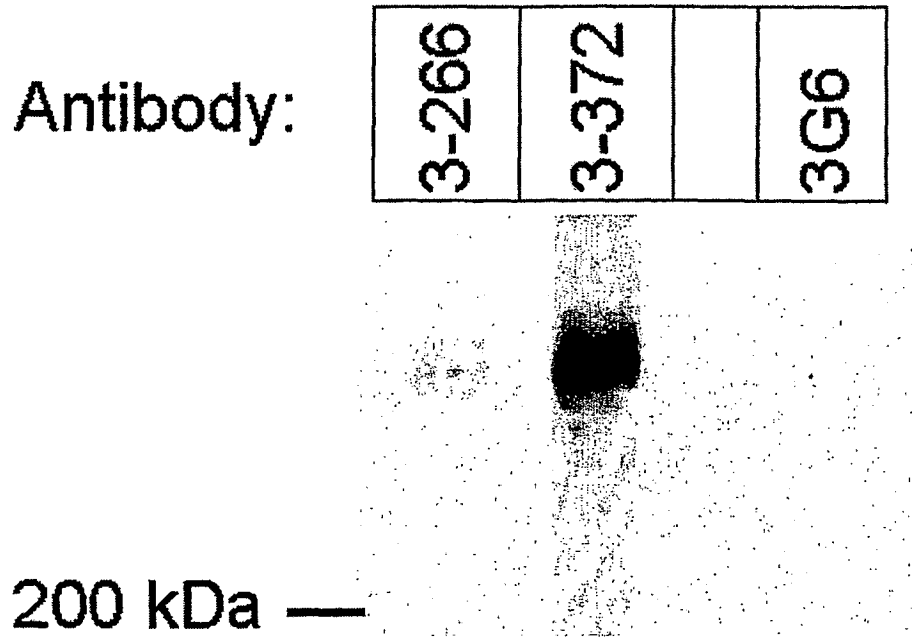
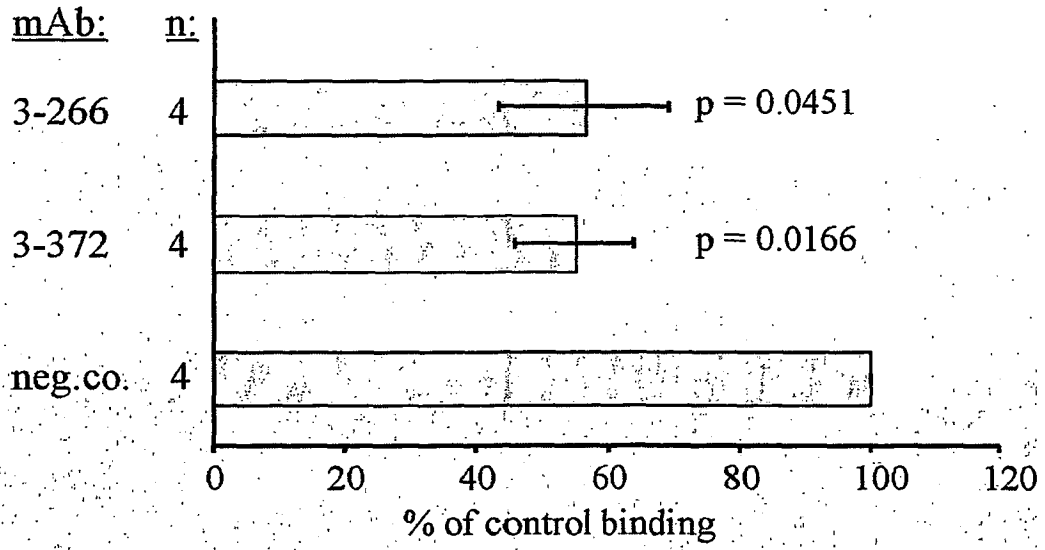


Fig. 2

A Lymphocyte binding to HEVs:



B Lymphocyte binding to lymph endothelium:

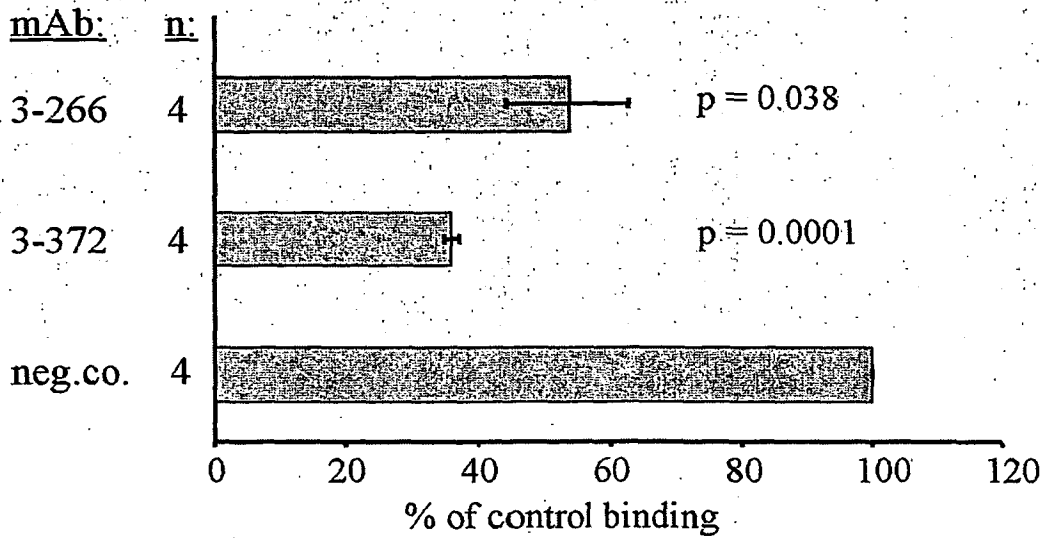
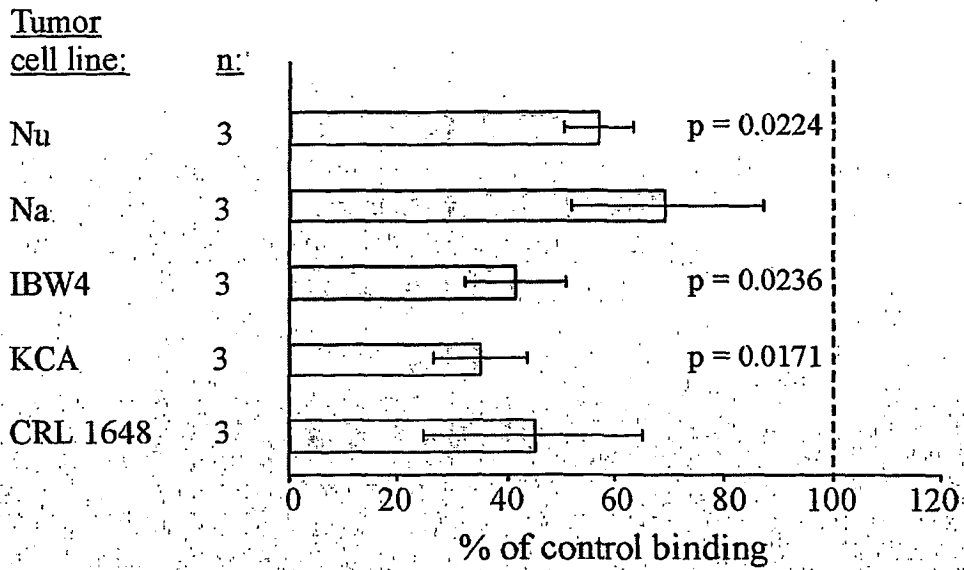


Fig. 3

A

Tumor cell binding to HEVs:



B

Tumor cell binding to lymphatic endothelium:

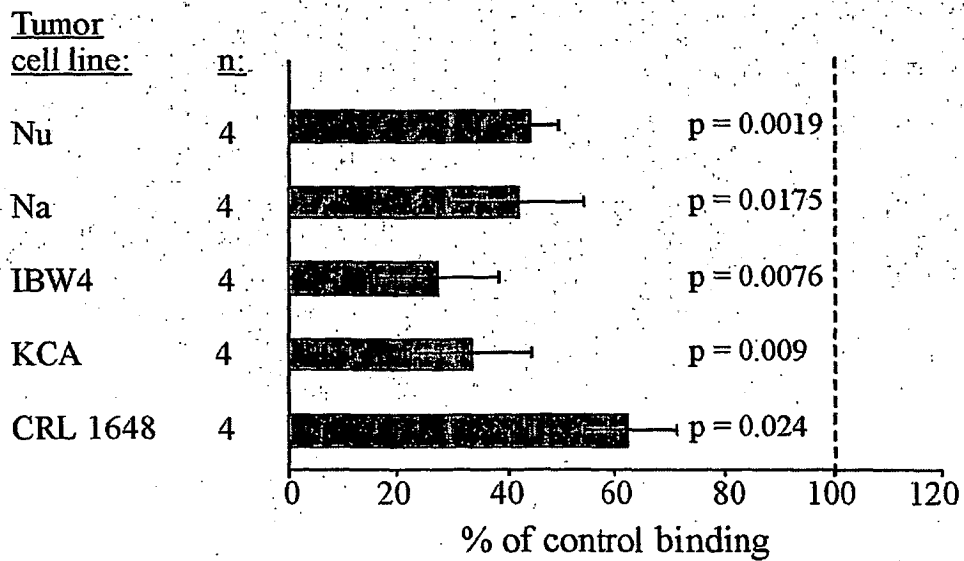


Fig. 4

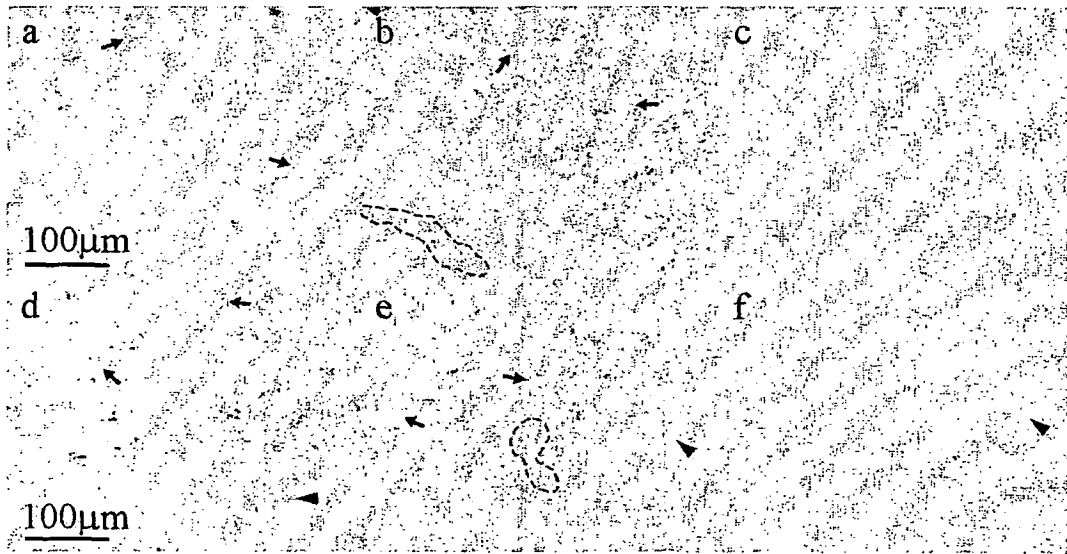


Fig. 5

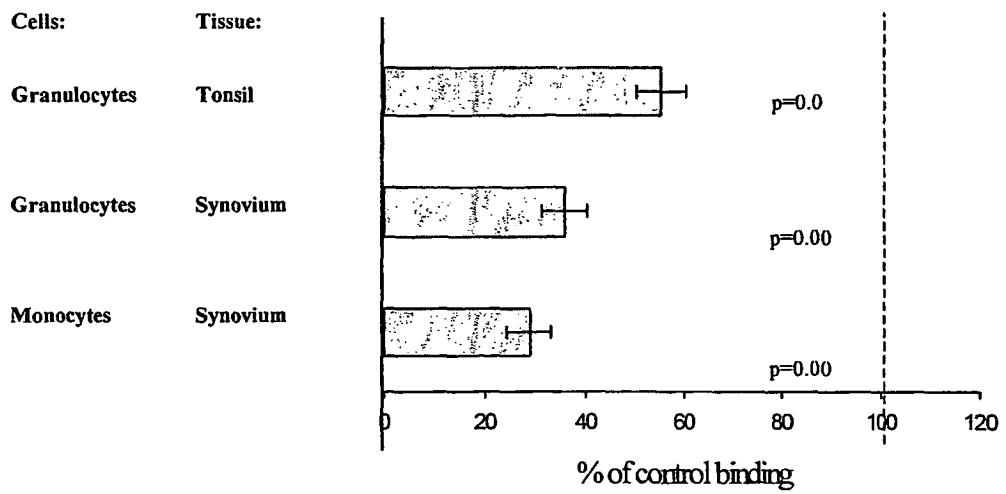


Fig. 6

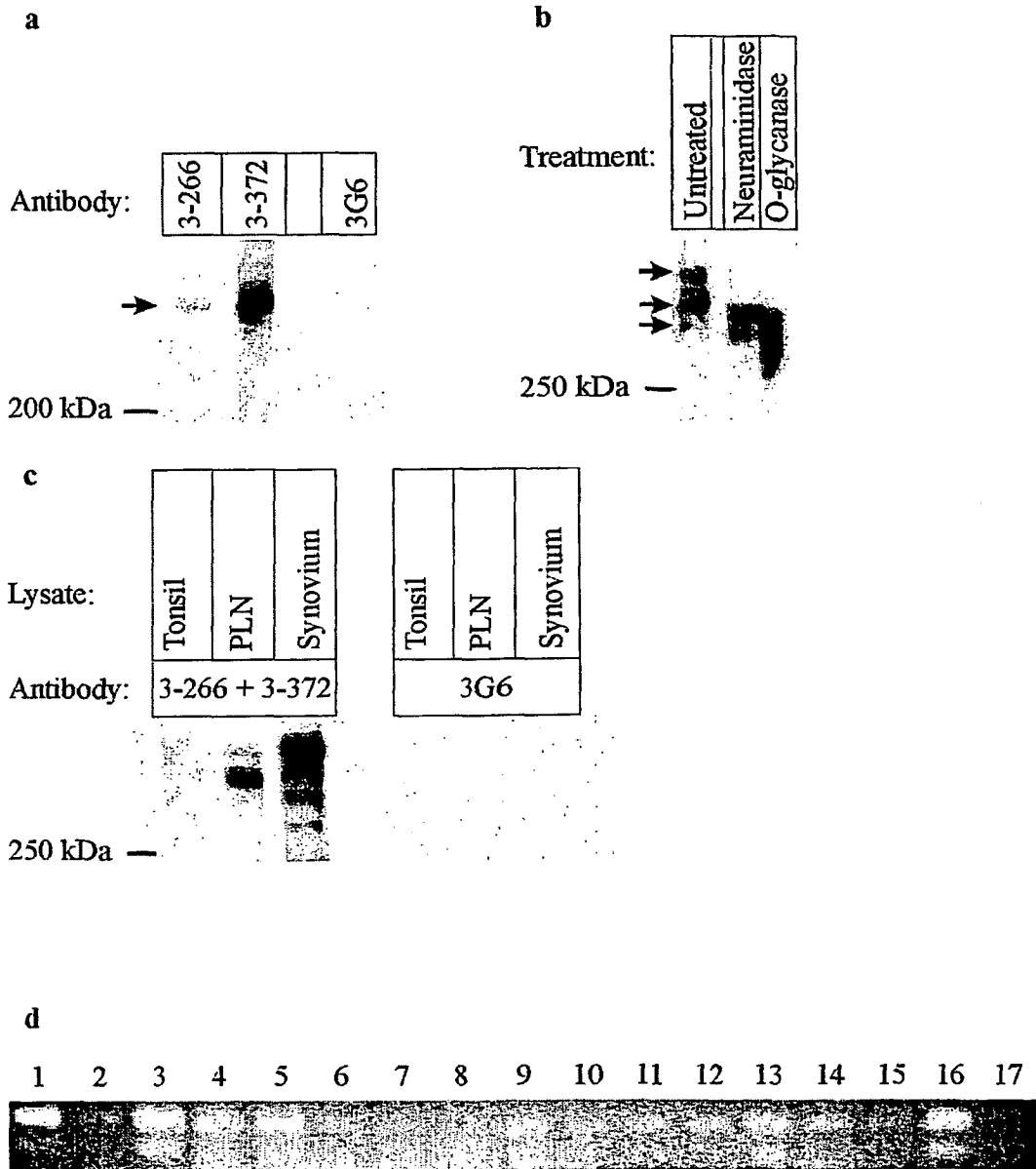


Fig. 7

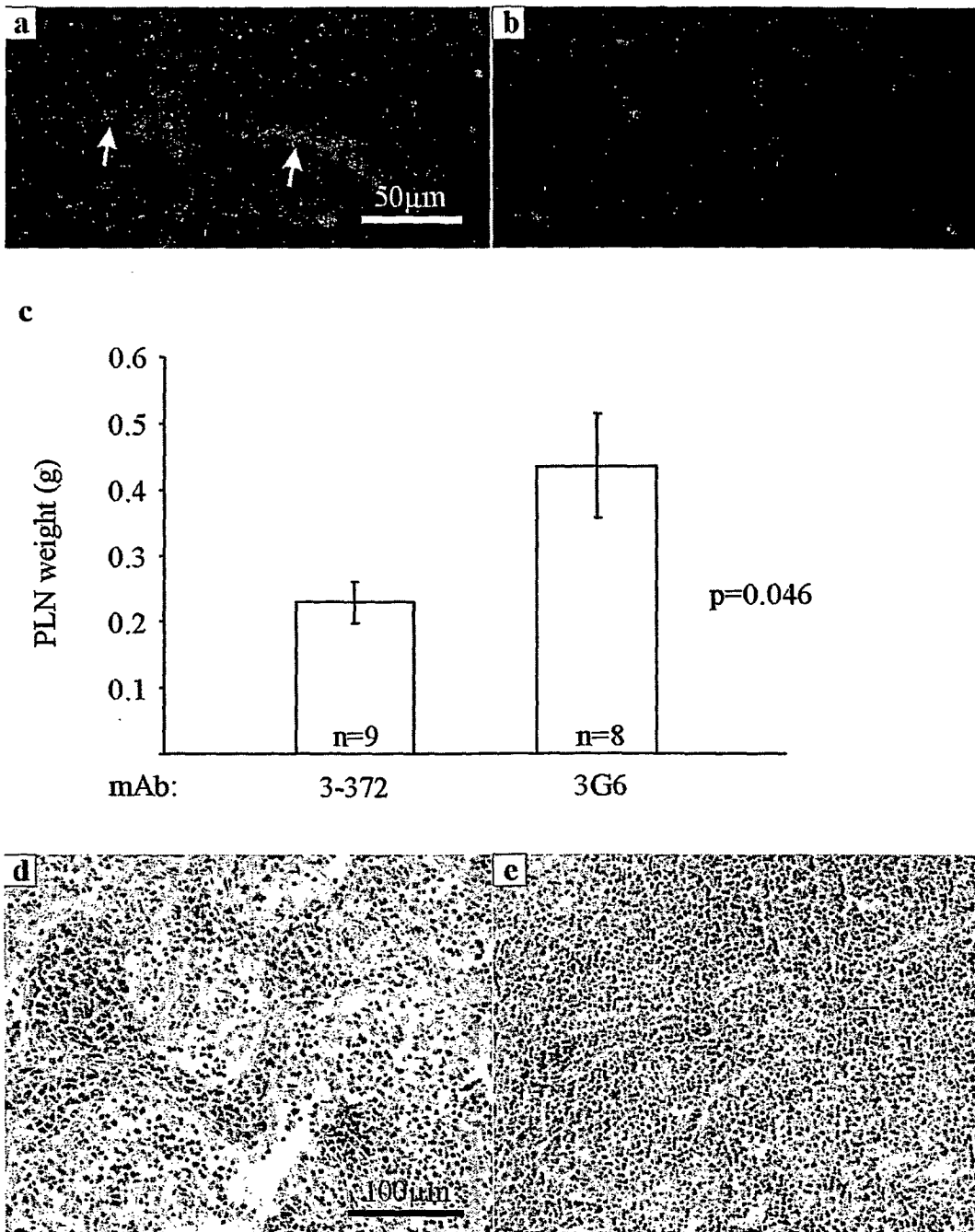


Fig. 8

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Fig. 9

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ggc ttt gcc ggg gat ggc tac cag tgc agc ccc atc gac ccc tgc cgg	2886
gca ggc aat ggc ggc tgc cac ggc <u>ctg gcc acc tgc cgg gca gtg ggg</u>	2934
<u>gga ggt cag cgg ttc tgc acg tgc ccc cct ggc ttt ggg ggt gat ggc</u>	2982
<u>ttc agc tgt tat gga gac atc ttc cgg</u> gag ctg gag gca aat gcc cac	3030
ttc tcc atc ttc tac caa tgg ctt aag agt gcc ggc atc acg ctt	3075
cct gcc gac cgc cga gtc aca gcc ctg gtg ccc tcc gag gct gca	3120
gtc cgt cag ctg agc ccc gag gac cga gct ttc tgg ctg cag cca	3165
agg acy ctg ccg aac ctg gtc agg gcc cat ttt ctc cag ggt gcc	3210
ctc ttc gag gag gag ctg gcc cgg ctg ggt ggg cag gaa gtg gcc	3255
acc ctg aac ccc acc aca cgc tgg gag att cgc aac att agt ggg	3300
agg gtc tgg gtg cag aat gcc agc gtg gat gtg gct gac ctc ctt	3345
gcc acc aac ggt gtc cta cac atc ctc agc cag gtc tta ctg ccc	3390
ccc cga ggg gat gtg ccc ggt ggg cag ggg ttg ctg cag cag ctg	3435
gac ttg gtg cct gcc ttc agc ctc ttc cgg gaa ttg ctg cag cac	3480
cat ggg ttg gtg ccc cag att gag gct gcc act gcc tac acc atc	3525
ttt gtg ccc acc aac cgc tcc ctg gag gcc cag ggc aac agc agt	3570
cac ctg gac gca gac aca gtg cgg cac cat gtg gtc ctg ggg gag	3615
gcc ctc tcc atg gaa acc ctg cgg aag ggt gga cac cgc aac tcc	3660
ctc ctg ggc cct gcc cac tgg atc gtc ttc tac aac cac agt ggc	3705
cag cct gag gtg aac cat gtg cca ctg gaa ggc ccc atg ctg gag	3750
gcc cct ggc cgc tgc ctg att ggt ctg tgc ggg gtc ctg acg gtg	3795
ggc tca agt cgc tgc ctg cat agc cac gct gag gcc ctg cgg gag	3840
aaa tgt gta aac tgc acc agg aga ttc cgc tgc act cag ggc ttc	3885
cag ctg cag gac aca ccc agg aag agc tgt gtc tac cga tct ggc	3930
ttc tcc ttc tcc cgg ggc tgc tct tac aca tgt gcc aag aag atc	3975
cag gtg ccg gac tgc tgc cct ggt ttc ttt ggc acg ctg tgt gag	4020
cca tgc cca ggg ggt cta ggg ggg gtg tgc tca ggc cat ggg cag	4065
tgc cag gac agg ttc ctg ggc agc ggg gag tgc cac tgc cac gag	4110
ggc ttc cat gga acg gcc tgt gag gtg tgt gag ctg ggc cgc tac	4155
ggg ccc aac tgc acc gga gtg tgt gac tgt gcc cat ggg ctg tgc	4200
cag gag ggg ctg caa ggg gac gga agc tgt gtc tgt aac gtg ggc	4245
tgg cag ggc ctc cgc tgt gac cag aaa atc acc agc cct cag tgc	4290
cct agg aag tgc gac ccc aat gcc aac tgc gtg cag gac tgc gcc	4335
gga gcc tcc acc tgc gcc tgt gct gcg gga tac tcc ggc aat ggc	4380
atc ttc tgt tca gag gtg gac ccc tgc gcc cac ggc cat ggg ggc	4425
tgc tcc cct cat gcc aac tgt acc aag gtg gca cct ggg cag cgg	4470
aca tgc acc tgc cag gat ggc tac atg ggc gac ggg gag ctg tgc	4515
cag gaa att aac agc tgt ctc atc cac cac ggg ggc tgc cac att	4560
cac gcc gag tgc atc ccc act ggc ccc cag cag gtc tcc tgc agc	4605
tgc cgt gag ggt tac agc ggg gat ggc atc cgg acc tgc gag ctc	4650
ctg gac ccc tgc tct aag aac aat gga gga tgc agc cca tat gcc	4695

Fig. 9 (cont.)

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acc tgc aaa agc aca	ggg gat ggc cag agg	aca tgt acc tgc gac	4740
aca gcc cac acc gtg	ggg gac ggc ctc acc	tgc cgt gcc cga gtc	4785
ggc ctg gag ctc ctg	agg gat aag cat gcc	tca ttc ttc agc ctc	4830
cgc ctc ctg gaa tat	aag gag ctc aag ggc	gat ggg cct ttc acc	4875
atc ttc gtg ccg cac	gca gat cta atg agc	aac ctg tcc cag gat	4920
gag ctg gcc cgg att	cgt gcg cat cgc cag	ctg gtg ttt cgc tac	4965
cac gtg gtt ggc tgt	cgg cgg ctg cgg agc	gag gac ctg ctg gag	5010
cag ggg tac gcc acg	gcc ctc tca ggg cac	cca ctg cgc ttc agc	5055
gag agg gag ggc agc	ata tac ctc aat gac	ttc gcg cgc gtg gtg	5100
agc agc gac cat gag	gcc gtg aac ggc atc	ctg cac ttc att gac	5145
cgt gtc ctg ctg ccc	ccc gag gcg ctg cac	tgg gag cct gat gat	5190
gct ccc atc cgg agg	aga aat gtc acc gcc	gcc gcc cag ggc ttc	5235
ggg tac aag atc ttc	agc ggc ctc ctg aag	gtg gcc ggc ctc ctg	5280
ccc ctg ctt cga gag	gca tcc cat agg ccc	ttc aca atg ctg tgg	5325
ccc aca gag gcc gcc	ttt cga gct ctg cct	ccg gat cgc cag gcc	5370
tgg ctg tac cat gag	gac cac cgt gac aag	cta gca gcc att ctg	5415
cgg ggc cac atg att	cgc aat gtc gag gcc	ttg gca tct gac ctg	5460
ccc aac ctg ggc cca	ctt cga acc atg cat	ggg acc ccc atc tct	5505
ttc tcc tgc agc cga	acg cgg ccc ggt gag	ctc atg gtg ggt gag	5550
gat gat gct cgc att	gtg cag cgg cac ttg	ccc ttt gag ggt ggc	5595
ctg gcc tat ggc atc	gac cag ctg ctg gag	cca cct ggc ctt ggt	5640
gct cgc tgt gac cac	ttt gag acc cgg ccc	ctg cga ctg aac acc	5685
tgc agc atc tgt ggg	ctg gag cca ccc tgt	cct gag ggg tca cag	5730
gag cag ggc agc cct	gag gcc tgc tgg cgc	ttc tac ccg aag ttc	5775
tgg acg tcc cct ccg	ctg cac tct ttg gga	tta cgc agc gtc tgg	5820
gtc cac ccc agc ctt	tgg ggt agg ccc caa	ggc ctg ggc agg ggc	5865
tgc cac cgc aat tgt	gtc acc acc acc tgg	aag ccc agc tgc tgc	5910
cct ggt cac tat ggc	agt gag tgc caa gct	tgc cct ggc ggc ccc	5955
agc agc cct tgt agt	gac cgt ggc gtg tgc	atg gac ggc atg agt	6000
ggc agt ggg cag tgt	ctg tgc cgt tca ggt	ttt gct ggg aca gcc	6045
tgt gaa ctc tgt gct	cct ggt gcc ttt ggg	ccc cat tgt caa gcc	6090
tgc cgc tgc act gtg	cat ggc cgc tgt gat	gag ggc ctt ggg ggc	6135
tct ggc tcc tgc ttc	tgt gat gaa ggc tgg	act ggg cca cgc tgt	6180
gag gtg caa ctg gag	ctg cag cct gtg tgt	acc cca ccc tgt gca	6225
ccc gag gct gtg tgc	cgt gca ggc aac agc	tgt gag tgc agc ctg	6270
ggc tat gaa ggg gat	ggc cgc gtg tgt aca	gtg gca gac ctg tgc	6315
cag gac ggg cat ggt	ggc tgc agt gag cac	gcc aac tgt agc cag	6360
gta gga aca atg gtc	act tgt acc tgc ctg	ccc gac tac gag ggt	6405
gat ggc tgg agc tgc	cgg gcc cgc aac ccc	tgc aca gat ggc cac	6450
cgc ggg ggc tgc agc	gag cac gcc aac tgc	ttg agc acc ggc ctg	6495
aac aca cgg cgc tgt	gag tgc cac gca ggc	tac gta ggc gat gga	6540
ctg cag tgt ctg gag	gag tcc gaa cca cct	gtg gac cgc tgc ttg	6585
ggc cag cca ccg ccc	tgc cac tca gat gcc	atg tgc act gac ctg	6630
cac ttc cag gag aaa	cgg gct ggc gtt ttc	cac ctc cag gcc acc	6675
agc ggc cct tat ggt	ctg aac ttt tcc gag	gct gag gcg gca tgc	6720
gaa gca cag gga gcc	gtc ctt gct tca ttc	cct cag ctc tct gct	6765
gcc cag cag ctg ggc	ttc cac ctg tgc ctc	atg ggc tgg ctg gcc	6810
aat ggc tcc act gcc	cac cct gtg gtt ttc	cct gtg gcg gac tgt	6855
ggc aat ggt cgg gtg	ggc gta gtc agc ctg	ggt gcc cgc aag aac	6900
ctc tca gaa cgc tgg	gat gcc tac tgc ttc	cgt gtg caa gat gtg	6945
gcc tgc cga tgc cga	aat ggc ttc gtg ggt	gac ggg atc agc acg	6990

Fig. 9 (cont.)

tgc aat ggg aag ctg ctg gat gtg ctg gct gcc act gcc aac ttc	7035
tcc acc ttc tat ggg atg cta ttg ggc tat gcc aat gcc acc cag	7080
cgg ggt ctc gac ttc ctg gac ttc ctg gat gat gag ctc acg tat	7125
aag aca ctc ttc gtc cct gtc aat gaa ggc ttt gtg gac aac atg	7170
acg ctg agt ggc cca gac ttg gag ctg cat gcc tcc aac gcc acc	7215
ctc cta agt gcc aac gcc agc cag ggg aag ttg ctt ccg gcc cac	7260
tca ggc ctc agc ctc atc atc agt gac gca ggc cct gac aac agt	7305
tcc tgg gcc cct gtg gcc cca ggg aca gtt gtg gtt agc cgt atc	7350
att gtg tgg gac atc atg gcc ttc aat ggc atc atc cat gct ctg	7395
gcc agc ccc ctc ctg gca ccc cca cag ccc cag gca gtg ctg gcg	7440
cct gaa gcc cca cct gtg gcg gca ggc gtg ggg gct gtg ctt gcc	7485
gct gga gca ctg ctt ggc ttg gtg gcc gga gct ctc tac ctc cgt	7530
gcc cga ggc aag ccc acg ggc ttt ggc ttc tct gcc ttc cag gcg	7575
gaa gat gat gct gac gac gac ttc tca ccg tgg caa gaa ggg acc	7620
aac ccc acc ctg gtc tct gtc ccc aac cct gtc ttt ggc agc gac	7665
acc ttt tgt gaa ccc ttc gat gac tca ctg ctg gag gag gac ttc	7710
cct gac acc cag agg atc ctc aca gtc aag tgacgaggct ggggctgaaa	7760
gcagaagcat gcacagggag gagaccactt ttattgcttg tctgggtgga tggggcagga	7820
ggggctgagg gcctgtccca gacaataaag tgccctcagc ggatgtgggc catgtcacc	7879

Fig. 9 (cont.)

REFERENCES CITED IN THE DESCRIPTION

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专利名称(译)	常见的淋巴管内皮和血管内皮受体-1 (聪明-1) 及其用途		
公开(公告)号	EP1463760B1	公开(公告)日	2007-09-05
申请号	EP2003729265	申请日	2003-01-08
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发明人	JALKANEN, SIRPA IRJALA, HEIKKI SALMI, MARKO		
IPC分类号	C07K14/705 A61K38/17 A61K39/395 A61P29/00 A61P35/04 G01N33/50 A61K45/00 A61P35/00 A61P43/00 C07K16/28 C12N15/02 C12P21/08 C12Q1/02 G01N33/15 G01N33/53 G01N33/574		
CPC分类号	A61K2039/505 A61P29/00 A61P35/00 A61P35/04 A61P43/00 C07K14/7056 C07K16/28		
优先权	60/346288 2002-01-09 US		
其他公开文献	EP1463760A2		
外部链接	Espacenet		

摘要(译)	<pre> actctgtcct ggacagcgtg cccaccagcc atg gcg ggg ccc cgg ggc ctc ctc Met Ala Gly Pro Arg Gly Leu Leu 1 5 cca ctc tgc ctc ctg gcc ttc tgc ctg gca agc ttc agc ttc gtc agg Pro Leu Cys Leu Leu Ala Phe Cys Leu Ala Gly Phe Ser Phe Val Arg 10 15 20 ggg cag gtg ctg ttc aaa ggc tgt gat gtg aaa acc acg ttt gtc act Gly Gln Val Leu Phe Lys Gly Cys Asp Val Lys Thr Thr Phe Val Thr 25 30 35 40 cat gta ccc tgc acc tcg tgc gcg gcc atc aag aag cag acg tgt ccc His Val Pro Cys Thr Ser Cys Ala Ala Ile Lys Lys Gln Thr Cys Pro 45 50 55 tca agc tgg ctg cgg aag ctc ccg gat cag ata acc cag gac tgc cgc Ser Gly Trp Leu Arg Glu Leu Pro Asp Gln Ile Thr Gln Asp Cys Arg 60 65 70 tac gaa gta cag ctg ggg ggc tct atg gtg tcc atg agc ggc tgc aga Tyr Glu Val Gln Leu Gly Gly Ser Met Val Ser Met Ser Gly Cys Arg 75 80 85 cgg aag tgc cgg aag caa gtg gtg cag aag gcc tgc tgc cct gcc tac Arg Lys Cys Arg Lys Gln Val Val Gln Lys Ala Cys Cys Pro Gly Tyr 90 95 100 tgg agt tcc cgg tgc cat gaa tgc cct ggg ggc gct gag acc cca tgc Trp Gly Ser Arg Cys His Glu Cys Pro Gly Gly Ala Glu Thr Pro Cys 105 110 115 aat ggc cac ggg acc tgc ttg gat ggc atg gac agg aat ggg acc tgt Asn Gly His Gly Thr Cys Leu Asp Gly Met Asp Arg Asn Gly Thr Cys 120 125 gtg tgc cag gaa aac ttc cgc ggc tca gcc tgc cag gag tgc caa gac Val Cys Gln Glu Asn Phe Arg Gly Ser Ala Cys Gln Glu Cys Gln Asp 130 135 140 145 150 ccc aac cgg ttc ggg cct gac tgc caa tcg gtg tgc agc tgt gtg cac 54 </pre>	54
描述了一种新的蛋白质共同淋巴内皮和血管内皮细胞受体-1 (CLEVER-1)。CLEVER-1介导白细胞和恶性细胞与血管和淋巴内皮细胞的结合。CLEVER-1是第一种报道介导淋巴结流入和流出的蛋白质。还提供了通过提供CLEVER-1结合抑制剂来治疗炎症和预防恶性细胞转移的方法。	<pre> cca ctc tgc ctc ctg gcc ttc tgc ctg gca agc ttc agc ttc gtc agg Pro Leu Cys Leu Leu Ala Phe Cys Leu Ala Gly Phe Ser Phe Val Arg 10 15 20 ggg cag gtg ctg ttc aaa ggc tgt gat gtg aaa acc acg ttt gtc act Gly Gln Val Leu Phe Lys Gly Cys Asp Val Lys Thr Thr Phe Val Thr 25 30 35 40 cat gta ccc tgc acc tcg tgc gcg gcc atc aag aag cag acg tgt ccc His Val Pro Cys Thr Ser Cys Ala Ala Ile Lys Lys Gln Thr Cys Pro 45 50 55 tca agc tgg ctg cgg aag ctc ccg gat cag ata acc cag gac tgc cgc Ser Gly Trp Leu Arg Glu Leu Pro Asp Gln Ile Thr Gln Asp Cys Arg 60 65 70 tac gaa gta cag ctg ggg ggc tct atg gtg tcc atg agc ggc tgc aga Tyr Glu Val Gln Leu Gly Gly Ser Met Val Ser Met Ser Gly Cys Arg 75 80 85 cgg aag tgc cgg aag caa gtg gtg cag aag gcc tgc tgc cct gcc tac Arg Lys Cys Arg Lys Gln Val Val Gln Lys Ala Cys Cys Pro Gly Tyr 90 95 100 tgg agt tcc cgg tgc cat gaa tgc cct ggg ggc gct gag acc cca tgc Trp Gly Ser Arg Cys His Glu Cys Pro Gly Gly Ala Glu Thr Pro Cys 105 110 115 aat ggc cac ggg acc tgc ttg gat ggc atg gac agg aat ggg acc tgt Asn Gly His Gly Thr Cys Leu Asp Gly Met Asp Arg Asn Gly Thr Cys 120 125 gtg tgc cag gaa aac ttc cgc ggc tca gcc tgc cag gag tgc caa gac Val Cys Gln Glu Asn Phe Arg Gly Ser Ala Cys Gln Glu Cys Gln Asp 130 135 140 145 150 ccc aac cgg ttc ggg cct gac tgc caa tcg gtg tgc agc tgt gtg cac 534 </pre>	102
		150
		198
		246
		294
		342
		390
		438
		486
		534