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(54) Title: LYMPHATIC AND BLOOD ENDOTHELIAL CELL GENES

(57) Abstract: The invention provides polynucleotides and genes that are differentially expressed in lymphatic versus blood vascular endothelial cells. These genes are useful for treating diseases involving lymphatic vessels, such as lymphedema, various inflammatory diseases, and cancer metastasis via the lymphatic system.

LYMPHATIC AND BLOOD ENDOTHELIAL CELL GENES

BACKGROUND OF THE INVENTION

Field of the Invention

The invention relates to polynucleotides and proteins specifically expressed in lymphatic endothelial cells.

Description of the Related Art

Recent evidence on the association of lymphangiogenic growth factors with intralymphatic growth and metastasis of cancers (Mandriota, et al., *EMBO J. 20*:672-682 (2001); Skobe, et al., *Nat. Med. 7*:192-198 (2001); Stacker, et al., *Nat. Med. 7*:186-191 (2001); Karpanen, et al., *Cancer Res. 61*:1786-1790 (2001)) has raised hopes that lymphatic vessels could be used as an additional target for tumor therapy. Cancer cells spread within the body by direct invasion to surrounding tissues, spreading to body cavities, invasion into the blood vascular system (hematogenous metastasis), as well as spread via the lymphatic system (lymphatic metastasis). Regional lymph node dissemination is the first step in the metastasis of several common cancers and correlates highly with the prognosis of the disease. The lymph nodes that are involved in draining tissue fluid from the tumor area are called sentinel nodes, and diagnostic measures are in place to find these nodes and to remove them in cases of suspected metastasis. However, in spite of its clinical relevance, little is known about the mechanisms leading to metastasis via the bloodstream or via the lymphatics.

Until recently, the lymphatic vessels have received much less attention than blood vessels, despite their importance in medicine. Lymphatic vessels collect protein-rich fluid and white blood cells from the interstitial space of most tissues and transport them as a whitish opaque fluid, the lymph, into the blood circulation. Small lymphatic vessels coalesce into larger vessels, which drain the lymph through the thoracic duct into large veins in the neck region. Lymph nodes serve as filtering stations along the lymphatic vessels and lymph movement is propelled by the

contraction of smooth muscles surrounding collecting lymphatic vessels and by bodily movements, the direction of flow being secured by valves as it is in veins. The lymphatic capillaries are lined by endothelial cells, which have distinct junctions with frequent large interendothelial gaps. The lymphatic capillaries also lack a continuous basement membrane, and are devoid of pericytes. Anchoring filaments connect the abluminal surfaces of lymphatic endothelial cells to the perivascular extracellular matrix and pull to maintain vessel patency in the presence of tissue edema. The absence or obstruction of lymphatic vessels, which is usually the result of an infection, surgery, or radiotherapy and in rare cases, a genetic defect, causes accumulation of a protein-rich fluid in tissues, lymphedema. The lymphatic system is also critical in fat absorption from the gut and in immune responses. Bacteria, viruses, and other foreign materials are taken up by the lymphatic vessels and transported to the lymph nodes, where the foreign material is presented to immune cells and where dendritic cells traverse via the lymphatics. There has been slow progress in the understanding of and ability to manipulate the lymphatic vessels.

Abnormal development or function of the lymphatic ECs can result in tumors or malformations of the lymphatic vessels, such as lymphangiomas or lymphangiectasis. Witte, et al., *Regulation of Angiogenesis* (eds. Goldber, I.D. & Rosen, E.M.) 65-112 (Birkäuser, Basel, Switzerland, 1997). The VEGFR-3 tyrosine kinase receptor is expressed in the normal lymphatic endothelium and is upregulated in many types of vascular tumors, including Kaposi's sarcomas. Jussila, et al., *Cancer Res 58*, 1955-1604 (1998); Partanen, et al., *Cancer 86*:2406-2412 (1999). Absence or dysfunction of lymphatic vessels which can result from an infection, surgery, radiotherapy or from a genetic defect, causes lymphedema, which is characterized by a chronic accumulation of protein-rich fluid in the tissues that leads to swelling. The importance of VEGFR-3 signaling for lymphangiogenesis was revealed in the genetics of familial lymphedema, a disease characterized by a hypoplasia of cutaneous lymphatic vessels, which leads to a disfiguring and disabling swelling of the extremities. Witte, et al., *Regulation of Angiogenesis* (eds. Goldber, I.D. & Rosen, E.M.) 65-112 (Birkäuser, Basel, Switzerland, 1997); Rockson, S.G, *Am. J. Med. 110*, 288-295 (2001). Some members of families with lymphoedema are heterozygous for

missense mutations of the *VEGFR3* exons encoding the tyrosine kinase domain, which results in an inactive receptor protein. Karkkainen, et al., *Nature Genet. 25*:153-159 (2000); Irrthum, et al., *Am. J. Hum. Genet. 67*:295-301 (2000).

There is a need in the art for information on the transcriptional program which controls the diversity of endothelial cells, and into the mechanisms of angiogenesis and lymphangiogenesis. There is also a need in the art for new vascular markers, which may be used as valuable targets in the study of a number of diseases involving the lymphatic vessels, including tumor metastasis.

SUMMARY OF THE INVENTION

[TO BE REVISED UPON FINALIZATION OF CLAIMS]

The compositions of the present invention include isolated polynucleotides, in particular, lymphatic endothelial genes, polypeptides, isolated polypeptides encoded by these polynucleotides, recombinant DNA molecules, cloned genes or degenerate variants thereof, especially naturally occurring variants such as allelic variants, and antibodies that specifically recognize one or more epitopes present on such polypeptides.

The compositions of the present invention additionally include vectors, including expression vectors, containing the polynucleotides of the invention, cells genetically engineered to contain such polynucleotides and cells genetically engineered to express such polynucleotides.

In selected embodiments, such isolated polynucleotides of the invention represent a polynucleotide comprising a nucleotide sequence set forth in the sequence listing, e.g., any of SEQ ID NOS:1-30.

The polynucleotides of the present invention also include, but are not limited to, a polynucleotide that hybridizes to the complement of the nucleotide sequence of SEQ ID NOS:1-30 under highly stringent hybridization conditions; a polynucleotide that hybridizes to the complement of the nucleotide sequence of SEQ ID NOS:1-30 under moderately stringent hybridization conditions; a polynucleotide which is an allelic variant of any polynucleotide recited above; a polynucleotide which encodes a species homologue of any of the proteins recited above; of a

polynucleotide that encodes a polypeptide comprising a specific domain or truncation of the polypeptide encoded by any one of SEQ ID NOS:1-30. Exemplary high stringency hybridization conditions are hybridization at 42°C for 20 hours in a solution containing 50% formamide, 5xSSPE, 5x Denhardt's solution, 0.1% SDS and 0.1 mg/ml denatured salmon sperm DNA, with a wash in 1xSSC, 0.1% SDS for 30 minutes at 65°C.

Another aspect of the invention is drawn to LEC and BEC polypeptides, including polypeptides encoded by the polynucleotides described above. In some embodiments, the polypeptides are the mature forms of the polypeptides of the invention. Expressly contemplated is a purified and isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391; and a purified and isolated polypeptide comprising an amino acid sequence selected from the group consisting of: (a) SEQ ID NOS: 31-34, 46, 48, 207, 676, 859, and 861; and (b) an extracellular domain fragment of at least 10 amino acids of an amino acid sequence of (a). Further, this aspect of the invention includes a purified and isolated, soluble polypeptide as described immediately above, comprising an extracellular domain fragment of an amino acid sequence selected from the group consisting of : SEQ ID NOS: 31-34, 46, 48, 207, 676, 859, and 861, wherein the polypeptide lacks any transmembrane domain. Such a polypeptide may further lack any intracellular domain. Also, the invention contemplates a fusion protein comprising a polypeptide as described above fused to an immunoglobulin fragment comprising an immunoglobulin constant region.

In a related aspect, the invention provides a composition comprising a polypeptide or protein as described above and a pharmaceutically acceptable diluent, carrier or adjuvant. Polypeptide compositions of the invention may comprise an acceptable carrier, such as a hydrophilic, *e.g.*, pharmaceutically acceptable, carrier. Further provided is a kit comprising such a composition and a protocol for administering the pharmaceutical composition to a mammalian subject to modulate the lymphatic system in the subject. The invention also provides an antibody that specifically binds to a polypeptide as described above, and that antibody is humanized

in some embodiments. Still further, the invention provides a protein comprising an antigen binding domain of an antibody that specifically binds to a polypeptide as described hereinabove, wherein the protein specifically binds to the polypeptide.

The invention also relates to methods for producing a polypeptide comprising growing a culture of the cells of the invention in a suitable culture medium, and purifying the protein from the culture or from an extract of the cells. In particular, the invention contemplates a method for producing a LEC polypeptide comprising steps of growing a host cell transformed or transfected with an expression vector as described herein under conditions in which the cell expresses the polypeptide encoded by the polynucleotide.

Methods of identifying the products and compositions described herein are also provided by the invention. In particular, the invention provides a method of identifying a LEC nucleic acid comprising: (a) contacting a biological sample containing a candidate LEC nucleic acid with a polynucleotide comprising a fragment of at least 14 contiguous nucleotides of a sequence selected from the group consisting of SEQ ID NOS:1-30, 45, 47, 49, 51, 82, 93, 111, 188, 208, 212, 236, 242, 294, and 392, or a complement thereof, under the following stringent hybridization conditions: (i) hybridization at 42°C for 20 hours in a solution containing 50% formamide, 5xSSPE, 5x Denhardt's solution, 0.1% SDS and 0.1 mg/ml denatured salmon sperm DNA, and (ii) washing for 30 minutes at 65°C in 1xSSC, 0.1% SDS; and (b) detecting hybridization of the candidate LEC nucleic acid and the polynucleotide, thereby identifying a LEC nucleic acid.

The invention also provides a method of identifying a LEC protein comprising: (a) contacting a biological sample containing a candidate LEC protein with a LEC protein binding partner selected from the group consisting of an antibody as described herein or a protein or polypeptide as described herein, under conditions suitable for binding therebetween; and (b) detecting binding between the candidate LEC protein and the LEC binding partner, thereby identifying a LEC protein.

Another related aspect of the invention is a method of identifying a LEC comprising: (a) contacting a biological sample comprising cells with a LEC binding partner under conditions suitable for binding therebetween, wherein the LEC

binding partner comprises an antibody that binds to a polypeptide comprising a sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 48, 207, 676, 859, and 861, or comprises an antigen binding fragment of the antibody; and (b) identifying a LEC by detecting binding between a cell and the LEC binding partner, where binding of the LEC binding partner to the cell identifies a LEC.

Polynucleotides according to the invention have numerous applications in a variety of techniques known to those skilled in the art of molecular biology. These techniques include use as hybridization probes, use as primers for PCR, use for chromosome and gene mapping, use in the recombinant production of protein, and use in generation of anti-sense DNA or RNA, their chemical analogs and the like. For example, when the expression of an mRNA is largely restricted to a particular cell or tissue type, such as a lymphatic endothelial cell, polynucleotides of the invention can be used as hybridization probes to detect the presence of the particular cell or tissue mRNA in a sample using, *e.g.*, *in situ* hybridization.

In another aspect, the invention provides a composition comprising an isolated polynucleotide that comprises a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391; and a pharmaceutically acceptable diluent, carrier or adjuvant. In some embodiments, the composition comprises a polynucleotide that comprises a nucleotide sequence selected from the group consisting of SEQ ID NOS: 14-30, 45, 47, 49, 51, 82, 93, 111, 188, 208, 212, 222, 236, 242, 294, and 392, or a fragment thereof that encodes the polypeptide.

Still another aspect of the invention is an expression vector comprising an expression control sequence operably linked to a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391. In some embodiments, the expression vector is a replication-deficient adenoviral or adeno-associated viral vector containing the polynucleotide. A related aspect of the invention is a composition comprising an expression vector as described above and a pharmaceutically acceptable diluent,

carrier, or adjuvant. Further, the invention provides a kit comprising the composition containing either the above-described polynucleotide or vector and a pharmaceutically acceptable diluent, carrier or adjuvant, packaged with a protocol for administering the composition to a mammalian subject to modulate the lymphatic system in the subject.

The invention further provides a host cell transformed or transfected with an expression vector as described above.

The polypeptides according to the invention can be used in a variety of conventional procedures and methods that are currently applied to other proteins. In addition, a polypeptide of the invention can be used to generate an antibody that specifically binds the polypeptide.

In one aspect of the invention, a method is provided for differentially modulating the growth or differentiation of blood endothelial cells (BEC) or lymphatic endothelial cells (LEC), comprising contacting endothelial cells with a composition comprising an agent that differentially modulates blood or lymphatic endothelial cells, said agent selected from the group consisting of: (a) a polypeptide that comprises an amino acid sequence of a BEC polypeptide or a LEC polypeptide, or an active fragment of the polypeptide ; (b) a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide according to (a); (c) an antibody that specifically binds to a polypeptide according to (a); (d) a polypeptide comprising a fragment of the antibody of (c), wherein the fragment and the antibody bind to the polypeptide; (e) an antisense nucleic acid to a human gene or mRNA encoding the polypeptide of (a); (f) an interfering RNA (RNAi) to a human gene or mRNA encoding the polypeptide of (a). The method may involve endothelial cell contact with the composition *ex vivo* or *in vivo*. The composition may comprise a pharmaceutically acceptable diluent, adjuvant, or carrier, and the contacting step may comprise administering the composition to a mammalian subject to differentially modulate BECs or LECs in the mammalian subject.

Further, the method may comprise identifying a human subject with a disorder characterized by hyperproliferation of LECs; and administering to the human subject the composition, wherein the agent differentially inhibits LEC growth compared to BEC growth; alternatively the method may comprise identifying a

human subject with a disorder characterized by hyperproliferation of LECs; screening LECs of the subject to identify overexpression of a polypeptide set forth in Table 3; and administering to the human subject the composition, wherein the agent differentially inhibits LEC growth compared to BEC growth by inhibiting expression of the polypeptide identified by the screening step.

This aspect of the invention also contemplates a method of modulating the growth of lymphatic endothelial cells in a human subject, comprising steps of identifying a human subject with a hypoproliferative lymphatic disorder; screening the subject to identify underexpression or underactivity of a LEC polypeptide set forth in Table 3, wherein the protein is not set forth in Table 1 or 2; administering to the human subject the composition, wherein the agent comprises the LEC polypeptide (a) identified by the screening step or an active fragment of the polypeptide, or comprises the polynucleotide (b) that comprises a nucleotide sequence that encodes the polypeptide.

A related aspect of the invention is drawn to a use of an agent for the manufacture of a medicament for the differential modulation of blood vessel endothelial cell (BEC) or lymphatic vessel endothelial cell (LEC) growth or differentiation, the agent selected from the group consisting of: (a) a polypeptide that comprises an amino acid sequence of a BEC polypeptide or a LEC polypeptide, or an active fragment of the polypeptide ; (b) a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide according to (a); (c) an antibody that specifically binds to a polypeptide according to (a); (d) a polypeptide comprising a fragment of the antibody of (c), wherein the fragment and the antibody bind to the polypeptide; (e) an antisense nucleic acid to a human gene or mRNA encoding the polypeptide of (a); (f) an interfering RNA (RNAi) to a human gene or mRNA encoding the polypeptide of (a).

In another aspect, the invention provides a method of identifying compounds that modulate growth of endothelial cells, comprising culturing endothelial cells in the presence and absence of a compound; and measuring expression of at least one BEC or LEC gene in the cells, wherein the BEC or LEC gene is selected from the genes encoding polypeptides set forth in Tables 3 and 4,

wherein a change in expression of at least one BEC gene in the presence compared to the absence of the compound identifies the compound as a modulator of BEC growth, and wherein a change in expression of at least one LEC gene in the presence compared to the absence of the compound identifies the compound as a modulator of LEC growth. The method may be used to screen for a compound that selectively modulates BEC or LEC growth or differentiaion, wherein the measuring step comprises measuring expression of at least one BEC gene and at least one LEC gene in the cells, and wherein the method comprises screening for a compound that selectively modulates BEC or LEC growth or differentiation by selecting a compound that differentially modulates expression of the at least one BEC gene compared to expression of the at least one LEC gene.

Further, the invention comprehends a method or use according to the aspects of the invention described above, wherein the polypeptide is a LEC polypeptide selected from the LEC polypeptides set forth in Table 3, and the agent differentially modulates LEC growth or differentiation over BEC growth or differentiation. In some embodiments, the LEC polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 81, 187, 207, 211, 221, 235, 241, 293, and 391; in other embodiments, the LEC polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-34, 46, and 48. In these embodiments, an agent may be an antibody that specifically binds to a LEC polypeptide as described above, or a polypeptide fragment of such an antibody. Further, the agent may be an extracellular domain of a polypeptide described above, a polynucleotide encoding an extracellular domain, or an antisense molecule or nucleic acid. Alternatively, the polypeptide is a BEC polypeptide selected from the BEC polypeptides set forth in Table 4, and the agent differentially modulates BEC growth or differentiation over LEC growth or differentiation. Preferably, the polypeptides are not set forth in Tables 1 or 2.

The methods of the present invention further relate to methods for detecting the presence of the polynucleotides or polypeptides of the invention in a sample. Such methods can, for example, be utilized as part of prognostic and diagnostic evaluation of disorders as recited above and for the identification of

subjects exhibiting a predisposition to such conditions. Furthermore, the invention provides methods for evaluating the efficacy of drugs, and monitoring the progress of patients, involved in clinical trials for the treatment of disorders related to lymphatic endothelial cells.

The invention also provides methods for the identification of compounds that modulate the expression of the polynucleotides and/or polypeptides of the invention. Such methods can be utilized, for example, for the identification of compounds that can ameliorate symptoms of disorders related to expression of proteins encoded by any one of SEQ ID NOS:1-30 as recited above. Such methods can include, but are not limited to, assays for identifying compounds and other substances that interact with (*e.g.*, bind to) the polypeptides of the invention.

Further, the invention provides a method of assaying for risk of developing hereditary lymphedema, comprising (a) assaying nucleic acid of a human subject for a mutation that correlates with a hereditary lymphedema phenotype and alters the encoded amino acid sequence of at least one gene allele of the human subject when compared to the amino acid sequence of the polypeptide encoded by a corresponding wild-type gene allele, wherein the wild-type polypeptide is a polypeptide identified in Table 3. Alternatively, a method of assaying for risk of developing hereditary lymphedema, comprises (a) assaying nucleic acid of a human subject for a mutation that correlates with a hereditary lymphedema phenotype and alters the encoded amino acid sequence of at least one gene allele of the human subject when compared to the amino acid sequence of the polypeptide encoded by a corresponding wild-type gene allele, wherein the wild-type polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 52, 54, 207, 676, 859, and 861; (b) correlating the presence or absence of the mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of the mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of the mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema.

In another method of assaying for risk of developing hereditary lymphedema, the steps comprise (a) assaying nucleic acid of a human subject for a

mutation that alters the encoded amino acid sequence of at least one transcription factor allele of the human subject and alters transcription modulation activity of the transcription factor polypeptide encoded by the allele, when compared to the transcription modulation activity of a transcription factor polypeptide encoded by a wild-type allele, wherein the wild-type transcription factor polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 211, SEQ ID NO: 241, and transcription factors encoded by sequences in Table 5; and (b) correlating the presence or absence of the mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of the mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of the mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema. In this method, the wild-type transcription factor allele may comprise the Sox18 amino acid sequence set forth as SEQ ID NO:54. In some embodiments of this method, the assaying identifies a mutation altering a transactivating or DNA binding domain amino acid sequence of the protein encoded by the Sox18 allele; in some other embodiments of the method, the mutation reduces transcriptional activation of a SOX18-responsive gene compared to transcriptional activation of the gene by wild-type SOX18.

In a related aspect, the invention provides a method of assaying for risk of developing hereditary lymphedema, comprising (a) assaying nucleic acid of a human subject for a mutation that alters the encoded amino acid sequence of at least one LEC gene allele of the human subject and alters the binding affinity of the adhesion polypeptide encoded by the LEC gene allele, when compared to the binding affinity of an adhesion polypeptide encoded by a wild-type allele, wherein the wild-type adhesion polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 207, 676, 859, and 861; and (b) correlating the presence or absence of the mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of the mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of the mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema. In some embodiments of this method, the at least

one gene corresponds to the human Sox18 gene that encodes the amino acid sequence set forth in SEQ ID NO: 54.

In the methods of assaying for risk of developing hereditary lymphedema according to the invention, the assaying may identify the presence of the mutation, and the correlating step may identify the increased risk of the patient developing hereditary lymphedema.

A related method according to the invention is a method of screening a human subject for an increased risk of developing hereditary lymphedema comprising assaying nucleic acid of a human subject for a mutation that alters the encoded amino acid sequence of at least one polypeptide comprising an amino acid sequence of Table 3. In some embodiments of this method, the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, and 54, 207, 676, 859, and 861 in a manner that correlates with the risk of developing hereditary lymphedema, and it is expressly contemplated that the polypeptide may comprise the SOX18 amino acid sequence set forth in SEQ ID NO: 54.

A related aspect of the invention is drawn to methods of assaying or screening for risk of developing hereditary lymphedema as described above, wherein the method comprises at least one procedure selected from the group consisting of: (a) determining a nucleotide sequence of at least one codon of at least one polynucleotide of the human subject; (b) performing a hybridization assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; (c) performing a polynucleotide migration assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; and (d) performing a restriction endonuclease digestion to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences.

A related aspect of the invention provides methods of assaying or screening for risk of developing hereditary lymphedema as described above, wherein the method comprises: performing a polymerase chain reaction (PCR) to amplify

nucleic acid comprising the coding sequence of the LEC polynucleotide, and determining nucleotide sequence of the amplified nucleic acid.

Further provided by the invention is a method of screening for a hereditary lymphedema genotype in a human subject, comprising: (a) providing a biological sample comprising nucleic acid from said subject, and (b) analyzing the nucleic acid for the presence of a mutation altering the encoded amino acid sequence of the at least one allele of at least one gene in the human subject relative to a human gene encoding an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 54, 207, 676, 859, and 861, wherein the presence of a mutation altering the encoded amino acid sequence in the human subject in a manner that correlates with lymphedema in human subjects identifies a hereditary lymphedema genotype. In some embodiments of this method, the biological sample is a cell sample. In other embodiments of this method, the analyzing comprises sequencing a portion of the nucleic acid. In still further embodiments of this method, the human subject has a hereditary lymphedema genotype identified by the method of screening.

Another aspect of the invention provides a method of inhibiting lymphangiogenesis comprising administering to a subject an inhibitor of a LEC transmembrane polypeptide, wherein the LEC transmembrane polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-34, 46 48, 207, 676, 859, and 861, and wherein the inhibitor is selected from the group consisting of (a) a soluble extracellular domain fragment of the LEC transmembrane polypeptide; (b) an antibody that binds to the extracellular domain of the LEC transmembrane polypeptide; (c) a polypeptide comprising an antigen binding domain of the antibody according to (b); and (d) an antisense nucleic acid complementary to the nucleic acid encoding the LEC transmembrane polypeptide or its complement. In some embodiments of the method, the inhibitor is a polypeptide comprising an extracellular domain fragment of an LEC polypeptide, wherein the sequence of the extracellular domain is selected from the group consisting of amino acids 1-152 of SEQ ID NO:31, amino acids 1-695 of SEQ ID NO:32 and amino acids 1-248 of SEQ

ID NO:33. In some embodiments of the method, the subject is a human containing a tumor.

In a related aspect, the invention provides a method for modulating lymphangiogenesis in a mammalian subject comprising: administering to a mammalian subject in need of modulation of lymphangiogenesis an antisense molecule to a LEC polynucleotide, in an amount effective to inhibit transcription or translation of the poypeptide encoded by the LEC polynucleotide, wherein the LEC polynucleotide comprises a nucleotide sequence selected from the group consisting of SEQ ID NOS: 14-30, 45, 47, 49, AND 51, 208, 677, 860, and 862.

The methods of the invention also include methods for the treatment of disorders related to lymphatic endothelial cells as recited above which may involve the administration of such compounds to individuals exhibiting symptoms or tendencies related to such disorders.

In another aspect, the invention provides a method of treating hereditary lymphedema, comprising: (a) identifying a human subject with hereditary lymphedema and with a mutation that alters the encoded amino acid sequence of at least one polypeptide of the human subject, relative to the amino acid sequence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, 54, 207, 676, 859, and 861; and (b) administering to the subject a lymphatic growth factor selected from the group consisting of a VEGF-C polynucleotide, a VEGF-C polypeptide, a VEGF-D polynucleotide, and a VEGF-D polypeptide.

The invention also provides a method of treating hereditary lymphedema comprising: identifying a human subject with lymphedema and with a mutation in at least one allele of a gene encoding a LEC protein identified in Table 3, wherein the mutation correlates with lymphedema in human subjects, and with the proviso that the LEC protein is not VEGFR-3; and administering to the subject a composition comprising a lymphatic growth agent selected from the group consisting of VEGF-C polypeptides, VEGF-D polypeptides, VEGF-C polynucleotides, and VEGF-D polynucleotides. The invention also comprehends use of a lymphatic growth agent selected from the group consisting of VEGF-C polypeptides, VEGF-D

polypeptides, VEGF-C polynucleotides, and VEGF-D polynucleotides in the manufacture of a medicament for the treatment of hereditary lymphedema resulting from a mutation in a LEC gene identified in Table 3, with the proviso that the gene is not *VEGFR-3*.

In addition, the invention encompasses methods for treating such diseases or disorders by administering compounds and other substances that modulate the overall activity of the target gene products. Compounds and other substances can effect such modulation either at the level of target gene expression or target protein activity. These treatment methods include the administration of a polypeptide or a polynucleotide according to the invention to an endothelial cell, e.g., a LEC and/or a BEC, or to an organism such as a human patient. An exemplary method according to this aspect of the invention is the administration of a therapeutic selected from the group consisting of an antisense polynucleotide capable of modulating the expression of at least one polynucleotide according to the invention, a polypeptide according to the invention, a polynucleotide according to the invention, an antibody or antibody fragment specifically recognizing a polypeptide according to the invention, a VEGF-C polynucleotide, a VEGF-C polypeptide, a VEGF-D polynucleotide, a VEGF-D polypeptide and a soluble VEGFR-3 polypeptide.

In another aspect, the invention provides a method of screening for an endothelial cell disorder or predisposition to the disorder, comprising obtaining a biological sample containing endothelial cell mRNA from a human subject; and measuring expression of a BEC or LEC gene from the amount of mRNA in the sample transcribed from the gene, wherein the BEC or LEC gene encodes a polypeptide identified in Table 3 or 4.

The invention relates to a method of inhibiting the growth of a lymphatic endothelial cell, the method comprising contacting the cell with a composition comprising at least one antibody conjugated to an agent capable of inhibiting the growth, wherein the agent is selected from the group consisting of a cytotoxic agent and a cytostatic agent, and wherein the antibody specifically binds to a polypeptide encoded by a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NOS:14-17, 45, 47, 860 and 862. In specific

embodiments of this method, the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 48, 859 and 861.

The invention further relates to methods of detecting a lymphatic endothelial cell, the method comprising contacting the cell with a composition comprising at least one antibody conjugated to a detectable agent, such as a fluorescent molecule or a radiolabeled molecule. In specific embodiments, the antibody specifically binds to a polypeptide encoded by a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NOS: 14-17, 45, 47, 860 and 862. In further specific embodiments of this method, the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-34, 46, 48, 859 and 861.

The invention still further relates to methods of isolating a lymphatic endothelial cell, comprising contacting the cell with a solid matrix comprising at least one antibody capable of binding to a transmembrane protein in the cell membrane of the cell, and isolating cells specifically bound to the antibody matrix. In specific embodiments, the antibody specifically binds to a polypeptide encoded by a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NOS:14-17, 45, 47, 860 and 862. In further specific embodiments of this method, the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 48, 859 and 861.

The invention also relates to the administration of an agonist or antagonist to a lymphatic endothelial cell, comprising selecting an antibody, a peptide or a small molecular weight compound that is capable of specifically binding to a lymphatic endothelial cell-specific protein, wherein the antibody, peptide or small molecular weight compound is an agonist or antagonist for a growth factor receptor, a cytokine receptor, a chemokine receptor, or a hemopoietic receptor, and contacting the antibody, peptide or small molecular weight compound with the lymphatic endothelial cell in need of growth stimulation or inhibition. In specific embodiments, such lymphatic endothelial cells are involved in lymphedema, lymphangioma, lymphangiomyeloma, lymphangiomatosis, lymphangiectasis, lymphosarcoma, and lymphangiosclerosis.

The invention also relates to the administration of a cytotoxic or cytostatic drug to a lymphatic endothelial cell, comprising selecting an antibody, a peptide or a small molecular weight compound that is capable of specifically binding to a lymphatic endothelial cell-specific protein, wherein the antibody, peptide or small molecular weight compound is complexed to the cytotoxic or cytostatic drug. In specific embodiments, administration of such complexes is useful in the treatment of malignant tumor diseases prone to metastatic spread through the lymphatic system.

The invention also provides a method of monitoring the efficacy or toxicity of a drug on endothelial cells, comprising steps of measuring expression of at least one BEC or LEC gene in endothelial cells of a mammalian subject before and after administering a drug to the subject, wherein the at least one BEC or LEC gene encodes a polypeptide set forth in Table 3 or Table 4, and wherein changes in expression of the BEC or LEC gene correlates with efficacy or toxicity of the drug on endothelial cells.

The invention relates to a lymphatic endothelial cell marker protein comprising a polypeptide encoded by a polynucleotide selected from the group consisting of SEQ ID NOS:14-17; and a polynucleotide hybridizable under stringent conditions with any one of SEQ ID NOS:14-17. In specific embodiments, the lymphatic endothelial cell marker protein comprises a polypeptide selected from the group consisting of SEQ ID NOS:31-34.

The invention also relates to an antibody capable of specifically binding to a lymphatic endothelial cell marker protein comprising a polypeptide selected from the group consisting of SEQ ID NOS:31-34.

The invention further relates to a method of detecting a lymphatic endothelial cell, comprising contacting said cell with the antibody wherein said antibody is detectably labeled.

The invention still further relates to a method of inhibiting at least one biological activity of a lymphatic endothelial cell, comprising contacting the cell with an agent capable of binding to at least one polypeptide encoded by any one of SEQ ID NOS:14-17, 45, 47, 860 and 862, wherein the activity of the polypeptide is reduced relative to the activity of a polypeptide that is not contacted with the agent.

The invention also relates to a method of inhibiting the growth of a lymphatic endothelial cell, the method comprising contacting the cell with an antisense oligonucleotide capable of specifically binding to at least one polynucleotide selected from the group consisting of SEQ ID NOS:1-30, 45, 47, 860 and 862. In a specific embodiment, the antisense oligonucleotide consists essentially of about 12 to about 25 contiguous nucleotides of any one of SEQ ID NOS: 1-30, 45, 47, 860 and 862.

Additional features and variations of the invention will be apparent to those skilled in the art from the entirety of this application, and all such features are intended as aspects of the invention.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

Figure 1: Examples of differentially expressed genes in LECs and BECs. Northern blotting and hybridization for the indicated transcripts. Equal loading was verified by probing with GAPDH. For the microarray analyses, RNA was extracted from LECs which were cultured in the presence of VEGF-C (LEC/+C). When validating the array results, RNA was extracted as a control also from cultures of LECs in which VEGF-C was not added (LEC/-C).

Figure 2: Cytoskeletal structures, cadherin complexes and integrin α9 expression in BECs and LECs. Mixed cultures of LEC and BEC were double-stained for N-cadherin (a), VE-cadherin (c), ß-catenin (e), plakoglobin (g), F-actin (i) and integrin α9 (k), and for the LEC-specific marker podoplanin (green; b, d, f, h, j, l). Expression of integrin α9 in the lymphatic (arrow) but not in blood vessel endothelia (arrowhead). Adjacent sections of human skin were stained with antibodies against integrin α9 (m), VEGFR-3 (n) or blood vessel endothelial antigen PAL-E (o).

DETAILED DESCRIPTION OF THE INVENTION

A major role of the lymphatic vasculature is to remove an excess of the protein-rich interstitial fluid that continuously escapes from the blood capillaries, and to return it to the blood circulation (Witte, M.H., et al., *Microsc. Res. Tech.* 55:122-

145. 2001; Karpanen, T., et al., *J. Exp. Med. 194*:F37-F42. 2001; Karkkainen, M.J., *et al., Trends Mol. Med.* 7:18-22. 2001). In addition, the lymphatic system provides constant immune surveillance by filtering lymph and its antigens through the chain of lymph nodes, and also serves as one of the major routes for absorption of lipids from the gut. It has been known for a long time that in many types of cancer the lymphatic vessels provide a major pathway for tumor metastasis, and regional lymph node dissemination correlates with the progression of the disease. Hereditary lymphedema, post-surgical secondary lymphedema and lymphatic obstruction in filariasis, are all characterized by disabling and disfiguring swelling of the affected areas, linked to the insufficiency or obstruction of the lymphatics. Witte, M.J., et al., *Microsc. Res. Tech 55*:122-145 (2001).

In spite of the importance of lymphatic vessels in medicine, the cell biology of this part of the vascular system has received little attention until recently. Studies during the past four years have uncovered the existence of the lymphatic specific vascular endothelial growth factors VEGF-C and VEGF-D, which serve as ligands for the receptor tyrosine kinase VEGFR-3, and demonstrated their importance for the normal development of the lymphatic vessels (*See*, Jeltsch, M., *et al., Science 276*:1423-1425 (1997); Veikkola, T., *et al., EMBO J. 20*:1223-1231 (2001); Mäkinen, T., *et al., Nat. Med. 7*:199-205 (2001)). These molecules also appear to be involved in the development of lymphedema and lymphatic metastasis (Karpanen, T., *et al., J. Exp. Med. 194*:F37-F42 (2001); Karkkainen, M.J., et al., *Trends Mol. Med.* 7:18-22. 2001).

The growth factor Vascular Endothelial Growth Factor C (VEGF-C), as well as native human, non-human mammalian, and avian polynucleotide sequences encoding VEGF-C, and VEGF-C variants and analogs, have been described in detail in International Patent Application Number PCT/US98/01973, filed February 2, 1998 and published on August 6, 1998 as International Publication Number WO 98/33917; in Joukov *et al., J. Biol. Chem.*, 273(12): 6599-6602 (1998); and in Joukov *et al., EMBO J.*, 16(13): 3898-3911 (1997), all of which are incorporated herein by reference in their entirety. As explained therein in detail, human VEGF-C (SEQ ID NO: 863) is initially produced in human cells as a prepro-VEGF-C polypeptide of 419

amino acids. A cDNA encoding human VEGF-C (SEQ ID NO: 864) has been deposited with the American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, VA 20110-2209 (USA), pursuant to the provisions of the Budapest Treaty (Deposit date of 24 July 1995 and ATCC Accession Number 97231). VEGF-C sequences from other species also have been reported. See Genbank Accession Nos. MMU73620 (*Mus musculus*); and CCY15837 (*Coturnix coturnix*) for example, incorporated herein by reference.

The prepro-VEGF-C polypeptide is processed in multiple stages to produce a mature and most active VEGF-C polypeptide of about 21-23 kD, as assessed by SDS-PAGE under reducing conditions (SEQ ID NO: 863). Such processing includes cleavage of a signal peptide (residues 1-31); cleavage of a carboxyl-terminal peptide (corresponding approximately to amino acids 228-419 and having a pattern of spaced Cysteine residues reminiscent of a Balbiani ring 3 protein (BR3P) sequence [Dignam *et al., Gene*, 88:133-40 (1990); Paulsson *et al., J. Mol. Biol.*, 211:331-49 (1990)]) to produce a partially-processed form of about 29 kD; and cleavage (apparently extracellularly) of an amino-terminal peptide (corresponding approximately to amino acids 32-103) to produced a fully-processed mature form of about 21-23 kD. Experimental evidence demonstrates that partially-processed forms of VEGF-C (e.g., the 29 kD form) are able to bind VEGFR-3 (Flt4 receptor), whereas high affinity binding to VEGFR-2 occurs only with the fully processed forms of VEGF-C. It appears that VEGF-C polypeptides naturally associate as non-disulfide linked dimers.

It has been demonstrated that amino acids 103-227 of VEGF-C are not all critical for maintaining VEGF-C functions. A polypeptide consisting of amino acids 113-213 (and lacking residues 103-112 and 214-227) retains the ability to bind and stimulate VEGF-C receptors, and it is expected that a polypeptide spanning from about residue 131 to about residue 211 will retain VEGF-C biological activity. The Cysteine residue at position 156 has been shown to be important for VEGFR-2 binding ability. However, VEGF-C ΔC_{156} polypeptides (i.e., analogs that lack this Cysteine due to deletion or substitution) remain potent activators of VEGFR-3. The Cysteine at position 165 of VEGF-C polypeptide is essential for binding either

receptor, whereas analogs lacking the Cysteine at positions 83 or 137 compete with native VEGF-C for binding with both receptors and stimulate both receptors.

VEGF-D is structurally and functionally most closely related to VEGF-C [see U.S. Patent 6,235,713 and International Patent Publ. No. WO 98/07832, incorporated herein by reference]. See SEQ ID NO: 866 for the polynucleotide sequence of VEGF-D; the encoded amino acid sequence is set forth in SEQ ID NO: 865. Like VEGF-C, VEGF-D is initially expressed as a prepro-peptide that undergoes N-terminal and C-terminal proteolytic processing, and forms non-covalently linked dimers. VEGF-D stimulates mitogenic responses in endothelial cells *in vitro*. During embryogenesis, VEGF-D is expressed in a complex temporal and spatial pattern, and its expression persists in the heart, lung, and skeletal muscles in adults. Isolation of a biologically active fragment of VEGF-D designated VEGF-DΔNΔC, is described in International Patent Publication No. WO 98/07832, incorporated herein by reference. VEGF-DΔNΔC consists of amino acid residues 93 to 201 of VEGF-D (SEQ ID NO: 26) optionally linked to the affinity tag peptide FLAG®, or other sequences.

The prepro-VEGF-D polypeptide has a putative signal peptide of 21 amino acids and is apparently proteolytically processed in a manner analogous to the processing of prepro-VEGF-C. A "recombinantly matured" VEGF-D lacking residues 1-92 and 202-354 retains the ability to activate receptors VEGFR-2 and VEGFR-3, and appears to associate as non-covalently linked dimers. Thus, preferred VEGF-D polynucleotides include those polynucleotides that comprise a nucleotide sequence encoding amino acids 93-201. The guidance provided above for introducing function-preserving modifications into VEGF-C polypeptides is also suitable for introducing function-preserving modifications into VEGF-D polypeptides. As another aspect of the invention, practice of the invention methods is contemplated wherein VEGF-D polypeptides are employed in lieu of VEGF-C polypeptides.

When compared with the blood vascular endothelium, the lymphatic endothelium exhibits specific morphological and molecular characteristics. For example, the lymphatic capillaries are larger than blood capillaries, they have an irregular or collapsed lumen with no red blood cells, a discontinuous basal lamina, overlapping intercellular junctional complexes and anchoring filaments that connect

the lymphatic endothelial cells to the extracellular matrix (Witte, M.H., et al., *Microsc. Res. Tech. 55*:122-145 (2001)). Unlike the blood capillaries, the lymphatic capillaries lack pericyte coverage. At the molecular level several lymphatic specific markers have been identified, including VEGFR-3, the Prox-1 transcription factor, the hyaluronan receptor LYVE-1, the membrane mucoprotein podoplanin, the beta-chemokine receptor D6, the cytoskeletal proteins desmoplakin I and II and the macrophage mannose receptor I (Wigle, J.T. & Oliver, G., *Cell 98*:769-778 (1999); Banerji, S., et al., *J. Cell Biol. 144*:789-801 (1999): Breiteneder-Geleff, S., et al., *Am. J. Pathol. 154*:385-394 (1999): Nibbs, R.J., et al., *Am. J. Pathol. 158*:867-877 (2001); Ebata, N., et al., *Microvasc. Res. 61*:40-48. (2001); Irjala, H., et al., *J. Exp. Med. 194*:1033-1041 (2001)). The present invention relates to the genetic identity of lymphatic capillary endothelial cells versus blood vascular endothelial cells using a gene profiling approach.

"Stringent hybridization conditions" or "stringent conditions" refer to conditions under which a nucleic acid such as an oligonucleotide will specifically hybridize to its target sequence. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer nucleic acids hybridize specifically at higher temperatures than shorter sequences. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration conditions) at which 50% of the nucleic acids complementary to the target sequence hybridize to the target sequence at equilibrium. The term "complementary" refers to standard Watson-Crick base pairing between nucleotides of two nucleic acid molecules. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and at a temperature that is at least about 30°C for short probes, primers or oligonucleotides (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for longer probes, primers or oligonucleotides. Stringent conditions also can be achieved with the addition of destabilizing agents, such as formamide, as is known in the art Exemplary stringent hybridization conditions are hybridization at 42°C for 20 hours

in a solution containing 50% formamide, 5xSSPE, 5x Denhardt's solution, 0.1% SDS and 0.1 mg/ml denatured salmon sperm DNA, with a wash in 1xSSC, 0.1% SDS for 30 minutes at 65°C.

According to the invention, distinct gene expression profiles for blood vascular and lymphatic endothelial cells have been discovered. These results provide new insights into the phenotypic diversity of endothelial cells and reveal new potential lymphatic endothelial molecules, some of which could provide important targets for the therapy of diseases characterized by abnormal angiogenesis or lymphangiogenesis.

Differences in the expression of genes encoding proteins involved in inflammatory processes were found, as well as in those mediating cell-cell and cell-matrix interactions. Furthermore, several previously unknown genes were identified in the context of endothelial cell biology, which were differentially expressed in the two cell lineages. Several of the genes were originally cloned from neural tissues, including genes involved in the uptake of synaptic macromolecules and in synapse formation and remodeling (neuronal pentraxins I and II (Kirkpatrick, L.L., et al., *J. Biol. Chem. 275*:17786-17792. 2000), in the trafficking of synaptic vesicles (NAP-22 (Yamamoto, Y., et al., *Neurosci. Lett. 224*:127-130. 1997), piccolo (Fenster, S.D., et al., *Neuron 25*:203-214 (2000)) and in the axon growth and guidance (Nr-CAM (Grumet, M., *Cell Tissue Res. 290*:423-428 (1997), reelin (Rice, D.S. & Curran, T., *Annu. Rev. Neurosci. 24*:1005-1039 (2001)).

In addition, the LECs especially expressed a number of as yet uncharacterized genes, which were originally cloned and highly expressed in nervous tissues (KIAA genes (Kikuno, R., et al., *Nucleic Acids Res. 30*:166-168. 2002). The gene expression profiling data disclosed herein therefore support the view that the same molecular mechanisms that are involved in governing neural cell positioning, in guiding axonal growth cones to their specific targets and in synaptogenesis may also be commonly used in the development of the vascular system and in the establishment of BEC and LEC identity. Some other signaling molecules first described in the developing nervous system have already been implicated in the development of the vasculature and vice versa (Shima and Mailhos, *Curr. Opin. Genet. Dev. 10*:536-542

(2000); Oosthuyse, et al., *Nat. Genet. 28*:131-138 (2001); Sondell, et al., *Eur. J. Neurosci. 12*:4243-4254 (2000)).

In the LECs, expression of several genes previously shown to be expressed in smooth muscle cells (SMCs) and pericytes was observed, such as matrix Gla, a mineral binding extracellular matrix protein involved in the inhibition of vascular and tissue calcification (Luo, G, et al., *Nature 386*:78-81 (1997)), monoamine oxidase A, the main degradative enzyme of monoamine hormones and neurotransmitters (Rodriguez, M.J., et al., *Cell Tissue Res. 304*:215-220 (2001)), integrin α9 (Palmer, E.L., et al., *J. Cell Biol. 123*:1289-1297 (1993)) and apolipoprotein D (Hu, C.Y., et al., *J. Neurocytol. 30*:209-218 (2001)). Some similarity of gene expression patterns between LECs and SMCs could be related to the lack of SMC around lymphatic capillaries. Instead, LECs may carry out some SMC functions by themselves. For example, lymph flow is maintained due to the intrinsic contractility of the LECs (Witte, M.H., et al., *Microsc. Res. Tech. 55*:122-145 (2001)), reminiscent of the ability of vascular SMCs to contract.

Molecular discrimination of the lymphatic and blood vessels is essential in studies of diseases involving the blood and/or lymphatic vessels and in the targeted treatment of such diseases. To date, several lymphatic endothelial specific markers have been identified, but some of them are expressed only in a subset of the lymphatic vessels, while others also occur in some blood vessel endothelia or in other cell types and their expression patterns may change in pathological conditions (for example, VEGFR-3 (Valtola, R., et al., *Am. J. Pathol. 154*:1381-1390. 1999)). Identification of new vascular markers according to the invention should provide a more reliable analysis of the blood and lymphatic vessels in pathological situations and eventually better diagnosis and treatment. Furthermore, inhibition of the function of certain molecules involved in the regulation of angiogenesis and/or lymphangiogenesis is known to prevent tumor growth and metastasis, and stimulation of the growth of blood or lymphatic vessels has been shown to be beneficial in several pathological conditions. Thus the BEC and LEC specific molecular regulators identified according to the invention may provide new targets for the treatment of diseases characterized by abnormal angiogenesis and lymphangiogenesis.

Several of the new LEC genes encode transmembrane proteins that may be specific molecular markers for lymphatic endothelial cells (Table 6). These genes and encoded proteins are useful for targeted treatment of diseases that involve lymphatic vessels. They may also be useful for preparing antibodies, as antibodies against LEC-specific proteins can be used to discriminate between blood and lymphatic vessels in pathological and physiological situations. Antibodies may also be useful for the isolation of lymphatic endothelial cells. These proteins may also play a role in the regulation of lymphangiogenesis, and can provide new candidate genes for diseases that involve lymphatic vessels, such as lymphedema.

The lymphatic endothelial cell specific surface molecules can be used for molecular drug targeting with antibodies, peptides and small-molecular weight compounds, which can act as agonists or antagonists for growth factor receptor, cyto- and chemokine receptor, and hemopoietin receptor signaling, cell adhesion and cell interaction with extracellular matrix or with other cell surface molecules. Such molecules can also be used for targeting of cytotoxic or cytostatic drugs into the lymphatic endothelial cells and for the attachment of electron-dense, radio-opaque or radioactive markers for imaging of disease processes associated with the lymphatic vessels. Such diseases include lymphedema, lymphangioma, lymphangiomyoma, lymphangiomatosis, lymphangiectasis, lymphosarcoma and lymphangiosclerosis.

The lymphatic endothelial cell surface molecules may be used for targeting of gene therapy for example by antibody-coated liposomes (containing proteins or genes as cargo) or by viral transducing vectors such as adenoviruses, adeno-associated viruses or lentiviruses having modified capsid/envelope proteins. The manipulation of lymphatic endothelial cell specific molecules may be applicable to treatment of disease processes associated with tissue edema by increasing fluid transport across the lymphatic vessel wall for example by modulating endothelial cell-cell or cell-matrix interactions or via stimulating transendothelial transport. Targeting of the lymphatic endothelial cells for example with cytotoxic or cytostatic compounds is contemplated to be valuable in malignant tumor diseases prone to metastatic spread via the lymphatic system.

The lymphatic endothelial cell molecules may allow the improved *in vitro* growth of lymphatic endothelial cells as well as *in vitro* tissue engineering of lymphatic vessels for use in diseases where the lymphatics have been damaged, such as after surgery and in various forms of lymphedema. Ligands of the cell surface proteins may further be applied to coat various polymeric matrices for the adhesion of cells in, e.g., bioimplants.

The lymphatic endothelial-cell-specific molecules such as surface molecules can provide important tools for the modulation of inflammatory, autoimmune and infectious processes involving leukocyte migration and immune recognition as well as the stimulation of secondary immune responses. Such processes include the migration of antigen presenting cells into the lymphatic system including lymph nodes as well as transendothelial cell trafficking of lymphocytes and other leukocyte subclasses and the homing, survival and function of the various classes of leukocytes.

These molecules may allow one to modulate the metabolism of fatty acids including fatty acid/chylomicron absorption from the gut and regulation of fat accumulation in adipose tissue in various organs such as in the subcutaneous tissue and in the arterial wall.

Lymphatic endothelial-cell-specific molecules may further allow one to modulate the metabolism of fatty acids including fatty acid/chylomicron absorption from the gut and regulation of fat accumulation in adipose tissue in various organs such as in the skin subcutaneous tissue and in the arterial wall.

The lymphatic-cell-specific transmembrane proteins are expected to function in cell adhesion (e.g., adhesion between lymphatic endothelial cell-lymphatic endothelial cell, lymphatic endothelial cell-smooth muscle cell, lymphatic endothelial cell-immune system cell such as lymphocyte or dendritic cell), cell-extracellular matrix contacts, or as receptors such as growth factor, cytokine, chemokine or microbial receptors or ion channels. The transmembrane proteins connect to intracellular molecules that can induce cell growth, cell migration, cell apoptosis, cell differentiation or cell adhesion or other cellular functions specific for endothelial cells such as expression of adhesion receptors for leukocytes, release of nitric oxide,

antigoagulant proteins, uptake of fluid and proteins from surrounding tissues and fat from gut or adipose tissues. TM proteins with short intracellular domains can function as auxiliary receptors in complex with other TM proteins.

The transmembrane proteins and their intracellular binding partner molecules can be used as molecular markers for lymphatic endothelial cells in normal and disease conditions, and to discriminate between blood and lymphatic vessels in pathological and physiological situations.

Antibodies against lymphatic specific transmembrane proteins, as well as peptides and small molecular compounds binding to extracellular domains of lymphatic-specific TM proteins can be used for the attachment of electron-dense, radio-opaque or radioactive markers for imaging of disease processes associated with the lymphatic vessels. Such diseases include lymphedema, lymphangioma, lymphangiomyoma, lymphangiomatosis, lymphangiectasis, lymphosarcoma and lymphangiosclerosis. Similarly, the lymphatic vessels can be visualized, e.g., during therapy of patients suffering from insufficient lymphatic growth, such as in lymphedema, or alternatively during treatment aiming to prevent lymphatic growth, e.g., in tumors, thereby facilitating the monitoring of the therapeutic method of the invention.

Antibodies against LEC-specific TM proteins are also expected to be useful for the isolation of lymphatic endothelial cells.

Antibodies against lymphatic-specific transmembrane proteins, or peptides or small-molecule compounds binding to the extracellular domain of lymphatic-specific TM proteins are expected to be useful in targeting drug delivery to lymphatic endothelial cells, e.g., by coupling an antibody, peptide or small-molecule compound to a cytotoxic or cytostatic compound. Such coupled compounds are useful as therapeutics in the treatment of malignant tumor diseases prone to metastatic spread via the lymphatic system, as well as in ameliorating a symptom associated with any such disease. The antibodies, peptides or small-molecule compounds can also be coupled to stimulatory lymphatic endothelial molecules such as growth factors, cytokines and chemokines to promote stimulation.

Additionally, antibodies against lymphatic-specific TM proteins or peptides, or small-molecule compounds binding to the extracellular domain of lymphatic-specific TM proteins, may be used for targeting of gene therapy, for example, by antibody-coated liposomes (containing proteins, genes or other molecules as cargo) or by viral transducing vectors such as adenoviruses, adeno-associated viruses, lentiviruses, or the like, having modified capsid/envelope proteins. The manipulation of lymphatic endothelial-cell-specific molecules are expected to be applicable to the treatment of disease processes associated with tissue edema due to the relative absence, or relative dysfunction, of lymphatic vessels, which can result from an infection, surgery, radiotherapy or a genetic defect by increasing fluid transport across the lymphatic vessel wall, for example by modulating endothelial cell-cell or cell-matrix interactions or by stimulating transendothelial transport.

The lymphatic endothelial cell molecules are expected to improve the *in vitro* growth of lymphatic endothelial cells, as well as the *in vitro* tissue engineering of lymphatic vessels for use in treating disorders or diseases where the lymphatics have been damaged, such as after surgery, in various forms of lymphedema, and in other applications as described herein. Ligands of the cell-surface proteins may further be applied as a coating to various polymeric matrices for the adhesion of cells in, e.g., bioimplants.

Inflammatory, autoimmune and infectious processes involving leukocyte migration and immune recognition, such as migration of antigen-presenting cells into the lymphatic system, including lymph nodes, as well as transendothelial cell trafficking of lymphocytes and other leukocyte subclasses and the homing, survival and function of the various classes of leukocytes can be modulated by targeting endothelial-cell-specific TM proteins, which mediate these cell adhesion processes.

Upregulation of lymphatic-specific genes in, e.g., cancer are expected to be useful as diagnostic markers, and monitoring such upregulated expression with an antibody against a lymphatic endothelial-cell-specific protein, e.g., by immunostaining of tissue(s) or by using a probe hybridizable to a lymphatic

endothelial-cell-specific mRNA, e.g., under stringent hybridization conditions as described herein, is contemplated.

Lymphatic endothelial-cell-specific transcription factors are expected to be useful for the differentiation of lymphatic endothelial cells from embryonic stem cells, endothelial precursor cells, or blood vascular endothelial cells.

The lymphatic endothelial transcription factors are expected to improve the *in vitro* growth of lymphatic endothelial cells, as well as to facilitate *in vitro* tissue engineering of lymphatic vessels for use in treating disorders or diseases where the lymphatics have been damaged, such as after surgery, in various forms of lymphedema, and in other applications disclosed herein.

Intracellular signaling proteins participating in signaling pathways regulating lymphatic endothelial cell proliferation, differentiation, apoptosis, migration or adhesion are expected to be useful targets for small-molecule compounds inhibiting these signaling events, and cellular functions dependent on such signaling. Signaling proteins are also expected to participate in VEGFR-3 signaling pathways, and will be useful in modulating cellular activities controlled, at least in part, by VEGFR-3 signaling, such as lymphangiogenesis.

The lymphatic endothelial cell molecules are expected to improve the *in vitro* growth of lymphatic endothelial cells as well as *in vitro* tissue engineering of lymphatic vessels for use in treating diseases or disorders where the lymphatics have been damaged, such as after surgery, in various forms of lymphedema, and in other applications as described herein.

Lymphatic-specific transcription factors are also expected to be useful in modulating gene expression in endothelial cells to induce the expression of other lymphatic-specific genes in, for example, blood vascular endothelial cells or endothelial precursor cells.

Lymphatic-specific gene transcripts are expected to provide useful targets for RNA interference (RNAi)-induced inhibition of expression. RNAi technology is expected to be useful in the methods according to the invention, such as therapeutic methods effective in treating hyper- and hypo-proliferative endothelial-cell-associated diseases and disorders, as well as methods of ameliorating a symptom

of any such disease or disorder. RNAi methodologies are known in the art and known RNAi technologies are contemplated as useful in various aspects of the invention. *See* Fire et al., Nature 391:806-811. (1998) and Sharp, P., Genes and Dev. 13:139-141. (1999), each of which is incorporated herein by reference. It is preferred that RNAi compounds be double-stranded RNA molecules corresponding to part or all of a coding region of a desired target for expression.

As noted, several of the new LEC genes encode transcription factors, which may control cellular fate (iroquois-related homeobox gene), and may have an important role in the differentiation of lymphatic endothelial cells. Transcription factors disclosed herein may control transcription of genes involved for example in the proliferation of lymphatic endothelial cells, and may be important molecular regulators of lymphatic growth (Table 5). Lymphatic endothelial cell specific transcription factors can be used for the differentiation of lymphatic endothelial cells from embryonic stem cells, endothelial precursor cells or from blood vascular endothelial cells.

The lymphatic endothelial transcription factors may allow the improved in vitro growth of lymphatic endothelial cells as well as in vitro tissue engineering of lymphatic vessels for use in diseases where the lymphatics have been damaged, such as after surgery and in various forms of lymphedema.

Polynucleotides of the Invention

In general, the isolated polynucleotides of the invention include the LEC and BEC polynucleotides exhibiting differential expression and identified in Tables 3, 4, 14, 15 and 16. The sequences of these polynucleoides are provided in Table 16, associated with their known database accession numbers, where applicable. In Tables 14 and 15, these accession numbers are correlated with unique sequence identifiers, thus permitting identification by sequence idenfier of each citation to an accession number. The polynuleotide sequences may include a coding region and may include non-coding flanking sequences, which are readily identifiable by one of skill in the art. The invention contemplates polynucleotides comprising part, or all, of a coding region, with or without flanking regions, e.g., poly A sequences, 5' non-

coding sequences, and the like. The polynucleotides of the present invention also include, but are not limited to, a polynucleotide that hybridizes to the complement of the nucleotide sequence of any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 under highly stringent hybridization conditions; a polynucleotide that hybridizes to the complement of the nucleotide sequence of any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 under moderately stringent hybridization conditions; a polynucleotide which is an allelic variant of any polynucleotide recited above; a polynucleotide which encodes a species homologue of any of the proteins recited above; or a polynucleotide that encodes a polypeptide comprising a specific domain or truncation of the polypeptide of any one of SEQ ID NOS: 31-44, 46, 48, 50 and 52. Such polynucleotides hybridize under the above conditions to the complement of any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 or to a fragment of any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 wherein the fragment is greater than at least about 10 bp, and, in alternate embodiments, is about 20 to about 50 bp, or is greater than about 100 bp, 200 bp, 300 bp, 400 bp, 500 bp, 600 bp, 700 bp, or 800 bp, where appropriate.

The polynucleotides of the invention also provide polynucleotides that are variants of the polynucleotides recited above. Typically, such a variant sequence varies from one of those listed herein by no more than about 20%, *i.e.*, the number of individual nucleotide substitutions, additions, and/or deletions in a similar sequence, as compared to the corresponding reference sequence, divided by the total number of nucleotides in the variant sequence is about 0.2 or less. Such a sequence is said to have 80% sequence identity to the listed sequence. Such a variant sequence can be routinely identified by applying the foregoing algorithm.

In one embodiment, a variant polynucleotide sequence of the invention varies from a listed sequence by no more than 10%, *i.e.*, the number of individual nucleotide substitutions, additions, and/or deletions in a variant sequence, as compared to the corresponding reference sequence, divided by the total number of nucleotides in the variant sequence is about 0.1 or less. Such a sequence is said to have 90% sequence identity to the listed sequence. Such a variant sequence can be routinely identified by applying the foregoing algorithm.

In an alternate embodiment a variant sequence of the invention varies from a listed sequence by no more than by no more than 5%, *i.e.*, the number of individual nucleotide substitutions, additions, and/or deletions in a variant sequence, as compared to the corresponding reference sequence, divided by the total number of nucleotides in the variant sequence is about 0.05 or less. Such a sequence is said to have 95% sequence identity to the listed sequence. Such a variant sequence can be routinely identified by applying the foregoing algorithm.

In yet another alternate embodiment, a variant sequence of the invention varies from a listed sequence by no more than 2%, *i.e.*, the number of individual nucleotide substitutions, additions, and/or deletions in a variant sequence, as compared to the corresponding reference sequence, divided by the total number of nucleotides in the variant sequence is about 0.02 or less. Such a sequence is said to have 98% sequence identity to the listed sequence. Such a variant sequence can be routinely identified.

A polynucleotide according to the invention can be joined to any of a variety of other nucleotide sequences by well-established recombinant DNA techniques (see Sambrook J et al. (2d Ed.; 1989) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). Useful nucleotide sequences for joining to polypeptides include an assortment of vectors, *e.g.*, plasmids, cosmids, lambda phage derivatives, phagemids, and the like, that are well known in the art. Accordingly, the invention also provides a vector including a polynucleotide of the invention and a host cell containing the polynucleotide. In general, the vector contains an origin of replication functional in at least one organism, convenient restriction endonuclease sites, and a selectable marker for the host cell. Vectors according to the invention include expression vectors, replication vectors, probe generation vectors, sequencing vectors, and retroviral vectors. A host cell according to the invention can be a prokaryotic or eukaryotic cell and can be a unicellular organism or part of a multicellular organism. Large numbers of suitable vectors and promoters are known to those of skill in the art and are commercially available for generating the recombinant constructs of the present invention.

The sequences falling within the scope of the present invention are not limited to the specific sequences herein described, but also include allelic variations thereof. Allelic variations can be routinely determined by comparing the sequence provided in any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51, a representative intermediate fragment thereof, or a nucleotide sequence at least 99.9% identical to any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 with a sequence from another isolate of the same species. Furthermore, to accommodate codon variability, the invention includes nucleic acid molecules coding for the same amino acid sequences as do the specific open reading frames (ORFs) disclosed herein. In other words, in the coding region of an ORF, substitution of one codon for another which encodes the same amino acid is expressly contemplated.

Unless provided for otherwise here, all terms are defined as is known in the art, for example as employed in U.S. Patent No. 6,350,447, incorporated herein by reference.

Also contemplated are antisense polynucleotides based on the sequence of any of the LEC or BEC polynucleotides according to the invention. Such antisense polynucleotides are substantially complementary (e.g., at least 90% complementarity), and preferably perfectly complementary, to sequences of the polynucleotides of the invention, or fragments thereof, set out in the sequence listing, Tables 3, 4, 14-16, and throughout this disclosure that are differentially expressed in LECs and BECs. These polynucleotide sequences include any of SEQ ID NOS: 1-30, 45, 47, 49 and 51, or a fragment thereof comprising at least 10 contiguous nucleotides. Antisense nucleic acid comprises a nucleotide sequence that is complementary to a "sense" nucleic acid encoding a protein (e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence). Methods for designing and optimizing antisense nucleotides are described in Lima *et al.*, (*J Biol Chem,* ;272:626-38. 1997) and Kurreck *et al.*, (*Nucleic Acids Res.*, ;30:1911-8. 2002). In one aspect, antisense nucleic acid molecules are provided that comprise a sequence complementary to at least about 10, 25, 50, 100, 250 or 500 nucleotides or an entire coding strand. An antisense nucleic

acid of the invention can be constructed using chemical synthesis or enzymatic ligation reactions using procedures known in the art.

In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding a polypeptide of the invention. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues. In another embodiment, the antisense nucleic acid molecule is antisense to a "conceding region" of the coding strand of a nucleotide sequence encoding the polynucleotide. The term "conceding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (i.e., also referred to as 5' and 3' untranslated regions).

Antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick or Hoogsteen base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of the mRNA of the polynucleotide of the invention, but more preferably is an oligonucleotide that is antisense to only a portion of the coding or noncoding region of the mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45, or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis or enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids (e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize or bind to cellular mRNA and/or genomic DNA encoding the complementary polynucleotide, thereby inhibiting expression of the protein (e.g., by inhibiting transcription and/or translation). The hybridization can reflect conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid

molecule that binds to DNA duplexes, through specific interactions in the major groove of the double helix.

An example of a route of administration of antisense nucleic acid molecules of the invention includes direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface (e.g., by linking the antisense nucleic acid molecules to peptides or antibodies that bind to cell surface receptors or antigens). Additional routes of antisense therapy may be used in the invention, e.g., topical administration, transdermal administration [reviewed by Brand in *Curr. Opin. Mol. Ther.* 3:244-8. 2001] antisense administration using nanoparticulate systems [Lambert et al., *Adv. Drug. Deliv. Rev.* 47:99-112. 2001], or administration of antisense nucleotides conjugated with peptide [Juliano et al., *Curr. Opin. Mol. Ther.* 2:297-303. 2000].

The invention further contemplates use of the polynucleotides of the invention for gene therapy or in recombinant expression vectors which produce polynucleotides or polypeptides of the invention that can regulate an activity of LEC genes, and are useful in therapy of LEC disorders such as lymphedema. Delivery of a functional gene encoding a polypeptide of the invention to appropriate cells is effected *ex vivo*, *in situ*, or *in vivo* by use of vectors, including viral vectors (e.g., adenovirus, adeno-associated virus, or a retrovirus), or *ex vivo* by use of physical DNA transfer methods (e.g., liposomes or chemical treatments). See, for example, Anderson, *Nature*, supplement to vol. 392, no. 6679, pp. 25-20 (1998). For additional reviews of gene therapy technology see Friedmann, (*Science,* 244: 1275-1281. 1989); Verma, (*Scientific American*: 263:68-72, 81-84. 1990); and Miller, (*Nature,* 357: 455-460. 1992). Introduction of any one of the nucleotides of the present invention or a gene encoding a polypeptide of the invention can also be accomplished with extrachromosomal substrates (transient expression) or artificial chromosomes (stable expression). Cells may also be cultured *ex vivo* in the presence of proteins of the present invention in order to proliferate or to produce a desired effect on, or activity

in, such cells. In another embodiment, cells comprising vectors expressing the polynucleotides or polypeptides of the invention may be cultured *ex vivo* and administered to an individual in need of treatment for an LEC disease or disorder.

Given the foregoing disclosure of the nucleic acid constructs, it is possible to produce the gene product of any of the genes comprising the sequence of any of SEQ ID NOS: 1-30, 45, 47, 49 and 51 by routine recombinant DNA/RNA techniques. A variety of expression vector/host systems may be utilized to contain and express the coding sequence. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, phagemid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (*e.g.*, baculovirus); plant cell systems transfected with virus expression vectors (*e.g.*, Cauliflower Mosaic Virus, CaMV; Tobacco Mosaic Virus, TMV) or transformed with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmid); or even animal cell systems. Mammalian cells that are useful in recombinant protein productions include, but are not limited to, VERO cells, HeLa cells, Chinese hamster ovary (CHO) cells, COS cells (such as COS-7), WI38, BHK, HepG2, 3T3, RIN, MDCK, A549, PC12, K562 and HEK 293 cells.

Polypeptides of the invention

In general, the isolated LEC and BEC polypeptides of the invention are encoded by the above-described differentially expressed LEC and BEC polynuleotides of the invention. The sequences of the LEC and BEC polypeptides are provided in Table 16, associated with their known database accession numbers, where applicable. In Tables 14 and 15, these accession numbers are correlated with unique sequence identifiers, thus permitting identification by sequence idenfier of 4each citation to an accession number. The isolated polypeptides of the invention include, but are not limited to, a polypeptide comprising: the amino acid sequences set forth as any one of SEQ ID NOS.: 31-44, 46, 48, 50 and 52 or an amino acid sequence encoded by any one of the nucleotide sequences set forth in SEQ ID NOS.: 1-30, 45, 47, 49 and 51, or the corresponding full length or mature protein. The invention also

provides biologically active or immunologically active variants of any of the amino acid sequences set forth as SEQ ID NOS.: 31-44, 46, 48, 50 and 52, or the corresponding full length or mature protein suitable variant polypeptides have sequences that are at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, 86%, 87%, 88%, 89%, at least about 90%, 91%, 92%, 93%, 94%, typically at least about 95%, 96%, 97%, more typically at least about 98%, or most typically at least about 99% amino acid identity, that retain biological activity. Fragments of the proteins of the present invention which comprise at least 10 contiguous amino acids of a sequence disclosed herein and that are capable of exhibiting a biological activity of the corresponding full length protein are also encompassed by the present invention.

The protein coding sequence is identified in the sequence listing by translation of the disclosed nucleotide sequences. The mature form of such protein may be obtained by expression of a full-length polynucleotide in a suitable mammalian cell or other host cell. The sequence of the mature form of the protein is also determinable from the amino acid sequence of the full-length form. Where proteins of the present invention are membrane bound, soluble forms of the proteins are also provided. In such forms, part or all of the regions causing the proteins to be membrane bound are deleted so that the proteins are capable of being fully secreted from the cell in which it is expressed.

A variety of methodologies known in the art can be utilized to obtain any one of the isolated polypeptides or proteins of the present invention. At the simplest level, the amino acid sequence can be synthesized using commercially available peptide synthesizers. The polypeptides and proteins of the present invention can alternatively be purified from cells which have been altered to express the desired polypeptide or protein. As used herein, a cell is said to be altered to express a desired polypeptide or protein when the cell, through genetic manipulation, is made to produce a polypeptide or protein which it normally does not produce or which the cell normally produces at a lower level. One skilled in the art can readily adapt procedures for introducing and expressing either recombinant or synthetic sequences

into eukaryotic or prokaryotic cells in order to generate a cell which produces one of the polypeptides or proteins of the present invention.

A "fragment" of a polypeptide is meant to refer to any portion of the molecule, such as the peptide core, a variant of the peptide core, or an extracellular region of the polypeptide. A "variant" of a polypeptide is meant to refer to a molecule substantially similar in structure and biological activity to either the entire molecule, or to a fragment thereof. Thus, provided that two molecules possess a similar activity, they are considered variants as that term is used herein even if the composition or secondary, tertiary, or quaternary structure of one of the molecules is not identical to that found in the other, or if the sequence of amino acid residues is not identical. An "analogue" of a polypeptide or genetic sequence is meant to refer to a protein or genetic sequence substantially similar in function and structure to the isolated polypeptide or genetic sequence.

It is understood herein that conservative amino acid substitutions can be performed to a purified and isolated polypeptide comprising any one of the sequences of SEQ ID NOS.: 31-44, 46, 48, 50 and 52 which are likely to result in a polypeptide that retains biological or immunological activity, especially if the number of such substitutions is small. By "conservative amino acid substitution" is meant substitution of an amino acid with an amino acid having a side chain of a similar chemical character. Similar amino acids for making conservative substitutions include those having an acidic side chain (glutamic acid, aspartic acid); a basic side chain (arginine, lysine, histidine); a polar amide side chain (glutamine, asparagine); a hydrophobic, aliphatic side chain (leucine, isoleucine, valine, alanine, glycine); an aromatic side chain (phenylalanine, tryptophan, tyrosine); a small side chain (glycine, alanine, serine, threonine, methionine); or an aliphatic hydroxyl side chain (serine, threonine).

Microarrays

Another aspect of the invention is a composition comprising a plurality of polynucleotide probes for use in detecting gene expression pattern(s) characteristic of particular cell type(s) and for detecting changes in the expression pattern of a

particular cell type, e.g., lymphatic endothelial cells. For example, the invention comprehends an array, such as a microarray, comprising polynucleotides having at least 10 contiguous nucleotides selected from the polynucleotide sequences presented in the sequence listing.

Also contemplated are microarrays comprising polynucleotides having at least 10 contiguous nucleotides selected from the group of SEQ ID NOS: 1-30, 45, 47, 49 and 51. Microarrays of the invention comprise at least 3 polynucleotides, wherein each enumerated polynucleotide has a distinct sequence selected from the group consisting of SEQ ID NOS:1-30, 45, 47, 49 and 51. Such microarrays may also have duplicate polynucleotides and additional polynucleotides, e.g., control polynucleotides for use in hybridization-based assays using the microarray. Arrays, including microarrays, having more than three distinct polynucleotides according to the invention, such as at least five, seven, nine, 20, 50 or more such polynucleotides, will be recognized as arrays according to the invention having the capability of yielding subtle distinctions between biological samples such as various endothelial cell types, or of providing a different, and typically greater, level of confidence in the various uses of such arrays, e.g., in screening for particular endothelial cells, in screening for abnormal or diseases cells and tissues, and the like.

The term "microarray" refers to an ordered arrangement of hybridizable array elements. The array elements are arranged so that there are preferably at least three or more different array elements, more preferably at least 100 array elements, and most preferably at least 1,000 array elements, on a solid support. Preferably, the solid support is a 1 cm^2 substrate surface, bead, paper, nylon or other type of membrane, filter, chip, glass slide, or any other suitable solid support. The hybridization signal from each of the array elements is individually distinguishable. In a preferred embodiment, the array elements comprise polynucleotide probes.

Hybridization means contacting two or more nucleic acids under conditions suitable for base pairing. Hybridization includes interaction between partially or perfectly complementary nucleic acids. Suitable hybridization conditions are well known to those of skill in the art. In certain applications, it is appreciated that lower stringency conditions may be required. Under these conditions,

hybridization may occur even though the sequences of the interacting strands are not perfectly complementary, being mismatched at one or more positions. Conditions may be rendered less stringent by adjusting conditions in accordance with the knowledge in the art, e.g., increasing salt concentration and/or decreasing temperature. Suitable hybridization conditions are those conditions that allow the detection of gene expression from identifiable expression units such as genes. Preferred hybridization conditions are stringent hybridization conditions, such as hybridization at 42°C in a solution (*i.e.*, a hybridization solution) comprising 50% formamide, 1% SDS, 1 M NaCl, 10% dextran sulfate, and washing for 30 minutes at 65°C in a wash solution comprising 1 X SSC and 0.1% SDS. It is understood in the art that conditions of equivalent stringency can be achieved through variation of temperature and buffer, or salt concentration, as described in Ausubel, *et al.* (Eds.), Protocols in Molecular Biology, John Wiley & Sons (1994), pp. 6.0.3 to 6.4.10. Modifications in hybridization conditions can be empirically determined or precisely calculated based on the length and the percentage of guanosine/cytosine (GC) base pairing of the probe. The hybridization conditions can be calculated as described in Sambrook, *et al.*, (Eds.), Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York (2d. Ed.; 1989), pp. 9.47 to 9.51.

One method of using probes and primers of the invention is in the detection of gene expression in human cells. Normally, the target will be expressed RNAs, although genomic DNA or a cDNA library may be screened. By varying the stringency of hybridization and the target binding site (i.e., the sequence of the probe, corresponding to a subset of one of the sequences set forth at SEQ ID NOS: 1-30, 45, 47, 49 and 51), different degrees of homology are expected to result in hybridization.

The microarray can be used for large-scale genetic or gene expression analysis of a large number of target polynucleotides. The microarray can also be used in the diagnosis of diseases and in the monitoring of treatments. Further, the microarray can be employed to investigate an individual's predisposition to a disease. Furthermore, the microarray can be employed to investigate cellular responses to infection, drug treatment, and the like.

The nucleic acid probes can be genomic DNA or cDNA or mRNA polynucleotides or oligonucleotides, or any RNA-like or DNA-like material, such as peptide nucleic acids, branched DNAs, and the like. The probes can be sense or antisense nucleotide probes. Where target polynucleotides are double-stranded, the probes may be either sense or antisense strands. Where the target polynucleotides are single-stranded, the probes are complementary single strands. In one embodiment, the probes are cDNAs. The size of the DNA sequence of interest may vary and is preferably from 100 to 10,000 nucleotides, more preferably from 150 to 3,500 nucleotides.

The probes can be prepared using a variety of synthetic or enzymatic techniques, which are well known in the art. The probes can be synthesized, in whole or in part, using chemical methods well known in the art (Caruthers et al., Nucleic Acids Res., Symp. Ser., 215-233, 1980).

Pharmaceutical Formulations and Routes of Administration

A protein of the present invention (from whatever source derived, such as from recombinant and non-recombinant sources) may be administered to a patient in need, by itself, or in pharmaceutical compositions where it is mixed with suitable carriers, diluents, adjuvants or excipients at doses to treat or ameliorate a variety of disorders. Such a composition may also contain (in addition to protein and a carrier) diluents, fillers, salts, buffers, stabilizers, solubilizers, and other materials well known in the art. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier will depend on the route of administration. The pharmaceutical composition of the invention may also contain cytokines, chemokines, lymphokines, growth factors, or other hematopoietic factors such as a PDGF, a VEGF (particularly a VEGF-C or a VEGF-D), VEGFR-3 (including soluble VEGFR-3 peptides comprising an extracellular domain), M-CSF, GM-CSF, TNF, IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, IL-13, IL-14, IL-15, IFN, TNF0, TNF1, TNF2, G-CSF, Meg-CSF, thrombopoietin, stem cell factor, and erythropoietin. Various forms of these polypeptides are

contemplated as well, such as isolated holoproteins, subunits, fragments (e.g., soluble fragments), and peptide fusions. The pharmaceutical composition may further contain other agents which either enhance the activity of the protein or complement its activity or use in treatment. Such additional factors and/or agents may be included in the pharmaceutical composition to produce a synergistic effect with a protein of the invention, or to minimize side effects. Conversely, a protein of the present invention may be included in formulations of the particular cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent to minimize side effects of the cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent. A protein of the present invention may be active in multimers (*e.g.*, heterodimers or homodimers) or complexes with itself or other proteins. As a result, pharmaceutical compositions of the invention may comprise a protein of the invention in such multimeric or complexed form.

Techniques for formulation and administration of the compounds of the instant application may be found in "Remington's Pharmaceutical Sciences," Mack Publishing Co., Easton, Pa., latest edition. A therapeutically effective dose further refers to that amount of the compound sufficient to result in amelioration of symptoms, *e.g.*, treatment, healing, prevention or amelioration of the relevant medical condition, or an increase in rate of beneficial change, healing, prevention or amelioration of such conditions. When applied to an individual active ingredient, administered alone, a therapeutically effective dose refers to that ingredient alone. When applied to a combination, a therapeutically effective dose refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

In practicing methods of treatment or use of the present invention, a therapeutically effective amount of protein of the present invention is administered to a mammal having a condition or disorder to be treated. Protein of the present invention may be administered in accordance with the method of the invention either alone or in combination with other therapies such as treatments employing cytokines, lymphokines or other hematopoietic factors. When co-administered with one or more

cytokines, lymphokines or other hematopoietic factors, a protein of the invention may be administered either simultaneously with the cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors, or sequentially. If administered sequentially, the attending physician will decide on the appropriate sequence of administering a protein of the invention in combination with a cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors.

Routes of Administration

Suitable routes of administration may, for example, include oral, rectal, transmucosal, or intestinal administration; parenteral delivery, including intramuscular, subcutaneous, intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections. Administration of protein of the present invention used in the pharmaceutical composition or to practice the method of the present invention is carried out in a variety of conventional ways, such as oral ingestion, inhalation, topical application or cutaneous, subcutaneous, intraperitoneal, parenteral or intravenous injection. Intravenous administration to a mammal, such as a human patient, is preferred. Alternately, one may administer the compound in a local rather than systemic manner, for example, via injection of the compound at the site of intended action.

Compositions/Formulations

Pharmaceutical compositions for use in accordance with the present invention thus may be formulated in a conventional manner using one or more physiologically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. These pharmaceutical compositions may be manufactured in a manner that is itself known, *e.g.*, by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes. Proper formulation is dependent upon the route of administration chosen. When a therapeutically effective amount of protein of the

present invention is administered orally, protein of the present invention will be in the form of a tablet, capsule, powder, solution or elixir. When administered in tablet form, the pharmaceutical composition of the invention may additionally contain a solid carrier such as a gelatin or an adjuvant. The tablet, capsule, and powder contain from about 5 to 95% protein of the present invention, and preferably from about 25 to 90% protein of the present invention. When administered in liquid form, a liquid carrier such as water, petroleum, oils of animal or plant origin such as peanut oil, mineral oil, soybean oil, or sesame oil, or synthetic oils may be added. The liquid form of the pharmaceutical composition may further contain physiological saline solution, dextrose or other saccharide solution, or glycols such as ethylene glycol, propylene glycol or polyethylene glycol. When administered in liquid form, the pharmaceutical composition contains from about 0.5 to 90% by weight of protein of the present invention, and preferably from about 1 to 50% protein of the present invention.

When a therapeutically effective amount of protein of the present invention is administered by intravenous, cutaneous or subcutaneous injection, protein of the present invention will be in the form of a pyrogen-free, parenterally acceptable aqueous solution. The preparation of such parenterally acceptable protein solutions, having due regard to pH, isotonicity, stability, and the like, is within the skill in the art. A preferred pharmaceutical composition for intravenous, cutaneous, or subcutaneous injection should contain, in addition to protein of the present invention, an isotonic vehicle such as Sodium Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, Lactated Ringer's Injection, or other vehicle as known in the art. The pharmaceutical composition of the present invention may also contain stabilizers, preservatives, buffers, antioxidants, or other additives known to those of skill in the art. For injection, the agents of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiological saline buffer. For transmucosal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

For oral administration, the compounds can be formulated readily by combining the active compounds with pharmaceutically acceptable carriers well known in the art. Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for oral ingestion by a patient to be treated. Pharmaceutical preparations for oral use can be obtained by combination with a solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired, disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof such as sodium alginate. Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the active ingredients in admixture with filler such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added. All formulations for oral administration should be in dosages suitable for such administration. For buccal administration, the compositions may take the form of tablets or lozenges formulated in conventional manner.

For administration by inhalation, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant, *e.g.*, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, *e.g.*, gelatin for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch. The compounds may be formulated for parenteral administration by injection, *e.g.*, by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, *e.g.*, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents.

Pharmaceutical formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, *e.g.*, sterile pyrogen-free water, before use.

The compounds may also be formulated in rectal compositions such as suppositories or retention enemas, *e.g.*, containing conventional suppository bases such as cocoa butter or other glycerides. In addition to the formulations described previously, the compounds may also be formulated as a depot preparation. Such long-acting formulations may be administered by implantation (for example

subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

A pharmaceutical carrier for the hydrophobic compounds of the invention is a cosolvent system comprising benzyl alcohol, a nonpolar surfactant, a water-miscible organic polymer, and an aqueous phase. The cosolvent system may be the VPD co-solvent system. VPD is a solution of 3% w/v benzyl alcohol, 8% w/v of the nonpolar surfactant polysorbate 80, and 65% w/v polyethylene glycol 300, made up to volume in absolute ethanol. The VPD co-solvent system (VPD:5W) consists of VPD diluted 1:1 with a 5% dextrose-in-water solution. This co-solvent system dissolves hydrophobic compounds well, and itself produces low toxicity upon systemic administration. Naturally, the proportions of a co-solvent system may be varied considerably without destroying its solubility and toxicity characteristics. Furthermore, the identity of the co-solvent components may be varied: for example, other low-toxicity nonpolar surfactants may be used instead of polysorbate 80; the fraction size of polyethylene glycol may be varied; other biocompatible polymers may replace polyethylene glycol, *e.g.*, polyvinyl pyrrolidone; and other sugars or polysaccharides may substitute for dextrose. Alternatively, other delivery systems for hydrophobic pharmaceutical compounds may be employed. Liposomes and emulsions are well known examples of delivery vehicles or carriers for hydrophobic drugs. Certain organic solvents such as dimethylsulfoxide also may be employed, although usually at the cost of greater toxicity. Additionally, the compounds may be delivered using a sustained-release system, such as semipermeable matrices of solid hydrophobic polymers containing the therapeutic agent. Various sustained-release materials have been established and are well known by those skilled in the art. Sustained-release capsules may, depending on their chemical nature, release the compound over a time period of a few weeks up to over 100 days. Depending on the chemical nature and the biological stability of the therapeutic reagent, additional strategies for protein stabilization may be employed.

The pharmaceutical compositions also may comprise suitable solid or gel-phase carriers or excipients. Examples of such carriers or excipients include but are not limited to calcium carbonate, calcium phosphate, various sugars, starches, cellulose derivatives, gelatin, and polymers such as polyethylene glycols. Many of the proteinase-inhibiting compounds of the invention may be provided as salts with pharmaceutically compatible counterions. Such pharmaceutically acceptable base addition salts are those salts which retain the biological effectiveness and properties of the free acids and which are obtained by reaction with inorganic or organic bases such as sodium hydroxide, magnesium hydroxide, ammonia, trialkylamine, dialkylamine, monoalkylamine, dibasic amino acids, sodium acetate, potassium benzoate, triethanol amine, and the like.

The pharmaceutical compositions of the invention may be in the form of a complex of a protein(s) of the present invention along with protein or peptide antigens. The pharmaceutical compositions of the invention may be in the form of a liposome in which protein of the present invention is combined, in addition to other pharmaceutically acceptable carriers, with amphipathic agents such as lipids which exist in aggregated form as micelles, insoluble monolayers, liquid crystals, or lamellar layers in aqueous solution. Suitable lipids for liposomal formulation include, without limitation, monoglycerides, diglycerides, sulfatides, lysolecithin, phospholipids, saponin, bile acids, and the like. Preparation of such liposomal formulations is within the level of skill in the art, as disclosed, for example, in U.S. Pat. Nos. 4,235,871; 4,501,728; 4,837,028; and 4,737,323, each of which is incorporated herein by reference.

The amount of protein of the invention in the pharmaceutical composition will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of protein of the present invention with which to treat each individual patient. Initially, the attending physician will administer low doses of protein of the present invention and observe the patient's response. Larger doses of protein of the present invention may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is

not increased further. It is contemplated that the various pharmaceutical compositions used to practice the methods of the invention should contain about 0.01 μg to about 100 mg (preferably about 0.1 μg to about 10 mg, more preferably about 0.1 μg to about 1 mg) of protein of the invention per kg body weight. When administered, the therapeutic composition for use in this invention is in a pyrogen-free, physiologically acceptable form. Therapeutically useful agents other than a protein of the invention which may also optionally be included in the composition as described above, may alternatively or additionally, be administered simultaneously or sequentially with the composition in the methods of the invention.

Polynucleotides of the present invention can also be used for gene therapy. Such polynucleotides can be introduced either *in vivo* or *ex vivo* into cells for expression in a mammalian subject. Polynucleotides of the invention may also be administered by other known methods for introduction of nucleic acid into a cell or organism (including, without limitation, in the form of viral vectors or naked DNA). Cells may also be cultured *ex vivo* in the presence of proteins of the present invention in order to proliferate or to produce a desired effect on or activity in such cells. Treated cells can then be introduced *in vivo* for therapeutic purposes.

Effective Dosage

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective amount to achieve an intended purpose. More specifically, a therapeutically effective amount means an amount effective to prevent development of, or to alleviate the existing symptoms of, the subject being treated. Suitable properties that may be used in determining effective dosages include measurements of LEC and/or BEC growth stimulation or inhibition, rates or extent of cell differentiation into LECs and/or BECs, tendencies of cell expression patterns to shift towards or away from LEC- or BEC-specific expression patterns, and the like. Determination of the effective amounts is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein. For any compound used in a method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. For

example, for inhibitory methods, a dose can be formulated in animal models to achieve a circulating concentration range that includes the IC_{50} as determined in cell culture (*i.e.*, the concentration of the test compound which achieves a half-maximal inhibitory concentration). Such information can be used to more accurately determine useful doses in humans.

A therapeutically effective dose refers to that amount of the compound that results in amelioration of symptoms or, in the case of life-threatening conditions, a prolongation of survival in a patient. Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, for determining the LD_{50} (the dose lethal to 50% of the population) and the ED_{50} (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio between LD_{50} and ED_{50}. Compounds which exhibit high therapeutic indices are preferred. The data obtained from these cell culture assays and animal studies can be used in formulating a range of dosage for use in human. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED_{50} with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition. See, *e.g.*, Fingl et al., 1975, in "The Pharmacological Basis of Therapeutics", Ch. 1 p.1.

The amount of composition administered will, of course, be dependent on the subject being treated, on the subject's weight, the severity of the affliction, the manner of administration and the judgment of the prescribing physician.

Packaging

The compositions may, if desired, be presented in a pack or dispenser device which may contain one or more unit dosage forms containing the active ingredient. The pack may, for example, comprise metal or plastic foil, such as a blister pack. The pack or dispenser device may be accompanied by instructions for administration. Compositions comprising a compound of the invention formulated in

a compatible pharmaceutical carrier may also be prepared, placed in an appropriate container, and labelled for treatment of an indicated condition.

In addition, the invention comprehends a use of such a composition to manufacture a medicament for the treatment of a cell or an organism, such as a human patient, having a hyperproliferative or hypoproliferative disorder of a LEC and/or BEC, such as lymphedema, lymphangioma, lymphangiomyeloma, lymphangiomatosis, lymphangiectasis, lymphosarcoma, or lymphangiosclerosis, comprising administering an effective amount, or dose, of a composition according to the invention to the cell or organism. Suitable compositions include, but are not limited to, any polynucleotide according to the invention (e.g., an antisense polynucleotide), any polypeptide according to the invention, an antibody specifically recognizing a polynucleotide or polypeptide according to the invention, a small molecule compound effective in modulating the expression of a polynucleotide according to the invention, and the like. Also contemplated are uses of compositions according to the invention for the manufacture of a medicament to ameliorate a symptom associated with a LEC- or BEC-associated disease or disorder.

Antibodies

Antibodies are useful for modulating the polypeptides of the invention due to the ability to easily generate antibodies with relative specificity, and due to the continued improvements in technologies for adopting antibodies to human therapy. Thus, the invention contemplates use of antibodies (e.g., monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies, bifunctional/bispecific antibodies, humanized antibodies, human antibodies, and complementary determining region (CDR)-grafted antibodies, including compounds which include CDR sequences which specifically recognize a polypeptide of the invention), specific for polypeptides of interest to the invention. Preferred antibodies are human antibodies, such as those produced in transgenic animals, which are produced and identified according to methods described in WO93/11236, published June 20, 1993, which is incorporated herein by reference in its entirety. Antibody fragments, including Fab, Fab′, F(ab′)2, and Fv, are also provided by the invention. The term "specific for,"

when used to describe antibodies of the invention, indicates that the variable regions of the antibodies of the invention recognize and bind the polypeptide of interest at a detectably different, and greater, level that bind to other substances (i.e., able to distinguish the polypeptides of interest from other known polypeptides of the same family, by virtue of measurable differences in binding affinity, despite the possible existence of localized sequence identity, homology, or similarity between family members). It will be understood that specific antibodies may also interact with other proteins (for example, *S. aureus* protein A or other antibodies in ELISA techniques) through interactions with sequences outside the variable region of the antibodies, and in particular, in the constant region of the molecule. Screening assays to determine binding specificity of an antibody of the invention are well known and routinely practiced in the art. For a comprehensive discussion of such assays, see Harlow et al. (Eds.), Antibodies A Laboratory Manual; Cold Spring Harbor Laboratory; Cold Spring Harbor, NY (1988), Chapter 6.

Non-human antibodies may be humanized by any method known in the art. A preferred "humanized antibody" has a human constant region, while the variable region, or at least a complementarity-determining region (CDR), of the antibody is derived from a non-human species. Methods for humanizing non-human antibodies are well known in the art. (*see* U.S. Patent Nos. 5,585,089, and 5,693,762). Generally, a humanized antibody has one or more amino acid residues introduced into its framework region from a source which is non-human. Humanization can be performed, for example, using methods described in Jones *et al.* [*Nature* 321: 522-525, (1986)], Riechmann *et al.*, [*Nature*, 332: 323-327, (1988)] and Verhoeyen *et al.* [*Science* 239:1534-1536, (1988)], by substituting at least a portion of a rodent CDR for the corresponding regions of a human antibody. Numerous techniques for preparing engineered antibodies are described, *e.g.*, in Owens and Young, *J. Immunol. Meth., 168*:149-165 (1994). Further changes can then be introduced into the antibody framework to modulate affinity or immunogenicity.

The invention further provides a hybridoma that produces an antibody according to the invention. Antibodies of the invention are useful for detection and/or purification of the polypeptides of the invention.

Polypeptides and/or polynucleotides of the invention may also be used to immunize animals to obtain polyclonal and monoclonal antibodies which specifically react with the polypeptide. Such antibodies may be obtained using either the entire polypeptide or fragments thereof as an immunogen. The peptide immunogens additionally may contain a cysteine residue at the carboxyl terminus, and may be conjugated to a hapten such as keyhole limpet hemocyanin (KLH). Methods for synthesizing such peptides are known in the art, for example, as in R. P. Merrifield, *J. Amer. Chem. Soc. 85*:2149-2154 (1963); J. L. Krstenansky, et al., *FEBS Lett.* 211: 10 (1987). Monoclonal antibodies binding to the protein of the invention may be useful diagnostic agents for the immunodetection of the polypeptide. Neutralizing monoclonal antibodies binding to the polypeptide may also be useful therapeutics for both conditions associated with the polypeptide and also in the treatment of some forms of cancer where abnormal expression of the polypeptide is involved. In the case of cancerous cells or leukemic cells, neutralizing monoclonal antibodies against the polypeptide are useful in detecting and preventing the metastatic spread of the cancerous cells mediated by the polypeptide. In general, techniques for preparing polyclonal and monoclonal antibodies as well as hybridomas capable of producing the desired antibody are well known in the art (Campbell, A. M., Monoclonal Antibodies Technology: Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers, Amsterdam, The Netherlands (1984); St. Groth et al., *J. Immunol. 35*:1-21 (1990); Kohler and Milstein, *Nature 256*:495-497 (1975)), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., *Immunology Today 4*:72 (1983); Cole et al., in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc. (1985), pp. 77-96).

Any animal (mouse, rabbit, and the like) which is known to produce antibodies can be immunized with a peptide or polypeptide of the invention. Methods for immunization are well known in the art. Such methods include subcutaneous or intraperitoneal injection of the polypeptide. One skilled in the art will recognize that the amount of the polypeptide encoded by an ORF of the invention used for immunization will vary based on the animal which is immunized, the antigenicity of the peptide and the site of injection. The protein that is used as an immunogen may

be modified or administered in an adjuvant in order to increase the protein's antigenicity. Methods of increasing the antigenicity of a protein are well known in the art and include, but are not limited to, coupling the antigen with a heterologous protein (such as a globulin or β-galactosidase) or through the inclusion of an adjuvant during immunization.

For monoclonal antibodies, spleen cells from the immunized animals are removed, fused with myeloma cells, such as SP2/0-Ag14 myeloma cells, and allowed to become monoclonal-antibody-producing hybridoma cells. Any one of a number of methods well known in the art can be used to identify the hybridoma cell which produces an antibody with the desired characteristics. These include screening the hybridomas with an ELISA assay, Western blot analysis, or radioimmunoassay (Lutz *et al., Exp. Cell Research.* 175:109-124. 1988). Hybridomas secreting the desired antibodies are cloned and the class and subclass is determined using procedures known in the art (Campbell, A. M., Monoclonal Antibody Technology: Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers, Amsterdam, The Netherlands (1984)). Techniques described for the production of single-chain antibodies (U.S. Pat. No. 4,946,778) can be adapted to produce single-chain antibodies to polypeptide of the invention.

For polyclonal antibodies, antibody-containing antiserum is isolated from the immunized animal and is screened for the presence of antibodies with the desired specificity using one of the above-described procedures. The invention further provides the above-described antibodies in detectably labeled form. Antibodies can be detectably labeled through the use of radioisotopes, affinity labels (such as biotin, avidin, and the like), enzymatic labels (such as horseradish peroxidase, alkaline phosphatase, and the like) fluorescent labels (such as FITC or rhodamine, and the like), paramagnetic atoms, and the like. Procedures for accomplishing such labeling are well-known in the art; for example, see Sternberger, L. A. et al., *J. Histochem. Cytochem. 18*:315. 1970; Bayer, E. A. et al., *Meth. Enzym. 62*:308 (1979); Engval, E. et al., *Immunol. 109*:129. 1972; and Goding, J. W. *J. Immunol. Meth. 13*:215. (1976).

The labeled antibodies of the present invention can be used for *in vitro*, *in vivo*, and *in situ* assays to identify cells or tissues in which a fragment of the polypeptide of interest is expressed. The antibodies may also be used directly in therapies or other diagnostics. The present invention further provides the above-described antibodies immobilized on a solid support. Examples of such solid supports include plastics such as polycarbonate, complex carbohydrates such as agarose and sepharose, and acrylic resins such as polyacrylamide and latex beads. Techniques for coupling antibodies to such solid supports are well known in the art (Weir, D. M. et al., "Handbook of Experimental Immunology" 4th Ed., Blackwell Scientific Publications, Oxford, England, Chapter 10 (1986); Jacoby, W. D. et al., Meth. Enzym. 34 Academic Press, N.Y. (1974)). The immobilized antibodies of the present invention can be used for *in vitro*, *in vivo*, and *in situ* assays as well as for immuno-affinity purification of the proteins of the present invention.

Computer-Readable Sequences

In one application of this embodiment, a nucleotide sequence of the present invention can be recorded on computer-readable media. As used herein, "computer-readable media" refers to any medium which can be read and accessed directly by a computer. Such media include, but are not limited to, magnetic storage media, such as floppy discs, hard disc storage medium, and magnetic tape; optical storage media such as CD-ROM; electrical storage media such as RAM and ROM; and hybrids of these categories such as magnetic/optical storage media. A skilled artisan can readily appreciate how any of the presently known computer-readable media can be used to create a manufacture comprising computer-readable medium having recorded thereon a nucleotide sequence of the present invention. As used herein, "recorded" refers to a process for storing information on computer-readable medium. A skilled artisan can readily adopt any of the presently known methods for recording information on a computer-readable medium to generate manufactures comprising the nucleotide sequence information of the present invention.

A variety of data storage structures are available to a skilled artisan for creating a computer-readable medium having recorded thereon a nucleotide sequence

of the present invention. The choice of the data storage structure will generally be based on the means chosen to access the stored information. In addition, a variety of data processor programs and formats can be used to store the nucleotide sequence information of the present invention on computer readable medium. The sequence information can be represented in a word processing text file, formatted in commercially-available software such as WordPerfect and Microsoft Word, or represented in the form of an ASCII file, stored in a database application, such as DB2, Sybase, Oracle, or the like. A skilled artisan can readily adapt any number of data processor structuring formats (*e.g.*, text file or database) in order to obtain computer readable medium having recorded thereon the nucleotide sequence information of the present invention. By providing the nucleotide sequence of SEQ ID NO: 1-30, 45, 47, 49 and 51 or a representative fragment thereof, or a nucleotide sequence at least 99.9% identical to SEQ ID NO: 1-30, 45, 47, 49 and 51 in computer-readable form, a skilled artisan can routinely access the sequence information for a variety of purposes. Computer software is publicly available which allows a skilled artisan to access sequence information provided in a computer-readable medium. The examples which follow demonstrate how software which implements the BLAST (Altschul et al., *J. Mol. Biol. 215*:403-410. 1990) and BLAZE (Brutlag et al., *Comp. Chem. 17*:203-207 (1993)) search algorithms on a Sybase system is used to identify open reading frames (ORFs) within a nucleic acid sequence. Such ORFs may be protein-encoding fragments and may be useful in producing commercially important proteins such as enzymes used in fermentation reactions and in the production of commercially useful metabolites.

As used herein, "a computer-based system" refers to the hardware means, software means, and data storage means used to analyze the nucleotide sequence information of the present invention. The minimum hardware means of the computer-based systems of the present invention comprises a central processing unit (CPU), input means, output means, and data storage means. A skilled artisan can readily appreciate that any one of the currently available computer-based systems are suitable for use in the invention. As stated above, the computer-based systems of the present invention comprise a data storage means having stored therein a nucleotide

sequence of the present invention and the necessary hardware means and software means for supporting and implementing a search means. As used herein, "data storage means" refers to memory which can store nucleotide sequence information of the present invention, or a memory access means which can access manufactures having recorded thereon the nucleotide sequence information of the present invention.

As used herein, "search means" refers to one or more programs which are implemented on the computer-based system to compare a target sequence or target structural motif with the sequence information stored within the data storage means. Search means are used to identify fragments or regions of a known sequence which match a particular target sequence or target motif. A variety of known algorithms are disclosed publicly and a variety of commercially available software for conducting search means are and can be used in the computer-based systems of the present invention. Examples of such software includes, but is not limited to, MacPattern (EMBL), BLASTN and BLASTA (NPOLYPEPTIDEIA). A skilled artisan can readily recognize that any one of the available algorithms or implementing software packages for conducting homology searches can be adapted for use in the present computer-based systems. As used herein, a "target sequence" can be any nucleic acid or amino acid sequence of six or more nucleotides or two or more amino acids. A skilled artisan can readily recognize that the longer a target sequence is, the less likely a target sequence will be present as a random occurrence in the database. The most preferred sequence length of a target sequence is from about 10 to 100 amino acids or from about 30 to 300 nucleotide residues. However, it is well recognized that searches for commercially important fragments, such as sequence fragments involved in gene expression and protein processing, may be of shorter length.

As used herein, "a target structural motif," or "target motif," refers to any rationally selected sequence or combination of sequences in which the sequence(s) are chosen based on a three-dimensional configuration which is formed upon the folding of the target motif. There are a variety of target motifs known in the art. Protein target motifs include, but are not limited to, enzyme active sites and signal sequences. Nucleic acid target motifs include, but are not limited to, promoter

sequences, hairpin structures and inducible expression elements (protein binding sequences).

Diagnostic Assays and Kits

The present invention further provides diagnostic assays, and related kits, for hyper- and/or hypo-proliferative disorders or diseases of endothelial cells such as LECs or BECs. These assays comprise methods to identify the presence or expression of one of the ORFs of the present invention, or homolog thereof, in a test sample, using a nucleic acid probe or an antibody according to the invention.

In general, methods for detecting a polynucleotide of the invention can comprise contacting a sample with a compound that binds to and forms a complex with, the polynucleotide for a period sufficient to form the complex, and detecting the complex, so that if a complex is detected, a polynucleotide of the invention is detected in the sample.

Such methods can also comprise contacting a sample under stringent hybridization conditions with nucleic acid primers that anneal to a polynucleotide of the invention under such conditions, and amplifying annealed polynucleotides, so that if a polynucleotide is amplified, a polynucleotide of the invention is detected in the sample.

In general, methods for detecting a polypeptide of the invention can comprise contacting a sample with a compound that binds to and forms a complex with the polypeptide for a period sufficient to form the complex, and detecting the complex, so that if a complex is detected, a polypeptide of the invention is detected in the sample. In detail, such methods comprise incubating a test sample with one or more of the antibodies or one or more of the nucleic acid probes of the invention and assaying for binding of the nucleic acid probes or antibodies to components within the test sample.

Conditions for incubating a nucleic acid probe or antibody with a test sample vary. Incubation conditions depend on the format employed in the assay, the detection methods employed, and the type and nature of the nucleic acid probe or antibody used in the assay. One skilled in the art will recognize that any one of the

commonly available hybridization, amplification or immunological assay formats can readily be adapted to employ the nucleic acid probes or antibodies of the present invention. Examples of such assays can be found in Chard, T., An Introduction to Radioimmunoassay and Related Techniques, Elsevier Science Publishers, Amsterdam, The Netherlands (1986); Bullock, G. R. et al., Techniques in Immunocytochemistry, Academic Press, Orlando, Fla. Vol. 1 (1982), Vol. 2 (1983), Vol. 3 (1985); Tijssen, P., Practice and Theory of immunoassays: Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers, Amsterdam, The Netherlands (1985). The test samples of the present invention include cells, protein or membrane extracts of cells, or biological fluids such as sputum, blood, serum, plasma, or urine. The test sample used in the above-described method will vary based on the assay format, nature of the detection method and the tissues, and cells or extracts used as the sample to be assayed. Methods for preparing protein extracts or membrane extracts of cells are well known in the art and can be readily be adapted in order to obtain a sample which is compatible with the system utilized.

In another embodiment of the present invention, kits are provided which contain the necessary reagents to carry out the assays of the present invention. In one embodiment, the invention provides a compartment kit to receive, in close confinement, one or more containers which comprises: (a) a first container comprising one of the probes or antibodies of the present invention; and (b) one or more other containers comprising one or more of the following: wash reagents, reagents capable of detecting presence of a bound probe or antibody.

In detail, a compartment kit includes any kit in which reagents are contained in separate containers. Such containers include small glass containers, plastic containers or strips of plastic or paper. Such containers allows one to efficiently transfer reagents from one compartment to another compartment such that the samples and reagents are not cross-contaminated, and the agents or solutions of each container can be added in a quantitative fashion from one compartment to another. Such containers will include a container which will accept the test sample, a container which contains the antibody or antibodies used in the assay, containers which contain wash reagents (such as phosphate-buffered saline, Tris buffers, and the

like), and containers which contain the reagents used to detect the bound antibody or probe. Types of detection reagents include labeled nucleic acid probes, labeled secondary antibodies, or in the alternative, if the primary antibody is labeled, the enzymatic, or antibody binding reagents which are capable of reacting with the labeled antibody. One skilled in the art will readily recognize that the disclosed probes and antibodies of the present invention can be readily incorporated into one of the established kit formats which are well known in the art.

EXAMPLES

Methods used in the examples are as follows:

Antibodies

Monoclonal antibodies against human VEGFR-3 (clone 2E11D11; see International Patent Application No. PCT/US02/22164, published as WO 03/006104), PAL-E (Monosan), CD31 (Dako), N-cadherin, VE-cadherin, ß-catenin and plakoglobin and polyclonal rabbit anti-human podoplanin were used (Breiteneder-Geleff, S., et al., *Am. J. Pathol. 154*:385-394 (1999)). Mouse anti-human integrin α9 was provided by Dr. Dean Sheppard (University of California at San Francisco, San Francisco) and Dr. Curzio Rüegg (University of Lausanne Medical School, Lausanne, Switzerland). The fluorochrome-conjugated secondary antibodies were obtained from Jackson Immunoresearch.

Cell Culture and Transfection

Human amniotic epithelial cells were cultured in Med199 medium in the presence of 5% fetal calf serum. Human dermal microvascular endothelial cells were obtained from PromoCell (Heidelberg, Germany). Anti-Podoplanin antibodies, MACS colloidal super-paramagnetic MicroBeads conjugated to goat anti-rabbit IgG antibodies (Miltenyi Biotech, Bergisch Gladbach, Germany), LD and MS separation columns and Midi/MiniMACS separators (Miltenyi Biotech) were used for cell sorting according to the instructions of the manufacturer. The isolated cells were

cultured on fibronectin-coated (10 μg/ml, Sigma, St. Louis, MO) plates as described (Mäkinen, T., *et al.*, *EMBO J. 20*:4762-4773. 2001).

RNA isolation, Northern blotting and microarray analyses

Total RNA was isolated and DNAseI treated in RNeasy columns (Qiagen, Valencia, CA). ^{32}P-labeled probes for hybridization with the Atlas filters (Clontech) were prepared using 2-5 μg of total RNA according to the manufacturer's instructions with the exception that the probe was purified using Nick-25 columns (Pharmacia Biotech, Uppsala, Sweden). Following hybridizations and washes, the membranes were analyzed using a Fuji BAS 100 phosphoimager. For the Affymetrix® analysis, four independent BEC and LEC sample preparations and hybridizations were carried out using RNA extracted from four lots of cells isolated from different individuals. For the Affymetrix® expression analysis, 5 μg of total RNA was used for the synthesis of double-stranded cDNA using Custom SuperScript ds-cDNA Synthesis Kit (Invitrogen, Carlsbad, CA). Biotin-labeled cRNA was then prepared using the Enzo BioArrayTMHighYieldTMRNA Transcript Labelling Kit (Affymetrix, Santa Clara, CA), and the unincorporated nucleotides were removed using RNeasy columns (Qiagen, Valencia, CA). The hybridization, washing and staining of Human Genome 95Av2 microarrays (for Prox-1 experiments) and 9513-E microarrays, which mainly contain uncharacterized EST sequences, were done according to the instructions of the manufacturer (Affymetrix, GeneChip Expression Analysis Technical Manual). The probe arrays were scanned at 570 nm using an Agilent GeneArray® Scanner and the readings from the quantitative scanning were analyzed by the Affymetrix® Microarray Suite version 5.0 and Data Mining Tool version 3.0. For the comparison analyses, the hybridization intensities were calculated using a global scaling intensity of 100.

The differentially expressed sequences were used for searching EST contigs in the GenBank database of the National Center for Biotechnology Information and the National Library of Medicine. (NCBI/NLM), and open reading frames were predicted using the orf finder software available at NCBI/NLM. The SOSUI system was used for prediction of transmembrane helices and signal

sequences from the protein sequences, and other protein domain architectures were analysed using Pfam (Protein families database of alignments and HMMs).

Immunofluorescence and Immunohistochemistry

The cells were cultured on coverslips, fixed with 4% paraformaldehyde, permeabilized with 0.1% Triton-X100 in phosphate-buffered saline (PBS) and stained with the primary antibodies. For integrin α9, staining live cells were incubated with the antibody for 15 minutes on ice before fixation. The cells were further stained with FITC- or TRITC-conjugated secondary antibodies. F-actin was stained using TexasRed-conjugated phalloidin (Molecular Probes, Eugene, OR). Cells were counterstained with Hoechst 33258 fluorochrome (Sigma) and viewed using a Zeiss Axioplan 2 fluorescent microscope.

Normal human skin obtained after surgical removal was embedded in Tissue-Tek® (Sakura, The Netherlands), frozen and sectioned. The sections (6 μm) were fixed in cold acetone for 10 minutes and stained with the primary antibodies followed by peroxidase staining using Vectastain Elite ABC kit (Vector Laboratories, Burlingame, CA) and 3-amino-9-ethyl carbazole (Sigma, St. Louis, MO).

EXAMPLE 1

IDENTIFICATION OF DIFFERENTIALLY EXPRESSED GENES

Blood vascular and lymphatic endothelial cells (BEC and LEC, respectively) were isolated from cultures of human dermal microvascular endothelial cells using magnetic microbeads and antibodies against the lymphatic endothelial cell surface marker podoplanin (Breiteneder-Geleff, S., et al., *Am. J. Pathol. 154*:385-394 (1999); Mäkinen, T., et al., *EMBO J. 20*:4762-4773 (2001)). The purities of the isolated BEC and LEC populations were confirmed to be over 99% as assessed by immunofluorescence using antibodies against VEGFR-3 or podoplanin. The isolated cells were cultured for a couple of passages, and RNA was extracted from the cultures and used for hybridization with oligonucleotide microarrays containing sequences from about 12,000 known genes, *i.e.*, approximately 1/3 of the total number of all predicted human transcripts.

As expected, podoplanin, desmoplakin I/II and the macrophage mannose receptor, which are known lymphatic endothelial cell markers, were found specifically in the LECs. See, Breiteneder-Geleff, S., et al., *Am. J. Pathol. 154*:385-394 (1999); Ebata, N., et al., *Microvasc. Res. 61*:40-48. (2001); and Irjala, H., et al., *J. Exp. Med. 194*:1033-1041 (2001). Since these results were consistent with the known gene expression patterns *in vivo* and *in vitro*, further characterization of the gene expression profiles was carried out. When a reproducible signal $\log_2$ ratio of 1.0 (twofold difference) was selected in the replicate analyses, over 400 genes were found to be differentially expressed between LECs and BECs. Some examples of the differentially expressed genes have been functionally annotated in Table 1 and a complete list of the differentially expressed genes is provided in Tables 2-4. A complete list of differentially expressed genes containing the GenBank accession numbers and the variation between the expression levels between independently harvested BECs and LECs (signal $\log_2$ ratio ±s.d.) are provided in Tables 3 and 4. The microarray data were validated by Northern blotting or by immunofluorescence for 31 of the selected genes (see Figure 1).

Each gene listed in Tables 3 and 4 is identified by a gene accession number which correlates to the sequence of the gene as found in a public genome database such as the GenBank database maintained by NCBI. These sequences are incorporated herein by reference.

Table 1

Selected classes of genes differentially expressed in BECs and LECs.

	Blood vascular EC	Lymphatic EC
Adhesion molecules	**integrin alpha5** integrin ß5, ß4* **ICAM-1***, ICAM-2 **N-cadherin*** selectin P, selectin E* protocadherin 42* CD44* EphrinB1*	**integrin alpha9*** **integrin alpha1** macrophage mannose receptor I*

	Blood vascular EC	Lymphatic EC
Cytoskeletal proteins	**vinculin** claudin 7* actin, alpha 2 profilin 2	**desmoplakin I and II*** **adducin gamma** alpha-actinin-2 associated LIM protein *
ECM proteins	**collagens 8A1***, 6A1*, 4A2/13A1*, 1A2* laminin* versican* proteoglycan 1	**Matrix Gla protein***
ECM modulation	**MMP-1**, MMP-10, MMP-14* uPA*, tPA* cathepsin C	**TIMP-3**
Receptor tyrosine kinases and other protein kinases	VEGFR-1 (sVEGFR-1*)	VEGFR-3* **Lyn** Dyrk3
Transcription factors	**STAT6*** TFEC* MAD-3* HMGI-C* JUN* GATA2	**prox-1*** **MEF2C*** c-maf* forkhead box M1 CREM ear-3
Growth factors	VEGF-C* **Placenta growth factor**	**Angiopoietin-2**
Cytokines, chemokines and receptors	**IL-8***, IL-6* **stem cell factor*** Monocyte chemotactic protein 1 **UFO/axl*** **CXCR4** CCRL2/CKRX* IL-4 receptor	IL-7* SDF-1b*
Cell cycle	p27* p21 gadd45	**Cdk-inhibitor p57KIP2*** cyclin-dependent kinase inhibitor 3, CIP2 cyclin E2* cyclin B1, B2*
Oxidative stress	thioredoxin reductase beta*	selenoprotein P*

	Blood vascular EC	Lymphatic EC
Other	Neuropilin-1 HNMP-1* endothelial cell protein C/APC receptor Rnase A, pancreatic* TGF-ß LTBP-2 metallothionein I, II, III Cyclooxygenase 2* clusterin/Apolipoprotein J neuronal pentraxin I*	**podoplanin*** MRC OX2 **Apolipoprotein D** Semaphorin 3A* fatty acid binding protein 4 LITAF/Pig7* IGFBP-2* piccolo* monoamine oxidase A neuronal pentraxin II*
Total	**222 genes**	**187 genes**

Genes shown in bold were confirmed by Northern blotting or immunofluorescence, and those marked with an asterisk (*) were specifically expressed in only one of the two cell lineages.

Table 2

Known LEC-specific genes

Gene	Detection *	Accession numbers starting EST	Accession numbers possible gene
CD36 =COL1/TSP receptor, fatty-acid transport protein	Af (S/4,3)	R20784 H54254	M98399
beta1-syntrophin	Af (S/4,5)	AA447177	L31529
collectin sub-family member 12	Af (S/4,5)	R74387	NM_030781
a disintegrin and metalloprotease domain 12	Af (S/4,3)	AA147933	NM_003474
cytotoxic T-lymphocyte-associated protein 4	Af (S/4,0)	AI733018	NM_005214
niban protein NM_022083 niban protein	Af (S/3,7)	AA554814	NM_052966
multi-PDZ-domain-containing protein, LNX	Af (S/3,5)	AI738919	NM_032622
MAGE-E1 protein	Af (S/3,2)	AI435112	NM_030801
upstream stimulatory factor 1, USF1 (genomic match)	Af (S/2,6)	AA701033	AB017568
hairy/enhancer-of-split related with YRPW motif 1	Af (NS/2,6)	R61374	NM_012258
alpha-2,8-polysialyltransferase	Af (S/2,5)	AI422986	L41680
semaphorin 6A1	Af (S/2,4)	W21965	NM_020796
guanine nucleotide binding protein (G prot), gamma 2	Af (S/2,3)	AA738022	
integral membrane protein 3	Af (S/2,3)	AA128019	NM_030926
similar to mouse glucocorticoid-induced gene 1	Af (S/2,0)	AI678080	XM_070471
YAP65 (Yes-associated protein of 65 kDa MW)	Af (NS/2,0)	AL048399	X80507
17 kDa fetal brain protein	Af (NS/1,9)	H92988	NM_022343
Kruppel-like factor 5	Af (S/1,8)	AI815057	NM_001730
calcitonin receptor-like, CGRP type 1 receptor	Af (S/1,7)	AI741128, T94540	NM_005795, L76380
fibroblast growth factor 13, isoform 1A	Af (NS/1,7)	AW014749	NM_004114
tetraspan NET-6 protein	Af (NS/1,6)	W22687	NM_014399
ring finger protein 11	Af (S/1,6)	AL079648	BC020964

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent $\log_2$ ratio of the signal intensities between BEC and LEC

EXAMPLE 2

BEC-SPECIFIC EXPRESSION OF GENES INVOLVED IN INFLAMMATION

Endothelial cells play an important role in several steps of the inflammatory response. They recruit leukocytes to inflammatory foci and specialized endothelial cells (high endothelial venules) are responsible for the homing of lymphocytes to the secondary lymphoid organs. In addition, endothelial cells modulate leukocyte activation and vice versa, and they can become activated by molecules secreted by the leukocytes. Consistent with their activation in cell culture, the BECs expressed high levels of pro-inflammatory cytokines and chemokines (stem cell factor, interleukin-8, monocyte chemotactic protein 1 (MCP-1)) and receptors (UFO/axl, CXCR4, IL-4R) see Table I. CXCR4 and its ligand, stromal cell-derived factor-1 (SDF-1), play important roles in the trafficking of normal lymphocytes, monocytes, and hematopoietic stem- and progenitor cell; targeted inactivation of either CXCR4 or SDF-1 results in impaired cardiogenesis, hematopoiesis and vascular development (Tachibana, *et al.*, *Nature 393*:591-594. 1998). SDF-1b was mainly produced by the LECs, suggesting that this chemokine may be involved in LEC-initiated chemotaxis of the CXCR4-expressing cells. Moreover, the reciprocal pattern of expression of CXCR4 and SDF-1 on BECs and LECs suggest that the two cell types use these molecules for paracrine communication.

EXAMPLE 3

DIFFERENCES IN CELL ADHESION, CELL-CELL INTERACTION AND CYTOSKELETAL MOLECULES

The most striking differences detected between the BECs and LECs was the expression of genes involved in cytoskeletal and cell-cell or cell-matrix interactions (see Tables 3 and 4). For example, N-cadherin, which is involved in the interaction of endothelial cells with SMCs and pericytes (Gerhardt, *et al.*, *Dev. Dyn. 218*:472-479. 2000), was detected specifically in BECs. This is consistent with the fact that the lymphatic capillaries are not ensheathed by SMCs. In immunostaining, N-cadherin was detected exclusively in the BECs, whereas VE-cadherin was present

in both cell types (Figure 2*a-d*). The cytoplasmic domains of cadherins interact with ß-catenin, plakoglobin (γ-catenin) and p120^{ctn}, which link them to the actin cytoskeleton via α-actinin, vinculin, ZO-1, ZO-2 and spectrin (Provost, E. & Rimm, *Curr. Op. Cell Biol. 11*:567-572. 1999). BECs expressed significantly higher levels of ß-catenin (Figure 2*e,f*) and vinculin, whereas plakoglobin was mostly present on LECs (Figure 2*g,h*). Staining of LECs and BECs also revealed a strikingly different organization of the actin cytoskeleton. BECs displayed numerous stress fibers, which in LECs were almost totally absent, and instead a cortical distribution of actin was observed in LECs(Figure 2*i,j*).

Integrins are important mediators of cell adhesion (Giancotti & Ruoslahti, *Science 285*:1028-1032. 1999). They are transmembrane proteins consisting of two polypeptides, the α and ß subunits. Their ectodomains bind extracellular matrix proteins while the cytoplasmic domains interact with the cytoskeleton and with proteins involved in signal transduction. Integrin α5, which acts as a subunit of the fibronectin receptor, mainly was expressed in BECs. By contrast, integrins α1 and α9, which provide subunits for the receptors for laminin and collagen and for osteopontin and tenascin, respectively, were expressed in LECs (Figure 1*a* and Figure 2*k,l*). In human skin, antibodies against integrin α9 stained lymphatic capillaries specifically, while blood vessel endothelia were negative (Figure 2*m-o*). In addition, integrin α9 was detected in arterial smooth muscle cells as previously reported (Palmer, *et al., J. Cell Biol. 123*:1289-1297. 1993). Interestingly, integrin α9 has been shown to be important for the normal development of the lymphatic system. Mice lacking integrin α9ß1 develop respiratory failure due to the accumulation of a milky pleural (presumably lymphatic) effusion and die within 6 to 12 days after birth (Huang, *et al., Mol. Cell Biol.* 20:5208-5215. 2000).

BECs, but not LECs, produced both laminin and different types of collagens (Table 4). In co-culture these basement membrane components may be necessary for the adhesion and growth of the LECs (Mäkinen, T., *et al., EMBO J. 20*:4762-4773. 2001). In addition, many of the proteins involved in matrix degradation and remodeling, including several matrix metalloproteinases, tissue-type and urokinase plasminogen activator, as well as plasminogen activator inhibitor I were

detected mainly in BECs, while the tissue inhibitor of matrix metalloproteinases-3 (TIMP-3) was detected mainly in LECs (Table 3 and Figure 1). Unlike the other TIMPs, which are soluble, TIMP-3 is a component of the extracellular matrix. Recombinant TIMP-3 has been reported to inhibit endothelial cell migration and tube formation in response to angiogenic factors, and when expressed in a tumor model, it inhibited tumor growth most likely by preventing tumor expansion, release of growth factors from the extracellular matrix, or angiogenesis (Anand-Apte, *et al.*, *Biochemistry & Cell Biology 74*:853-862. 1996).

Additional previously unknown genes were identified in the microarray as LEC-specific transcription factors or transmembrane proteins. See Tables 5 and 6.

Table 5

Transcription Factors Identified

Gene	Detection *	Accession numbers starting EST	possible gene
Homologous to Iroquois related homeobox 2	Af (S/4,2)	AA936528	not cloned from human (
similar to mouse odd-skipped related 1 zinc-finger TF	Af (S/3,3)	AI809953	(19)
PAC clone RP4-751H13 from 7q35-qter	Af (S/2,3)	AC004877	
similar to mouse glucocorticoid-induced gene 1	Af (NS/2)	AI678080	XM_070471

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent log2 ratio of the signal intensities between BEC and LEC

Table 6

Transmembrane Proteins Identified

Gene	Detection *	Accession numbers starting EST	possible gene	
KIAA0626	Af (S/4,7)	AB014526	NM_021647	(14)
KIAA0644	Af (S/3,9)	AB014544	NM_014817	(15)

unknown protein	Af (S/3,5)	AI333655	XM_059074	(16)
hypothetical protein FLJ20898	Af (NS/1,8)	AI733570	NM_024600	(862)
similar to layilin, unnamed protein product	Af (NS/1,7)	AA447940	AK055654, XM_084655	
hypothetical protein FLJ23403	Af (NS/3,2)	AI681538	NM_022068	(860)
KIAA0062		D31887	XM_046677	(47)
mesenchymal stem cell protein DSCD75	Af (S/1,8)	AW009871	NM_016647	(17)

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent log2 ratio of the signal intensities between BEC and LEC

Additionally, Tables 10 and 11 describe the known LEC genes identified and their accession numbers, and the differentially expressed genes and their accession numbers, respectively, while Table 12 describes other unknown proteins identified in the screen.

EXAMPLE 4

DIFFERENTIAL REGULATION OF LEC GENES BY PROX-1

The mechanisms responsible for the lymphatic differentiation program were investigated. The Prox-1 homeobox transcription factor was found to be expressed specifically in LECs and targeted disruption of Prox-1 in mice was reported to result in the arrest of lymphatic vessel development (Wigle *et al., Cell*, 98:769-778. 1999). Despite the fact that the *prox-1* gene was discovered nearly ten years ago, Prox-1 target genes have not been identified. To determine whether the homeodomain transcription factor Prox-1 contributes to the differentiated LEC and BEC phenotypes, the genes identified above were analyzed for expression in primary BECs and LECs, in the presence and absence of Prox-1 over-expression.

Adenovirus-mediated gene transfer of *prox-1* in primary endothelial cells was used to induce gene expression in the BEC cells. In order to eliminate gene expression changes caused by adenoviral infection, AdLacZ (encoding ß-galactosidase) was introduced into BECs as a control.

A *prox-1* cDNA was amplified by RT-PCR using total RNA from human endothelial cells and the primers 5'-

GCCATCTAGACTACTCATGAAGCAGCT-3' (SEQ ID NO: 61) and 5'-GCGCAGAATTCGGCCCTGACCATGACAGCACA-3' (SEQ ID NO: 62). The PCR product was cloned into the pAMC expression vector, producing N-terminally Myc-tagged Prox-1. The construct was then subcloned into pAdCMV to yield AdProx-1 for adenovirus production. AdProx-1 and AdLacZ virus stocks were produced as described (Laitinen *et al., Hum. Gene Ther.* 9:1481-1486. 1998). Adenovirally produced Prox-1 migrated with a molecular weight of about 85 kDa and it was also recognized by antibodies against a Prox-1 C-terminal peptide. Mutant Prox-1 N625A/R627A, (asparagine to alanine change at codon 625, arginine to alanine change at codon 627) was made using the QuikChange site-directed mutagenesis kit (Stratagene, La Jolla, CA) and the following primers:

5'-CTCATCAAGTGGTTTAGCGCTTTCCGTAGTTTTACTAC-3' (SEQ ID NO: 63) and

5'-GTAGTAAAACTCACGGAAGCGCTAAACCACTTGATGAG-3. (SEQ ID NO: 64).

Human dermal microvascular endothelial cells, coronary artery endothelial cells (CAECs), saphenous vein endothelial cells (SAVECs), BECs and LECs were plated 24 hours before adenoviral infection at a density of 8,000 cells/cm^2 and infected for 1 hour in serum-free medium at 50-100 PFU/cell. At the end of the incubation period the cells were washed and then cultured in complete medium for 20-24 hours. Total RNA isolation and array hybridization were performed as described above.

Titration experiments showed that infection of human microvascular endothelial cells with AdProx-1 or AdLacZ led to nuclear expression of the adenovirus-encoded protein in >90% of the cells at 24 hours post-infection. To investigate the changes in gene expression induced by Prox-1, human cDNA filter arrays were used, which contain about 1,000 genes known to be important for general cellular metabolism as wells as genes specifically implicated in the regulation of cardiovascular function or hematopoiesis. AdProx-1 up-regulated the expression of 28 LEC genes and down-regulated 63 BEC genes, (see Table 7 below), which was confirmed by Northern blotting for 10 of 11 selected genes. When compared with

genes differentially expressed in LECs and BECs, 15 genes (i.e., about 30%) modulated by Prox-1 were found to be differentially expressed between cultured LECs and BECs, suggesting that Prox-1 is a major regulator of lymphatic endothelial cell identity.

Table 7
Prox-1 Regulated LEC/BEC Genes

Gene	Accession numbers		[1] signal log ratio	[2] s.d.
LEC-specific induced by AdProx-1 (28 genes)				
cyclin E2	AF091433	NM_057735	4.95	1.17
cysteine and glycine-rich protein 2	U57646	NM_001321	4.58	0.36
Cdk-inhibitor p57KIP2	U22398	NM_000076	3.77	0.68
paternally expressed 10	AB028974	NM_015068	3.54	0.95
thromboxane A2 receptor	D38081	NM_001060	2.32	0.13
B-myb	X13293	NM_002466	2.11	0.28
retinoblastoma-associated protein HEC	AF017790	NM_006101	1.86	0.13
cholesterol 25-hydroxylase	AF059214	NM_003956	1.86	0.56
G protein-coupled receptor, family C, group 5, member B	AC004131		1.83	0.32
thymidine kinase 1	M15205	NM_003258	1.80	0.39
CREM (cAMP responsive element modulator)	S68134	NM_001881	1.78	0.30
alpha-actinin-2-associated LIM protein	AF002282	NM_014476	1.77	0.42
desmoplakin (DPI, DPII)	AL031058		1.74	1.03
MCM6 minichromosome maintenance deficient 6	D84557	NM_005915	1.72	0.06
erythrocyte membrane protein band 4.9 (dematin)	U28389	NM_001978	1.71	0.25
GTP cyclohydrolase 1	U19523	NM_000161	1.61	0.04
KIAA0186 gene product	D80008	NM_021067	1.47	0.11
cell division cycle 2 protein	X05360	NM_001786	1.35	0.43
hypothetical protein from clone 643	AF091087	NM_020467	1.25	0.22
ubiquitin carrier protein E2-C	U73379	NM_007019	1.23	0.12
mitotic checkpoint kinase Mad3L	AF053306	NM_001211	1.22	0.47
V-Erba Related Ear-3 Protein	HG3510-HT3704		1.20	0.20
glycogen phosphorylase (PYGL)	AF046798		1.16	0.54

Gene	Accession numbers		[1] signal log ratio	[2] s.d.
fms-related tyrosine kinase 4, VEGFR-3	X69878	NM_002020	1.10	0.00
BTB (POZ) domain containing 3	AB023169	NM_014962	1.10	0.08
SMC4 structural maintenance of chromosomes 4-like 1 (yeast)	AB019987	NM_005496	1.09	0.59
high-mobility group protein 2	X62534	NM_002129	1.07	0.04
alpha topoisomerase	L47276		1.04	0.49

Gene	Accession numbers		[1] signal log ratio	[2] s.d.
BEC-specific suppressed by AdProx-1 (63 genes)				
neuropilin-1	AF016050	NM_003873	-3.99	0.42
ras-related C3 botulinum toxin substrate 2, RAC2	M64595	NM_002872	-3.87	0.47
tripartite motif-containing 22	X82200	NM_006074	-3.56	0.28
small inducible cytokine A2 (monocyte chemotactic protein 1)	M26683	NM_002982	-3.56	0.03
zinc finger protein 238	AJ223321	NM_006352	-3.08	0.13
uPA	X02419		-3.05	0.02
transcription factor EC	D43945	NM_012252	-3.04	0.08
RNase A, pancreatic	D26129	NM_002933	-2.72	0.02
vitamin A responsive; cytoskeleton regulated	AF070523	NM_006407	-2.51	0.6
interleukin 6	X04430	NM_000600	-2.42	0.63
Rho GDP dissociation inhibitor (GDI) beta	X69549	NM_001175	-2.42	0.03
matrix metalloproteinase 14	X83535	NM_004995	-2.37	0.08
E3 ubiquitin ligase SMURF2	AA630312	NM_022739	-2.22	0.06
death receptor 6	AF068868	NM_014452	-2.16	0.61
protein C receptor, endothelial (EPCR)	L35545	NM_006404	-2.09	0.14

Gene	Accession numbers		[1] signal log ratio	[2] s.d.
hematopoietic and neural membrane protein (HNMP-1)	U87947	NM_001425	-2.08	0.63

Gene	Accession numbers		[1] signal log ratio	[2] s.d.
KIAA0836	AB020643		-2.07	0.44
chondroitin sulfate proteoglycan 2 (versican)	X15998	NM_004385	-1.99	0.65
regulator of G-protein signaling 4	AI267373	NM_005613	-1.93	0.54
phosphofructokinase, muscle	U24183	NM_000289	-1.93	0.11
IGF-II mRNA-binding protein 3	U97188	NM_006547	-1.9	0.23
neuronal cell adhesion molecule Nr-CAM/hBRAVO	AB002341	NM_005010	-1.89	0.13
cell surface glycoprotein CD44	L05424		-1.84	0.12
plasminogen activator inhibitor-1	J03764	NM_000602	-1.83	0.33
AF1Q protein	U16954	NM_006818	-1.79	0.23
Homo sapiens clone 24674 mRNA sequence	AF070578		-1.76	0.01
nicotinamide N-methyltransferase	U08021	NM_006169	-1.74	0.49
lactate dehydrogenase B	X13794		-1.73	0.08
KIAA0537 gene product	AB011109	NM_014840	-1.73	0.08
LIM domain protein	X93510	NM_003687	-1.67	0.11
lymphocyte antigen 75, DEC-205	AF011333	NM_002349	-1.61	0.08
natural killer cell transcript 4	AA631972	NM_004221	-1.59	0.05
phospholipase A2	M72393		-1.58	0.41
R-ras	M14949		-1.56	0.1
adenylyl cyclase-associated protein 2	N90755	NM_006366	-1.55	0.08
leupaxin	AF062075	NM_004811	-1.53	0.3
signal transducer and activator of transcription 6 (STAT6)	AF067575		-1.51	0.45
LYL-1	M22637		-1.51	0.14
selectin P	M25322	NM_003005	-1.47	0.37
protein kinase, cAMP-dependent, catalytic, beta	M34181	NM_002731	-1.43	0.49
TRAM-like protein	D31762	NM_012288	-1.42	0.43
guanylate binding protein 2, interferon-inducible	M55543	NM_004120	-1.41	0.51
intercellular adhesion molecule 2	X15606	NM_000873	-1.38	0.13
proteoglycan 1, secretory granule	X17042	NM_002727	-1.35	0.47

tropomyosin 1 (alpha)	Z24727	NM_000366	-1.32	0.1
fibroblast activation protein, alpha subunit	U09278	NM_004460	-1.25	0.12
hypothetical protein DKFZp564D0462	AL033377		-1.25	0.23
mitogen-activated protein kinase-activated protein kinase 3	U09578	NM_004635	-1.2	0.35
amyloid beta (A4) precursor protein-binding	U62325		-1.2	0.18
AXL receptor tyrosine kinase	M76125	NM_001699	-1.19	0.3
integrin alpha 5	X06256	NM_002205	-1.18	0.02
prion protein (PrP)	U29185		-1.18	0.07
TRAF family member-associated NFKB activator	U59863	NM_004180	-1.17	0.13
annexin VI	Y00097	NM_001155	-1.16	0.12
transcobalamin II	L02648	NM_000355	-1.16	0.12
sushi-repeat-containing protein, X chromosome	U61374	NM_006307	-1.13	0.09
bone morphogenetic protein 6	M60315	NM_001718	-1.13	0.39
hypothetical protein from clones 23549 and 23762	U90908	NM_021226	-1.1	0.6
retina cDNA randomly primed sublibrary, EST	W28438		-1.09	0.36
TU3A protein	AF035283		-1.06	0.29
keratin 7	AJ238246	NM_005556	-1.05	0.53
latent transforming growth factor beta binding protein 2	Z37976	NM_000428	-1.04	0.13
N-cadherin	M34064	NM_001792	-1.02	0.12
cDNA DKFZp564J0323 (from clone DKFZp564J0323)	AL049957		-1.01	0.22

[1] The change is expressed as the $\log_2$ ratio.

[2] Standard deviation of the change in the expression level.

The ability of recombinant Prox-1 expression in BECs (where it is normally absent) to modify the transcriptional program of these cells towards the lymphatic endothelial cell phenotype was also investigated. The control, AdLacZ, did not significantly alter the expression of BEC- or LEC-specific transcripts as determined by oligonucleotide microarray analyses. By contrast, AdProx-1 increased expression of many LEC-specific mRNAs, such as VEGFR-3, p57Kip2, desmoplakin I/II and alpha-actinin-associated LIM protein (see Table 8). Surprisingly, Prox-1 also suppressed the expression of about 40 % of genes characteristically expressed in BECs, such as the transcription factor STAT6, the UFO/axl receptor tyrosine kinase, neuropilin-1 (NRP-1), monocyte chemoattractant protein-1 (MCP-1) and integrin α5 (see Table 7 and Table 8). These gene expression results are in agreement with the *in vivo* studies of lymphatic vessels. For example, VEGFR-3 and desmoplakin I/II are found in the lymphatic endothelium (Ebata *et al., Microvasc. Res.* 61:40-48. 2001; Kaipainen *et al., Proc. Natl. Acad. Sci. U.S.A.* 92: 3566-70. 1995), and the VEGF co-receptor NRP-1, which was suppressed by Prox-1 in the BECs, was found to be expressed in blood vessels, but not in lymphatic vessels in mouse skin.

Table 8

Examples of LEC- and BEC-specific genes regulated by Prox-1

	LEC-specific, up-regulated	BEC-specific, down-regulated
Adhesion molecules		Integrin alpha 5 ICAM-2 CD44 Nr-CAM P-selectin
Cytoskeletal proteins	Desmoplakin I and II alpha-actinin-2 associated LIM protein	leupaxin
ECM proteins		versican proteoglycan 1
ECM proteins		versican proteoglycan 1

	LEC-specific, up-regulated	BEC-specific, down-regulated
ECM modulation		MMP-14 uPA PAI-I

Receptor tyrosine kinases	**VEGFR-3**	**UFO/axl**
Transcription factors	CREM ear-3	**STAT6**
Cytokines, chemokines and receptors		TFEC IL-6 MCP-1
Cell cycle control	**p57Kip2** **cyclin E2**	
Other	cholesterol 25-hydroxylase thromboxane A2 receptor	**Neuropilin-1** endothelial cell protein C receptor
Total	**28 genes** **(19% of LEC-specific genes)**	**63 genes** **(38% of BEC-specific genes)**

Genes shown in **bold** were confirmed by Northern blotting or RT-PCR.

In order to determine whether the Prox-1-induced changes in gene expression were cell-type specific, changes in gene expression after AdProx-1 or AdLacZ infection were analyzed in additional endothelial cell types, i.e., coronary artery endothelial cells (CAECs) and saphenous vein endothelial cells (SAVECs), as well as a non-endothelial cell type, i.e., amniotic epithelial cells (AEC). In all of these cell types, AdProx-1 strongly up-regulated Cyclins E1 and E2, Histone H2B, and PCNA. However, AdProx-1 induced VEGFR-3 expression only in CAECs and SAVECs, and not in AECs.

These results are consistent with the lack of lymphatic differentiation in Prox-1-deficient embryos. Interestingly, the expression of Prox-1 in primary endothelial cells leads to up-regulation of VEGFR-3 receptor tyrosine kinase, which is specific for the lymphatic endothelium after midgestation and is essential for proper lymphatic growth and function (Karkkainen and Petrova, *Oncogene* 19:5598-5605. 2000). For example, inactivating mutations of VEGFR-3 in humans and mice lead to lymphatic hypoplasia and lymphedema (Jeltsch *et al., Science* 276:1423-1425. 1997; Karkkainen *et al., Nat. Genet.*, 25:153-159. 2000; Karkkainen *et al, Trends Mol. Med.* 7:18-22. 2001). The results described above therefore suggest that the up-regulation of VEGFR-3 expression by Prox-1 is one of the key pathways involved in the establishment of lymphatic endothelial cell identity and also suggest that the distinct

phenotypes of cells in the adult vascular endothelium are plastic and sensitive to transcriptional reprogramming, which is useful in the therapeutic methods of the invention affecting endothelial cells.

EXAMPLE 5

EX-VIVO CELL STIMULATION AND GENE THERAPY FOR LYMPHEDEMA WITH ADPROX-1 TRANSFECTED CELLS

The ability of Prox-1 to regulate genes specifically involved in LEC development provides a means for treatment of individuals exhibiting a LEC disorder or condition resulting from either an increase or decrease in LEC gene expression levels. Prox-1 upregulation is useful in promoting LEC development as a treatment for LEC disorders characterized by an under-developed lymphatic system of a condition characterized by a risk of wider-development such as lymphedema. Conversely, Prox-1 inhibition is useful in downregulating LEC development as a treatment for LEC disorders characterized by an over-developed lymphatic system such as lymphedema. It is known in the art that *ex vivo* transfection of cells and subsequent transfer of these cells to patients is an effective method to upregulate *in vivo* levels of the specific gene transferred and to provide relief from a disease resulting from under-expression of the gene(s) (Gelse et al., *Arthritis Rheum.* 48:430-41. 2003; Huard *et al, Gene Ther.* 9:1617-26. 2002; Kim *et al., Mol. Ther.* 6:591-600 2002).

To develop a therapy for treating irregularities of LEC development, endothelial cells, such as CAECs, SAVECs, LECs or BECs, are isolated from individuals experiencing an LEC disorder (e.g. lymphedema) and then placed in an appropriate culture medium (see above) to promote the growth and viability of the cells. The cells are then transfected as described with the AdProx-1 vector as described above to initiate LEC differentiation of the non-LECs *in vitro* and to promote growth of the LECs in culture. These transfected cells are then transferred into an affected patient in therapeutically effective numbers to promote LEC expansion *in vivo*. In preferred embodiments, the manipulated cells are autologous cells. These cells are delivered by one or more administrations typically involving

injection. The cells are delivered at a local site of an LEC disease or disorder such as lymphedema or systemically.

Addition of the Prox-1 transfected cells to patients with lymphedema provides supplementary LECs that are incorporated into the lymphatic network to promote lymphatic development and effectuate lymph clearance to relieve the symptoms of lymphedema. It is contemplated that a method comprising AdProx-1 transfection into endothelial cells and administration of transfected cells is useful in the treatment of any disease characterized by an alteration in LEC numbers or activity, such as lymphedema, lymphangioma, lymphangiomyeloma, lymphangiomatosis, lymphangiectasis, lymphosarcoma, and lymphangiosclerosis. Additionally, such methods are useful in ameliorating a symptom (e.g., lymph-induced swelling in the case of lymphedema) associated with such diseases.

EXAMPLE 6

CHARACTERIZATION OF LEC-SPECIFIC GENES

LEC-specific genes were further analyzed using a subtraction library between the LEC and BEC genes. To construct the library, total RNA was isolated as previously described and 5μg of total RNA was pre-amplified using a SMART™ PCR cDNA synthesis kit (BD Biosciences Clontech). After RsaI-digestion, PCR-Select cDNA subtraction was carried out in both directions, resulting in selective amplification of differentially expressed sequences, and subtracted LEC and BEC cDNA libraries were prepared (BD Biosciences Clontech). Subtractive hybridization was performed with a 1 (tester): 30 (driver) ratio in both directions and subtracted cDNA pools were amplified by PCR. Forty ng of the purified PCR-amplified product were cloned into the pAtlas vector (PUC-based vector) for the construction of subtracted libraries, although a number of other vectors could be used in the construction, as would be known in the art.

Differential screening of the subtracted LEC-specific library was carried out as described in the PCR-Select Differential Screening Kit User Manual (BD Biosciences Clontech). The LEC-specific subtracted library was plated and individual bacterial clones were picked and grown. After DNA extraction, the inserts

were amplified by PCR and used for sequencing. An aliquot of each PCR-amplified insert was also arrayed onto a nylon membrane and used for hybridization with ^{32}P-labeled cDNA probes. The results from the hybridizations with subtracted LEC-specific (tester) and subtracted BEC-specific (driver) cDNA probes were used for the differential expression analyses.

BLAST (The Basic Local Alignment Search Tool) was used to compare the sequences against nucleotide, protein and EST sequence databases. For unknown sequences, EST contigs were searched and open reading frames were predicted using ORF finder. Protein domain architectures were analyzed using Pfam (Protein families database of alignments and HMMs) and Smart (Simple Modular Architecture Research Tool).

The nucleotide sequences of clones that were differentially expressed in LECs versus BECs were analyzed in the manner described above. Several of the EST or unknown gene fragments detected in the first screen have been investigated further to determine their sequence similarities to known gene sequences and to identify any open reading frames and functional domain similarities. The results are collected in Table 9.

Table 9

Clone designation and (SEQ ID NO)	EST	Human Genome Accession #	KIAA Designation # and (SEQ ID NO)	Expected Gene Function
LE000100001_A06 (SEQ ID NO: 61)	AB014526	NM_021647	KIAA0626 SEQ ID NO: 14	Ig domain motif, likely cell adhes function
LE0000100050_A01 (SEQ ID NO: 59) LE0000100055_H05	AB014544	NM_014817	KIAA0644 SEQ ID NO: 15	Leucine rich moti cell adhesion activity
	AI333655	XM_059074	no KIAA, designated hLyrp SEQ ID NO: 16	Leucine rich repeats, cell adhesion protein
	AI681538	NM_016647	SEQ ID NO: 17	Similar to mesenchymal stem cell protein
	AA447940	XM_084655	SEQ ID NO: 45	similar to layili likely cell adhes function
LE000100017_C02 (SEQ ID NO: 55)	XM_04667 7	D31887	KIAA0062 SEQ ID NO: 47	Zinc transporter motif, metal ion transport
LE0000100049_E10 LE0000100054_F09 LE0000100056_F07 SEQ ID NOs:	XM_04767 2	XM_047672	KIAA1673 SEQ ID NO: 26	RNA-binding regio similar to RNA binding protein
LE0000100053_A06 SEQ ID NO: 56	AI761647	NM_015147	KIAA0582 SEQ ID NO: 49	
LE0000100055_G10 LE0000100046_C12	D14657	NM_014736	KIAA0101 SEQ ID NO: 51	

SEQ ID NOs: 57-58

Several of the LEC-specific genes have been found to correspond to KIAA gene sequences, which are large nucleotide EST clones encoding unknown human proteins. (Kazusa DNA Research Institute, 1532-3, Yana Kisarazu, Chiba, 292-0812, Japan). These LEC-specific genes were further analyzed in several available databases to determine the existence of species homologs and the percent similarity in these homologs and also to reveal amino acid sequences that demonstrate similarity to conserved protein domains.

Analyses of the LEC clone sequences was performed using the HomoloGene database maintained by the U.S. National Center for Biotechnology Information offered by the National Institutes of Health to determine species homologs and orthologs and their percent similarity to the newly isolated human LEC-specific genes. Analyses of the sequences was performed using a resource of curated and calculated homologs for genes as represented by UniGene or by annotation of genomic sequences, generally comparing EST and mRNA sequences from UniGene, as well as transcripts extracted from annotated genomic sequences. (Zhang, *et al., J. Comp. Biol.* 7:203-14. 2000). The best match for a nucleotide sequence in one organism to a nucleotide sequence in a second organism is based on the degree of similarity between the two sequences, with a minimum alignment of 100 base pairs. The similarity between the two sequences was determined by an alignment score. The alignment score for a sequence pair is the sum of the similarity scores of the sections of the two sequences that aligned.

HomoloGene analyses indicate that human LEC genes corresponding to KIAA0626, KIAA0644, and KIAA0062, are homologous to EST and unknown gene sequences in mouse (all), rat (KIAA0062, KIAA0644), cow (KIAA0062), pig (KIAA0626, KIAA0644) and Xenopus (KIAA0644). The clones showed approximately 80% (± 3%) similarity to the genes identified as homologs by HomoloGene, with KIAA0644 demonstrating as high as 86% homology to pig EST sequence BE233028.1 and as low as 72% similarity to an *X. laevis* gene.

Analyses of the LEC genes using Pfam comparison revealed that nucleotide sequences corresponding to KIAA0626 (SEQ ID NO: 14), KIAA0644 (SEQ ID NO: 15), hLyrp (SEQ ID NO: 16), XM_084655 (SEQ ID NO: 45) and KIAA0062 (SEQ ID NO: 47), showed nucleotide sequence motifs characteristic of encoded transmembrane domains, indicating that the corresponding polypeptides (whose amino acid sequences are set out in SEQ ID NOS: 31, 32, 33, 46 and 48, respectively) are expressed on the cell surface. KIAA1673, KIAA0582 and KIAA0101 do not demonstrate an apparent transmembrane domain and are expected to be cytoplasmic or nuclear proteins. Tissue expression assayed by Northern blot reveals that KIAA0101 is detectable in kidney, thymus, colon and small intestine while KIAA0582 is expressed strongly in heart, skeletal muscle, and ovary, less in kidney and placenta, and more weakly in brain, lung, thymus, small intestine and prostate.

Northern blot analysis of the KIAA0626 transcript indicates that KIAA0626 is expressed specifically in LEC and is found in heart, skeletal muscle and kidney. *In situ* analysis demonstrates KIAA0626 expression in mouse embryonic day 11 (E11) embryos in the intersomitic tissue and pericytes surrounding the blood vessels, and in the yolk sac vessels, endothelial cells and in the surrounding pericytes. The polynucleotide sequence of KIAA0626 (SEQ ID NO: 14) encodes a 409 amino acid (409 aa) protein (SEQ ID NO: 31) possessing a signal sequence (at amino acids 1-29), an Ig superfamily domain (approximately aa 61-127), a short transmembrane region (about aa 153-175) and a long 234-amino-acid cytoplasmic domain from about amino acids 176-409. The presence of an Ig domain is expected to assist in binding of the protein to its ligand while the long cytoplasmic domain indicates that KIAA0626 may be involved in intracellular signaling in LECs.

KIAA0644 (SEQ ID NO: 15) is detected by Northern blot analysis primarily in heart and brain tissue. *In situ* assay of E10 mouse embryos shows KIAA0644 expression throughout the embryo. The KIAA0644 polynucleotide encodes a 811-amino-acid polypeptide (SEQ ID NO: 32) demonstrating a total of 13 leucine rich regions. Leucine-rich regions comprise a short sequence motif of approximately 20-28 amino acids which are present in proteins functioning as cell-

adhesion and receptor molecules. Leucine-rich regions, designated below as LRRNT and LRRCT are often flanked by cysteine-rich domains. The KIAA0644 protein contains a leucine-rich N-terminal region (LRRNT: aa 26-54), 11 internal leucine-rich regions (LRR1: aa84-107, LRR2: aa108-131, LRR3: aa132-155, LRR4: aa156-179, LRR5: aa180-203, LRR6: aa204-223, LRR7: aa230-253, LRR8: aa254-277, LRR9: aa278-301, LRR10: aa302-325, and LRR11: aa326-349) and a C-terminal leucine-rich region (LRRCT) from about amino acids 359-404. The KIAA0644 transmembrane domain spans approximately amino acids 696-718, leaving a cytoplasmic domain of about 95 amino acids, from aa719-811. The leucine-rich regions of the KIAA0644 gene implicate it in protein-protein interactions characteristic of cell-adhesion or ligand binding.

The hLyrp (SEQ ID NO: 16) mRNA is detectable in skeletal muscle tissue and is localized by *in situ* hybridization to the lymphatic vessels when compared to Prox-1 staining in E11 and yolk sac of mouse embryos. Similar to KIAA0644, the hLyrp protein (SEQ ID NO: 33) contains a series of leucine-rich regions beginning at the leucine-rich N-terminal region (LRRNT: aa27-55) extending through 5 internal leucine -rich regions (LRR1: aa57-80, LRR2: aa81-104, LRR3: aa105-128, LRR4: aa129-153, LRR5: aa154-176) and ending with a C-terminal leucine-rich region (LRRCT) from approximately aa186-240. The hLyrp polypeptide also contains a transmembrane domain from amino acids 249-272, leaving a short cytoplasmic domain of 22 amino acids. The presence of several consecutive leucine-rich regions in the hLyrp polypeptide indicates that the polypeptide functions as a cell-adhesion molecule and/or a cell surface receptor.

Several additional sequences shown in Table 3 were isolated with full-length mRNA sequences which are expressed specifically in LECs. Domain prediction of these sequences indicates that KIAA0711 (SEQ ID NO: 81 and 82) contains a BPB/POZ domain spanning approximately amino acids 171-269, this domain is expected to function in protein-protein interactions. POZ domains appear in transcriptional co-factors such as zinc-finger proteins that mediate transcriptional repression and interact with components of histone deacetylase complexes. KIAA0711 also has three Kelch repeats, spanning amino acids 386-437, 439-480, and

484-525, and Kelch motifs have been implicated in the formation of beta sheet structures. Additionally, KIAA0711 mRNA is expressed in a variety of tissues. From highest expression levels to lowest, KIAA0711 mRNA is found in brain and kidney; liver; spleen; lung; ovary, pancreas and heart; smooth muscle and testis. Because this expression pattern was obtained from a single run of RT-PCR ELISA, the expression profile has a chance to include significant run-to- run variations. Accordingly, the expression profiles are most suitable for screening genes for tissue-specific expression on a qualitative level. If more accurate quantitative expression profiles are required, more statistically reliable approaches should be employed (e.g., multiple RT-PCR-ELISA measurements, DNA chip analyses, RNA blot analyses, and the like).

Domain mapping of the sequence corresponding to cDNA DKFZp5640222 (SEQ ID NO: 93) indicates the presence of an N-terminal signal peptide (amino acids 1-23), two internal repeat domains and an olfactomedin domain (amino acids 361-616), which is detected in proteins such as myocilin, pancortin, and latrophilin. Mutations in the OLF domain of myocilin are associated with glaucoma.

Domain mapping of KIAA1233 (SEQ ID NO: 111) indicates that the KIAA sequence contains six thrombospondin type I repeats, which are found in extracellular matrix proteins and are implicated generally in cell-cell interactions, and more specifically in the complement pathway, in the inhibition of angiogenesis, and in apoptosis. KIAA1233 also contains three immunoglobulin C-2 type domains, similar to many glycoproteins. Proteins possessing both thrombospondin repeats and immunoglobulin domains are also involved in intracellular interactions, such as cell - adhesion and apoptosis. From highest expression levels to lowest, KIAA1233 mRNA is found in the spinal cord; heart, general brain, lung, liver, kidney, pancreas, various regions of the brain (amygdala, corpus callosum, caudate nucleus, hippocampus, substantia nigra, thalamus, and subthalamic nucleus) and fetal liver; fetal brain; spleen; and testis.

The KIAA0846 (SEQ ID NO: 188) protein contains motifs found in guanine nucleotide exchange factors and is thus probably an intracellular protein, perhaps a signaling protein. KIAA0846 also exhibits two EF-hand motifs found in signalling proteins (e.g. calmodulin, S100B), which undergo a calcium-dependent

conformational change and are also found in buffering/transport proteins. From highest expression levels to lowest, KIAA0846 mRNA is found in kidney; heart, brain and lung; liver, spleen and ovary; pancreas, smooth muscle and testis.

Protein FLJ13110 (SEQ ID NOS: 207 and 208) exhibits a TB2/DP1, HVA22 family protein domain and two short transmembrane regions (amino acids 4-22 and 43-65 of SEQ ID NO: 207). The HVA22 family includes members from a wide variety of eukaryotes, including the TB2/DP1 (deleted in severe familial adenomatous polyposis) protein which is deleted in severe forms of familial adenomatous polyposis, an autosomal dominant oncological inherited disease.

The LEC-specific gene screen also identified protein KIAA0937 (SEQ ID NOS: 211 and 212). KIAA0937 contains WWE domains (from approximately amino acids 30-112, and 113-189 of SEQ ID NO: 211) which is named after three of its conserved residues and is predicted to mediate specific protein-protein interactions in ubiquitin and ADP ribose conjugation systems. KIAA0937 is also predicted to contain a zinc finger domain (from amino acids 443-501 of SEQ ID NO: 211) and is expected to be an intracellular transcription factor. From highest expression levels to lowest, KIAA0937 mRNA is found in the spinal cord; the subthalamic nucleus and cerebellum of the brain; the brain in general (including the amygdale, corpus callosum and fetal brain) and ovary; fetal liver, heart, lung, kidney, spleen and parts of the brain (caudate nucleus and hippocampus); testis and pancreas; and smooth muscle.

KIAA0952 (SEQ ID NO: 241 and 242) contains a Broad-Complex, Tramtrack and a Bric-a-brac domain, also known as a POZ (poxvirus and zinc finger) domain. These domains are known to be protein-protein interaction domains found at the N-termini of several C2H2-type transcription factors, as well as Shaw-type potassium channels. The known structure of these domains reveals a tightly intertwined dimer formed via interactions between an N-terminal polypeptide strand and helix structures.

The protein designated KIAA0429 (SEQ ID NOS: 391 and 392) is similar to metastasis suppressor protein and contains an actin-binding WH2 domain from approximately amino acids 467-484, as well as a proline-rich region from amino acids 348-466.

Protein FLJ23403 (amino acid sequence, SEQ ID NO:859; polynucleotide sequence, SEQ ID NO:860) shows approximately 85% homology to an unknown mouse protein (GenBank Acc. No. XM_129000) and contains a series of four transmembrane domains spanning amino acids 44-66, 86-108, 115-137 and 452-474.

Additional LEC-specific, upregulated genes include previously unidentified proteins KIAA0186 (SEQ ID NOS: 221 and 222), KIAA0513 (SEQ ID NOS: 235 and 236) and the protein designated FLJ13910 (SEQ ID NOS: 293 and 294).

The manipulation of lymphatic endothelial-cell-specific molecules is expected to be applicable to treatments of LEC diseases disorders associated with tissue edemas. Without wishing to be bound by theory, manipulation of such molecules is expected to modulate endothelial cell-cell or cell-matrix protein interactions or to affect transendothelial transport thereby altering the state of fluid transport across the lymphatic vessel wall. Further, such molecules provide targets for the delivery of therapeutic compounds, such as growth factors, mitogens, and the like, as well as cytostatic or cytotoxic agents known in the art. These therapeutic compounds are targeted to such cells by associating a therapeutic agent with, e.g., a binding partner (such as an antibody) of the LEC surface marker. The transmembrane proteins identified herein, in particular the leucine-rich proteins, also provide useful targets for modulating cell adhesion events integral to lymph clearance.

EXAMPLE 7

MICROARRAY ANALYSIS TO DETECT LEC-AND LYMPH-RELATED DISORDERS

The LEC-specific genes identified herein are useful in the detection of LEC *in vivo* and in determining the extent of the lymphatic vasculature in a sample. The LEC-specific genes are also expected to be useful in diagnosing lymphedema and other LEC-related disorders.

Another aspect of the invention is a composition comprising a plurality of polynucleotide probes for use in detecting gene expression pattern(s) characteristic of particular cell type(s) and for detecting changes in the expression pattern of a

particular cell type, *e.g.*, lymphatic endothelial cells. The term "polynucleotide probe" is used herein to refer to any one of the nucleic acid sequences listed in SEQ ID NO: 1-30, 45, 47, 49 and 51, or any fragment thereof or a nucleic acid sequence encoding an amino acid sequence listed in SEQ ID NOS: 31-44, 46, 48, and 50, or a fragment thereof. Preferably, the fragment is at least 10 nucleotides in length; more preferably, it is at least 20 nucleotides in length. Such a composition is employed for the diagnosis and treatment of any condition or disease in which the dysfunction or non-function of lymphatic endothelial cells is implicated or suspected. In one embodiment, the present invention provides a composition comprising a plurality of polynucleotide probes, wherein at least a subset of the polynucleotide probes comprises at least a portion of an expressed gene isolated from a population of LEC-specific genes identified above. Also contemplated is a composition comprising a plurality of polynucleotide probes, with at least a subset of such probes each comprising a unique sequence selected from the group of SEQ ID NOs: 1-30, 45, 47, 49 and 51. Preferably, the composition comprises a subset of at least 3 polynucleotides, each having a different sequence selected from the group of SEQ ID NOs: 1-30, 45, 47, 49 and 51. Also preferred are compositions comprising at least 5, at least 7, at least 9, at least 15, at least 20, or at least 25 distinct polynucleotides having sequences selected from the group of SEQ ID NOs: 1-30, 45, 47, 49 and 51.

The composition is particularly useful as a set of hybridizable array elements in a microarray for monitoring the expression of a plurality of target polynucleotides. The microarray comprises a substrate and the hybridizable array elements. The microarray is used, for example, in the diagnosis and prognosis of a disease derived from aberrant lymphatic endothelial cell activity, such as lymphedema, lymphangioma, lymphangiomyeloma, lymphangiomatosis, lymphangiectasis, lymphosarcoma, and lymphangiosclerosis. Compositions may be useful in identifying more than one cell type and may be useful in the diagnosis and prognosis of more than one disease, disorder or condition. Further, useful information is obtained from those probes yielding a signal and from those probes not yielding a signal.

A polynucleotide comprising the sequence of any one of SEQ ID NOS: 1-30, 45, 47, 49 and 51 may be used for the diagnosis of conditions or diseases with which the abnormal expression of any one of the genes encoded by SEQ ID NOS: 1-30, 45, 47, 49 and 51 is associated. For example, a polynucleotide comprising any one of the sequences set forth in SEQ ID NOS: 1-30, 45, 47, 49 and 51 may be used in hybridization or PCR assays of fluids or tissues (e.g., obtained from biopsies) to detect abnormal gene expression in patients with lymphedema or another lymph-associated disease. In addition, a polynucleotide comprising a sequence encoding any of the amino acid sequences set forth in SEQ ID NOS: 31-44, 46, 48 or 50 is useful for the diagnosis of conditions or diseases associated with aberrant expression of a polypeptide having any one of those amino acid sequences. Fragments comprising at least 10 nucleotides are also useful in these diagnostic methods.

Expression profiles may be generated using the compositions of the invention comprising SEQ ID NOs: 1-30, 45, 47, 49 and 51. The expression profile generated from the microarray is used to detect changes in the expression of genes implicated in disease.

EXAMPLE 8

TRANSCRIPTION FACTORS IN BECS AND LECS

Transcription factors preferentially expressed in the LECs included the zinc finger factor c-maf and the MADS-family transcription factor MEF2C (Figure 1). Targeted mutagenesis of *MEF2C* leads to embryonic death at E9.5-10 due to defects in the remodeling of the primary vasculature and abnormal endocardiogenesis (Bi, *et al.*, *Dev. Biol. 211*:255-267. 1999). MEF2C has been reported to bind the transcription factor Sox18 and to potentiate its activity in endothelial cells (Hosking, *et al., Biochem. Biophys. Res. Commun. 287*:493-500. 2001). Mouse pups with a homozygous mutation in Sox18 that disrupts the MEF2C complex develop chylous ascites in some genetic backgrounds (Pennisi, D., et al., *Nat. Genet. 24*:434-437. 2000), suggesting that both proteins may be involved in the regulation of lymphatic development. In line with this hypothesis, RT-PCR analysis of *MEF2C*$^{-/-}$ embryos showed decreased VEGFR-3 expression (Bi, *et al., Dev. Biol. supra*).

The STAT6 transcription factor, which is activated in response to IL-4, was expressed specifically in the BECs. Consistent with this observation, the results herein show that the IL-4 receptor was expressed preferentially in BECs, as were some of the IL-4 target chemokines and receptors such as MCP-1 and CXCR4. VEGF stimulation and activation of VEGFR-2 is also known to lead to STAT6 phosphorylation and activation in endothelial cells (Bartoli, *et al.*, *J. Biol. Chem.* *275*:33189-33192. 2000). The absence of STAT6 in LECs, therefore, suggests that the downstream signaling pathways of VEGFR-2 differ in BECs and LECs. Expression patterns of other transcription factors are shown in Table 5.

EXAMPLE 9

SOX18 AND HEREDITARY LYMPHEDEMA

Expression of the transcription factor MEF2C is upregulated in LECs. *Sox18* (SEQ ID NO: 53, and encoding SOX18, SEQ ID NO: 54), which was reported to interact with MEF2C in mice, was also shown to play a potential role in lymphatic endothelial cell development. To investigate the role of *Sox18* in human lymphedema, the correlation of human *Sox18* mutants with human hereditary lymphedema was investigated.

The SOX proteins, homologs of the family of SRY transcription factors, are ubiquitous transcription factors which contain a putative high-mobility-group (HMG) DNA binding domain. (Wegner, M., *Nucl. Acids Res.* 27:1409-20. 1999). SOX proteins bind their DNA targets at a heptameric SOX consensus binding sequence [5'- (A/T)(A/T)CAA(A/T)G-3'] (Pennisi *et al.*, *Mol. Cell Bio.* 20:9331-36. 2000) and generally bind DNA in the minor groove rather than the major groove of the double helix, which results in transcriptional regulation of the target gene. SOX proteins may also be involved in recruiting other DNA binding proteins to a DNA-protein complex, thereby assisting in transcription regulation (Wegner, *supra*). SOX18 shares homology with both SOX7 and SOX17, all members of the Group F Sox genes.

SOX18 is involved in vascular development and has been localized to the developing cardiovascular system and sites of angiogenic activity. Mice

homozygous for the Ragged (*Ra*) mutation in *Sox18* exhibit chylous ascites and edema (Pennisi *et al., Nat. Genet.* 24:434-37. 2000), similar to the *Chy* mouse model of lymphedema (Lyon *et al., Mouse News Lett. 71*: 26. 1984). The mutation in *Ra* mice has been determined to be a frameshift mutation that causes truncation of the transactivating domain (Pennisi *et al., Nat. Genet.* 24:434-37. 2000). *Sox18* null mice, however, demonstrate only a slight phenotypic change in hair follicle development and show no signs of edema or irregular vascular development (Downes and Koopman, *Trends Cardio. Med.* 11:318-24. 2001). This phenotype may be due to redundancy among the Group F Sox members, SOX7 and SOX17. These proteins may substitute for SOX18 function in its absence, but cannot overcome a *Sox18* dominant negative mutant such as the *Ra* mutations. Hence, knocking out the entire Group F family may produce a lymphedema phenotype similar to the *Ragged* mice.

Mouse and human SOX18 are homologous proteins containing a DNA binding HMG-box of approximately 80 amino acids (97% homologous), a transactivating domain which in mouse is about 93 amino acids (90% homologous), and a C-terminal domain (92% homologous) (Downes and Koopman, *supra*). The human SOX18 HMG-box has been localized to nucleotides 395-598, corresponding to amino acids 84-151. The mouse HMG-box is encoded by nucleotides 320-532, corresponding to amino acids 78-148. The human transactivation domain has not been delineated to date, but one of skill in the art could readily obtain the human transactivating domain using the homologous mouse sequence, which is found at amino acids 252-346 of mouse SOX18 (Hosking *et al., Gene* 262:239-47. 2001). Although the human SOX18 protein exhibits similarities to mouse SOX18 at the primary structural level, there is no known association of a human *Sox18* mutant with a disease or condition, such as hereditary lymphedema.

Human *Sox18* has been mapped to chromosome 20q.13.3 (Stanojcic *et al., Biochem. Biophys. Acta.* 1492:237-41. 2000). Elucidation of an inheritable mutation at or near this chromosomal location that correlates with hereditary lymphedema is useful in confirming the genetic basis of the disease, in the screening of patients affected by hereditary lymphedema, in the screening of patients for a pre-

disposition to develop hereditary or other forms of lymphedema, and also as a basis for target treatment regimens directed to overcoming the inherited mutation.

To determine the linkage of *Sox18* with lymphedema, families with inherited lymphedema are identified for the purpose of conducting linkage and positional candidate gene analyses. Family members are considered affected with hereditary lymphedema if they exhibit asymmetry or obvious swelling of one or both legs or if they have received a medical diagnosis of lymphedema or if there are personal or family reports of extremity swelling or asymmetry.

Biological samples are obtained from members of the families to conduct the genetic analyses. DNA is isolated from EDTA-anticoagulated whole blood by the method of Miller *et al.*, (*Nucleic Acids Res.* 16:1215. 1998), and from cytobrush specimens using the Puregene DNA isolation kit (Gentra Systems, Minneapolis, MN). Analysis of the markers used in the genome scan are performed by methods recognized in the art. See Browman *et al., Am. J. Hum. Genetic.*, 63:861-869 (1998); see also the NHLBI Mammalian Genotyping Service.

To explore the potential role of *Sox18* in lymphedema, probands from the lymphedema families are screened for variation by direct sequencing of portions of the *Sox18* gene. The sequencing strategy uses amplification primers generated based upon the *Sox18* cDNA sequence (SEQ ID NO: 53) and information on the genomic organization (intron-exon data, identified domain motifs) of the related *Sox* genes. Variable positions (single nucleotide polymorphisms) and unique sequence primers are used to amplify sequences flanking each variable site located in the domains used for analysis.

The *Sox18* genomic DNA from both the normal and lymphedema affected individuals is sequenced and a map of mutations detected in the *Sox18* gene of lymphedema patients as compared to unaffected individuals is generated. Commonly detected mutations in lymphedema patients, such as a conservative or non-conservative nucleotide change, a deletion, or an insertion, indicates that a mutation in that particular nucleotide confers a pre-disposition to developing lymphedema. Analysis of the genomic DNA of the affected individuals will correlate mutations in the *Sox18* genomic sequence and lymphedema.

To confirm the correlation of Sox18 mutations and the development of lymphedema, genetic linkage studies are performed, as set out in the method of identifying genetic polymorphisms described in U.S. patent application number US2003026759 and PCT/US99/06133, each of which is incorporated herein by reference.

Two-point linkage analysis is conducted using an autosomal dominant model predicting 80% penetrance in the heterozygous state, 99% penetrance in the homozygous state, and a 1% phenocopy rate. The frequency of the disease allele is set at 1/10,000. Microsatellite marker allele frequencies are calculated by counting founder alleles, with the addition of counts of non-transmitted alleles. Multipoint analysis is carried out using distances from the Location Database provided by the University of Southampton School of Medicine. Multipoint and 2-point analyses are facilitated using the VITESSE (v1.1) program. (O'Connell, and Weeks, *Nature Genet.*, 11:402-408. 1995).

Analysis of the markers used in the genome scan are performed by methods recognized in the art. [See Browman *et al.*, *Am. J. Hum. Genetic., 63*:861-869 (1998); see also the NHLBI Mammalian Genotyping Service and databases offered by the Center for Molecular Genetics (Marshfield, WI). One of skill in the art readily chooses genetic linkage markers identified in chromosome 20 (specifically 20q13.3), where Sox18 has been localized (Stanojcic *et al.*, *supra*).

Linkage simulation is performed using SLINK (Weeks *et al.*, *Am. J. Hum. Genet. 47:*A204. 1990) and linkage is analyzed using MSIM (Ott, J., *Proc. Nat. Acad. Sci. USA, 86:*4175-4178. 1989) to estimate the potential power of two point linkage analysis in the family being assessed. Marker genotypes are simulated for a marker with heterozygosity of 0.875 under a linked (θ=0) and unlinked (θ=0.5) model using the available individuals. The simulation is set such that the power to detect linkage is greater than 90% for a LOD score threshold of Z(θ) 2.0 and the false positive rate is less than 5%.

Mutations that correlate strongly with a heritable lymphedema are expected to be mutations in functional domains of the SOX18 protein, e.g., the HMG-Box domain or the transactivating domain. Exemplary mutations include missense

mutations that cause non-conservative substitutions, nucleotide deletions or insertions that cause frameshifts in the *Sox18* coding region, in-frame deletions or insertions such as those affecting a functional domain(s), or alterations of control regions affecting the level of Sox18 expression.

Upon identification of the *Sox18* lymphedema-correlated mutations, *Sox18* mutant expression vectors containing an isolated mutant *Sox18* allele is expressed in, e.g., 293T or endothelial cells. The *Sox18* mutant DNA can also be integrated into a plasmid useful in the mammalian two-hybrid system, such as pGAL4, to measure SOX18 interaction with its binding partners, such as MEF2C (Hosking *et al., Biochem. Biophys. Res. Comm.* 287: 493-500. 2001) or to screen for SOX18 binding partners. For example, pGAL4*Sox18* vector links the *Sox18* gene to the yeast Gal4 DNA binding domain and a transcriptional activator is linked to a SOX18 binding partner in a separate vector. Co-introduction of these vectors into a host cell will result in detectable reporter gene expression resulting from SOX18 interactions with the binding partner or candidate binding partner. The pCMV-BD and pCMV-AD vectors, which contain a GAL4 DNA binding domain and the NF-κB transcriptional domain, respectively, are useful in this assay (BD Biosciences Clontech) for constructing and expressing gene fusions, with *SOX18* binding activity detected using the luciferase reporter system.

In such a di-hybrid assay, a *Sox18* lymphedema-correlated mutant that contains a mutation affecting SOX18 binding via the transactivating domain will decrease the amount of luciferase reporter activity, indicating that the *Sox18* lymphedema-correlated mutation may result in lymphedema through a defect in its ability to bind its binding partner through its transactivating domain.

A *Sox18* allele is also assessed for a mutation in its HMG-box DNA binding domain through several techniques. DNA binding is assessed in a one-hybrid assay in which the DNA sequence bound by SOX18, e.g. 5'- (A/T)(A/T)CAA(A/T)G-3' and permutations thereof, is placed in front of (i.e., upstream of or 5' to) a promoter/reporter gene construct similar to the target plasmid in a two-hybrid assay. The reporter assay then detects binding between a SOX18 protein and its putative DNA binding sequence. DNA binding is also assessed using a gel shift assay

performed by incubating a purified SOX18 protein with a ^{32}P end-labeled DNA fragment containing the SOX18 DNA-binding sequence. The reaction products are then analyzed on a non-denaturing polyacrylamide gel to measure the mobility of DNA-bound or free SOX18. The specificity of a SOX18 polypeptide for the putative binding site is established by competition experiments using DNA fragments or oligonucleotides containing a binding site for SOX18 or other unrelated DNA sequences.

Additionally, fluorescence-based assays for detection of DNA/protein binding are used. SOX18 DNA binding is detected by fluorescence measurement of single fluorophores which are bound to either the DNA or protein. In these assays, protein binding is determined by a change in fluorescence intensity or polarization when DNA-protein complexes form. Alternatively, two DNA fragments, each containing half of the protein binding site, are generated. The two double-stranded DNA fragments have complementary single-strand overhangs that comprise part of the protein binding site. One DNA fragment is labeled with a fluorescence donor while the other is labeled with an acceptor, with fluorescence detected only upon fluorescence resonance energy transfer (FRET). Upon protein binding, the overhangs of the two DNA fragments anneal and bring the fluorescence donor and acceptor into proximity, resulting in transfer of the fluorescence energy, which results in detectable fluorescence of the acceptor. See Heyduk, *et al., Nat. Biotechnol.* 20:171-6. 2002.

Correlation of a mutation in the human *Sox18* genome with the risk of developing lymphedema provides another method for diagnosis and/or treatment of individuals affected by hereditary lymphedema. Elucidation of a *Sox18* mutation associated with lymphedema allows for the determination of the SOX18 protein activity is disturbed by the mutation, e.g., DNA binding or protein binding, and provides direction for treatment of patients with lymphedema.

Additionally contemplated is the treatment of patients with *Sox18*-induced lymphedema with a lymphatic growth factor such as VEGF-C and/or VEGF-D to overcome impaired lymphatic vascular development. For example, treatment of VEGFR-3 defective animals with VEGF-C and/or VEGF-D overcomes the inability of VEGFR-3 to signal, thereby promoting lymphangiogenesis and ameliorating

symptoms of lymphedema. *Sox18*-induced lymphedema patients are treated with a therapeutically effective amount of VEGF-C and/or VEGF-D. In an additional embodiment, VEGF-C and/or VEGF-D are administered to the above patients in conjunction with other therapies designed to relieve the symptoms of lymphedema.

EXAMPLE 10

VEGF-C and VEGF-D knockout mice demonstrate aberrant vascular development which can be overcome by administration of exogenous VEGF-C and/or VEGF-D polypeptide. To determine if Sox18 transcriptional regulation can overcome this defect due to its potential interaction with, and transcriptional effect on, the VEGFR-3 promoter, VEGF-C or VEGF-D knockout mice are genetically crossed by interbreeding with mice overexpressing *Sox18* from a cell-specific-promoter (e.g. K-14 keratin promoter) or a retroviral vector. The effects of Sox18 activity on lymphedema are assessed through measurement of lymphedema and vascular development, as described in Example 10.

Survival of the knockout mice and detection of lymphatic development in the VEGF-C and/or VEGF-D knockout/Sox18-overexpressing mice indicates that Sox18 induces VEGFR-3 signaling and plays a key role in lymphangiogenesis.

VEGF-C overexpressing mice (K-14-VEGF-C Tg) exhibit an extensive network of lymphatic vasculature, are prone to tumor metastasis, and demonstrate upregulated VEGFR-3 expression and symptoms of lymphedema (US Patent No. 6,361,946). To determine if *Sox18* regulates VEGF-C signaling through VEGFR-3, K-14-VEGF-C Tg mice are crossed to animals which express a naturally mutated *Sox18* (*Ragged* mutation) or a laboratory-designed mutant constructed using site-directed mutagenesis and standard knockout techniques known in the art to generate a mutation in either the DNA-binding or transactivating domain of the SOX protein, resulting in a K-14-VEGF-C Tg/Sox18$^{-/-}$ mouse.

Decreased lymphangiogenesis, decreased incidence of tumor metastasis, and decreased levels of VEGFR-3 exhibited by the K-14-VEGF-C Tg/*Sox18*$^{-/-}$ double mutant animals as compared to the K-14-VEGF-C Tg single mutant animal indicates that the Sox18 molecule interferes with VEGF-C signaling

through VEGFR-3 and that inhibition of the VEGFR-3 signaling in the *Sox18* mutant downregulates the lymphangiogenic effects of activated VEGFR-3.

Alternatively, K-14-VEGF-C Tg mice are crossed to mice transgenic for a *Sox18* allele that is overexpressed (see above) and the effects of Sox18 upregulation are measured. A decrease in lymphangiogenesis, decreased incidence of tumor metastasis, and decreased levels of VEGFR-3 exhibited by the K-14-VEGF-C Tg/*Sox18* overexpressing double mutant animals as compared to the K-14-VEGF-C Tg single mutation indicates that Sox18 transcriptional regulation inhibits VEGFR-3 signaling and is likely a factor in negatively regulating lymphangiogenesis.

A result indicating that *Sox18* is a negative regulator of lymphangiogenesis provides a method of treating disorders mediated by extensive lymphatic vasculature, such as lymphangiogenesis in tumor development or lymphangiosarcoma, by administration of a vector providing the SOX18 transcription factor in excess thereby preventing the induction of lymphangiogenic signals.

EXAMPLE 11

SOX18 IN LYMPHATIC DEVELOPMENT

Lymphatic endothelial cells show a unique development pattern that is highly regulated by several LEC-specific genes such as VEGFR-3 and Prox-1. Sox18, as a DNA binding protein and transcription factor, is expected to be involved in the regulation of these LEC-specific genes, contributing to the elaboration of a LEC cellular fate. Several lines of evidence indicate that Sox18 may be involved in VEGFR-3 transcription regulation: SOX18 binds to the transcription factor MEF2C in mice, both *Sox18*-mutant and MEF2C-deficient mice exhibit lymphedema symptoms similar to VEGFR-3 mutant mice, and the VEGFR-3 promoter contains a MEF2C binding site (Iljin *et al, FASEB J.* 15:1028-36. 2001). These observations support a role for SOX18 in lymphatic development.

To analyze the ability of *Sox18* to affect the transcription of LEC-specific growth factors, blood vascular endothelial cells are induced to develop into LECs by the addition of an AdProx-1 vector. *Sox18* mRNA and protein levels are measured before and after the addition of the Prox-1 vector. Upregulation of *Sox18*

after the addition of the Prox-1 vector is expected to correlate with the development of lymphatic endothelial cells, indicating that *Sox18* is a factor in LEC differentiation. Alternatively, either the DNA binding or transactivation activity of *Sox18* is disrupted via site-directed mutagenesis, thereby resulting in either a dominant negative or inactive SOX18 protein. The plasmid containing the *Sox18*-disrupted allele is co-transfected into BECs with the AdProx-1 vector to assess LEC development in the presence of a dysfunctional *Sox18* gene. Detection of LEC-specific markers such as LYVE-1 and podoplanin are also used in these experiments to measure the ability of *Sox18* to modulate lymphatic development. Additionally, mutant *Sox18* is also co-transfected with vectors encoding LEC-specific proteins (e.g., VEGFR-3, Prox-1, LYVE-1) into 293T cells and the ability of the mutated *Sox18* to regulate the activities of those genes is assessed. For example, signaling in VEGFR-3 co-transfected 293T cells stimulated with VEGF-C in the presence and absence of *Sox18* is assessed using a phosphorylation assay.

Development of the lymphatic vasculature can also be evaluated in *Sox18* mutant mice, including *Ra* mice, *Sox18* null mice, and *Sox18* mice transgenic for a mutation described herein that correlates with a pre-disposition to lymphedema. Transgenic *Sox18* mice exhibiting a symptom of lymphedema are engineered to express a mutation in the mouse gene homologous to the human mutation or are engineered to express the human *Sox18* gene containing a lymphedema-specific mutation. Development of the vasculature in these animals is analyzed, as set out in US Patent No. 6,361,946 (see also Kaipainen *et al., Proc. Natl. Acad. Sci. (USA)*, 92:3566-70. 1995), using techniques known in the art, such as *in situ* hybridization, to detect VEGF-C and/or VEGFR-3 mRNA expression, antibody detection of VEGF-C and/or VEGFR-3 proteins *in vivo*, and Evan's blue dye detection to determine the extent of LEC development and to visualize effective lymph drainage *in vivo*.

An increase in VEGFR-3 signaling in a dominant negative *Sox18* mutant transfectant indicates that *Sox18* expression has a detrimental effect on VEGFR-3-mediated activity. The invention contemplates a therapy to overcome this type of mutation comprising administering to mammal, such as a human patient, a composition comprising a SOX18 inhibitor, such as a dominant negative gene or

dominant negative Sox18 ligand which interferes with the ability of SOX18 to interfere with VEGFR-3 signaling. Alternatively, if *Sox18* activation promotes VEGFR-3 activity this provides an indication that a therapy for lymphedema comprises a composition which promotes SOX18 transcriptional activity, such as cells given *ex-vivo* which overexpress *Sox18*.

EXAMPLE 12

SOX18 DIRECTED THERAPY IN LYMPHEDEMA

Another aspect of the invention is the use of *Sox18* to produce cell-based therapeutic compositions, particularly LEC cell-based compositions. In one embodiment, the cells are autologous cells, *i.e.*, cells of the organism (*e.g.*, human patient) receiving treatment for a disease or disorder of the lymphatic system. The invention contemplates elevating the endogenous expression of *Sox18,* for example by the modification of expression control regions, e.g., promoters, through recombinant techniques such as homologous recombination. Alternatively, the cells are transformed or transfected with an isolated *Sox18*, e.g., a heterologous *Sox18*, for heterologous *Sox18* expression, either *in vivo* or *ex vivo*.

For example, SOX18 interacts with transcription factor MEF2C, with the complex binding to the VEGFR-3 promoter, thereby inducing VEGFR-3 transcription and affecting VEGFR-3 protein expression and signaling levels. It is contemplated that insertion of a *Sox18* gene driven by a retroviral or adenoviral vector into an LEC expressing VEGFR-3 will upregulate VEGFR-3-mediated signaling.

These *Sox18*-expressing cells are then used as a therapeutic composition in the treatment of patients with an LEC disease or disorder, such as hereditary lymphedema or trauma-induced lymphedema. These cells are used to treat any disease or condition associated with a decrease in expression of VEGFR-3, such as lymphangioma, lymphangiomyeloma, lymphangiomatosis, lymphangiectasis, lymphosarcoma, and lymphangiosclerosis.

Additionally, a SOX18 polypeptide or polypeptide fragment is administered to a patient experiencing lymphedema to relieve the symptoms of lymphedema. It is contemplated that administration of either a full-length SOX18

polypeptide or a fragment of SOX18, which contains either the DNA binding domain or the transactivating domain, will bind to its cognate binding partner *in vivo* and promote VEGFR-3 signaling, or will initiate downstream events in the lymphangiogenic process, thus bypassing a defect in VEGFR-3 signaling or VEGF-C ligand binding involved in lymphedema.

In a related aspect, if SOX18 expression inhibits VEGFR-3 signaling via decreased transcription factor binding or DNA binding, it is expected that inhibition of SOX18 will result in a compensatory upregulation of VEGFR-3, ameliorating deleterious symptoms associated with VEGFR-3 under-expression. Administration of antisense therapy specific for the Sox 18 gene in instances where Sox 18 negatively regulates VEGFR-3 activity will inhibit SOX18 activity thereby allowing VEGFR-3-mediated signaling and lymphatic growth. Due to the potential functional redundancy of the Group F SOX proteins (SOX7/17/18), however, it may be necessary to inactivate all three proteins through a mechanism that inhibits the DNA binding activity of all Group F proteins. This is done, e.g., by targeting the DNA binding domain, which is highly homologous among all the proteins. It is contemplated that recombinant SOX7/17/18 proteins expressing a mutated DNA binding domain, when administered as a pharmaceutical composition (containing all three mutant peptides), will inhibit SOX18 downregulation of VEGFR-3 and induce or promote VEGFR-3 signaling activity. From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention.

All of the above U.S. patents, U.S. patent application publications, U.S. patent applications, foreign patents, foreign patent applications and non-patent publications referred to in this specification and/or listed in the **[Application Data Sheet]** are incorporated herein by reference, in their entirety.

Table 3

Lymphatic EC (187 genes)

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
lung type-I cell membrane-associated protein, podoplanin	AF030428	NM_006474	41870_at	A	P	7.4	0.925		+	
lung type-I cell membrane-associated protein, podoplanin	*AI660929*	*NM_006474*	*41871_at*	*A*	*P*	6.1	0.150		+	
cellular retinol-binding protein	M11433	NM_002899	38634_at	A	P	7.3	0.255			
macrophage mannose receptor (MRC1)	M93221		36908_at	A	P	7.1	0.682			
transcription factor C-MAF	AF055376	NM_005360	41504_s_at	P	P	5.1	0.522			
transcription factor C-MAF	*AF055376*	*NM_005360*	*41505_r_at*	*A*	*P*	3.9	0.588			
selenoprotein P	Z11793	NM_005410	34363_at	A	P	5.0	0.331			
KIAA0466, immunoglobulin superfamily, member 3	AB007935	NM_001542	38086_at	A	P	4.9	2.028			
type II membrane protein similar to HIV gp120-binding C-type lectin, CD209 antigen-like	AB015629	NM_014257	39270_at	A	P	4.9	0.846			
KIAA0626	AB014526	NM_021647	33241_at	A	P	4.7	0.212			
KIAA0711	AB018254	NM_014867	36453_at	A	P	4.6	0.055	+		
integrin alpha 9	D25303	NM_002207	1508_at	A	P	4.3	0.712	+	+	
integrin alpha 9	*D25303*	*NM_002207*	*35948_at*	*A*	*P*	3.6	1.086	+	+	
relaxin H2	X00948	NM_005059	31732_at	A	P	4.1	0.432			
KIAA0644	AB014544	NM_014817	34214_at	A	P	3.9	0.803	+		
Cdk-inhibitor p57KIP2 (KIP2)	U22398	NM_000076	1787_at	A	P	3.8	2.020	+		3.3

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
Cdk-inhibitor p57KIP2 (KIP2)	*U22398*	*NM_000076*	*39545_at*	*A*	*P*	3.0	0.150	+		*1.4*
transient receptor potential channel TRPC6	AJ006276	NM_004621	36365_at	A	P	3.8	0.988			
cDNA DKFZp564O222 (from clone DKFZp564O222)	AL050002		38312_at	A	P	3.6	0.876			
subtilisin-like protein (PACE4), paired basic amino acid cleaving system 4	M80482	NM_002570	32001_s_at	P	P	3.6	0.334			
regulator of G-protein signalling 16, A28-RGS14p	U70426	NM_002928	41779_at	A	P	3.6	0.673			
dihydropyrimidinase related protein-1, collapsin response mediator protein 1	D78012	NM_001313	40272_at	A	P	3.5	1.192			
desmoplakin (DPI, DPII)	AL031058	NM_004415	36133_at	A	P	3.5	0.426		+	1.0
pendrin, solute carrier family, member 4	AF030880	NM_000441	36376_at	A	P	3.3	1.156			
reelin (RELN)	U79716	NM_005045	37530_s_at	P	P	3.3	0.142			
integrin, alpha 1	X68742		120_at	P	P	3.3	0.080	+		
integrin alpha 1	*X68742*		*37484_at*	*M*	*P*	2.4	0.345	+		
cholesterol 25-hydroxylase	AF059214	NM_003956	32363_at	A	P	3.3	0.137			1.5
inhibin beta-B-subunit precursor	M31682	NM_002193	38545_at	P	P	3.2	0.056			3.0
KIAA1233	AL109724		38856_at	A	P	3.1	1.540			
pre-B cell stimulating factor homologue (*SDF1b*)	L36033	NM_000609	33834_at	A	P	3.0	0.860	RT-PCR		
V-Erba Related Ear-3 Protein	HG3510-HT3704		1147_at	P	P	2.9	0.398			1.7
antigen identified by monoclonal antibody MRC OX-2	X05323		37716_at	P	P	2.9	0.283	+		1.0
apolipoprotein D	J02611	NM_001647	36681_at	M	P	2.9	0.150	+		

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
TIMP3, tissue inhibitor of matrix metalloproteinases	U14394	NM_000362	1035_g_at	A	P	2.8	0.528	+		
TIMP3	*U14394*	*NM_000362*	*1034_at*	*P*	*P*	1.9	0.224	+		
aldehyde dehydrogenase 1	K03000	NM_000689	37015_at	P	P	2.8	0.275			
prospero-related homeobox 1 (prox 1)	U44060	NM_002763	31918_at	A	P	2.8	0.299	+	+	(5,6)
matrix Gla protein	AI953789	NM_000900	36683_at	A	P	2.6	0.250	+		
neuronal pentraxin II (NPTX2)	U29195		35663_at	A	P	2.6	1.267			2.9
histatin 2 (HIS2)	M26665	NM_000200	41148_at	A	P	2.6	1.009			
ADDL mRNA for adducin-like protein, adducin 3 (gamma)	D67031	NM_016824	33102_at	P	P	2.6	0.277	+		
adducin 3 (gamma)	*U37122*	*NM_016824*	*33103_s_at*	*M*	*P*	*2.4*	*0.397*	+		
MADS box transcription enhancer factor 2, polypeptide C (myocyte enhancer factor 2C)	L08895	NM_002397	37710_at	A	P	2.5	0.540	+		
MADS box transcription enhancer factor 2, (myocyte enhancer factor 2C)		*NM_002397*	*37712_g_at*	*A*	*P*	*1.9*	*0.197*	+		
MADS box transcription enhancer factor 2, polypeptide C (myocyte enhancer factor 2C)	*S57212*	*NM_002397*	*37711_at*	*A*	*P*	1.4	0.442	+		
phosphoglucomutase 5	L40933	NM_021965	33694_at	A	P	2.5	0.431			
cyclin E2	AF102778	NM_004702	35249_at	A	P	2.5	0.906			5.8
interleukin 7 (IL7)	M29053		33966_at	A	P	2.4	0.191			
interleukin 7	*J04156*	*NM_000880*	*1159_at*	*A*	*P*	*1.9*	*0.921*			
cDNA DKFZp586L0120 (from clone DKFZp586L0120)	AL050154		38351_at	P	P	2.4	0.135			
peroxisome proliferative activated receptor, gamma, PPARG	L40904	NM_005037	37104_at	A	P	2.4	0.502			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
fatty acid binding protein 4	AA128249	NM_001442	38430_at	P	P	2.4	0.132			
protein kinase C zeta	Z15108	NM_002744	362_at	P	P	2.4	0.008			
46 kDa coxsackievirus and adenovirus receptor (CAR) protein	Y07593	NM_001338	37534_at	P	P	2.3	0.137			2.2
PAC clone RP4-751H13 from 7q35-qter, zinc finger-like	AC004877		39837_s_at	A	P	2.3	0.714			
thymidine kinase 1, soluble	M15205	NM_003258	910_at	P	P	2.3	0.205			2.1
thymidine kinase 1	*K02581*	*NM_003258*	*41400_at*	*M*	*P*	1.7	0.193			*1.4*
Pig7 (PIG7), LPS-induced TNF-alpha factor	AF010312	NM_004862	37024_at	A	P	2.3	0.233			4.6
LPS-induced TNF-alpha factor	*AL120815*	*NM_004862*	*37025_at*	*P*	*P*	1.3	0.327			*3.8*
lipase A, lysosomal acid, cholesterol esterase	X76488	NM_000235	38745_at	P	P	2.3	0.281			
ubiquitin specific protease 13 (isopeptidase T-3)	U75362	NM_003940	40701_at	A	P	2.2	0.334			
carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein) CEACAM1	X16354	NM_001712	988_at	P	P	2.2	0.048			
cDNA DKFZp586D0918 (from clone DKFZp586D0918)	AL049370		41856_at	P	P	2.1	0.385			
KIAA0598, B cell RAG associated protein	AB011170	NM_014863	35350_at	P	P	2.1	0.154			
RAMP2 (receptor (calcitonin) activity modifying protein 2)	AJ001015	NM_005854	38177_at	P	P	2.1	0.361			
cholesteryl ester transfer protein precursor	M30185	NM_000078	40741_at	A	P	2.1	0.191			
epithelial membrane protein 2	U52100	NM_001424	39631_at	P	P	2.0	0.141			1.0
MHC class II lymphocyte antigen (HLA-DP) beta chain	M83664	NM_002121	38095_i_at	A	P	2.0	1.368			
MHC class II lymphocyte antigen	*M83664*	*NM_002121*	*38096_f_at*	*A*	*P*	1.2	0.034			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
(HLA-DP) beta chain										
beta-arrestin 2	AF106941	NM_004313	33283_at	A	P	2.0	0.273			
mitotic checkpoint kinase Bub1 (BUB1)	AF053305	NM_004336	41081_at	A	P	2.0	0.195			
KIAA0229, similar to human ankyrin 1(S08275)	D86982		40971_at	P	P	2.0	0.195			1.6
Sprouty 1 homolog (antagonist of FGF signaling)	AF041037		38767_at	P	P	2.0	0.209			
guanine nucleotide exchange factor for Rap1; M-Ras-regulated GEF, KIAA0277		NM_012294	38062_at	P	P	2.0	0.497			2.1
translin	X78627	NM_004622	36177_at	A	P	2.0	0.140			
erythrocyte membrane protein band 4.9 (dematin)	U28389	NM_001978	37192_at	P	P	2.0	0.265			1.9
KIAA0846 protein	AB020653	NM_015376	34748_at	A	P	2.0	0.457			
glia maturation factor, gamma	W07033	NM_004877	35261_at	P	P	1.9	0.083			1.0
insulin-like growth factor binding protein 2 (IGFBP-2)	X16302	NM_000597	40422_at	A	P	1.9	1.157			
smooth muscle myosin heavy chain isoform Smemb	S67247		32838_at	A	P	1.9	0.179			
TTG-2 (cysteine rich protein with LIM motif), LIM domain only 2 (rhombotin-like 1)	X61118	NM_005574	32184_at	P	P	1.9	0.221			1.0
cyclin B2	AL080146	NM_004701	32263_at	A	P	1.9	0.276			
KIAA0353	AB002351		39544_at	P	P	1.9	0.158			1.8
KIAA0559, piccolo (presynaptic cytomatrix protein)	AB011131		37780_at	A	P	1.9	0.330			
G protein-coupled receptor, family C, group 5, member B	AC004131	NM_016235	40240_at	P	P	1.9	0.047			2.1
G protein-coupled receptor, family C,	*AI801872*	*NM_016235*	*40239_g_at*	*P*	*P*	1.4	0.303			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
group 5, member B										
CREM (cyclic AMP-responsive element modulator beta isoform)	S68134	NM_001881	32066_g_at	P	P	1.9	0.098			1.7
CREM (cyclic AMP-responsive element modulator beta isoform)	*S68134*	*NM_001881*	*32065_at*	*P*	*P*	*1.8*	*0.241*			*2.0*
CREM (cyclic AMP-responsive element modulator beta isoform)	*S68271*	*NM_001881*	*32067_at*	*P*	*P*	1.5	0.182			*2.0*
hypothetical protein FLJ13110	AL080222	NM_022912	36096_at	A	P	1.9	0.387			
inositol(myo)-1(or 4)-monophosphatase 2	AF014398	NM_014214	36496_at	A	P	1.8	0.590			3.2
KIAA0937 protein	AB023154		35369_at	P	P	1.8	0.185			
mitotic spindle coiled-coil related protein	AF063308	NM_006461	32120_at	A	P	1.8	0.257			
cysteine and glycine-rich protein 2 (CSRP2)	U57646	NM_001321	41401_at	A	P	1.8	0.431			4.3
topoisomerase (DNA) II alpha (170kD)	AI375913	NM_001067	40145_at	P	P	1.8	0.239			1.0
DNA topoisomerase II	*J04088*	*NM_001067*	*1592_at*	*P*	*P*	1.2	0.162			
protein phosphatase inhibitor 2 (PPP1R2)	U68111		33180_at	P	P	1.8	0.319			
KIAA0186	D80008	NM_021067	39677_at	A	P	1.8	0.269			1.4
dual-specificity tyrosine-(Y)-phosphorylation regulated kinase 3 (Dyrk3)	Y12735	NM_003582	39931_at	P	P	1.8	0.146			
kinesin-like spindle protein HKSP (HKSP)	U37426	NM_004523	40726_at	M	P	1.8	0.439			
huntingtin-associated protein interacting protein (duo)	U94190	NM_003947	40655_at	P	P	1.8	0.529			1.4
diubiquitin	AL031983	NM_006398	39959_at	A	P	1.8	0.841			
bikunin, serine protease inhibitor,	U78095	NM_021102	34348_at	A	P	1.8	0.398			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
Kunitz type, 2										
cytochrome P-450-1 (TCDD-inducible)	K03191	NM_000499	36767_at	A	P	1.7	0.165			2.0
cytochrome P(1)-450	*X02612*	*NM_000499*	*1025_g_at*	*P*	*P*	1.1	0.125			*1.7*
KIAA0513		NM_014732	38735_at	A	P	1.7	0.297			
protein phosphatase inhibitor 2 (PPP1R2)	U68111		812_at	P	P	1.7	0.185			
RAMP3 (receptor (calcitonin) activity modifying protein 3)	AJ001016	NM_005856	35152_at	P	P	1.7	0.228			
B-myb	X13293	NM_002466	1854_at	M	P	1.7	0.455			2.3
KIAA0952	AB023169	NM_014962	37755_at	P	P	1.7	0.254			1.0
interferon stimulated gene (20kD), HEM45	U88964	NM_002201	33304_at	A	P	1.7	0.178			
GS3955	D87119	NM_021643	717_at	P	P	1.7	0.107			1.6
GS3955	*D87119*	*NM_021643*	*40113_at*	*P*	*P*	1.3	0.098			*1.3*
GRB2-related adaptor protein (Grap)	U52518	NM_006613	805_at	A	P	1.7	0.147			
KIAA1071 protein	AB028994		38286_at	A	P	1.7	0.625			
RNA-binding protein gene with multiple splicing, RBP-MS/type 5	D84111	NM_006867	34162_at	P	P	1.7	0.347			
RNA-binding protein gene with multiple splicing, RBP-MS/type 5	*D84111*	*NM_006867*	*34163_g_at*	*P*	*P*	1.6	0.147			
RBP-MS/type 4, RNA-binding protein gene with multiple splicing	*D84110*	*NM_006867*	*1276_g_at*	*P*	*P*	1.5	0.263			
RBP-MS/type 4, RNA-binding protein gene with multiple splicing	*D84110*	*NM_006867*	*38049_g_at*	*P*	*P*	1.3	0.268			
RBP-MS/type 3, RNA-binding protein gene with multiple splicing	*D84109*	*NM_006867*	*38047_at*	*P*	*P*	1.2	0.225			
alpha-actinin-2-associated LIM protein	AF002282	NM_014476	39690_at	A	P	1.7	0.728			1.5

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
semaphorin-III (Hsema-I), semaphorin 3A	L26081	NM_006080	33461_at	A	P	1.6	0.445			
IQ motif containing GTPase activating protein 2	U51903	NM_006633	1647_at	M	P	1.6	0.395			
Arrestin, Beta 2	HG2059-HT2114		957_at	P	P	1.6	0.342			
retinoblastoma-associated protein HEC	AF017790	NM_006101	40041_at	P	P	1.6	0.153			2.0
LIM domain binding protein (LDB1)	AF052389	NM_001290	36065_at	P	P	1.6	0.153			1.9
dual specificity phosphatase 5	U15932	NM_004419	529_at	P	P	1.6	0.207			
Homo sapiens cDNA 3', mRNA sequence	AI557322		39611_at	P	P	1.6	0.081			
monoamine oxidase A (MAOA)	M68840	NM_000240	41772_at	P	P	1.6	0.148			
monoamine oxidase A	*AA420624*	*NM_000240*	*41771_g_at*	*P*	*P*	1.4	0.230			
NECDIN related protein	U35139	NM_002487	36073_at	P	P	1.6	0.245			
regulatory solute carrier protein, family 1, member 1	X82877	NM_006511	31695_g_at	A	P	1.6	0.916			
TTK protein kinase	M86699	NM_003318	572_at	P	P	1.6	0.196			
fms-related tyrosine kinase 4, VEGFR-3	X69878	NM_002020	403_s_at	A	P	1.5	0.403	+	+	1.1
TSC403, similar to lysosome-associated membrane glycoprotein	AB013924	NM_014398	37168_at	P	P	1.5	0.164			
HMG-2	X62534		38065_at	P	P	1.5	0.105			1.1
Homo sapiens clone 24416 mRNA sequence	AF052159		35342_at	P	P	1.5	0.253			
calcitonin receptor-like	L76380	NM_005795	34995_at	P	P	1.5	0.509			1.4
KIAA0582 protein	AI761647	NM_015147	40191_s_at	M	P	1.5	0.558			1.4
cDNA DKFZp434B102 (from clone DKFZp434B102)	AL080192		38630_at	A	P	1.5	0.719			
cDNA DKFZp586G1922 (from clone	AL080110		39600_at	P	P	1.5	0.160			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
DKFZp586G1922)										
Acyl-CoA synthetase 3	D89053	NM_004457	33880_at	P	P	1.5	0.264			
fatty-acid-Coenzyme A ligase, long-chain 3	*AA977580*	*NM_004457*	*33881_at*	*P*	*P*	1.0	0.120			*1.7*
STAT induced STAT inhibitor-2	AF037989		38994_at	A	P	1.5	0.391			
Homeotic Protein Hox5.4	HG3502-HT3696		696_at	P	P	1.5	0.181			
hypothetical protein FLJ13910, cDNA DKFZp586M141 (from clone DKFZp586M141)	AL050139	NM_022780	36580_at	P	P	1.5	0.228			
cDNA DKFZp586N012 (from clone DKFZp586N012)	AL049471		41690_at	P	P	1.4	0.320			
UbcH10, ubiquitin carrier protein E2-C	U73379	NM_007019	1651_at	P	P	1.4	0.022			1.1
cyclin-dependent kinase inhibitor 3, protein tyrosine phosphatase (CIP2)	L25876	NM_005192	1599_at	P	P	1.4	0.431			
glycogen phosphorylase (PYGL)	AF046798		37215_at	P	P	1.4	0.423			1.5
Angiopoietin-2	AF004327	NM_001147	1951_at	P	P	1.4	0.175	+		1.2
Angiopoietin-2	*AF004327*	*NM_001147*	*37461_at*	*P*	*P*	1.2	0.134	+		
forkhead box M1	U74612	NM_021953	34715_at	M	P	1.4	0.367			1.4
potentially prenylated protein tyrosine phosphatase hPRL-3	AF041434	NM_007079	36008_at	A	P	1.4	0.094			2.3
RAB31, Low Mr GTP-binding protein of the Rab subfamily	U59877	NM_006868	33371_s_at	P	P	1.4	0.299			
RAB31, member RAS oncogene family	*AI189226*	*NM_006868*	*33372_at*	*P*	*P*	1.2	0.444			
myosin VIIA	U39226	NM_000260	33197_at	P	P	1.4	0.038			1.2
Grb2-associated binder-1, docking protein related to IRS-1	U43885	NM_002039	1249_at	A	P	1.4	0.073			
lamin B1	L37747		37985_at	P	P	1.4	0.643			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
minichromosome maintenance deficient (mis5, S. pombe) 6 HsMcm6	D84557	NM_005915	40117_at	P	P	1.4	0.170			1.8
cyclin B1	M25753		1945_at	P	P	1.4	0.398			
cyclin B1	*M25753*		*34736_at*	*P*	*P*	*1.3*	*0.160*			
RTP, N-myc downstream regulated	D87953	NM_006096	36933_at	P	P	1.4	0.131			
alpha2,3-sialyltransferase	AB022918	NM_006100	39298_at	P	P	1.4	0.150			
ADP-ribosylation factor-like protein 4	U73960	NM_005738	33796_at	P	P	1.4	0.281			
centromere protein F (350/400kD, mitosin)	U30872	NM_016343	37302_at	A	P	1.4	0.245			
paternally expressed 10, KIAA1051	AB028974	NM_015068	39696_at	P	P	1.4	0.300			4.2
tubulin, alpha 1 (testis specific)	X06956		36591_at	M	P	1.4	0.300			1.8
KIAA0101	D14657	NM_014736	38116_at	P	P	1.4	0.409			
KIAA0128, septin 2	D50918		38826_at	P	P	1.4	0.381			1.8
protein phosphatase 2, regulatory subunit B (B56), gamma	Z69030	NM_002719	40785_g_at	P	P	1.4	0.453			
deoxycytidine kinase	M60527	NM_000788	886_at	A	P	1.3	0.455			
integrin beta 3 binding protein (beta3-endonexin)	U37139	NM_014288	38501_s_at	P	P	1.3	0.171			
TAL1 (SCL) interrupting locus	M74558	NM_003035	32767_at	M	P	1.3	0.150			
KIAA0666	AB014566		33753_at	P	P	1.3	0.356			
cAMP-specific phosphodiesterase 8A, PDE8A1	AF056490		37676_at	P	P	1.3	0.222			
mitotic checkpoint kinase Mad3L (MAD3L), BUB1B	AF053306	NM_001211	35699_at	P	P	1.3	0.216			1.6
ribosomal S6 kinase	X85106	NM_021135	32892_at	P	P	1.3	0.145			
HPTP epsilon (protein tyrosine phosphatase epsilon)	X54134	NM_006504	32916_at	P	P	1.3	0.100			1.0

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
Lyn tyrosine kinase, v-yes-1 Yamaguchi sarcoma viral related oncogene homolog	M79321	NM_002350	2024_s_at	P	P	1.3	0.054	+		
lyn tyrosine kinase, v-yes-1 Yamaguchi sarcoma viral related oncogene homolog	*M16038*	*NM_002350*	*1402_at*	*P*	*P*	*1.3*	*0.382*	+		
lyn tyrosine kinase	*M16038*	*NM_002350*	*32616_at*	*P*	*P*	1.2	0.066	+		
brachyury variant A (TBX1), T-box 1 transcription factor	AF012130	NM_005992	32285_g_at	P	P	1.3	0.352			
mki67a mRNA (long type) for antigen of monoclonal antibody Ki-67	X65550	NM_002417	418_at	A	P	1.3	0.357			1.5
protein tyrosine phosphatase receptor pi (PTPRP)	U81561	NM_002847	36160_s_at	P	P	1.3	0.193			
cbl-b	U26710	NM_004351	514_at	A	P	1.3	0.482			
Cyclin A2	X51688	NM_001237	1943_at	P	P	1.3	0.277			
nucleoside phosphorylase	X00737	NM_000270	430_at	P	P	1.3	0.272			
TNF-related apoptosis inducing ligand TRAIL	U37518	NM_003810	1715_at	P	P	1.3	0.316			
phosphodiesterase 4B, cAMP-specific	L20971	NM_002600	33705_at	P	P	1.3	0.275			
nidogen (enactin)	M30269	NM_002508	35366_at	P	P	1.3	0.050			
HYA22 protein	D88153	NM_005808	40196_at	P	P	1.3	0.150			1.3
phosphatidic acid phosphatase type 2A	AF014402	NM_003711	34797_at	P	P	1.3	0.191			
KIAA0512, ALEX2	AB011084	NM_014782	36057_at	P	P	1.2	0.268			
thromboxane A2 receptor	D38081	NM_001060	336_at	M	P	1.2	0.385			2.4
trefoil factor 3 (intestinal)	AI985964	NM_003226	37897_s_at	P	P	1.2	0.183			
G-2 and S-phase expressed 1	AL031588	NM_016426	41660_at	A	P	1.2	0.424			3.1

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
ADP-ribosyltransferase (NAD+; poly (ADP-ribose) polymerase)-like 2	AJ236876	NM_005484	34756_g_at	M	P	1.2	0.362			1.1
serine/threonine kinase 12	AF015254	NM_004217	33266_at	P	P	1.2	0.126			
Tubulin, Alpha 1, Isoform 44	HG2259-HT2348		330_s_at	P	P	1.2	0.096			1.1
lamin B receptor	L25931	NM_002296	288_s_at	P	P	1.2	0.141			
KIAA0429	AB007889	NM_014751	37363_at	P	P	1.2	0.150			2.5
transcription factor 4	M74719	NM_003199	36605_at	P	P	1.2	0.050			1.1
syndecan 3 (N-syndecan), KIAA0468	AB007937	NM_014654	32092_at	P	P	1.2	0.206			1.0
RECK protein precursor	AA099265	NM_021111	35236_g_at	P	P	1.2	0.173			
Putative prostate cancer tumor suppressor	U42349	NM_006765	36852_at	P	P	1.1	0.082			1.1
protein phosphatase 1, regulatory (inhibitor) subunit	AB020630		41577_at	P	P	1.1	0.082			
PDZ and LIM domain 1 (elfin)	U90878	NM_020992	36937_s_at	P	P	1.1	0.096			
hypothetical protein from clone 643	AF091087	NM_020467	34176_at	P	P	1.1	0.096			1.1
p53-regulated DDA3	AA926959		37347_at	P	P	1.1	0.058			
KIAA0062	D31887		38797_at	P	P	1.1	0.058			
medium-chain acyl-CoA dehydrogenase	M91432		37532_at	P	P	1.1	0.308			
gap junction protein, alpha 1, 43kD (connexin 43)	M65188	NM_000165	2018_at	P	P	1.1	0.329			
MyoD family inhibitor	U78313	NM_005586	38156_at	P	P	1.1	0.381			
endo/exonuclease Mre11 (MRE11A)	AF073362	NM_005591	32869_at	P	P	1.1	0.642			
nuclear receptor subfamily 2, group F, member 1	X16155	NM_005654	39294_at	A	P	1.0	0.446			

accession numbers	Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio

[1] A measurement indicating whether the transcript was detected (present, P), not detected (absent, A) or marginally detected (marginal, M; also if P in one experiment but A in another)

[2] The change in expression level for a transcript between two independently harvested BECs and LECs (= total of 4 comparisons). The change is expressed as the log2 ratio.

[3] Standard deviation of the change in the expression level (in 4 comparisons)

NB= Northern blot, IF= immunofluorescence

Table 4

Blood Vascular EC (222 genes)

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
p27 mRNA, interferon alpha-inducible protein 27	X67325	NM_005532	425_at	P	A	8.3	0.620			1.5
ribonuclease A (RNase A), pancreatic	D26129	NM_002933	37402_at	P	A	7.2	0.208			2.7
hematopoietic neural membrane protein (HNMP-1)	U87947	NM_001425	39182_at	P	A	5.9	0.381			1.6
N-cadherin	M34064	NM_001792	2054_g_at	P	A	5.7	1.345		+	
N-cadherin	*M34064*	*NM_001792*	*2053_at*	*P*	*P*	3.7	0.514		+	
interleukin 8 (IL8)	M28130	NM_000584	1369_s_at	P	A	5.3	1.477	+		
interleukin 8, beta-thromboglobulin-like protein precursor	*M17017*	*NM_000584*	*35372_r_at*	*P*	*P*	2.8	0.406	+		*1.1*
tyrosine kinase receptor (axl)	M76125	NM_001699	38433_at	P	A	5.1	1.112	+		1.0
Tyrosine Kinase, Receptor Axl, Alt. Splice 2	*HG162-HT3165*		*1278_at*	*P*	*A*	4.7	0.937	+		*1.1*
cell surface glycoprotein CD44 (CD44)	L05424		1126_s_at	P	A	4.9	1.527			1.9
cell adhesion molecule (CD44)	*M59040*	*NM_000610*	*2036_s_at*	*P*	*A*	1.9	0.402			*2.6*
hyaluronate receptor (CD44)	*L05424*		*40493_at*	*P*	*P*	1.9	0.136			*2.0*
vascular endothelial growth factor related protein VRP, VEGF-C	U43142	NM_005429	159_at	P	A	4.6	0.850	+		
Vascular endothelial growth factor C	*X94216*	*NM_005429*	*1934_s_at*	*P*	*A*	4.4	1.342	+		

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
collagen type XIII, alpha 1 (=COL4A2)	M33653	NM_005203	38952_s_at	P	A	4.5	0.213			2.7
collagen type XIII, alpha-1	*M59217*	*NM_005203*	*38951_at*	*P*	*A*	3.6	1.683			*1.1*
collagen alpha-2 type I	K01079		32306_g_at	P	A	4.5	2.161			
collagen alpha-2 type I	*K01079*		*32305_at*	*P*	*A*	2.8	1.464			
collagen, type I, alpha 2	*V00503*	*NM_000089*	*32307_s_at*	*P*	*A*	2.4	1.394			
proteoglycan 1	X17042	NM_002727	32227_at	P	P	4.3	0.385			1.0
phospholipase A2, group IVA, calcium-dependent phospholipid-binding protein (PLA2)	M72393		35938_at	P	A	4.3	2.398			1.9
carbohydrate (keratan sulfate Gal-6) sulfotransferase	AB003791	NM_003654	41395_at	P	P	4.2	0.232			
tropomyosin 2 (beta), fibroblast tropomyosin	M12125	NM_003289	32314_g_at	P	A	4.2	1.845			
chondroitin sulfate proteoglycan 2 (versican)	X15998	NM_004385	38111_at	P	A	4.1	1.746			1.5
chondroitin sulfate proteoglycan 2 (versican)	*X15998*	*NM_004385*	*38112_g_at*	*P*	*A*	2.0	1.219			
latent transforming growth factor-beta binding protein (LTBP-2)	Z37976	NM_000428	37906_at	P	P	4.1	0.381			1.1
interleukin 6 (interferon, beta 2)	X04430	NM_000600	38299_at	P	A	4.0	0.776			2.0
bone morphogenetic protein-4 (hBMP-4)	U43842	NM_001202	40333_at	P	A	4.0	0.883			
bone morphogenetic protein 2B, BMP-4	*M22490*	*NM_001202*	*1114_at*	*P*	*A*	2.6	1.146			
sarcolectin, keratin 7	AJ238246	NM_005556	41294_at	P	P	3.9	0.631			
neuronal cell adhesion molecule, KIAA0343	AB002341	NM_005010	37286_at	P	A	3.9	1.642			1.8
neuronal cell adhesion molecule, hBRAVO/Nr-CAM precursor	*U55258*	*NM_005010*	*37288_g_at*	*P*	*A*	1.5	0.309			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
matrix metalloproteinase 1 (interstitial collagenase), skin collagenase	M13509	NM_002421	38428_at	P	P	3.9	0.521	+		
stem cell factor, KIT ligand	M59964	NM_000899	597_at	P	A	3.9	0.554	+		
uPA	X02419	NM_002658	37310_at	P	A	3.8	0.282			3.0
plasminogen activator inhibitor-1	J03764	NM_000602	672_at	P	P	3.7	0.161			1.6
plasminogen activator inhibitor 1	*M14083*	*NM_000602*	*38125_at*	*P*	*P*	2.9	0.118			*1.9*
selectin P, CD62, granule membrane protein-140 (GMP-140) precursor	M25322	NM_003005	40366_at	P	P	3.6	1.869			1.2
latrophilin-2	AJ131581	NM_012302	34174_s_at	P	A	3.6	0.098			1.1
actin, alpha 2	X13839	NM_001613	32755_at	P	P	3.6	1.067			
fibroblast activation protein, alpha	U09278	NM_004460	39945_at	P	A	3.6	0.789			1.2
regulator of G-protein signalling 20	AF060877	NM_003702	41086_at	P	A	3.5	0.615			
IGF-II mRNA-binding protein 3	U97188	NM_006547	37558_at	P	P	3.5	0.528			2.1
retina cDNA randomly primed sublibrary, EST	W28438		36497_at	P	A	3.5	0.414			
brain acid-soluble protein 1, neuronal tissue-enriched acidic protein (NAP-22)	AF039656	NM_006317	32607_at	P	A	3.4	0.104			1.6
profilin 2	AL096719	NM_002628	38839_at	P	P	3.4	0.111			1.6
profilin 2	*L10678*	*NM_002628*	*38840_s_at*	*P*	*P*	*3.1*	*0.076*			*1.6*
Na,K-ATPase beta-1 subunit	U16799	NM_001677	37669_s_at	P	A	3.4	0.249			
Claudin-7	AJ011497	NM_001307	38482_at	P	A	3.4	0.798			
normal gingiva	U51712		39698_at	P	A	3.4	0.391			1.1
a disintegrin and metalloproteinase domain 23	AB009672	NM_003812	40350_at	P	A	3.4	0.600			
COL8A1 mRNA for alpha 1(VIII) collagen	X57527	NM_001850	37459_at	P	A	3.3	0.819	+		1.7

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
signal transducer and activator of transcription 6 (STAT6)	AF067575		41222_at	P	P	3.3	0.338	+	+	1.8
transcription factor IL-4 Stat, STAT6	*U16031*	*NM_003153*	*845_at*	*P*	*A*	2.1	0.493	+	+	*1.6*
lipocortin-III, annexin A3	M20560	NM_005139	31792_at	P	P	3.3	0.265			1.1
intercellular adhesion molecule 1 (CD54), major group rhinovirus receptor precusor	M24283	NM_000201	32640_at	P	A	3.2	0.184	+		
solute carrier family 1 (neuronal/epithelial high affinity glutamate transporter, system Xag)	U08989	NM_004170	38268_at	P	A	3.2	0.687			1.3
solute carrier family 1 (neuronal/epithelial high affinity glutamate transporter, system Xag)	*AI928365*	*NM_004170*	*38267_at*	*P*	*A*	*2.6*	*0.524*			*1.2*
p53 inducible protein	L47738		37579_at	P	A	3.2	0.249			1.0
dihydropyrimidine dehydrogenase, DPYD	U20938	NM_000110	38220_at	P	A	3.2	0.306			
natural killer cell transcript 4	AA631972	NM_004221	39119_s_at	P	P	3.1	0.082			1.6
PFTAIRE protein kinase 1, KIAA0834	AB020641	NM_012395	36502_at	P	A	3.1	0.841			
RGP4, regulator of G-protein signalling 4	U27768	NM_005613	34272_at	P	A	3.0	0.396			1.5
regulator of G-protein signalling 4	*AI267373*	*NM_005613*	*34273_at*	*P*	*A*	2.6	0.473			*1.5*
Oncogene Aml1-Evi-1, Fusion Activated	HG4058-HT4328		1882_g_at	P	M	2.9	0.590			1.5
Oncogene Aml1-Evi-1, Fusion Activated	*HG4058-HT4328*		*1881_at*	*P*	*A*	2.0	0.374			
adenylyl cyclase-associated protein 2	N90755	NM_006366	33405_at	P	A	2.8	0.596			1.5
clusterin (complement lysis inhibitor, SP-40,40, sulfated	M25915	NM_001831	36780_at	P	P	2.8	0.161			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
glycoprotein 2, apolipoprotein J)										
ADP ribosylation factor-like 7	AB016811	NM_005737	39829_at	P	A	2.7	0.531			
H factor (complement)-like 1	M65292	NM_002113	32249_at	P	A	2.7	0.589			
RNA helicase-related protein, metallothionein-If	H68340	NM_007372	41446_f_at	P	P	2.7	0.296			
stimulated trans-acting factor (50 kDa) Staf50	X82200	NM_006074	36825_at	P	A	2.7	0.730			6.2
cyclooxygenase-2 (hCox-2)	U04636	NM_000963	1069_at	P	A	2.6	1.281			
GRO1 oncogene, melanoma growth stimulatory activity (MGSA)	X54489	NM_001511	408_at	P	A	2.6	0.369			
NRGN, neurogranin	X99076	NM_006176	33925_at	M	A	2.6	1.237			
homologue of mouse dkk-1	AB020315		35977_at	P	A	2.5	0.398			
gastrointestinal tumor-associated antigen GA733-1, tumor-associated calcium signal transducer 2	J04152	NM_002353	291_s_at	P	P	2.5	0.139			
laminin	Z15008	NM_005562	35280_at	P	A	2.5	0.824			
transgelin, 22kDa smooth muscle protein (SM22)	M95787	NM_003186	36931_at	P	A	2.5	0.980			
JE gene encoding a monocyte secretory protein	M28225		34375_at	P	P	2.4	0.186			3.6
zinc finger protein 238, RP58	AJ223321	NM_006352	35824_at	P	A	2.4	0.498			3.2
cathepsin C	X87212	NM_001814	133_at	P	P	2.4	0.244			
tissue-type plasminogen activator (t-PA)	M15518	NM_000930	33452_at	P	A	2.4	0.479			
sushi-repeat protein	AF060567	NM_014467	37805_at	P	A	2.4	0.670			
annexin A6	D00510	NM_001155	39082_at	P	A	2.4	0.181			1.2
EphrinB1	U09303	NM_004429	39721_at	P	A	2.4	0.946			
EphrinB1	*U09303*	*NM_004429*	*188_at*	*P*	*A*	1.2	0.489			
TFEC isoform (transcription factor	D43945	NM_012252	34470_at	P	A	2.4	0.028			3.1

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
EC)										
small inducible cytokine A2, (monocyte chemotactic protein1)	M26683	NM_002982	875_g_at	P	P	2.3	0.283			3.5
small inducible cytokine A2 (monocyte chemotactic protein 1)	*M26683*	*NM_002982*	*874_at*	*P*	*A*	*1.2*	*0.385*			
endothelial cell protein C/APC receptor (EPCR)	L35545	NM_006404	647_at	P	P	2.3	0.259			2.2
transglutaminase 2 (TGase)	M55153	NM_004613	231_at	P	M	2.3	0.413			
transglutaminase (TGase)	*M55153*	*NM_004613*	*38404_at*	*P*	*P*	1.6	0.093			
human metallothionein-If	M10943		31622_f_at	P	M	2.3	0.230			
transforming growth factor beta-induced (BIGH3)	M77349	NM_000358	1385_at	P	A	2.3	0.951			
ENO2 gene for neuron specific (gamma) enolase	X51956		40193_at	P	A	2.3	0.121			1.6
FAT tumor suppressor (Drosophila) homolog	X87241	NM_005245	40454_at	P	A	2.3	1.204			
malignant cell expression-enhanced gene/tumor progression-enhanced gene	S82470	NM_024298	181_g_at	P	P	2.2	0.469			
malignant cell expression-enhanced gene/tumor progression-enhanced gene	*S82470*	*NM_024298*	*180_at*	*P*	*A*	1.7	0.420			
cDNA DKFZp566G0746 (from clone DKFZp566G0746)	AL050078		39324_at	P	A	2.2	1.281			
lysyl oxidase-like 2	U89942	NM_002318	33127_at	P	P	2.2	0.274			
ras-related C3 botulinum toxin substrate 2 (rho family, small GTP binding protein Rac2)	M64595	NM_002872	32737_at	P	P	2.2	0.143			3.5
endothelial leukocyte adhesion molecule 1 (ELAM-1), selectin E	M24736	NM_000450	265_s_at	P	A	2.2	0.448			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
laminin, alpha 5, KIAA0533	AB011105		41610_at	P	P	2.2	0.146			
placenta growth factor (PlGF)	X54936	NM_002632	793_at	P	P	2.2	0.301	+		1.0
ALL1-fused gene from chromosome 1q, AF1q	U16954	NM_006818	36941_at	P	P	2.2	0.592			
stromelysin-2, MMP-10	X07820	NM_002425	1006_at	P	P	2.1	0.159			1.0
metallothionein-I-A	K01383		31623_f_at	P	P	2.1	0.238			
collagen VI alpha-1	X15880		38722_at	P	A	2.1	0.898			
mad protein homolog (hMAD-3)	U68019	NM_005902	1433_g_at	P	A	2.1	0.269			1.1
mad protein homolog (hMAD-3)	*U68019*	*NM_005902*	*38944_at*	*P*	*A*	1.8	0.367			*1.4*
mad protein homolog (hMAD-3)	*U68019*	*NM_005902*	*1454_at*	*P*	*A*	2.0	0.304			
integral membrane protein 2A	AL021786		40775_at	P	P	2.1	0.570			
interleukin 1 receptor-like 1	D12763	NM_003856	40322_at	P	A	2.1	0.718			
high-mobility group (nonhistone chromosomal) protein isoform I-C (HMGI-C)	X92518		35200_at	P	A	2.0	0.177			
epidermal growth factor receptor kinase substrate (Eps8)	U12535	NM_004447	1467_at	P	A	2.0	0.710			1.0
lactate dehydrogenase B	X13794	NM_002300	33819_at	P	P	2.0	0.029			1.6
mRNA for unknown product	D29810		40227_at	P	A	2.0	1.170			
hypothetical protein DKFZp564D0462	AL033377		36014_at	P	A	2.0	0.137			1.1
lysyl hydroxylase isoform 2 (PLOD2)	U84573	NM_000935	34795_at	P	P	2.0	0.157			
follistatin-like 3, follistatin-related protein (FLRG)	U76702	NM_005860	33900_at	P	A	2.0	0.075			2.6
Homo sapiens clone 24674 mRNA sequence	AF070578		36758_at	P	A	2.0	0.612			1.8
L-iditol-2 dehydrogenase	L29254		38763_at	P	A	1.9	0.140			
neuronal pentraxin 1	U61849	NM_002522	37921_at	P	A	1.9	0.744			2.5

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
hypothetical protein from clones 23549 and 23762	U90908	NM_021226	34010_at	P	A	1.9	0.870			
UDP-N-acetylglucosamine pyrophosphorylase	AB011004	NM_003115	41242_at	P	P	1.9	0.342			
zinc finger protein 185 (LIM domain)	Y09538	NM_007150	32139_at	P	A	1.8	0.062			1.6
four and a half LIM domains 2, heart protein (FHL-2)	U29332	NM_001450	38422_s_at	P	P	1.8	0.229			
mitogen-activated protein kinase-activated protein kinase 3, MAPKAP kinase (3pK)	U09578	NM_004635	1637_at	P	A	1.8	0.454			1.4
metallothionein 1E (functional)	R92331		36130_f_at	P	P	1.8	0.131			
TU3A protein	AF035283	NM_007177	38044_at	P	A	1.8	0.298			1.8
metallothionein 1H	R93527	NM_005951	39594_f_at	P	P	1.8	0.415			
guanylate binding protein isoform II (GBP-2)	M55543	NM_004120	32700_at	P	P	1.8	0.304			1.1
soluble vascular endothelial cell growth factor receptor 1 (sVEGFR-1)	U01134	NM_002019	1964_g_at	P	M	1.8	0.384	+		
R-Ras	M14949		38338_at	P	P	1.8	0.119			
R-ras	*M14949*		*1879_at*	*P*	*P*	*1.2*	*0.330*			*1.5*
creatine transporter (SLC6A8), solute carrier family 6, member 8	U36341	NM_005629	40926_at	P	A	1.8	0.305			
target of myb1 (chicken) homolog, Heme Oxygenase 1 (HO-1)	Z82244	NM_005488	33802_at	P	P	1.8	0.149			
procollagen-lysine, 2-oxoglutarate 5-dioxygenase, lysyl hydroxylase (PLOD)	L06419	NM_000302	36184_at	P	P	1.8	0.310			
KIAA0836	AB020643		33296_at	P	P	1.8	0.203			1.8
cDNA DKFZp434C171 (from clone	AL080169		34183_at	P	A	1.8	0.606			1.2

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
DKFZp434C171)										
IL-4-R mRNA for the interleukin 4 receptor	X52425	NM_000418	404_at	P	P	1.7	0.244			
chemokine (C-C motif) receptor-like 2 (CCRL2), chemokine receptor X (CKRX)	AF014958	NM_003965	1445_at	P	A	1.7	0.386			1.8
phospholipase C, beta 3 (phosphatidylinositol-specific)	Z16411	NM_000932	364_s_at	P	A	1.7	0.098			
LIM domain protein	X93510	NM_003687	32610_at	P	P	1.7	0.067			1.9
protein kinase (cAMP-dependent, catalytic) inhibitor beta	M34181	NM_002731	36215_at	P	A	1.7	0.315			1.0
rho GDP-dissociation Inhibitor 2	X69549	NM_001175	1984_s_at	P	P	1.7	0.163			2.4
KIAA0975, imidazoline receptor candidate	AB023192	NM_007184	33916_at	P	A	1.7	0.343			
poliovirus receptor	X64116	NM_006505	32698_at	P	P	1.7	0.173			
poliovirus receptor	*X64116*	*NM_006505*	*32699_s_at*	*P*	*P*	1.3	0.102			
immediate early response 3	S81914	NM_003897	1237_at	P	P	1.7	0.171			
metallothionein 2A	AI547258	NM_005953	39081_at	P	A	1.7	0.247			
tropomyosin 1 (alpha)	M19267	NM_000366	36791_g_at	P	P	1.6	0.184			1.0
tropomyosin 1 (alpha)	*Z24727*	*NM_000366*	*36792_at*	*P*	*P*	1.4	0.094			*1.3*
tropomyosin 1 (alpha)	*M19267*	*NM_000366*	*36790_at*	*P*	*P*	1.2	0.213			
TRAM-like protein	D31762	NM_012288	40051_at	P	P	1.6	0.244			1.1
E3 ubiquitin ligase SMURF2	AA630312	NM_022739	33354_at	P	P	1.6	0.244			2.2
EGF-containing fibulin-like extracellular matrix protein 1	U03877	NM_004105	32551_at	P	P	1.6	0.152			
G protein-coupled receptor 56	AJ011001	NM_005682	35769_at	P	P	1.6	0.075			
c-jun proto oncogene (JUN)	J04111	NM_002228	32583_at	P	A	1.6	0.377			
regulator of G-protein signalling 10, RGS10	AF045229	NM_002925	33121_g_at	P	A	1.6	0.064			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
amyloid beta (A4) precursor protein-binding, family B, member 2 (Fe65-like)	U62325		40148_at	P	P	1.6	0.312			1.3
ras-related rho protein	M12174	NM_004040	1826_at	P	P	1.6	0.281			1.4
proteasome (prosome, macropain) 26S subunit, non-ATPase, 10	AL031177	NM_002814	37957_at	P	A	1.5	0.265			
KIAA0537	AB011109	NM_014840	33787_at	P	P	1.5	0.135			1.8
lysosome-associated membrane protein-2	X77196	NM_002294	38403_at	P	P	1.5	0.432			
phospholipid transfer protein	L26232	NM_006227	40081_at	P	P	1.5	0.046			
N-myristoyltransferase 2	AF043325	NM_004808	41656_at	P	P	1.5	0.038			
phosphofructokinase (PFKM)	U24183	NM_000289	36196_at	P	P	1.5	0.374			2.0
integrin, beta 4	X53587	NM_000213	406_at	P	A	1.5	0.195			
leupaxin	AF062075	NM_004811	36062_at	P	A	1.5	0.231			1.3
endothelin-converting-enzyme 1	Z35307	NM_001397	41726_at	P	P	1.5	0.180			
wild-type p53 activated fragment-1 (WAF1), cyclin-dependent kinase inhibitor 1A (p21, Cip1)	U03106	NM_000389	2031_s_at	P	P	1.5	0.399			
ICAM-2, cell adhesion ligand for LFA-1	X15606	NM_000873	38453_at	P	P	1.5	0.024			1.5
ICAM-2, cell adhesion ligand for LFA-1	*X15606*	*NM_000873*	*38454_g_at*	*P*	*P*	1.5	0.153			*2.0*
intercellular adhesion molecule 2 (ICAM-2)	*M32334*		*590_at*	*P*	*P*	1.4	0.152			*1.5*
eukaryotic translation initiation factor 2B, eIF-2B beta subunit	AF035280	NM_014239	40515_at	P	P	1.5	0.108			
uridine phosphorylase	X90858	NM_003364	37351_at	P	M	1.5	0.064			
integrin, beta 5	X53002	NM_002213	39754_at	P	P	1.5	0.068			
N-sulfoglucosamine sulfohydrolase (sulfamidase)	U30894	NM_000199	35626_at	P	P	1.5	0.077			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
synaptojanin 2	AF039945		36532_at	P	A	1.5	0.164			1.1
metallothionein 1L	AA224832	NM_002450	39120_at	P	A	1.4	0.664			
macrophage capping protein, gelsolin-like	M94345	NM_001747	38391_at	P	P	1.4	0.281			
HSPC022 protein	W68830	NM_014029	32736_at	P	P	1.4	0.062	-		2.7
Human clone 137308 mRNA, partial cds	AW006742		38207_at	P	A	1.4	0.442			
protocadherin 42, PC42, protocadherin 1 (cadherin-like 1)	L11370	NM_002587	37562_at	P	A	1.4	0.166			
caspase-like apoptosis regulatory protein 2 (CLARP2)	AF005775	NM_003879	1867_at	P	P	1.4	0.363			1.5
caspase-like apoptosis regulatory protein 2 (CLARP2)	*AF005775*	*NM_003879*	*1868_g_at*	*P*	*P*	1.2	0.325			*2.2*
major vault protein, lrp	X79882	NM_005115	38064_at	P	P	1.4	0.252			1.0
Fanconi anemia, complementation group G	AC004472	NM_004629	33842_at	P	A	1.4	0.233			
prion protein (PrP)	U29185	NM_000311	36159_s_at	P	P	1.4	0.342			1.1
interferon-stimulated protein, 15 kDa	AA203213	NM_005101	38432_at	P	A	1.4	0.244			
serine (or cysteine) proteinase inhibitor, clade B (ovalbumin), cytoplasmic antiproteinase 2 (CAP2)	L40377	NM_002640	36312_at	P	A	1.3	0.360			2.2
biglycan	J04599	NM_001711	38126_at	P	P	1.3	0.101			
chemokine (C-X-C motif), receptor 4 (fusin)	L06797	NM_003467	649_s_at	P	P	1.3	0.177	+		
ubiquitin carboxyl-terminal esterase L1 (ubiquitin thiolesterase)	X04741	NM_004181	36990_at	P	P	1.3	0.117			
KIAA0469	AB007938	NM_014851	37230_at	P	P	1.3	0.124			
TNF (ligand) superfamily, member	AL022310	NM_003326	32319_at	P	A	1.3	0.349			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
4 (tax-transcriptionally activated glycoprotein 1, 34kD)										
KIAA1053	AB028976		40855_at	P	P	1.3	0.242			1.6
NAD(P)H-quinone oxireductase	M81600		38066_at	P	P	1.3	0.058			1.8
sushi-repeat-containing protein	U61374	NM_006307	31855_at	P	P	1.3	0.610			1.2
integrin, alpha 5	X06256	NM_002205	39753_at	P	P	1.3	0.179		+	1.2
enigma (LIM domain protein)	L35240	NM_005451	39530_at	P	P	1.3	0.396			
ectonucleoside triphosphate diphosphohydrolase 1	AJ133133	NM_001776	32826_at	P	A	1.3	0.412			1.5
transforming growth factor-beta (tgf-beta), bone morphogenetic protein 6	M60315	NM_001718	39279_at	P	P	1.3	0.206			
transforming growth factor-beta (tgf-beta), bone morphogenetic protein 6	*M60315*	*NM_001718*	*1733_at*	*P*	*P*	1.0	0.308			
nicotinamide N-methyltransferase, NNMT	U08021	NM_006169	37032_at	P	P	1.2	0.083			2.1
cDNA DKFZp564J0323 (from clone DKFZp564J0323)	AL049957		39170_at	P	P	1.2	0.264			1.2
thioredoxin reductase beta	AB019694	NM_006440	41711_at	P	A	1.2	0.206			
f-box and leucine-rich repeat protein 2	AL049953		36525_at	P	A	1.2	0.300			1.2
transcobalamin II (TCN2)	L02648	NM_000355	37922_at	P	A	1.2	0.342			1.2
aldehyde dehydrogenase 2, mitochondrial	X05409	NM_000690	32747_at	P	P	1.2	0.117			
GTP-binding protein ragB	X90530	NM_006064	39989_at	P	M	1.2	0.602			
lymphocyte antigen 75	AF011333	NM_002349	38160_at	P	A	1.2	0.132			1.7
GM2 activator protein	X62078		35820_at	P	P	1.2	0.101			1.5
type 3 inositol 1,4,5-trisphosphate receptor (ITPR3)	U01062	NM_002224	182_at	P	P	1.2	0.052			1.5

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
KIAA0284	AI828210		38592_s_at	P	P	1.2	0.078			1.9
metallothionein I-B	M13485		609_f_at	P	P	1.2	0.266			
BTG2	U72649	NM_006763	36634_at	P	P	1.2	0.210			
adenylate kinase 1	J04809	NM_000476	36997_at	P	A	1.2	0.246			
tumor necrosis factor receptor superfamily, member 12, WSL-LR, WSL-S1 and WSL-S2 proteins	Y09392	NM_003790	41189_at	P	P	1.2	0.350			
aminopeptidase N/CD13	M22324	NM_001150	39385_at	P	P	1.2	0.398			
growth arrest and DNA-damage-inducible protein (gadd45)	M60974	NM_001924	1911_s_at	P	P	1.2	0.177			
KIAA0638 protein	AB014538		37375_at	P	P	1.2	0.680			
vinculin	M33308	NM_003373	36601_at	P	P	1.2	0.078		+	
procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha polypeptide II	U90441	NM_004199	34390_at	P	P	1.1	0.347			
msg1-related gene 1 (mrg1), Cbp/p300-interacting transactivator	U65093	NM_006079	33113_at	P	P	1.1	0.164			1.2
microsomal glutathione S-transferase 3	AF026977	NM_004528	39018_at	P	P	1.1	0.191			1.1
vitamin A responsive; cytoskeleton related	AF070523	NM_006407	39091_at	P	P	1.1	0.216			2.9
17-kDa protein, interferon-stimulated protein, 15 kDa	M13755	NM_005101	1107_s_at	P	P	1.1	0.119			
matrix metalloproteinase 14 (membrane-inserted)	X83535	NM_004995	34747_at	P	A	1.1	0.487			2.4
4F2 cell-surface antigen, solute carrier family 3, member 2	J02939	NM_002394	38029_at	P	P	1.1	0.143			
metallothionein-III	M93311	NM_005954	870_f_at	P	P	1.1	0.334			
protein kinase (cAMP-dependent, catalytic) inhibitor alpha	S76965	NM_006823	36202_at	P	P	1.1	0.046			

	accession numbers		Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
protein kinase (cAMP-dependent, catalytic) inhibitor alpha	*S76965*	*NM_006823*	*546_at*	*P*	*P*	1.0	0.367			
reticulocalbin 1, EF-hand calcium binding domain	D42073	NM_002901	40556_at	P	P	1.1	0.035			1.1
lipin 1, KIAA0188	D80010		38098_at	P	P	1.1	0.080			1.4
protease, serine, 23	AF015287	NM_007173	40078_at	P	P	1.0	0.099			1.3
hect domain and RLD 2	AF041080	NM_004667	40877_s_at	P	P	1.0	0.104			
GATA-binding protein (GATA2)	M68891	NM_002050	37194_at	P	P	1.0	0.325			1.0
agrin precursor	AF016903		33454_at	P	P	1.0	0.272			
equilibrative nucleoside transporter 1 (hENT1)	U81375	NM_004955	33901_at	P	P	1.0	0.352			
coronin, actin-binding protein 2B, KIAA0925	AB023142		34772_at	P	A	1.0	0.459			
f-box and WD-40 domain protein 3	U07000	NM_012165	537_f_at	P	M	1.0	0.212			
nonsyndromic hearing impairment protein (DFNA5)	AF073308	NM_004403	41872_at	P	P	1.0	0.535			1.0
actin filament associated protein	D25248	NM_021638	37578_at	P	P	1.0	0.218			
TNFR-related death receptor-6 (DR6)	AF068868	NM_014452	35402_at	P	A	1.0	0.235			1.7
serum/glucocorticoid regulated kinase	Y10032	NM_005627	973_at	P	A	1.0	0.174			
DNase X	X90392	NM_006730	37214_g_at	P	P	1.0	0.507			
DNase X	*X90392*	*NM_006730*	*37213_at*	*P*	*P*	1.0	0.376			
fatty acid desaturase 3	AC004770	NM_021727	34224_at	P	P	1.0	0.294			
LYL-1	M22637		39971_at	P	P	1.0	0.313			1.4
ATP-binding cassette, sub-family C (CFTR/MRP), member 1	X78338	NM_004996	34016_s_at	P	A	1.0	0.258			2.3
transmembrane protein (CD59)	M84349		39351_at	P	P	1.0	0.141			1.1
fms-related tyrosine kinase 1,	S77812	NM_002019	1545_g_at	P	P	1.0	0.535	+		1.9

	accession numbers	Affymetrix ID	Detection[1] BEC	LEC	Gene expression analysis signal log ratio[2]	s.d.[3]	Confirmed by: NB	IF	signal log ratio
VEGFR-1									

[1] A measurement indicating whether the transcript was detected (present, P), not detected (absent, A) or marginally detected (marginal, M; also if P in one experiment but A in another)

[2] The change in expression level for a transcript between two independently harvested BECs and LECs (= total of 4 comparisons). The change is expressed as the log2 ratio.

[3] Standard deviation of the change in the expression level (in 4 comparisons)

NB= Northern blot, IF= immunofluorescence

Table 10

Known LEC-specific genes

Gene	Detection *	Accession numbers starting EST	Accession numbers possible gene
CD36 =COL1/TSP receptor, fatty-acid transport protein	Af (S/4,3)	R20784 H54254	M98399
beta1-syntrophin	Af (S/4,5)	AA447177	L31529
collectin sub-family member 12	Af (S/4,5)	R74387	NM_030781
a disintegrin and metalloprotease domain 12	Af (S/4,3)	AA147933	NM_003474
cytotoxic T-lymphocyte-associated protein 4	Af (S/4,0)	AI733018	NM_005214
niban protein NM_022083 niban protein	Af (S/3,7)	AA554814	NM_052966
multi-PDZ-domain-containing protein, LNX	Af (S/3,5)	AI738919	NM_032622
MAGE-E1 protein	Af (S/3,2)	AI435112	NM_030801
upstream stimulatory factor 1, USF1 (genomic match)	Af (S/2,6)	AA701033	AB017568
hairy/enhancer-of-split related with YRPW motif 1	Af (NS/2,6)	R61374	NM_012258
alpha-2,8-polysialyltransferase	Af (S/2,5)	AI422986	L41680
semaphorin 6A1	Af (S/2,4)	W21965	NM_020796
guanine nucleotide binding protein (G prot), gamma 2	Af (S/2,3)	AA738022	
integral membrane protein 3	Af (S/2,3)	AA128019	NM_030926
similar to mouse glucocorticoid-induced gene 1	Af (S/2,0)	AI678080	XM_070471
YAP65 (Yes-associated protein of 65kDa MW)	Af (NS/2,0)	AL048399	X80507
17 kDa fetal brain protein	Af (NS/1,9)	H92988	NM_022343
Kruppel-like factor 5	Af (S/1,8)	AI815057	NM_001730
calcitonin receptor-like, CGRP type 1 receptor	Af (S/1,7)	AI741128, T94540	NM_005795, L76380
fibroblast growth factor 13, isoform 1A	Af (NS/1,7)	AW014749	NM_004114
tetraspan NET-6 protein	Af (NS/1,6)	W22687	NM_014399
ring finger protein 11	Af (S/1,6)	AL079648	BC020964

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent log2 ratio of the signal intensities between BEC and LEC

Table 11

Differentially expressed genes identified by accession number

Gene	Detection *	starting EST	SEQ ID NO:
EST	Af (S/4,9)	AL079386	1
EST	Af (S/3,7)	N21555	2
EST	Af (S/3,2)	AL119027	3
EST	Af (S/2,9)	H05299	4
EST	Af (S/2,8)	AA973128	5
EST	Af (NS/2,6)	AI128820	
EST	Af (S/2,3)	AW044647	6
EST	Af (S/2,2)	AI333058	7
EST	Af (S/2,1)	AI536067	8
EST	Af (NS/2,0)	AA156409	
EST	Af (S/1,9)	AI770080	9
EST	Af (NS/1,9)	AA456099	
EST	Af (S/1,8)	AI692645	10
EST	Af (S/1,7)	AL119265	11
EST	Af (S/1,6)	AI478114	12
EST	Af (S/1,6)	AI817448	13

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent log2 ratio of the signal intensities between BEC and LEC

Table 12

Other Proteins Identified

Gene	Detection *	Accession numbers starting EST	possible gene	
KIAA1392, hypothetical protein DKFZp762K222	Af (S/5,3)	N50545	XM_048721	(20)
similar to phosphoglucomutase 5	Af (S/4,5)	AL046941	XM_047649	(21)
Similar to transmembrane receptor Unc5H1	Af (S/4,5)	R56359	XM_030300	(22)
hypothetical protein MGC21854	Af (S/3,7)	AI659418	NM_052862	(23)
KIAA1877	Af (S/3,4)	AW004016		
similar to unnamed protein product	Af (NS/3,1)	AA036952	XM_085235	
unknown protein	Af (S/2,9)	AA846091	XM_038314	(24)
KIAA1058 (+ missing N-term from ests)	Af (S/2,6)	AA007697	AB028981	(25)
similar to KIAA1673	Af (S/2,3)	AI948598	XM_059607	(26)
similar to lysosomal amino acid transporter 1	Af (S/2,3)	AI692279	XM_058449	(27)
Homo sapiens similar to KIAA1673 protein	Af (S/2,3)	AI948598	XM_059607	
KIAA0493	Af (S/2,3)	AA532655	AB007962	(28)
hypothetical protein MGC2780	Af (S/2,3)	AI734962	NM_025266	(29)
transmembrane protein 2	Af (NS/2,3)	NM_013390	57094_at	
Novel human gene mapping to chomosome 1	Af (S/2,2)	AA651889	HS455J72	(30)

* Af=Affymetrix, S=specific for LEC, NS=nonspecific (also expressed in BEC), numbers represent log2 ratio of the signal intensities between BEC and LEC

In Tables 5, 6 and 12, the numbers in parentheses refer to the SEQ ID NO: in the Sequence Listing. Table 13 below correlates these sequences with polypeptide sequences SEQ ID NO:31-44 and 46 (Open reading frames, ORF's).

Table 13

Polypeptides corresponding to LEC-specific polynucleotides

Accession number	Polynucleotide	Polypeptide
NM_021647	SEQ ID NO:14	SEQ ID NO:31
NM_014817	SEQ ID NO:15	SEQ ID NO:32
XM_059074	SEQ ID NO:16	SEQ ID NO:33
NM_016647	SEQ ID NO:17	SEQ ID NO:34
XM_048721	SEQ ID NO:20	SEQ ID NO:35
XM_047649	SEQ ID NO:21	SEQ ID NO:36
XM_030300	SEQ ID NO:22	SEQ ID NO:37
NM_052862	SEQ ID NO:23	SEQ ID NO:38
XM_039314	SEQ ID NO:24	SEQ ID NO:39
AB028981	SEQ ID NO:25	SEQ ID NO:40
XM_059607	SEQ ID NO:26	SEQ ID NO:41
XM_058449	SEQ ID NO:27	SEQ ID NO:42
NM_025266	SEQ ID NO:29	SEQ ID NO:43
AL137762/HS455J72	SEQ ID NO:30	SEQ ID NO:44
XM_084655	SEQ ID NO:45	SEQ ID NO:46

Table 14

Sequence identifiers for sequences in Table 3

	accession numbers		aa SEQ ID NO:	nt SEQ ID NO:
lung type-I cell membrane-associated protein, podoplanin	AF030428	NM_006474	SEQ ID NO: 65	SEQ ID NO: 66
lung type-I cell membrane-associated protein, podoplanin	*AI660929*	*NM_006474*	duplicate	
cellular retinol-binding protein	M11433	NM_002899	SEQ ID NO: 67	SEQ ID NO: 68
macrophage mannose receptor (MRC1)	M93221		SEQ ID NO: 69	SEQ ID NO: 70
transcription factor C-MAF	AF055376	NM_005360	SEQ ID NO: 71	SEQ ID NO: 72
transcription factor C-MAF	*AF055376*	*NM_005360*	duplicate	
selenoprotein P	Z11793	NM_005410	SEQ ID NO: 73	SEQ ID NO: 74
KIAA0466, immunoglobulin superfamily, member 3	AB007935	NM_001542	SEQ ID NO: 75	SEQ ID NO: 76
type II membrane protein similar to HIV gp120-binding C-type lectin, CD209 antigen-like	AB015629	NM_014257	SEQ ID NO: 77	SEQ ID NO: 78
KIAA0626	AB014526	NM_021647	SEQ ID NO: 79	SEQ ID NO: 80
KIAA0711	AB018254	NM_014867	SEQ ID NO: 81	SEQ ID NO: 82
integrin alpha 9	D25303	NM_002207	SEQ ID NO: 83	SEQ ID NO: 84
integrin alpha 9	*D25303*	*NM_002207*	duplicate	
relaxin H2	X00948	NM_005059	SEQ ID NO: 85	SEQ ID NO: 86
KIAA0644	AB014544	NM_014817	SEQ ID NO: 87	SEQ ID NO: 88
Cdk-inhibitor p57KIP2 (KIP2)	U22398	NM_000076	SEQ ID NO: 89	SEQ ID NO: 90
Cdk-inhibitor p57KIP2 (KIP2)	*U22398*	*NM_000076*	duplicate	
transient receptor potential channel TRPC6	AJ006276	NM_004621	SEQ ID NO: 91	SEQ ID NO: 92
cDNA DKFZp564O222 (from clone DKFZp564O222)	AL050002			SEQ ID NO: 93
	M80482	NM_002570	SEQ ID NO: 94	SEQ ID NO: 95
regulator of G-protein signalling 16, A28-RGS14p	U70426	NM_002928	SEQ ID NO: 96	SEQ ID NO: 97

dihydropyrimidinase related protein-1, collapsin response mediator protein 1	D78012	NM_001313	SEQ ID NO: 98	SEQ ID NO: 99
desmoplakin (DPI, DPII)	AL031058	NM_004415	SEQ ID NO: 100	SEQ ID NO: 101
pendrin, solute carrier family, member 4	AF030880	NM_000441	SEQ ID NO: 102	SEQ ID NO: 103
reelin (RELN)	U79716	NM_005045	SEQ ID NO: 104	SEQ ID NO: 105
integrin, alpha 1	X68742			SEQ ID NO: 106
integrin alpha 1	*X68742*		duplicate	
cholesterol 25-hydroxylase	AF059214	NM_003956	SEQ ID NO: 107	SEQ ID NO: 108
inhibin beta-B-subunit precursor	M31682	NM_002193	SEQ ID NO: 109	SEQ ID NO: 110
KIAA1233	AL109724			
pre-B cell stimulating factor homologue (SDF1b)	L36033	NM_000609	SEQ ID NO: 112	SEQ ID NO: 113
V-Erba Related Ear-3 Protein	HG3510-HT3704			SEQ ID NO: 114
antigen identified by monoclonal antibody MRC OX-2	X05323		SEQ ID NO: 115	SEQ ID NO: 116
apolipoprotein D	J02611	NM_001647	SEQ ID NO: 117	SEQ ID NO: 118
TIMP3, tissue inhibitor of matrix metalloproteinases	U14394	NM_000362	SEQ ID NO: 119	SEQ ID NO: 120
TIMP3	*U14394*	*NM_000362*	duplicate	
aldehyde dehydrogenase 1	K03000	NM_000689	SEQ ID NO: 121	SEQ ID NO: 122
prospero-related homeobox 1 (prox 1)	U44060	NM_002763	SEQ ID NO: 123	SEQ ID NO: 124
matrix Gla protein	AI953789	NM_000900	SEQ ID NO: 125	SEQ ID NO: 126
neuronal pentraxin II (NPTX2)	U29195		SEQ ID NO: 127	SEQ ID NO: 128
histatin 2 (HIS2)	M26665	NM_000200	SEQ ID NO: 129	SEQ ID NO: 130
ADDL mRNA for adducin-like protein, adducin 3 (gamma)	D67031	NM_016824	SEQ ID NO: 131	SEQ ID NO: 132
adducin 3 (gamma)	*U37122*	*NM_016824*	duplicate	
MADS box transcription enhancer factor 2, polypeptide C (myocyte enhancer factor 2C)	L08895	NM_002397	SEQ ID NO: 133	SEQ ID NO: 134
MADS box transcription enhancer factor 2, (myocyte enhancer factor 2C)		*NM_002397*	duplicate	
MADS box transcription enhancer factor 2, polypeptide C (myocyte enhancer factor 2C)	*S57212*	*NM_002397*	duplicate	
phosphoglucomutase 5	L40933	NM_021965	SEQ ID NO: 135	SEQ ID NO: 136
cyclin E2	AF102778	NM_004702	SEQ ID NO: 137	SEQ ID NO: 138
interleukin 7 (IL7)	M29053		SEQ ID NO: 139	SEQ ID NO: 140
interleukin 7	*J04156*	*NM_000880*	duplicate	

cDNA DKFZp586L0120 (from clone DKFZp586L0120)	AL050154			SEQ ID NO: 141
peroxisome proliferative activated receptor, gamma, PPARG	L40904	NM_005037	SEQ ID NO: 142	SEQ ID NO: 143
fatty acid binding protein 4	AA128249	NM_001442	SEQ ID NO: 144	SEQ ID NO: 145
protein kinase C zeta	Z15108	NM_002744	SEQ ID NO: 146	SEQ ID NO: 147
46 kDa coxsackievirus and adenovirus receptor (CAR) protein	Y07593	NM_001338	SEQ ID NO: 148	SEQ ID NO: 149
PAC clone RP4-751H13 from 7q35-qter, zinc finger-like	AC004877		SEQ ID NO: 150	SEQ ID NO: 151
thymidine kinase 1, soluble	M15205	NM_003258	SEQ ID NO: 152	SEQ ID NO: 153
thymidine kinase 1	*K02581*	*NM_003258*	duplicate	
Pig7 (PIG7), LPS-induced TNF-alpha factor	AF010312	NM_004862	SEQ ID NO: 154	SEQ ID NO: 155
LPS-induced TNF-alpha factor	*AL120815*	*NM_004862*	duplicate	
lipase A, lysosomal acid, cholesterol esterase	X76488	NM_000235	SEQ ID NO: 156	SEQ ID NO: 157
ubiquitin specific protease 13 (isopeptidase T-3)	U75362	NM_003940	SEQ ID NO: 158	SEQ ID NO: 159
carcinoembryonic antigen-related cell adhesion molecule 1 (biliary glycoprotein) CEACAM1	X16354	NM_001712	SEQ ID NO: 160	SEQ ID NO: 161
cDNA DKFZp586D0918 (from clone DKFZp586D0918)	AL049370			SEQ ID NO: 162
KIAA0598, B cell RAG associated protein	AB011170	NM_014863	SEQ ID NO: 163	SEQ ID NO: 164
RAMP2 (receptor (calcitonin) activity modifying protein 2)	AJ001015	NM_005854	SEQ ID NO: 165	SEQ ID NO: 166
cholesteryl ester transfer protein precursor	M30185	NM_000078	SEQ ID NO: 167	SEQ ID NO: 168
epithelial membrane protein 2	U52100	NM_001424	SEQ ID NO: 169	SEQ ID NO: 170
MHC class II lymphocyte antigen (HLA-DP) beta chain	M83664	NM_002121	SEQ ID NO: 171	SEQ ID NO: 172
MHC class II lymphocyte antigen (HLA-DP) beta chain	*M83664*	*NM_002121*	duplicate	
beta-arrestin 2	AF106941	NM_004313	SEQ ID NO: 173	SEQ ID NO: 174
mitotic checkpoint kinase Bub1 (BUB1)	AF053305	NM_004336	SEQ ID NO: 175	SEQ ID NO: 176
KIAA0229, similar to human ankyrin 1(S08275)	D86982		SEQ ID NO: 177	SEQ ID NO: 178
Sprouty 1 homolog (antagonist of FGF signaling)	AF041037		SEQ ID NO: 179	SEQ ID NO: 180
guanine nucleotide exchange factor for Rap1; M-Ras-regulated GEF, KIAA0277		NM_012294	SEQ ID NO: 181	SEQ ID NO: 182
translin	X78627	NM_004622	SEQ ID NO: 183	SEQ ID NO: 184
erythrocyte membrane protein band 4.9 (dematin)	U28389	NM_001978	SEQ ID NO: 185	SEQ ID NO: 186

KIAA0846 protein	AB020653	NM_015376	SEQ ID NO: 187	SEQ ID NO: 188
glia maturation factor, gamma	W07033	NM_004877	SEQ ID NO: 189	SEQ ID NO: 190
insulin-like growth factor binding protein 2 (IGFBP-2)	X16302	NM_000597	SEQ ID NO: 191	SEQ ID NO: 192
smooth muscle myosin heavy chain isoform Smemb	S67247		SEQ ID NO: 193	SEQ ID NO: 194
TTG-2 (cysteine rich protein with LIM motif), LIM domain only 2 (rhombotin-like 1)	X61118	NM_005574	SEQ ID NO: 195	SEQ ID NO: 196
cyclin B2	AL080146	NM_004701	SEQ ID NO: 197	SEQ ID NO: 198
KIAA0353	AB002351		SEQ ID NO: 199	SEQ ID NO: 200
KIAA0559, piccolo (presynaptic cytomatrix protein)	AB011131		SEQ ID NO: 201	SEQ ID NO: 202
G protein-coupled receptor, family C, group 5, member B	AC004131	NM_016235	SEQ ID NO: 203	SEQ ID NO: 204
G protein-coupled receptor, family C, group 5, member B	*AI801872*	*NM_016235*	duplicate	dup
CREM (cyclic AMP-responsive element modulator beta isoform)	S68134	NM_001881	SEQ ID NO: 205	SEQ ID NO: 206
CREM (cyclic AMP-responsive element modulator beta isoform)	*S68134*	*NM_001881*	duplicate	dup
CREM (cyclic AMP-responsive element modulator beta isoform)	*S68271*	*NM_001881*	duplicate	dup
hypothetical protein FLJ13110	AL080222	NM_022912	SEQ ID NO: 207	SEQ ID NO: 208
inositol(myo)-1(or 4)-monophosphatase 2	AF014398	NM_014214	SEQ ID NO: 209	SEQ ID NO: 210
KIAA0937 protein	AB023154		SEQ ID NO: 211	SEQ ID NO: 212
mitotic spindle coiled-coil related protein	AF063308	NM_006461	SEQ ID NO: 213	SEQ ID NO: 214
cysteine and glycine-rich protein 2 (CSRP2)	U57646	NM_001321	SEQ ID NO: 215	SEQ ID NO: 216
topoisomerase (DNA) II alpha (170kD)	AI375913	NM_001067	SEQ ID NO: 217	SEQ ID NO: 218
DNA topoisomerase II	*J04088*	*NM_001067*	duplicate	dup
protein phosphatase inhibitor 2 (PPP1R2)	U68111		SEQ ID NO: 219	SEQ ID NO: 220
KIAA0186	D80008	NM_021067	SEQ ID NO: 221	SEQ ID NO: 222
dual-specificity tyrosine-(Y)-phosphorylation regulated kinase 3 (Dyrk3)	Y12735	NM_003582	SEQ ID NO: 223	SEQ ID NO: 224
kinesin-like spindle protein HKSP (HKSP)	U37426	NM_004523	SEQ ID NO: 225	SEQ ID NO: 226
huntingtin-associated protein interacting protein (duo)	U94190	NM_003947	SEQ ID NO: 227	SEQ ID NO: 228
diubiquitin	AL031983	NM_006398	SEQ ID NO: 229	SEQ ID NO: 230
bikunin, serine protease inhibitor, Kunitz type, 2	U78095	NM_021102	SEQ ID NO: 231	SEQ ID NO: 232

cytochrome P-450-1 (TCDD-inducible)	K03191	NM_000499	SEQ ID NO: 233	SEQ ID NO: 234
cytochrome P(1)-450	*X02612*	*NM_000499*	duplicate	dup
KIAA0513		NM_014732	SEQ ID NO: 235	SEQ ID NO: 236
protein phosphatase inhibitor 2 (PPP1R2)	U68111		duplicate	
RAMP3 (receptor (calcitonin) activity modifying protein 3)	AJ001016	NM_005856	SEQ ID NO: 237	SEQ ID NO: 238
B-myb	X13293	NM_002466	SEQ ID NO: 239	SEQ ID NO: 240
KIAA0952	AB023169	NM_014962	SEQ ID NO: 241	SEQ ID NO: 242
interferon stimulated gene (20kD), HEM45	U88964	NM_002201	SEQ ID NO: 243	SEQ ID NO: 244
GS3955	D87119	NM_021643	SEQ ID NO: 245	SEQ ID NO: 246
GS3955	*D87119*	*NM_021643*	duplicate	dup
GRB2-related adaptor protein (Grap)	U52518	NM_006613	SEQ ID NO: 247	SEQ ID NO: 248
KIAA1071 protein	AB028994		SEQ ID NO: 249	SEQ ID NO: 250
RNA-binding protein gene with multiple splicing, RBP-MS/type 5	D84111	NM_006867	SEQ ID NO: 251	SEQ ID NO: 252
RNA-binding protein gene with multiple splicing, RBP-MS/type 5	*D84111*	*NM_006867*	duplicate	dup
RBP-MS/type 4, RNA-binding protein gene with multiple splicing	*D84110*	*NM_006867*	duplicate	
RBP-MS/type 4, RNA-binding protein gene with multiple splicing	*D84110*	*NM_006867*	duplicate	dup
RBP-MS/type 3, RNA-binding protein gene with multiple splicing	*D84109*	*NM_006867*	duplicate	dup
alpha-actinin-2-associated LIM protein	AF002282	NM_014476	SEQ ID NO: 253	SEQ ID NO: 254
semaphorin-III (Hsema-I), semaphorin 3A	L26081	NM_006080	SEQ ID NO: 255	SEQ ID NO: 256
IQ motif containing GTPase activating protein 2	U51903	NM_006633	SEQ ID NO: 257	SEQ ID NO: 258
Arrestin, Beta 2	HG2059-HT2114		duplicate	
retinoblastoma-associated protein HEC	AF017790	NM_006101	SEQ ID NO: 259	SEQ ID NO: 260
LIM domain binding protein (LDB1)	AF052389	NM_001290	SEQ ID NO: 261	SEQ ID NO: 262
dual specificity phosphatase 5	U15932	NM_004419	SEQ ID NO: 263	SEQ ID NO: 264
Homo sapiens cDNA 3', mRNA sequence	AI557322			SEQ ID NO: 265
monoamine oxidase A (MAOA)	M68840	NM_000240	SEQ ID NO: 266	SEQ ID NO: 267
monoamine oxidase A	*AA420624*	*NM_000240*	duplicate	

NECDIN related protein	U35139	NM_002487	SEQ ID NO: 268	SEQ ID NO: 269
regulatory solute carrier protein, family 1, member 1	X82877	NM_006511	SEQ ID NO: 270	SEQ ID NO: 271
TTK protein kinase	M86699	NM_003318	SEQ ID NO: 272	SEQ ID NO: 273
fms-related tyrosine kinase 4, VEGFR-3	X69878	NM_002020	SEQ ID NO: 274	SEQ ID NO: 275
TSC403, similar to lysosome-associated membrane glycoprotein	AB013924	NM_014398	SEQ ID NO: 276	SEQ ID NO: 277
HMG-2	X62534		SEQ ID NO: 278	SEQ ID NO: 279
Homo sapiens clone 24416 mRNA sequence	AF052159		SEQ ID NO: 280	SEQ ID NO: 281
calcitonin receptor-like	L76380	NM_005795	SEQ ID NO: 282	SEQ ID NO: 283
KIAA0582 protein	AI761647	NM_015147	SEQ ID NO: 284	SEQ ID NO: 285
cDNA DKFZp434B102 (from clone DKFZp434B102)	AL080192			SEQ ID NO: 286
cDNA DKFZp586G1922 (from clone DKFZp586G1922)	AL080110		SEQ ID NO: 287	SEQ ID NO: 287
Acyl-CoA synthetase 3	D89053	NM_004457	SEQ ID NO: 288	SEQ ID NO: 289
fatty-acid-Coenzyme A ligase, long-chain 3	*AA977580*	*NM_004457*	duplicate	
STAT induced STAT inhibitor-2	AF037989		SEQ ID NO: 290	SEQ ID NO: 291
Homeotic Protein Hox5.4	HG3502-HT3696			SEQ ID NO: 292
hypothetical protein FLJ13910, cDNA DKFZp586M141 (from clone DKFZp586M141)	AL050139	NM_022780	SEQ ID NO: 293	SEQ ID NO: 294
cDNA DKFZp586N012 (from clone DKFZp586N012)	AL049471			SEQ ID NO: 295
UbcH10, ubiquitin carrier protein E2-C	U73379	NM_007019	SEQ ID NO: 296	SEQ ID NO: 297
cyclin-dependent kinase inhibitor 3, protein tyrosine phosphatase (CIP2)	L25876	NM_005192	SEQ ID NO: 298	SEQ ID NO: 299
glycogen phosphorylase (PYGL)	AF046798		SEQ ID NO: 300	SEQ ID NO: 301
Angiopoietin-2	AF004327	NM_001147	SEQ ID NO: 302	SEQ ID NO: 303
Angiopoietin-2	*AF004327*	*NM_001147*	duplicate	dup
forkhead box M1	U74612	NM_021953	SEQ ID NO: 304	SEQ ID NO: 305
potentially prenylated protein tyrosine phosphatase hPRL-3	AF041434	NM_007079	SEQ ID NO: 306	SEQ ID NO: 307
RAB31, Low Mr GTP-binding protein of the Rab subfamily	U59877	NM_006868	SEQ ID NO: 308	SEQ ID NO: 309
RAB31, member RAS oncogene family	*AI189226*	*NM_006868*		
myosin VIIA	U39226	NM_000260	SEQ ID NO: 310	SEQ ID NO: 311
Grb2-associated binder-1, docking protein related to IRS-1	U43885	NM_002039	SEQ ID NO: 312	SEQ ID NO: 313

lamin B1	L37747		SEQ ID NO: 314	SEQ ID NO: 315
minichromosome maintenance deficient (mis5, S. pombe) 6 HsMcm6	D84557	NM_005915	SEQ ID NO: 316	SEQ ID NO: 317
cyclin B1	M25753			SEQ ID NO: 318
cyclin B1	*M25753*		duplicate	dup
RTP, N-myc downstream regulated	D87953	NM_006096	SEQ ID NO: 319	SEQ ID NO: 320
alpha2,3-sialyltransferase	AB022918	NM_006100	SEQ ID NO: 321	SEQ ID NO: 322
ADP-ribosylation factor-like protein 4	U73960	NM_005738	SEQ ID NO: 323	SEQ ID NO: 324
centromere protein F (350/400kD, mitosin)	U30872	NM_016343	SEQ ID NO: 325	SEQ ID NO: 326
paternally expressed 10, KIAA1051	AB028974	NM_015068	SEQ ID NO: 327	SEQ ID NO: 328
tubulin, alpha 1 (testis specific)	X06956		SEQ ID NO: 329	SEQ ID NO: 330
KIAA0101	D14657	NM_014736	SEQ ID NO: 331	SEQ ID NO: 332
KIAA0128, septin 2	D50918		SEQ ID NO: 333	SEQ ID NO: 334
protein phosphatase 2, regulatory subunit B (B56), gamma	Z69030	NM_002719	SEQ ID NO: 335	SEQ ID NO: 336
deoxycytidine kinase	M60527	NM_000788	SEQ ID NO: 337	SEQ ID NO: 338
integrin beta 3 binding protein (beta3-endonexin)	U37139	NM_014288	SEQ ID NO: 339	SEQ ID NO: 340
TAL1 (SCL) interrupting locus	M74558	NM_003035	SEQ ID NO: 341	SEQ ID NO: 342
KIAA0666	AB014566		SEQ ID NO: 343	SEQ ID NO: 344
cAMP-specific phosphodiesterase 8A, PDE8A1	AF056490		SEQ ID NO: 345	SEQ ID NO: 346
mitotic checkpoint kinase Mad3L (MAD3L), BUB1B	AF053306	NM_001211	SEQ ID NO: 347	SEQ ID NO: 348
ribosomal S6 kinase	X85106	NM_021135	SEQ ID NO: 349	SEQ ID NO: 350
HPTP epsilon (protein tyrosine phosphatase epsilon)	X54134	NM_006504	SEQ ID NO: 351	SEQ ID NO: 352
Lyn tyrosine kinase, v-yes-1 Yamaguchi sarcoma viral related oncogene homolog	M79321	NM_002350	SEQ ID NO: 353	SEQ ID NO: 354
lyn tyrosine kinase, v-yes-1 Yamaguchi sarcoma viral related oncogene homolog	*M16038*	*NM_002350*	duplicate	
lyn tyrosine kinase	*M16038*	*NM_002350*	duplicate	
brachyury variant A (TBX1), T-box 1 transcription factor	AF012130	NM_005992	SEQ ID NO: 355	SEQ ID NO: 356
mki67a mRNA (long type) for antigen of monoclonal antibody Ki-67	X65550	NM_002417	SEQ ID NO: 357	SEQ ID NO: 358
protein tyrosine phosphatase receptor pi (PTPRP)	U81561	NM_002847	SEQ ID NO: 359	SEQ ID NO: 360
cbl-b	U26710	NM_004351	SEQ ID NO: 361	SEQ ID NO: 362
Cyclin A2	X51688	NM_001237	SEQ ID NO: 363	SEQ ID NO: 364

nucleoside phosphorylase	X00737	NM_000270	SEQ ID NO: 365	SEQ ID NO: 366
TNF-related apoptosis inducing ligand TRAIL	U37518	NM_003810	SEQ ID NO: 367	SEQ ID NO: 368
phosphodiesterase 4B, cAMP-specific	L20971	NM_002600	SEQ ID NO: 369	SEQ ID NO: 370
nidogen (enactin)	M30269	NM_002508	SEQ ID NO: 371	SEQ ID NO: 372
HYA22 protein	D88153	NM_005808	SEQ ID NO: 373	SEQ ID NO: 374
phosphatidic acid phosphatase type 2A	AF014402	NM_003711	SEQ ID NO: 375	SEQ ID NO: 376
KIAA0512, ALEX2	AB011084	NM_014782	SEQ ID NO: 377	SEQ ID NO: 378
thromboxane A2 receptor	D38081	NM_001060	SEQ ID NO: 379	SEQ ID NO: 380
trefoil factor 3 (intestinal)	AI985964	NM_003226	SEQ ID NO: 381	SEQ ID NO: 382
G-2 and S-phase expressed 1	AL031588	NM_016426	SEQ ID NO: 383	SEQ ID NO: 384
ADP-ribosyltransferase (NAD+; poly (ADP-ribose) polymerase)-like 2	AJ236876	NM_005484	SEQ ID NO: 385	SEQ ID NO: 386
serine/threonine kinase 12	AF015254	NM_004217	SEQ ID NO: 387	SEQ ID NO: 388
Tubulin, Alpha 1, Isoform 44	HG2259-HT2348		duplicate	
lamin B receptor	L25931	NM_002296	SEQ ID NO: 389	SEQ ID NO: 390
KIAA0429	AB007889	NM_014751	SEQ ID NO: 391	SEQ ID NO: 392
transcription factor 4	M74719	NM_003199	SEQ ID NO: 393	SEQ ID NO: 394
syndecan 3 (N-syndecan), KIAA0468	AB007937	NM_014654	SEQ ID NO: 395	SEQ ID NO: 396
RECK protein precursor	AA099265	NM_021111	SEQ ID NO: 397	SEQ ID NO: 398
Putative prostate cancer tumor suppressor	U42349	NM_006765	SEQ ID NO: 399	SEQ ID NO: 400
protein phosphatase 1, regulatory (inhibitor) subunit	AB020630		SEQ ID NO: 401	SEQ ID NO: 402
PDZ and LIM domain 1 (elfin)	U90878	NM_020992	SEQ ID NO: 403	SEQ ID NO: 404
hypothetical protein from clone 643	AF091087	NM_020467	SEQ ID NO: 405	SEQ ID NO: 406
p53-regulated DDA3	AA926959			SEQ ID NO: 407
KIAA0062	D31887		SEQ ID NO: 408	SEQ ID NO: 409
medium-chain acyl-CoA dehydrogenase	M91432		SEQ ID NO: 410	SEQ ID NO: 411
gap junction protein, alpha 1, 43kD (connexin 43)	M65188	NM_000165	SEQ ID NO: 412	SEQ ID NO: 413
MyoD family inhibitor	U78313	NM_005586	SEQ ID NO: 414	SEQ ID NO: 415

endo/exonuclease Mre11 (MRE11A)	AF073362	NM_005591	SEQ ID NO: 416	SEQ ID NO: 417
nuclear receptor subfamily 2, group F, member 1	X16155	NM_005654	SEQ ID NO: 418	SEQ ID NO: 419

Table 15

Sequence identifiers for sequences in Table 4

	accession numbers		aa SEQ ID NO:	nt SEQ ID NO:
p27 mRNA, interferon alpha-inducible protein 27	X67325	NM_005532	420	421
ribonuclease A (RNase A), pancreatic	D26129	NM_002933	422	423
hematopoietic neural membrane protein (HNMP-1)	U87947	NM_001425	424	425
N-cadherin	M34064	NM_001792	426	427
N-cadherin	*M34064*	*NM_001792*	duplicate	
interleukin 8 (IL8)	M28130	NM_000584	428	429
interleukin 8, beta-thromboglobulin-like protein precursor	*M17017*	*NM_000584*	430	431
tyrosine kinase receptor (axl)	M76125	NM_001699	432	433
	HG162-HT3165		duplicate	
cell surface glycoprotein CD44 (CD44)	L05424		434	435
cell adhesion molecule (CD44)	*M59040*	*NM_000610*	duplicate	
hyaluronate receptor (CD44)	*L05424*		duplicate	
vascular endothelial growth factor related protein VRP, VEGF-C	U43142	NM_005429	436	437
Vascular endothelial growth factor C	*X94216*	*NM_005429*	duplicate	
collagen type XIII, alpha 1 (=COL4A2)	M33653	NM_005203	438	439
collagen type XIII, alpha-1	*M59217*	*NM_005203*	duplicate	
collagen alpha-2 type I	K01079			440
collagen alpha-2 type I	*K01079*		duplicate	
collagen, type I, alpha 2	*V00503*	*NM_000089*	duplicate	
proteoglycan 1	X17042	NM_002727	441	442

phospholipase A2, group IVA, calcium-dependent phospholipid-binding protein (PLA2)	M72393		443	444
carbohydrate (keratan sulfate Gal-6) sulfotransferase	AB003791	NM_003654	445	446
tropomyosin 2 (beta), fibroblast tropomyosin	M12125	NM_003289	447	448
chondroitin sulfate proteoglycan 2 (versican)	X15998	NM_004385	449	450
chondroitin sulfate proteoglycan 2 (versican)	*X15998*	*NM_004385*	duplicate	
latent transforming growth factor-beta binding protein (LTBP-2)	Z37976	NM_000428	451	452
interleukin 6 (interferon, beta 2)	X04430	NM_000600	453	454
bone morphogenetic protein-4 (hBMP-4)	U43842	NM_001202	455	456
bone morphogenetic protein 2B, BMP-4	*M22490*	*NM_001202*	duplicate	
sarcolectin, keratin 7	AJ238246	NM_005556	457	458
neuronal cell adhesion molecule, KIAA0343	AB002341	NM_005010	459	460
neuronal cell adhesion molecule, hBRAVO/Nr-CAM precursor	*U55258*	*NM_005010*	duplicate	
matrix metalloproteinase 1 (interstitial collagenase), skin collagenase	M13509	NM_002421	461	462
stem cell factor, KIT ligand	M59964	NM_000899	463	464
uPA	X02419	NM_002658	465	466
plasminogen activator inhibitor-1	J03764	NM_000602	467	468
plasminogen activator inhibitor 1	*M14083*	*NM_000602*	duplicate	
selectin P, CD62, granule membrane protein-140 (GMP-140) precursor	M25322	NM_003005	469	470
latrophilin-2	AJ131581	NM_012302	471	472
actin, alpha 2	X13839	NM_001613	473	474
fibroblast activation protein, alpha	U09278	NM_004460	475	476
regulator of G-protein signalling 20	AF060877	NM_003702	477	478
IGF-II mRNA-binding protein 3	U97188	NM_006547	479	480
retina cDNA randomly primed sublibrary, EST	W28438			481
brain acid-soluble protein 1, neuronal tissue-enriched acidic	AF039656	NM_006317	482	483

protein (NAP-22)				
profilin 2	AL096719	NM_002628	484	485
profilin 2	*L10678*	*NM_002628*	duplicate	
Na,K-ATPase beta-1 subunit	U16799	NM_001677	486	487
Claudin-7	AJ011497	NM_001307	488	489
normal gingiva	U51712			490
a disintegrin and metalloproteinase domain 23	AB009672	NM_003812	491	492
COL8A1 mRNA for alpha 1(VIII) collagen	X57527	NM_001850	493	494
signal transducer and activator of transcription 6 (STAT6)	AF067575		495	496
transcription factor IL-4 Stat, STAT6	*U16031*	*NM_003153*	duplicate	
lipocortin-III, annexin A3	M20560	NM_005139	497	498
intercellular adhesion molecule 1 (CD54), major group rhinovirus receptor precusor	M24283	NM_000201	499	500
solute carrier family 1 (neuronal/epithelial high affinity glutamate transporter, system Xag)	U08989	NM_004170	501	502
solute carrier family 1 (neuronal/epithelial high affinity glutamate transporter, system Xag)	*AI928365*	*NM_004170*	duplicate	
p53 inducible protein	L47738		503	504
dihydropyrimidine dehydrogenase, DPYD	U20938	NM_000110	505	506
natural killer cell transcript 4	AA631972	NM_004221	507	508
PFTAIRE protein kinase 1, KIAA0834	AB020641	NM_012395	509	510
RGP4, regulator of G-protein signalling 4	U27768	NM_005613	511	512
regulator of G-protein signalling 4	*AI267373*	*NM_005613*	duplicate	
Oncogene Aml1-Evi-1, Fusion Activated	HG4058-HT4328		513	514
Oncogene Aml1-Evi-1, Fusion Activated	*HG4058-HT4328*		duplicate	
adenylyl cyclase-associated protein 2	N90755	NM_006366	515	516
clusterin (complement lysis inhibitor, SP-40,40, sulfated glycoprotein 2, apolipoprotein J)	M25915	NM_001831	517	518
ADP ribosylation factor-like 7	AB016811	NM_005737	519	520

H factor (complement)-like 1	M65292	NM_002113	521	522
RNA helicase-related protein, metallothionein-If	H68340	NM_007372	523	524
stimulated trans-acting factor (50 kDa) Staf50	X82200	NM_006074	525	526
cyclooxygenase-2 (hCox-2)	U04636	NM_000963	527	528
GRO1 oncogene, melanoma growth stimulatory activity (MGSA)	X54489	NM_001511	529	530
NRGN, neurogranin	X99076	NM_006176	531	532
homologue of mouse dkk-1	AB020315		533	534
gastrointestinal tumor-associated antigen GA733-1, tumor-associated calcium signal transducer 2	J04152	NM_002353	535	536
laminin	Z15008	NM_005562	537	538
transgelin, 22kDa smooth muscle protein (SM22)	M95787	NM_003186	539	540
JE gene encoding a monocyte secretory protein	M28225		541	542
zinc finger protein 238, RP58	AJ223321	NM_006352	543	544
cathepsin C	X87212	NM_001814	545	546
tissue-type plasminogen activator (t-PA)	M15518	NM_000930	547	548
sushi-repeat protein	AF060567	NM_014467	549	550
annexin A6	D00510	NM_001155	551	552
EphrinB1	U09303	NM_004429	553	554
EphrinB1	*U09303*	*NM_004429*	duplicate	
TFEC isoform (transcription factor EC)	D43945	NM_012252	555	556
small inducible cytokine A2, (monocyte chemotactic protein1)	M26683	NM_002982	557	558
small inducible cytokine A2 (monocyte chemotactic protein 1)	*M26683*	*NM_002982*	duplicate	
endothelial cell protein C/APC receptor (EPCR)	L35545	NM_006404	559	560
transglutaminase 2 (TGase)	M55153	NM_004613	561	562
transglutaminase (TGase)	*M55153*	*NM_004613*	duplicate	
human metallothionein-If	M10943		563	564
transforming growth factor beta-induced (BIGH3)	M77349	NM_000358	565	566

ENO2 gene for neuron specific (gamma) enolase	X51956		567	568
FAT tumor suppressor (Drosophila) homolog	X87241	NM_005245	569	570
malignant cell expression-enhanced gene/tumor progression-enhanced gene	S82470	NM_024298	571	572
malignant cell expression-enhanced gene/tumor progression-enhanced gene	*S82470*	*NM_024298*	duplicate	574
cDNA DKFZp566G0746 (from clone DKFZp566G0746)	AL050078			575
lysyl oxidase-like 2	U89942	NM_002318	576	577
ras-related C3 botulinum toxin substrate 2 (rho family, small GTP binding protein Rac2)	M64595	NM_002872	578	579
endothelial leukocyte adhesion molecule 1 (ELAM-1), selectin E	M24736	NM_000450	580	581
laminin, alpha 5, KIAA0533	AB011105		582	583
placenta growth factor (PlGF)	X54936	NM_002632	584	585
ALL1-fused gene from chromosome 1q, AF1q	U16954	NM_006818	586	587
stromelysin-2, MMP-10	X07820	NM_002425	588	589
metallothionein-I-A	K01383		590	591
collagen VI alpha-1	X15880		592	593
mad protein homolog (hMAD-3)	U68019	NM_005902	594	595
mad protein homolog (hMAD-3)	*U68019*	*NM_005902*	duplicate	
mad protein homolog (hMAD-3)	*U68019*	*NM_005902*	duplicate	
integral membrane protein 2A	AL021786			596
interleukin 1 receptor-like 1	D12763	NM_003856	597	598
high-mobility group (nonhistone chromosomal) protein isoform I-C (HMGI-C)	X92518		599	600
epidermal growth factor receptor kinase substrate (Eps8)	U12535	NM_004447	601	602
lactate dehydrogenase B	X13794	NM_002300	603	604
mRNA for unknown product	D29810		605	606
hypothetical protein DKFZp564D0462	AL033377			607
lysyl hydroxylase isoform 2 (PLOD2)	U84573	NM_000935	608	609
follistatin-like 3, follistatin-related protein (FLRG)	U76702	NM_005860	610	611

Homo sapiens clone 24674 mRNA sequence	AF070578			612
L-iditol-2 dehydrogenase	L29254		613	614
neuronal pentraxin 1	U61849	NM_002522	615	616
hypothetical protein from clones 23549 and 23762	U90908	NM_021226	617	618
UDP-N-acetylglucosamine pyrophosphorylase	AB011004	NM_003115	619	620
zinc finger protein 185 (LIM domain)	Y09538	NM_007150	621	622
four and a half LIM domains 2, heart protein (FHL-2)	U29332	NM_001450	623	624
mitogen-activated protein kinase-activated protein kinase 3, MAPKAP kinase (3pK)	U09578	NM_004635	625	626
metallothionein 1E (functional)	R92331			627
TU3A protein	AF035283	NM_007177	628	629
metallothionein 1H	R93527	NM_005951	630	631
guanylate binding protein isoform II (GBP-2)	M55543	NM_004120	632	633
soluble vascular endothelial cell growth factor receptor 1 (sVEGFR-1)	U01134	NM_002019	634	635
R-Ras	M14949		636	637
R-ras	*M14949*		638	639
creatine transporter (SLC6A8), solute carrier family 6, member 8	U36341	NM_005629	640	641
target of myb1 (chicken) homolog, Heme Oxygenase 1 (HO-1)	Z82244	NM_005488	642	643
procollagen-lysine, 2-oxoglutarate 5-dioxygenase, lysyl hydroxylase (PLOD)	L06419	NM_000302	644	645
KIAA0836	AB020643		646	647
cDNA DKFZp434C171 (from clone DKFZp434C171)	AL080169		648	649
IL-4-R mRNA for the interleukin 4 receptor	X52425	NM_000418	650	651
chemokine (C-C motif) receptor-like 2 (CCRL2), chemokine receptor X (CKRX)	AF014958	NM_003965	652	653
phospholipase C, beta 3 (phosphatidylinositol-specific)	Z16411	NM_000932	654	655
LIM domain protein	X93510	NM_003687	656	657
protein kinase (cAMP-dependent, catalytic) inhibitor beta	M34181	NM_002731	658	659
rho GDP-dissociation Inhibitor 2	X69549	NM_001175	660	661

KIAA0975, imidazoline receptor candidate	AB023192	NM_007184	662	663
poliovirus receptor	X64116	NM_006505	664	665
poliovirus receptor	*X64116*	*NM_006505*	duplicate	
immediate early response 3	S81914	NM_003897	666	667
metallothionein 2A	AI547258	NM_005953	668	669
tropomyosin 1 (alpha)	M19267	NM_000366	670	671
tropomyosin 1 (alpha)	*Z24727*	*NM_000366*	duplicate	
tropomyosin 1 (alpha)	*M19267*	*NM_000366*	duplicate	
TRAM-like protein	D31762	NM_012288	672	673
E3 ubiquitin ligase SMURF2	AA630312	NM_022739	674	675
EGF-containing fibulin-like extracellular matrix protein 1	U03877	NM_004105	676	677
G protein-coupled receptor 56	AJ011001	NM_005682	678	679
c-jun proto oncogene (JUN)	J04111	NM_002228	680	681
regulator of G-protein signalling 10, RGS10	AF045229	NM_002925	682	683
amyloid beta (A4) precursor protein-binding, family B, member 2 (Fe65-like)	U62325		684	685
ras-related rho protein	M12174	NM_004040	686	687
proteasome (prosome, macropain) 26S subunit, non-ATPase, 10	AL031177	NM_002814	688	689
KIAA0537	AB011109	NM_014840	690	691
lysosome-associated membrane protein-2	X77196	NM_002294	692	693
phospholipid transfer protein	L26232	NM_006227	694	695
N-myristoyltransferase 2	AF043325	NM_004808	696	697
phosphofructokinase (PFKM)	U24183	NM_000289	698	699
integrin, beta 4	X53587	NM_000213	700	701
leupaxin	AF062075	NM_004811	702	703
endothelin-converting-enzyme 1	Z35307	NM_001397	704	705

wild-type p53 activated fragment-1 (WAF1), cyclin-dependent kinase inhibitor 1A (p21, Cip1)	U03106	NM_000389	706	707
ICAM-2, cell adhesion ligand for LFA-1	X15606	NM_000873	708	709
ICAM-2, cell adhesion ligand for LFA-1	*X15606*	*NM_000873*	duplicate	
intercellular adhesion molecule 2 (ICAM-2)	*M32334*		710	711
eukaryotic translation initiation factor 2B, eIF-2B beta subunit	AF035280	NM_014239	712	713
uridine phosphorylase	X90858	NM_003364	714	715
integrin, beta 5	X53002	NM_002213	716	717
N-sulfoglucosamine sulfohydrolase (sulfamidase)	U30894	NM_000199	718	719
synaptojanin 2	AF039945		720	721
metallothionein 1L	AA224832	NM_002450	722	723
macrophage capping protein, gelsolin-like	M94345	NM_001747	724	725
HSPC022 protein	W68830	NM_014029	726	727
Human clone 137308 mRNA, partial cds	AW006742		no	728
protocadherin 42, PC42, protocadherin 1 (cadherin-like 1)	L11370	NM_002587	729	730
caspase-like apoptosis regulatory protein 2 (CLARP2)	AF005775	NM_003879	731	732
caspase-like apoptosis regulatory protein 2 (CLARP2)	*AF005775*	*NM_003879*	duplicate	
major vault protein, lrp	X79882	NM_005115	733	734
Fanconi anemia, complementation group G	AC004472	NM_004629	735	736
prion protein (PrP)	U29185	NM_000311	737	738
interferon-stimulated protein, 15 kDa	AA203213	NM_005101	739	740
serine (or cysteine) proteinase inhibitor, clade B (ovalbumin), cytoplasmic antiproteinase 2 (CAP2)	L40377	NM_002640	741	742
biglycan	J04599	NM_001711	743	744
chemokine (C-X-C motif), receptor 4 (fusin)	L06797	NM_003467	745	746
ubiquitin carboxyl-terminal esterase L1 (ubiquitin thiolesterase)	X04741	NM_004181	747	748
KIAA0469	AB007938	NM_014851	749	750

TNF (ligand) superfamily, member 4 (tax-transcriptionally activated glycoprotein 1, 34kD)	AL022310	NM_003326	751	752
KIAA1053	AB028976		753	754
NAD(P)H-quinone oxireductase	M81600		755	756
sushi-repeat-containing protein	U61374	NM_006307	757	758
integrin, alpha 5	X06256	NM_002205	759	760
enigma (LIM domain protein)	L35240	NM_005451	761	762
ectonucleoside triphosphate diphosphohydrolase 1	AJ133133	NM_001776	763	764
transforming growth factor-beta (tgf-beta), bone morphogenetic protein 6	M60315	NM_001718	765	766
transforming growth factor-beta (tgf-beta), bone morphogenetic protein 6	*M60315*	*NM_001718*	duplicate	
nicotinamide N-methyltransferase, NNMT	U08021	NM_006169	767	768
cDNA DKFZp564J0323 (from clone DKFZp564J0323)	AL049957		no	769
thioredoxin reductase beta	AB019694	NM_006440	770	771
f-box and leucine-rich repeat protein 2	AL049953		772	773
transcobalamin II (TCN2)	L02648	NM_000355	774	775
aldehyde dehydrogenase 2, mitochondrial	X05409	NM_000690	776	777
GTP-binding protein ragB	X90530	NM_006064	778	779
lymphocyte antigen 75	AF011333	NM_002349	780	781
GM2 activator protein	X62078		782	783
type 3 inositol 1,4,5-trisphosphate receptor (ITPR3)	U01062	NM_002224	784	785
KIAA0284	AI828210		no	786
metallothionein I-B	M13485		787	788
BTG2	U72649	NM_006763	789	790
adenylate kinase 1	J04809	NM_000476	791	792
tumor necrosis factor receptor superfamily, member 12, WSL-LR, WSL-S1 and WSL-S2 proteins	Y09392	NM_003790	793	794
aminopeptidase N/CD13	M22324	NM_001150	795	796
growth arrest and DNA-damage-inducible protein (gadd45)	M60974	NM_001924	797	798

KIAA0638 protein	AB014538		799	800
vinculin	M33308	NM_003373	801	802
procollagen-proline, 2-oxoglutarate 4-dioxygenase (proline 4-hydroxylase), alpha polypeptide II	U90441	NM_004199	803	804
msg1-related gene 1 (mrg1), Cbp/p300-interacting transactivator	U65093	NM_006079	805	806
microsomal glutathione S-transferase 3	AF026977	NM_004528	807	808
vitamin A responsive; cytoskeleton related	AF070523	NM_006407	809	810
17-kDa protein, interferon-stimulated protein, 15 kDa	M13755	NM_005101	811	812
matrix metalloproteinase 14 (membrane-inserted)	X83535	NM_004995	813	814
4F2 cell-surface antigen, solute carrier family 3, member 2	J02939	NM_002394	815	816
metallothionein-III	M93311	NM_005954	817	818
protein kinase (cAMP-dependent, catalytic) inhibitor alpha	S76965	NM_006823	819	820
protein kinase (cAMP-dependent, catalytic) inhibitor alpha	*S76965*	*NM_006823*	duplicate	
reticulocalbin 1, EF-hand calcium binding domain	D42073	NM_002901	821	822
lipin 1, KIAA0188	D80010		823	824
protease, serine, 23	AF015287	NM_007173	825	826
hect domain and RLD 2	AF041080	NM_004667	827	828
GATA-binding protein (GATA2)	M68891	NM_002050	829	830
agrin precursor	AF016903		831	832
equilibrative nucleoside transporter 1 (hENT1)	U81375	NM_004955	833	834
coronin, actin-binding protein 2B, KIAA0925	AB023142		835	836
f-box and WD-40 domain protein 3	U07000	NM_012165	837	838
nonsyndromic hearing impairment protein (DFNA5)	AF073308	NM_004403	839	840
actin filament associated protein	D25248	NM_021638	841	842
TNFR-related death receptor-6 (DR6)	AF068868	NM_014452	843	844
serum/glucocorticoid regulated kinase	Y10032	NM_005627	845	846
DNase X	X90392	NM_006730	847	848

DNase X	*X90392*	*NM_006730*	duplicate	
fatty acid desaturase 3	AC004770	NM_021727	849	850
LYL-1	M22637		851	852
ATP-binding cassette, sub-family C (CFTR/MRP), member 1	X78338	NM_004996	853	854
transmembrane protein (CD59)	M84349		855	856
fms-related tyrosine kinase 1, VEGFR-1	S77812		857	858
Hypothetical protein FLJ23403	AI681538	NM_022068	859	860
hypothetical protein FLJ20898	AI733570	NM_024600	861	862

CLAIMS

We claim:

1. A method for differentially modulating the growth or differentiation of blood endothelial cells (BEC) or lymphatic endothelial cells (LEC), comprising contacting endothelial cells with a composition comprising an agent that differentially modulates blood or lymphatic endothelial cells, said agent selected from the group consisting of:

(a) a polypeptide that comprises an amino acid sequence of a BEC polypeptide or a LEC polypeptide, or an active fragment of said polypeptide ;

(b) a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide according to (a);

(c) an antibody that specifically binds to a polypeptide according to (a);

(d) a polypeptide comprising a fragment of the antibody of (c), wherein the fragment and the antibody bind to the polypeptide;

(e) an antisense nucleic acid to a human gene or mRNA encoding the polypeptide of (a);

(f) an interfering RNA (RNAi) to a human gene or mRNA encoding the polypeptide of (a).

2. A method according to claim 1, wherein the endothelial cells are contacted with the composition *ex vivo*.

3. A method according to claim 1, wherein the composition comprises a pharmaceutically acceptable diluent, adjuvant, or carrier, and the contacting step comprises administering the composition to a mammalian subject to differentially modulate BECs or LECs in the mammalian subject.

4. A method according to claim 3, comprising:

identifying a human subject with a disorder characterized by hyperproliferation of LECs; and

administering to the human subject the composition, wherein the agent differentially inhibits LEC growth compared to BEC growth.

5. A method according to claim 3, comprising:

identifying a human subject with a disorder characterized by hyperproliferation of LECs;

screening LECs of the subject to identify overexpression of a polypeptide set forth in Table 3; and

administering to the human subject the composition, wherein the agent differentially inhibits LEC growth compared to BEC growth by inhibiting expression of the polypeptide identified by the screening step.

6. A method according to claim 3 of modulating the growth of lymphatic endothelial cells in a human subject, comprising steps of:

identifying a human subject with a hypoproliferative lymphatic disorder;

screening the subject to identify underexpression or underactivity of a LEC polypeptide set forth in Table 3, wherein said protein is not set forth in Table 1 or 2;

administering to the human subject said composition, wherein the agent comprises the LEC polypeptide (a) identified by the screening step or an active fragment of said polypeptide, or comprises the polynucleotide (b) that comprises a nucleotide sequence that encodes the polypeptide.

7. Use of an agent for the manufacture of a medicament for the differential modulation of blood vessel endothelial cell (BEC) or lymphatic vessel endothelial cell (LEC) growth or differentiation, said agent selected from the group consisting of:

(a) a polypeptide that comprises an amino acid sequence of a BEC polypeptide or a LEC polypeptide, or an active fragment of said polypeptide ;

(b) a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide according to (a);

(c) an antibody that specifically binds to a polypeptide according to (a);

(d) a polypeptide comprising a fragment of the antibody of (c), wherein the fragment and the antibody bind to the polypeptide;

(e) an antisense nucleic acid to a human gene or mRNA encoding the polypeptide of (a);

(f) an interfering RNA (RNAi) to a human gene or mRNA encoding the polypeptide of (a).

8. A method or use according to any one of claims 1-7, wherein the polypeptide is a LEC polypeptide selected from the LEC polypeptides set forth in Table 3, and the agent differentially modulates LEC growth or differentiation over BEC growth or differentiation..

9. A method or use according to any one of claims 1-7, wherein the polypeptide is a BEC polypeptide selected from the BEC polypeptides set forth in Table 4, and the agent differentially modulates BEC growth or differentiation over LEC growth or differentiation.

10. A method or use according to any one of claims 8 and 9, wherein the polypeptide is not set forth in Tables 1 or 2.

11. A method or use according to claim 8, wherein the LEC polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 81, 187, 207, 211, 221, 235, 241, 293, and 391.

12. A method or use according to claim 8, wherein the LEC polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-34, 46, and 48.

13. A method or use according to claim 12, wherein the agent comprises an antibody according to (c) or polypeptide according to (d).

14. A method according to claim 12, wherein the agent comprises an extracellular domain fragment of the polypeptide of (a), or a polynucleotide encoding said extracellular domain fragment.

15. A method or use according to any one of claims 1-10, wherein the agent comprises an antisense molecule.

16. A method of treating hereditary lymphedema comprising:

identifying a human subject with lymphedema and with a mutation in at least one allele of a gene encoding a LEC protein identified in Table 3, wherein the mutation correlates with lymphedema in human subjects, and with the proviso that said LEC protein is not VEGFR-3; and

administering to said subject a composition comprising a lymphatic growth agent selected from the group consisting of VEGF-C polypeptides, VEGF-D polypeptides, VEGF-C polynucleotides, and VEGF-D polynucleotides.

17. Use of a lymphatic growth agent selected from the group consisting of VEGF-C polypeptides, VEGF-D polypeptides, VEGF-C polynucleotides, and VEGF-D polynucleotides in the manufacture of a medicament for the treatment of hereditary lymphedema resulting from a mutation in a LEC gene identified in Table 3, with the proviso that said gene is not *VEGFR-3*.

18. A method of screening for an endothelial cell disorder or predisposition to said disorder, comprising'

obtaining a biological sample containing endothelial cell mRNA from a human subject; and

measuring expression of a BEC or LEC gene from the amount of mRNA in the sample transcribed from said gene, wherein the BEC or LEC gene encodes a polypeptide identified in Table 3 or 4.

19. A method of monitoring the efficacy or toxicity of a drug on endothelial cells, comprising steps of:

measuring expression of at least one BEC or LEC gene in endothelial cells of a mammalian subject before and after administering a drug to the sujbect, wherein the at least one BEC or LEC gene encodes a polypeptide set forth in Table 3 or Table 4, and wherein changes in expression of the BEC or LEC gene correlates with efficacy or toxicity of the drug on endothelial cells.

20. A method of identifying compounds that modulate growth of endothelial cells, comprising

culturing endothelial cells in the presence and absence of a compound; and

measuring expression of at least one BEC or LEC gene in the cells, wherein the BEC or LEC gene is selected from the genes encoding polypeptides set forth in Tables 3 and 4, wherein a change in expression of at least one BEC gene in the presence compared to the absence of the compound identifies the compound as a modulator of BEC growth, and wherein a change in expression of at least one LEC gene in the presence compared to the absence of the compound identifies the compound as a modulator of LEC growth.

21. A method according to claim 20 of screening for a compound that selectively modulates BEC or LEC growth or differentiaion,

wherein the measuring step comprises measuring expression of at least one BEC gene and at least one LEC gene in the cells, and

wherein the method comprises screening for a compound that selectively modulates BEC or LEC growth or differentiation by selecting a compound that differentially modulates expression of the at least one BEC gene compared to expression of the at least one LEC gene.

22. A composition comprising

an isolated polynucleotide that comprises a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391; and

a pharmaceutically acceptable diluent, carrier or adjuvant.

23. A composition according to claim 22, comprising a polynucleotide that comprises a nucleotide sequence selected from the group consisting of SEQ ID NOS: 14-30, 45, 47, 49, 51, 82, 93, 111, 188, 208, 212, 222, 236, 242, 294, and 392, or a fragment thereof that encodes the polypeptide.

24. An expression vector comprising an expression control sequence operably linked to a polynucleotide that comprises a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391.

25. An expression vector according to claim 24 that is a replication-deficient adenoviral or adeno-associated viral vector containing the polynucleotide.

26. A composition comprising an expression vector according to claim 24 or 25 and a pharmaceutically acceptable diluent, carrier, or adjuvant.

27. A kit comprising the composition according to any one of claims 22, 23, or 26 packaged with a protocol for administering the composition to a mammalian subject to modulate the lymphatic system in said subject.

28. A host cell transformed or transfected with an expression vector according to claim 24.

29. A method for producing a LEC polypeptide comprising steps of growing a host cell according to claim 28 under conditions in which the cell expresses the polypeptide encoded by the polynucleotide.

30. A purified and isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, 81, 187, 207, 211, 221, 235, 241, 293, and 391.

31. A purified and isolated polypeptide comprising an amino acid sequence selected from the group consisting of:

(a) SEQ ID NOS: 31-34, 46, 48, 207, 676, 859, and 861; and

(b) an extracellular domain fragment of at least 10 amino acids of an amino acid sequence of (a).

32. A purified and isolated, soluble polypeptide according to claim 31 comprising an extracellular domain fragment of an amino acid sequence selected from the group consisting of : SEQ ID NOS: 31-34, 46, 48, 207, 676, 859, and 861, wherein the polypeptide lacks any transmembrane domain.

33. A polypeptide according to claim 32 that lacks any intracellular domain.

34. A fusion protein comprising a polypeptide according to claim 32 or 33 fused to an immunoglobulin fragment comprising an immunoglobulin constant region.

35. A composition comprising a polypeptide or protein according to any one of claims 30-34 and a pharmaceutically acceptable diluent, carrier or adjuvant.

36. A kit comprising the composition according to claim 35 and a protocol for administering said pharmaceutical composition to a mammalian subject to modulate the lymphatic system in said subject.

37. An antibody that specifically binds to a polypeptide according to any one of claims 30-34.

38. An antibody according to claim 37 that is a humanized antibody.

39. A protein comprising an antigen binding domain of an antibody that specifically binds to a polypeptide according to any one of claim 30-34, wherein said protein specifically binds to said polypeptide.

40. A method of identifying a LEC nucleic acid comprising:

(a) contacting a biological sample containing a candidate LEC nucleic acid with a polynucleotide comprising a fragment of at least 14 contiguous nucleotides of a sequence selected from the group consisting of SEQ ID NOS:1-30, 45, 47, 49, 51, 82, 93, 111, 188, 208, 212, 236, 242, 294, and 392, or a complement thereof, under the following stringent hybridization conditions:

(i) hybridization at 42°C for 20 hours in a solution containing 50% formamide, 5xSSPE, 5x Denhardt's solution, 0.1% SDS and 0.1 mg/ml denatured salmon sperm DNA, and

(ii) washing for 30 minutes at 65°C in 1xSSC, 0.1% SDS; and

(b) detecting hybridization of said candidate LEC nucleic acid and said polynucleotide, thereby identifying a LEC nucleic acid.

41. A method of identifying a LEC protein comprising:

(a) contacting a biological sample containing a candidate LEC protein with a LEC protein binding partner selected from the group consisting of an antibody according to claim 37 or a protein according to claim 39, under conditions suitable for binding therebetween; and

(b) detecting binding between said candidate LEC protein and said LEC binding partner, thereby identifying a LEC protein.

42. A method of identifying a LEC comprising:

(a) contacting a biological sample comprising cells with a LEC binding partner under conditions suitable for binding therebetween, wherein said LEC binding partner comprises an antibody that binds to a polypeptide comprising a sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 48, 207, 676, 859, and 861, or comprises an antigen binding fragment of said antibody; and

(b) identifying a LEC by detecting binding between a cell and said LEC binding partner, where binding of the LEC binding partner to the cell identifies a LEC.

43. A method of assaying for risk of developing hereditary lymphedema, comprising

(a) assaying nucleic acid of a human subject for a mutation that correlates with a hereditary lymphedema phenotype and alters the encoded amino acid sequence of at least one gene allele of the human subject when compared to the amino acid sequence of the polypeptide encoded by a corresponding wild-type gene allele, wherein the wild-type polypeptide is a polypeptide identified in Table 3.

44. A method of assaying for risk of developing hereditary lymphedema, comprising

(a) assaying nucleic acid of a human subject for a mutation that correlates with a hereditary lymphedema phenotype and alters the encoded amino acid sequence of at least one gene allele of the human subject when compared to the amino acid sequence of the polypeptide encoded by a corresponding wild-type gene allele, wherein the wild-type polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 52, 54, 207, 676, 859, and 861;

(b) correlating the presence or absence of said mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of said mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of said mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema.

45. A method of assaying for risk of developing hereditary lymphedema, comprising

(a) assaying nucleic acid of a human subject for a mutation that alters the encoded amino acid sequence of at least one transcription factor allele of the human subject and alters transcription modulation activity of the transcription factor polypeptide encoded by the allele, when compared to the transcription modulation activity of a transcription factor polypeptide encoded by a wild-type allele,

wherein the wild-type transcription factor polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 211, SEQ ID NO: 241, and transcription factors encoded by sequences in Table 5; and

(b) correlating the presence or absence of said mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of said mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of said mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema.

46. The method according to claim 45 wherein said wild-type transcription factor allele comprises the Sox18 amino acid sequence set forth as SEQ ID NO:54.

47. The method according to claim 46 wherein the assaying identifies a mutation altering a transactivating or DNA binding domain amino acid sequence of the protein encoded by the Sox18 allele.

48. The method according to claim 46, wherein said mutation reduces transcriptional activation of a SOX18-responsive gene compared to transcriptional activation of said gene by wild-type SOX18.

49. A method of assaying for risk of developing hereditary lymphedema, comprising

(a) assaying nucleic acid of a human subject for a mutation that alters the encoded amino acid sequence of at least one LEC gene allele of the human subject and alters the binding affinity of the adhesion polypeptide encoded by the LEC gene allele, when compared to the binding affinity of an adhesion polypeptide encoded by a wild-type allele,

wherein the wild-type adhesion polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS:31-34, 46, 207, 676, 859, and 861; and

(b) correlating the presence or absence of said mutation in the nucleic acid to a risk of developing hereditary lymphedema, wherein the presence of said mutation in the nucleic acid correlates with an increased risk of developing hereditary lymphedema, and wherein the absence of said mutation in the nucleic acid correlates with no increased risk of developing hereditary lymphedema.

50. The method according to any one of claims 43-49, wherein the assaying identifies the presence of the mutation, and the correlating step identifies the increased risk of said patient developing hereditary lymphedema.

51. A method of screening a human subject for an increased risk of developing hereditary lymphedema comprising assaying nucleic acid of a human subject for a mutation that alters the encoded amino acid sequence of at least one polypeptide comprising an amino acid sequence of Table 3.

52. A method of claim 51, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, and 54, 207, 676, 859, and 861 in a manner that correlates with the risk of developing hereditary lymphedema.

53. The method according to claim 52 wherein the polypeptide comprises the SOX18 amino acid sequence set forth in SEQ ID NO: 54.

54. The method according to any one of claims 43-53 wherein said method comprises at least one procedure selected from the group consisting of:

(a) determining a nucleotide sequence of at least one codon of at least one polynucleotide of the human subject;

(b) performing a hybridization assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences;

(c) performing a polynucleotide migration assay to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences; and

(d) performing a restriction endonuclease digestion to determine whether nucleic acid from the human subject has a nucleotide sequence identical to or different from one or more reference sequences.

55. The method according to any one of claims 43-53 wherein said method comprises: performing a polymerase chain reaction (PCR) to amplify nucleic

acid comprising the coding sequence of said LEC polynucleotide, and determining nucleotide sequence of the amplified nucleic acid.

56. A method of screening for a hereditary lymphedema genotype in a human subject, comprising:

(a) providing a biological sample comprising nucleic acid from said subject, and

(b) analyzing said nucleic acid for the presence of a mutation altering the encoded amino acid sequence of the at least one allele of at least one gene in the human subject relative to a human gene encoding an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-44, 46, 48, 50, 52, 54, 207, 676, 859, and 861, wherein the presence of a mutation altering the encoded amino acid sequence in the human subject in a manner that correlates with lymphedema in human subjects identifies a hereditary lymphedema genotype.

57. The method according to claim 56 wherein said biological sample is a cell sample.

58. The method according to claim 56 wherein said analyzing comprises sequencing a portion of said nucleic acid.

59. The method according to claim 56 wherein the human subject has a hereditary lymphedema genotype identified by the method of screening.

60. The method according to claim 49, wherein the at least one gene corresponds to the human Sox18 gene that encodes the amino acid sequence set forth in SEQ ID NO: 54.

61. A method of inhibiting lymphangiogenesis comprising

administering to a subject an inhibitor of a LEC transmembrane polypeptide,

wherein the LEC transmembrane polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs: 31-34, 46 48, 207, 676, 859, and 861, and

wherein the inhibitor is selected from the group consisting of

(a) a soluble extracellular domain fragment of the LEC transmembrane polypeptide;

(b) an antibody that binds to the extracellular domain of the LEC transmembrane polypeptide;

(c) a polypeptide comprising an antigen binding domain of the antibody according to (b); and

(d) an antisense nucleic acid complementary to the nucleic acid encoding the LEC transmembrane polypeptide or its complement.

62. A method according to claim 61, wherein the inhibitor is a polypeptide comprising an extracellular domain fragment of an LEC polypeptide, wherein the sequence of said extracellular domain is selected from the group consisting of amino acids 1-152 of SEQ ID NO:31, amino acids 1-695 of SEQ ID NO:32 and amino acids 1-248 of SEQ ID NO:33.

63. The method according to claim 61 or 62 wherein said subject is a human containing a tumor.

64. A method for modulating lymphangiogenesis in a mammalian subject comprising: administering to a mammalian subject in need of modulation of lymphangiogenesis an antisense molecule to a LEC polynucleotide, in an amount effective to inhibit transcription or translation of the poypeptide encoded by the LEC polynucleotide, wherein the LEC polynucleotide comprises a nucleotide sequence selected from the group consisting of SEQ ID NOS: 14-30, 45, 47, 49, AND 51, 208, 677, 860, and 862.

65. A method of treating hereditary lymphedema, comprising:

(a) identifying a human subject with hereditary lymphedema and with a mutation that alters the encoded amino acid sequence of at least one polypeptide of the human subject, relative to the amino acid sequence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOS: 31-44, 46, 48, 50, 52, 54, 207, 676, 859, and 861; and

(b) administering to said subject a lymphatic growth factor selected from the group consisting of a VEGF-C polynucleotide, a VEGF-C polypeptide, a VEGF-D polynucleotide, and a VEGF-D polypeptide.

66. A method of modulating the growth of endothelial cells or endothelial precursor cells, comprising contacting endothelial cells or endothelial precursor cells with a composition comprising an agent the modulates prox-1 transcription regulation in the cells, wherein the agent is selected from the group consisting of:

(a) a prox-1 polypeptide;

(b) a polynucleotide encoding a prox-1 polypeptide;

(c) an antisense molecule to prox-1.

67. A method according to claim 66, wherein the cells comprises cultured endothelial cells or endothelial precursor cells, and the contacting is performed *ex vivo*.

68. A method according to claim 67, wherein the contacting comprises including the agent in the culture medium.

69. A method according to any one of claims 66-68, wherein the cells comprise endothelial precursor cells.

70. A method according to any one of claims 66-69, wherein the cells are introduced into a mammalian subject after the contacting step.

71. A method according to claim 70, wherein the subject is human.

72. A method according to claim 71, wherein the human subject has a LEC disorder.

73. A method of increasing LEC function in a human subject, comprising:

isolating endothelial cells or endothelial precursor cells from a human subject;

transforming or transfecting the endothelial cells with an expression vector comprising a nucleotide sequence encoding a prox-1 polypeptide, to promote LEC differentiation and growth; and

administering the LEC cells to a human subject after the transforming or transfecting step.

74. A method according to claim 73, wherein the human subject of the isolating and administering steps is the same.

75. A method according to claim 73 or 74, wherein the human subject has lymphedema.

76. A method according to any one of claims 73-75, wherein the vector and transforming or transfecting method are selected for transient expression of the prox-1.

77. A method according to any one of claims 73-75, wherein the expression vector comprises a replication-deficient adenoviral vector.

78. An isolated polypeptide comprising an amino acid sequence at least 95% identical to amino acids 61-127 of SEQ ID NO: 31.

79. A polypeptide according to claim 78, comprising an amino acid sequence at least 95% identical to amino acids 30-152 of SEQ ID NO: 31.

80. A soluble polypeptide comprising a fragment of the amino acid sequence set forth in SEQ ID NO: 31, wherein said fragment lacks the transmembrane and intracellular amino acids of SEQ ID NO: 31.

81. An isolated polypeptide comprising at least one leucine-rich region of SEQ ID NO: 32.

82. An isolated polypeptide according to claim 81, wherein the polypeptide lacks transmembrane amino acids of SEQ ID NO: 32.

83. An isolated polypeptide comprising at least one leucine-rich region of SEQ ID NO: 33.

84. An isolated polypeptide according to claim 81, wherein the polypeptide lacks transmembrane amino acids of SEQ ID NO: 33.

85. An isolated polypeptide comprising an amino acid sequence at least 95% identical to a fragment of a polypeptide comprising the amino acid sequence set forth in SEQ ID NO: 111,

wherein said fragment includes at least one thrombospondin type I repeat sequence.

86. An isolated polypeptide according to claim 85, wherein said fragment includes the six thrombospondin type I repeat sequences of SEQ ID NO: 111.

87. An isolated polypeptide comprising an amino acid sequence at least 95% identical to a fragment of a polypeptide comprising the amino acid sequence set forth in SEQ ID NO: 111,

wherein said fragment includes at least one immunoglobulin C-2 type domain.

88. An isolated polypeptide according to claim 85, wherein said fragment includes the three immunoglobulin C-2 type domain sequences of SEQ ID NO: 111.

89. A fusion protein comprising a polypeptide according to any one of claims 78-88 and a heterologous polypeptide.

90. An antibody that specifically binds to a polypeptide according to any one of claims 78-88.

91. A polynucleotide comprising a nucleotide sequence that encodes a polypeptide according to any one of claims 78-89.

92. An expression vector comprising a polynucleotide according to claim 91 operatively linked to an expression control sequence.

93. An expression vector according to claim 92 that is a reprelication deficient adenoviral vector.

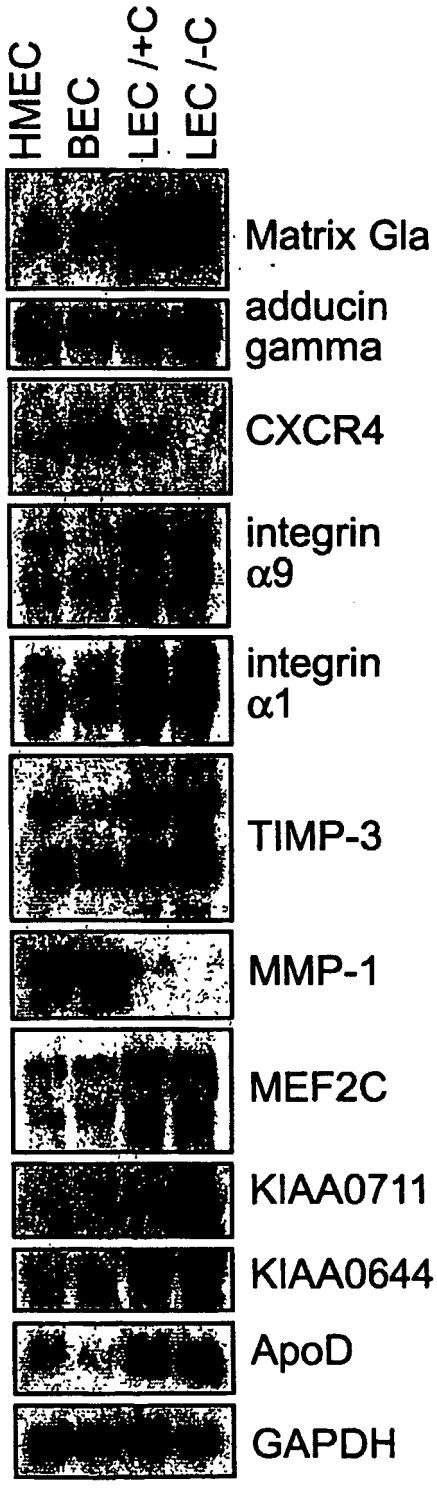
HMEC
BEC
LEC /+C
LEC /-C
Matrix Gla
adducin gamma
CXCR4
integrin α9
integrin α1
TIMP-3
MMP-1
MEF2C
KIAA0711
KIAA0644
ApoD
GAPDH

Fig. 1

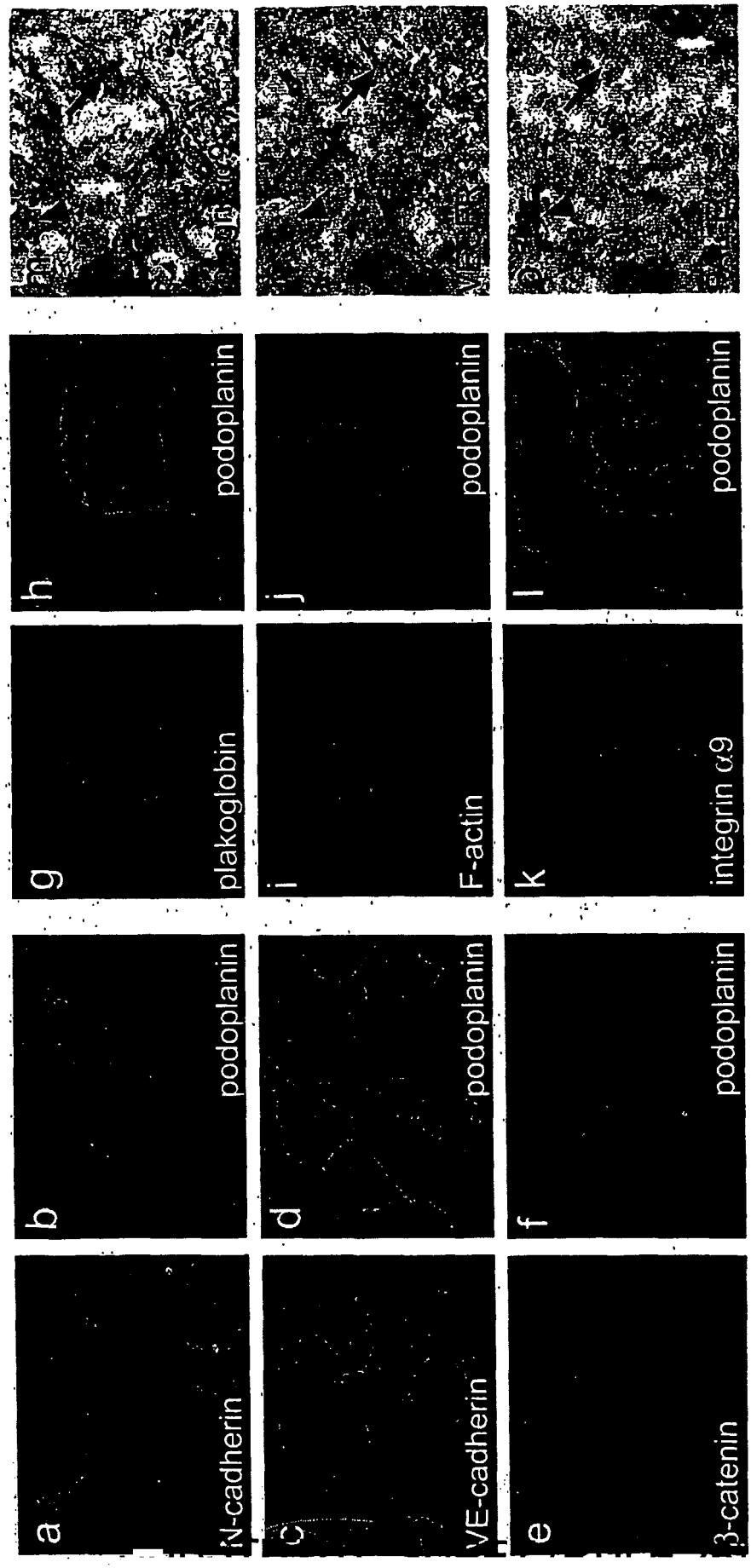
a
N-cadherin
b
podoplanin
c
VE-cadherin
d
podoplanin
e
β-catenin
f
podoplanin
g
plakoglobin
h
podoplanin
i
F-actin
j
podoplanin
k
integrin α9
l
podoplanin

Fig. 2

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US03/06900

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C07H 21/04; C12Q 1/68
US CL : 435/6, 325, 375, 91.1; 514/2, 44; 536/24.5, 24.3

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
U.S. : 435/6, 325, 375, 91.1; 514/2, 44; 536/24.5, 24.3

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Continuation Sheet

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	KARPANEN et al. Vascular Endothelial Growth Factor C Promotes Tumor Lymphangiogenesis and Intralymphatic Tumor Growth. Cancer Research. 01 March, 2001, Vol. 61, pages 1786-1790.	1-9, 13-14, 16-21, 25-29, 33-39, 41, 43, 47, 48, 50, 51, 57-59, 63, 66-75, and 89-93
A	MANDRIOTA et al. Vascular endothelial growth factor-C-mediated lymphangiogenesis promotes tumor metastasis. The EMBO Journal. 2001, Vol. 20, No. 4, pages 672-682.	1-9, 13-14, 16-21, 25-29, 33-39, 41, 43, 47, 48, 50, 51, 57-59, 63, 66-75, and 89-93
A	JOUKOV et al. A Recombinant Mutant Vascular Endothelial Growth Factor-C that Has Lost Vascular Endothelial Growth Factor Receptor-2 Bidning, Activation, and Vascular permeability Activities. Journal of Biological Chemistry. 20 March, 1998, Vol. 273, No. 12, pages 6599-6602.	1-9, 13-14, 16-21, 25-29, 33-39, 41, 43, 47, 48, 50, 51, 57-59, 63, 66-75, and 89-93
A	WO 98/33917 A1 (THE LUDWIG INSTITUTE FOR CANCER RESEARCH) 06 August 1998 (06.08.1998), see Abstract.	1-9, 13-14, 16-21, 25-29, 33-39, 41, 43, 47, 48, 50, 51, 57-59, 63, 66-75, and 89-93

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search
12 June 2003 (12.06.2003)

Date of mailing of the international search report
15 AUG 2003

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Form PCT/ISA/210 (second sheet) (July 1998)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US03/06900

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claim Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claim Nos.: 11,12,22-24,30-32,40,42,44-46,49,52,53,56,60-62,64,65 and 78-88
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
Please See Continuation Sheet

3. ☒ Claim Nos.: 10, 15, 54, 55, 76, 77
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

PCT/US03/06900

Continuation of Box I Reason 2:
Claims 11, 12, 22, 23, 24, 30, 31, 32, 40, 42, 44, 45, 46, 49, 52, 53, 56, 60, 61, 62, 64, 65, and 78-88 encompass SEQ ID NOs. where no meaningful search could be performed because the application does not contain a sequence listing or a disk containing the sequence listing.

Continuation of B. FIELDS SEARCHED Item 3:
Medline, Embase, CaPlus, CancerLit
search terms: VEGF-C vascular endothelial growth factor C, VEGFR-3, VEGF-D, VEGFR-2 and tumor growth

专利名称(译)	淋巴和血液内皮细胞基因		
公开(公告)号	EP1487857A1	公开(公告)日	2004-12-22
申请号	EP2003713942	申请日	2003-03-07
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摘要(译)

本发明提供了在淋巴管与血管内皮细胞中差异表达的多核苷酸和基因。
这些基因可用于通过淋巴系统治疗涉及淋巴管的疾病，例如淋巴水肿，
各种炎性疾病和癌症转移。