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(54) METHOD OF MEASURING CARDIAC RELATED PARAMETERS NON-INVASIVELY VIA THE LUNG DURING SPONTANEOUS AND CONTROLLED VENTILATION

(71) Applicant: THORNHILL SCIENTIFIC INC.,

Toronto, CA (US)

Inventors: Joseph Fisher, Thornhill (CA); David

Preiss, Thornhill (CA); Takafumi Asami, Nagoya (JP); Alex Vesely, Victoria (CA); Eltan Prisman, Toronto (CA); Ron Somogyi, Toronto (CA); Steve Iscoe, Kingston (CA)

Assignee: THORNHILL SCIENTIFIC INC.,

Toronto (CA)

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(57)ABSTRACT

An apparatus to measure pulmonary blood flow and cardiac output (Q) comprising: a) a breathing circuit which, at exhalation, keeps exhaled gas separate from inhaled gas and at inhalation, when V_E is greater than first gas set (FGS) flow, results in a subject inhaling FGS first and then a second gas set (SGS), for the balance of inhalation; b) a gas sensor for monitoring gas concentrations at the patient-circuit interface; c) a gas flow control means for controlling the rate of FGS flow into the breathing circuit; d) machine intelligence consisting of a computer or logic circuit capable of controlling the gas flow control means which receives the output of the gas sensor means and outputs pulmonary blood flow.

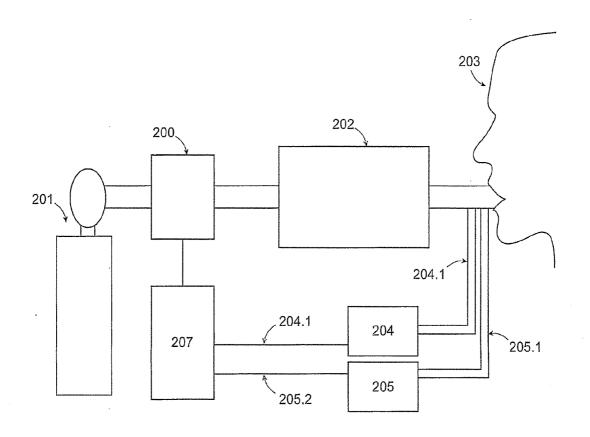


Figure 2

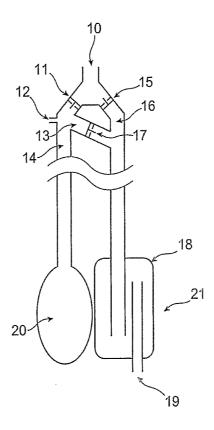


Figure 3

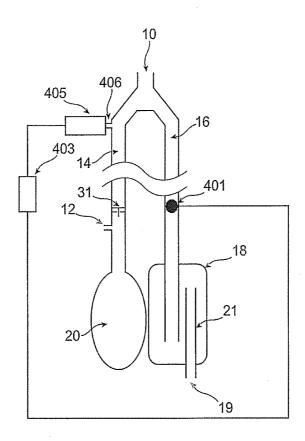


Figure 3B

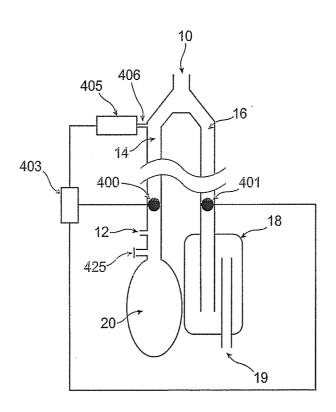


Figure 3C

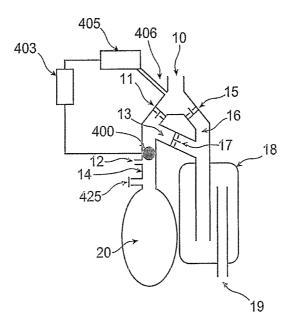


Figure 3D

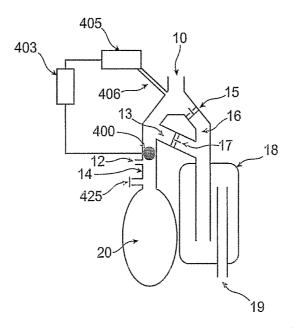


Figure 3E

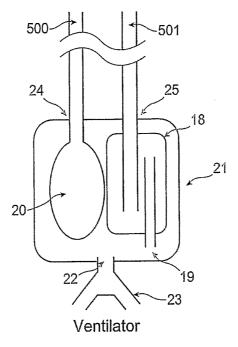


Figure 4

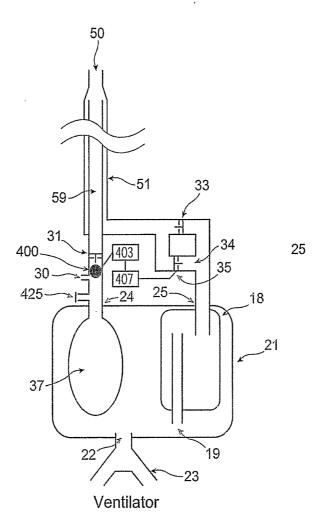


Figure 4B

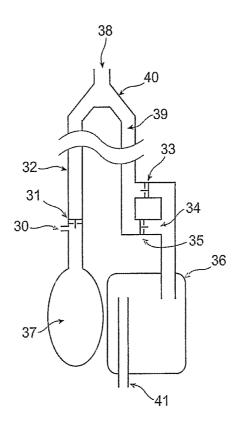


Figure 5

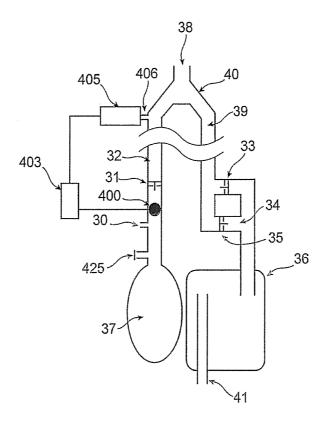


Figure 5A

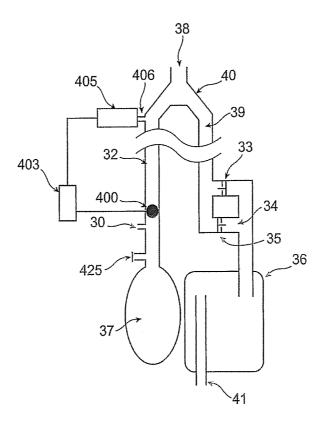


Figure 5B

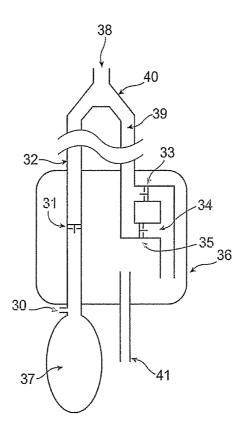


Figure 5C

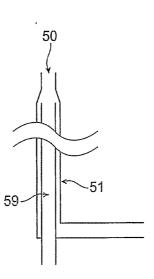


Figure 6

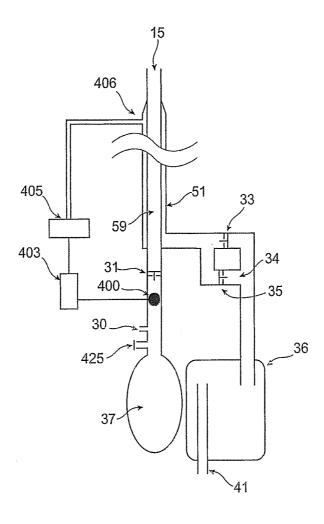


Figure 6A

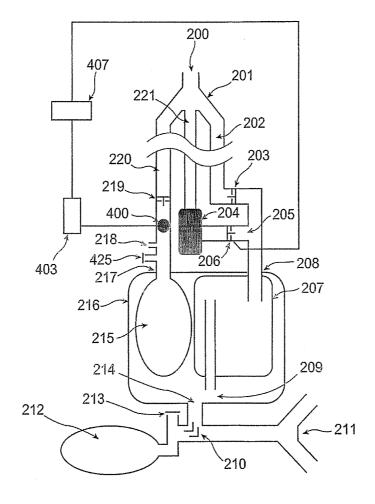
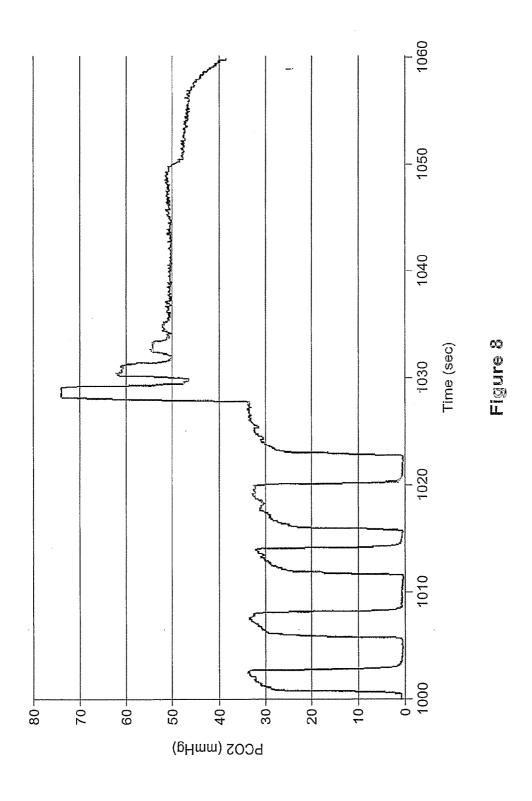
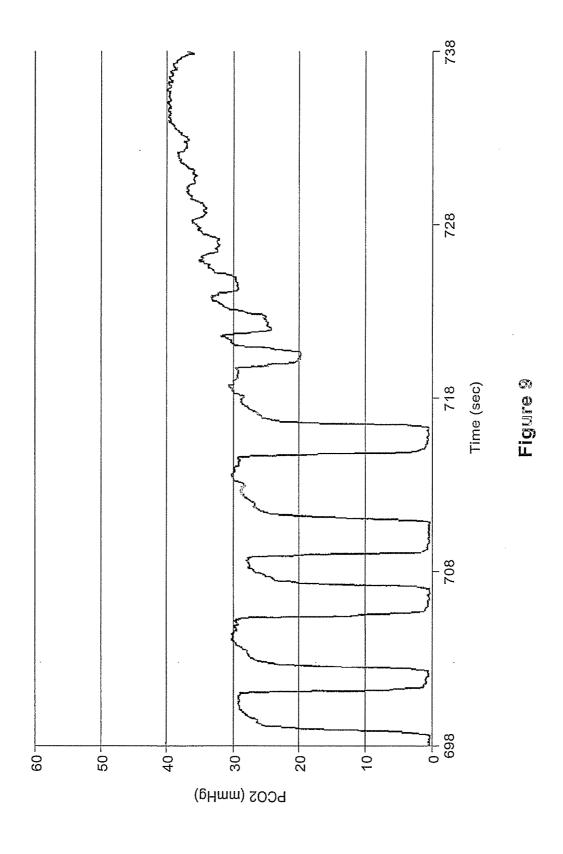
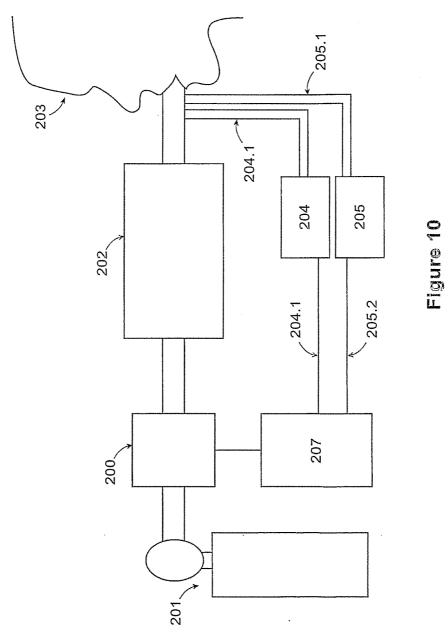


Figure 7







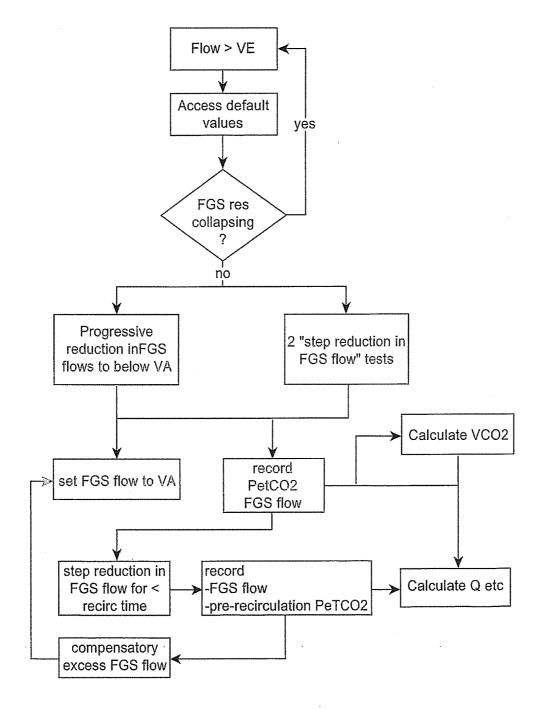


Figure 11

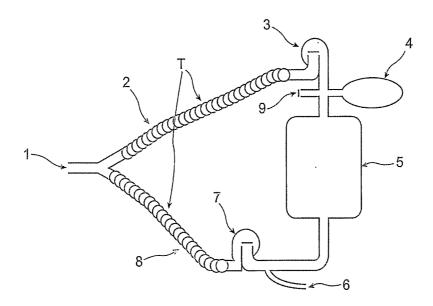


Figure 12

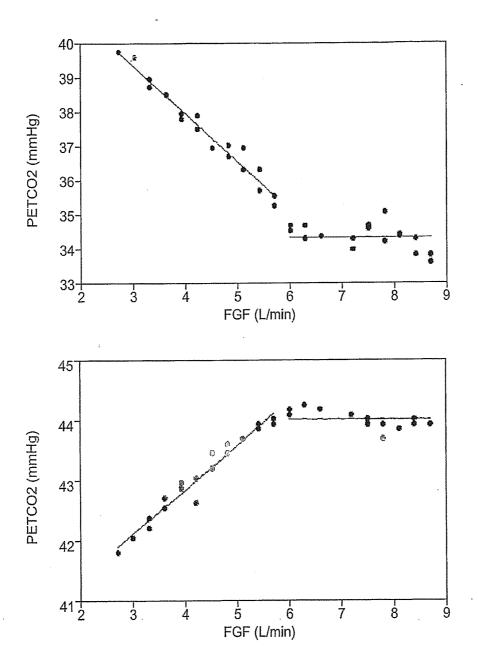


Figure 13

METHOD OF MEASURING CARDIAC RELATED PARAMETERS NON-INVASIVELY VIA THE LUNG DURING SPONTANEOUS AND CONTROLLED VENTILATION

FIELD OF THE INVENTION

[0001] This invention discloses a method that calculates noninvasively, via the lung, the total cardiac output, pulmonary blood flow, shunt flow, anatomical and alveolar deadspace, true mixed venous $\rm O_2$ saturation, true mixed venous $\rm PCO_2$, and $\rm PaCO_2$. Furthermore the method can be performed in ventilated subjects, subjects breathing spontaneously, even in the presence of variations in their tidal volume and breathing frequency. Subjects need not perform any respiratory manoeuvre such as hyperventilation or breath holding to perform the test.

BACKGROUND OF THE INVENTION

1. Importance of Cardiac Output

[0002] A physician's ability to determine a patient's cardiac output (Q, the volume of blood pumped by the heart each minute) is important in the assessment of critically ill patients. There are various devices and methods that provide a direct or indirect measure of \dot{Q} (see table 1). The most common method used in clinical practice is thermo-dilution, established by Ganz et al (1). Commercially manufactured catheters (referred to as Swan-Ganz catheters, named after the inventors) contain multiple lumina, an embedded thermister, and a balloon at the tip. The method requires the insertion of the catheter through the skin to access a large central vein such as the internal jugular, subclavian, cephalic or femoral. When the balloon at the end of the catheter is inflated, the catheter tip is carried along with the flow of blood to the right ventricle of the heart and then into the pulmonary artery. The part of the catheter that remains outside the body has connections that can be attached to electrical sensors that determine the pressure and temperature in the pulmonary artery where the tip of the catheter is positioned. Calculation of Q requires the injection of a fixed volume of cool liquid of known temperature into a lumen of the catheter that has its opening part way along its length (usually in a part of the catheter in the right atrium). The thermister at the tip of the catheter will register changes in temperature as the cool liquid, carried by the blood, passes. The extent of dilution of the cold bolus of liquid by warm blood will determine the temporal profile of the temperature change at the tip of the catheter. This is referred to as the thermodilution method of measuring cardiac output (TD Q).

[0003] The popularity of TD Q stems from ease of use once the catheter is in place. However, the placing and maintenance of the catheter entails considerable risk and expense. Insertion of the Swan-Ganz catheter is associated with complications that are frequently fatal such as puncture of the carotid or subclavian artery with associated internal haemorrhage or stroke, tension pneumothorax, rupture of the right ventricle, malignant arrhythmias (including fatal ventricular fibrillation), and rupture of the pulmonary artery. As a foreign body violating the skin barrier, a pulmonary artery catheter is a constant threat as a source of blood-born infection that is the greatest risk to heart valves, artificial joints, and other implants. Such infections are medical disasters leading to severe morbidity and death. Furthermore, the use of pulmo-

nary artery catheters to measure TD \dot{Q} is very expensive as it requires admission to an intensive care facility where there is continuous presence of critical care nursing and medical staff. Despite these risks, it is still not the ideal method to measure \dot{Q} as it tends to overestimate \dot{Q} by as much as 10% compared to the Fick method (see below) and, for greatest accuracy, requires repeated measurements as its precision is poor. The variability of repeated single measurements is about 22% and can be reduced to 10% by repeated averages of 3 measurements (2). A single thermodilution measurement is considered to be plus or minus 33% the true value.(3)

[0004] Because of the expense and risks of keeping the catheters in place, they are removed as soon as practical, often within 24-48 hours of major heart surgery. Often they are removed while the information they provide can still be clinically useful and well before the patient is no longer at significant risk for relapse. If the patient's health deteriorates, a decision must be made about re-inserting the catheter.

[0005] An automated non-invasive method of \dot{Q} monitoring would be very useful in the following clinical scenarios:

- [0006] a) Selected low risk patients now routinely undergoing pulmonary artery catheterization for intra- and postoperative monitoring.
- [0007] b) Patients whose Q would be clinically important to know but in whom the risks and costs of insertion of a pulmonary catheter cannot be justified; this includes ward patients, outpatients or patients in the emergency department or doctor's office.
- [0008] c) Patients who are too sick to warrant the added risk of pulmonary artery catheter insertion
- [0009] d) High and moderate cardiac risk patients undergoing minor and moderate non-cardiac surgical procedures
- [0010] e) Severely ill patients with non-cardiac disease.
 [0011] f) Relatively healthy patients undergoing major stressful surgery.
- [0012] g) Situations in which Q is clinically indicated but there is no access to the expertise and critical care facilities required for the use of the pulmonary artery catheters
- [0013] h) Means of monitoring response to cardiovascular therapy such as for hypertension and heart failure.
- [0014] i) As a non-invasive diagnostic test of cardiopulmonary status.

[0016] j) As a means of assessing cardiovascular fitness. [0016] Despite these many applications, non-invasive methods of Q measurements have not obtained widespread clinical acceptance. The most commonly researched methods include ECG bio-impedance (Imhoff, 2000 (4)), and pulsed-wave Doppler esophageal sonography. These methods have good repeatability (5-12) and good limits of agreement with either thermodilution or Fick-based methods but only in some populations of subjects. Each method fails in certain patients groups with such pathologies as very high or low Q states as occur in surgical patients, septic shock, exercise or cardiogenic shock.

2. Background Physiology and Definition of Terms

[0017] Venous blood returns to the right side of the heart from the muscles and organs with reduced oxygen (O_2) and increased carbon dioxide (CO_2) levels. Blood from various parts of the body is mixed in the right side of the heart and pumped to the lungs via the pulmonary artery. The blood in the pulmonary artery is known as the mixed venous blood. In

the lungs the blood vessels break up into a network of small vessels that surround tiny lung sacs known as alveoli. This network of vessels surrounding the alveoli provides a large surface area for the exchange of gases by diffusion along their partial pressure gradients. After a breath of air is inhaled into the lungs, it dilutes the CO₂ left in the alveoli at the end of the previous expiration, thereby establishing a pressure gradient between the partial pressure of CO₂ (PCO₂) in the mixed venous blood (Pv, CO₂) arriving at the alveoli and the alveolar PCO₂ (PACO₂). The CO₂ diffuses into the alveoli from the mixed venous blood diminishing the PCO₂ in the blood, and increasing the PCO₂ in the alveoli until equilibrium is established between the PCO2 in alveolar capillary blood and the PCO₂ in the alveoli. The blood then returns to the left side of the heart via the pulmonary vein and is pumped into the arterial system by the left ventricle. The PCO₂ in the arterial blood (PaCO₂) is now the same as that in the alveoli. When the subject exhales, the gas at the very end of exhalation is considered to have come from the alveoli and thus simultaneously reflects the PCO₂ in the pulmonary capillaries and the alveoli; the PCO₂ in this gas is called the end-tidal PCO₂ $(PETCO_2)$.

[0018] The volume of gas breathed per minute, or minute ventilation (\dot{V}_E), is measured at the airway opening (nose and/mouth) and is expressed in L/min. The volume of breathed gas distributed to the alveoli (and thus contributing to gas exchange) is termed the alveolar ventilation (\dot{V}_A) and is also expressed in L/min. The part of \dot{V}_E that does not contribute to gas exchange is termed dead space ventilation. This is divided into the anatomical dead space that consists of the trachea and other gas-conducting tubes leading from the nose and mouth to the alveoli, and the alveolar dead space that is collectively the alveoli that are ventilated but not perfused with blood.

[0019] The VE during normal breathing provides the VA that is required to eliminate the CO₂ brought to the lungs. VE is controlled by a feedback system to keep PaCO₂ at a set level of approximately 40 mmHg. Under steady state conditions, the rate at which CO_2 is exhaled from the lungs ($\dot{V}CO_2$) is equal to the rate that it is brought to the lungs, which in turn is equal to the metabolic CO₂ production. We define steady state as the condition in which the flux of CO₂ at the lungs is equal to the CO₂ production and the VCO₂, PvCO₂ and PaCO₂ remain steady. If the VCO₂ is diminished, the CO₂ extraction from the mixed venous blood passing by the alveoli will be reduced resulting in an increase in the PaCO₂ when that blood reaches the arterial system. As the blood traverses the body, it will pick up additional CO₂ and will return to the pulmonary artery with a higher PCO₂ than on its previous passage. The time between the change in VCO₂ and re reappearance of the blood with raised PCO₂ in the mixed venous circulation is termed the recirculation time which is generally taken as 20-30 s in resting subjects.

3. The Fick Equation

[0020] The approach for respiratory-based methods for measuring \dot{Q} non-invasively is described by the Fick equation, a mass balance of any substance across the lungs. The Fick method was originally described for O_2 as a method for determining pulmonary blood flow. The Fick relation states that the O_2 uptake by the lung is equal to the difference between the pulmonary artery and systemic arterial O_2 contents times the \dot{Q} . The blood contents originally had to be obtained invasively from blood samples. The same relation

holds with respect to CO_2 . The advantage of using CO_2 as the tracer is that mixed venous and arterial blood contents of CO_2 may be determined non-invasively. The Fick mass balance equation for CO_2 is:

$$\dot{Q} = \frac{\dot{V}CO_2}{(C\overline{v}CO_2 - CaCO_2)}$$

where \dot{Q} is the cardiac output, $\dot{V}CO_2$ is the rate of elimination of CO_2 at the lungs, $C\bar{v}CO_2$ and $CaCO_2$ are the mixed venous and systemic arterial contents of CO_2 , respectively. $\dot{V}CO_2$ can be measured by a timed collection of expired gas and measuring its volume and CO_2 concentration. The term $CaCO_2$ can be calculated using an estimate of arterial PCO_2 ($PaCO_2$) as derived from the PCO_2 of end tidal gas ($PetCO_2$). The hemoglobin concentration (easily obtained from a venous blood sample or a drop of blood from a finger prick) and the relation between blood PCO_2 and CO_2 content (available from standard physiology texts) are then used to calculate $CaCO_2$.

[0021] However, $C\overline{v}CO_2$ is difficult to estimate. The PCO₂ of mixed venous blood ($P\overline{\nu}CO_2$) is difficult to determine as true mixed venous blood is present only in the pulmonary artery, which is inaccessible from the surface. The air in the lungs is in intimate contact with mixed venous blood, but CO₂ diffuses rapidly from the mixed venous blood into the alveoli before an equilibrium is established. The PCO₂ of the expired gas therefore reflects this equilibrium PCO₂ and not the PCO₂ of mixed venous blood. The PvCO₂ can be determined from expired gas only when there has been full equilibration with continuously replenished mixed venous blood or partial equilibration under controlled conditions that allow for back calculation of PvCO₂ from the PCO₂ in expired gas. Hence during rebreathing, the alveolar gas is not refreshed and the mixed venous blood continuously passes the alveoli such that an equilibrium is established whereby the PetCO₂ reflects the PCO₂ in mixed venous blood.

[0022] However, even in this scenario, the PCO₂ is not that which exists in the pulmonary artery. Blood in the pulmonary artery has a relatively low PO₂. Because of the Haldane effect, the low PO₂ allows the CO₂ to be carried by the hemoglobin at a relatively low PCO₂. When the mixed venous blood is exposed to gas in the alveoli, O₂ diffuses into the blood, binds to the hemoglobin and increases the PCO₂ needed for a given CO₂ content on the hemoglobin (the complimentary aspect of the Haldane effect). All methods based on full or partial equilibration of alveolar gas with PvCO2 take into account that the equilibration is to a virtual PCO₂ that would exist if the CO₂ content of the hemoglobin were the same as in mixed venous blood but the hemoglobin were saturated with O₂. We refer to this as the oxygenated mixed venous PCO2 (P vCO₂-oxy). Because the relationship between PCO₂ and content of CO₂ in blood is known, CvCO₂ can be calculated from both the true $P\overline{v}CO_2$ (as obtained, for example, from a pulmonary arterial blood sample) and PvCO2-oxy (as obtained by some of the non-invasive methods described below)¹.

The Pv^-CO_2 -oxy does not really exist but is a virtual number created by instantaneously oxygenating mixed venous blood before and diffusion of CO_2 into the alveoli. The $C\overline{v}CO_2$ is the same in each.

4. Rebreathing-Equilibration Method One method of measuring $P\overline{v}CO_2$ -oxy was introduced by Collier in 1956, and is known as the equilibration method. A bag is pre-filled with a high concentration of CO_2 (~10-13%) and the subject exhales

and inhales rapidly to and from the bag and PCO_2 is monitored continuously at the mouth. The object of the test is to find the combination of bag volume and bag concentration of CO_2 such that once the gas in the bag mixes with that in the lungs (the concentration of CO_2 in the residual gas in the lung at the end of a breath in a healthy person is ~5.5%), the partial pressure of CO_2 in the lung is equal to that in mixed venous blood. A flat segment of the PCO_2 tracing segment indicates that inspired and expired PCO_2 are equal. To identify the true $P\overline{\nu}CO_2$ -oxy, the flat segment must occur within the first 3-4 breaths, before recirculation raises the $P\overline{\nu}CO_2$ -oxy (see FIG. 8).

[0023] 4.1.1 Advantages of the Equilibration Method:

[0024] The capnograph reading is of gas equilibrated with $P\overline{\nu}CO_2$ -oxy and can be considered a directly measured value as opposed to a value obtained from calculation or extrapolation.

[0025] 4.1.2 Limitations of the Equilibration Method:

[0026] 4.1.2.1 The CO₂ concentration in the bag depends on bag size, the patient's lung volume, and the P vCO₂-oxy—the last being the unknown value. Therefore, the concentration of CO₂ in the bag must be individualized to the patient and thus found by trial and error. The method is therefore difficult to automate fully.

[0027] 4.1.2.2 In practice, since the characteristic of a suitable endpoint (the plateau of PCO₂) is subjective, identification of a suitable plateau is difficult to automate.

[0028] 4.1.2.3 The manoeuvre of rebreathing from a bag is difficult to perform in mechanically ventilated patients and is therefore not suitable for such patients.

[0029] 4.1.2.4 Inhaling 10-13% CO₂ is very uncomfortable and most people cannot tolerate it. It is particularly uncomfortable to someone who is short of breath or exercising.

[0030] 4.1.2.5 The method requires an external source of CO₂. This makes testing equipment bulky and awkward.

[0031] 4.1.2.6 The method requires that the subject hyperventilate in order to mix thoroughly the gas in the bag and the lungs before recirculation of blood takes place. This requirement limits the test to those subjects who can perform this manoeuvre and who can provide this degree of cooperation. This excludes patients who have severe lung disease, those who are too young, too confused or too ill to cooperate.

[0032] 4.1.2.7 The test loads a considerable volume of CO₂ into the subject's lungs and at the same time prevents CO₂ from leaving the blood for the duration of the test. This has negative consequences for the subject:

[0033] 4.1.2.7.1 Following the test, the subject must hyperventilate to eliminate the applied CO₂ load as well as the volume of metabolically-produced CO₂ not eliminated during the test. This may pose a considerable burden for some subjects with lung disease or exercising subjects who are already expending considerable effort to cope with their existing metabolic CO₂ load.

[0034] 4.1.2.7.2 A period of hyperventilation following the test is required to eliminate the CO₂. This may be difficult for some subjects to perform and, consequently, they may experience respiratory distress for some time until their PCO₂ is decreased.

[0035] 4.1.2.7.3 Repeated tests must be delayed until the extra CO₂ is eliminated and the baseline state re-established.

Dec. 26, 2013

[0036] 4.1.2.7.4 The test itself may distress the subject and alter the Q.

5. Rebreathing—Exponential Method

[0037] In this technique, a small amount of CO_2 is placed in a bag and the subject asked to rebreathe from the bag. The $PetCO_2$ s of successive breaths will rise exponentially towards $P\overline{\nu}CO_2$ -oxy. A rising exponential curve is then fit to the $PetCO_2$ s of these breaths to predict an asymptotic value that is assumed to be the $P\overline{\nu}CO_2$ -oxy (See FIG. 9).

[0038] 5.1 Advantages of the Exponential Method

[0039] 5.1.1 There is no requirement for respiratory manoeuvres by the patient.

[0040] 5.1.2 A smaller CO₂ load is placed on the subject in order to perform the test

[0041] 5.2 Limitations of the Exponential Method

[0042] 5.2.1 This is an indirect test in which the P $\overline{v}CO_2$ -oxy is not measured directly but calculated from data generated by a test.

[0043] 5.2.2 As the metabolic production of CO₂ is small compared to the size of the lung and bag, the rise of PCO₂ occurs over a prolonged period. This severely limits the number of useful data points for accurate extrapolation from an exponential curve, before recirculation.

[0044] 5.2.3 The most important limitation of this and other methods that use partial equilibration during rebreathing to extrapolate to an asymptote using a single exponential is that the assumptions underlying the method are incorrect. In fact, the method produces two different mathematical profiles: the one describing the washout of CO₂ from the lung into the bag is a decreasing exponential whereas the second describing the build-up of CO₂ released from the blood into the lungbag mixture is an increasing exponential (13). Only after the gases in the lung-bag system have become well mixed do the two exponentials resolve to a single exponential. By then, very few breaths (if any) that can provide suitable data for extrapolation from a single exponential can be taken before recirculation.

[0045] 5.2.4 A continually rising level of CO₂ makes this test unpleasant in conscious patients, especially in those exercising or very ill.

[0046] 5.2.5 The manoeuvre of rebreathing from a bag is difficult to perform in mechanically ventilated patients and is therefore not suitable for such patients.

[0047] 5.2.6 The method requires an external source of CO₂. This makes testing equipment bulky and awkward.

[0048] 5.2.7 The test loads a volume of CO₂ into the subject's lungs and at the same time prevents CO₂ from leaving the blood for the duration of the test. Although the extent of the CO₂ load on the subject is less than with the equilibration method, the negative consequences for the subject, outlined in the section on the equilibration method discussed above, must be considered.

[0049] 5.2.8 Priming the rebreathing bag with some CO₂ improves the predictive qualities of the asymptote since every data point lies closer to the asymptote, but the increased CO₂ concentrations increase the discomfort and the limitations approach those outlined above for the equilibration method.

6.0 Calculating Q without First Calculating PvCO₂-Oxy

[0050] Gedeon in 1980 described a method of calculating Q in ventilated patients via a differential Fick method that circumvents the need to calculate PvCO₂-oxy. The underlying assumptions of the method are that Q and PvCO2 will remain unchanged during a step change in lung CO₂ elimination and alveolar PCO₂ (PACO₂) lasting less than a recirculation time (about 30 seconds). Gedeon proposed reducing lung CO₂ elimination by reducing either the tidal volume or respiratory frequency setting of the ventilator. As a modification of this method, Orr et al. proposed leaving the ventilator settings unchanged and reducing lung CO2 elimination by temporarily interposing a dead space between the ventilator and the patient's airway resulting in a transient period of rebreathing previously exhaled gas.

6.1 Theoretical Basis of Gedeon/Orr Method:

[0051] The method applies to a subject being ventilated under control conditions in which CO₂ elimination and PetCO₂ are measured. A test manoeuvre consisting of a transient alteration in the CO₂ elimination for a time less than a recirculation time is effected and the resulting "equilibrium" PetCO₂ is noted. It is assumed that the Q and $PvCO_2$ -oxy during the test are unchanged from control conditions. The Fick equation for these two conditions can be written as

$$\dot{Q} = \frac{\dot{V}CO_2}{CvCO_2 - CaCO_2}$$
$$\dot{Q} = \frac{\dot{V}CO_2'}{CvCO_2 - CaCO_2'}$$

$$\dot{Q} = \frac{V CO_2}{C\overline{V}CO_2 - CaCO_2^2}$$

where VCO2' is the CO2 flux at the lungs during the test and ${\rm CaCO_2}$ ' is the corresponding 'new' arterial content of ${\rm CO_2}$. These two equations can be combined to yield the differential form of Fick's equation:

$$\dot{Q} = \frac{\Delta \dot{V} \text{CO}_2}{\Delta Ca \text{CO}_2}$$

where Δ denotes a "difference in". Since the PaCO₂ and P $\overline{v}CO_2$ -oxy lie on the same CO_2 dissociation curve, partial pressures of CO₂ can be substituted for CO₂ content to yield the following relation:

$$\dot{Q} = \frac{\Delta \dot{V} \text{CO}_2}{S * \Delta Pa \text{CO}_2}$$

where S is the slope of the CO₂ dissociation curve. Like the conventional non-invasive CO₂-based Fick method, the differential Fick method relies on predicting PaCO2 through measurements of PetCO2. However, instead of requiring a calculation of PvCO₂-oxy, the differential Fick equation assumes no change in PvCO2-oxy over the duration of the test, and uses the measured quantities VCO2 and VCO2 and well as PaCO2' and PaCO2' (from PETCO2) to calculate the remaining unknown value in the equation: Q.

6.2 Advantages of Gedeon/Orr Method

[0052] 6.2.1 The main advantage is that PvCO₂ does not need to be calculated.

[0053] 6.2.2 If the deadspace method is used to alter the VCO₂, then no change in breathing pattern is required.

[0054] 6.2.3 The method can, theoretically, be fully automated. (In its present commercial form, the size of the interposed deadspace must still be altered manually).

6.3 Limitations of Gedeon/Orr Method

[0055] There are a number of limitations in applying Orr's method to spontaneously ventilating subjects.

[0056] 6.3.1 In spontaneously breathing subjects, there is considerable breath-to-breath variation in breath size and breathing frequency resulting in a variation in PetCO₂. This poses problems with respect to:

[0057] 6.3.1.1 Identification of PetCO₂ and PetCO₂'. Long periods of baseline measurements are needed in order to average the end tidal values and identify the PetCO₂ to be used as the baseline PetCO₂ in the differential Fick equation. The test phase cannot last for more than about 30 seconds (due to recirculation), typically 5 breaths. This leaves little time to determine an accurate average PetCO₂'. During prolonged baseline periods of observation, the condition of the patient may change.

[0058] 6.3.1.2 Calculation of $\dot{V}CO_2$. The variations in PetCO₂ are related to variations in CO₂ elimination but the relationship is not consistently reflected by the PetCO₂. For example, assuming a subject breathing at rest with an average resting breath size, an interposed smaller breath may result in a lower PetCO₂ (due to a smaller contribution of alveolar gas to the end tidal sample) but the CO₂ elimination from that breath will be diminished. Conversely, a larger breath may result in the same PetCO₂ as the resting breath but a greater volume of CO₂ is eliminated. The commercial automated Gedeon method (NICO2, Novametrics Medical Systems, Wallingford, Conn., U.S.A.) measures the CO₂ eliminated breath-by-breath and therefore must continuously average the values to measure VCO₂. The NICO2 method of calculating VCO₂ by realtime integration of continuous measurements of flow (with a pneumotachymeter) and CO₂ concentration (with a capnograph) is fraught with potential for errors: a small error in the integration of these two signals with different time delays and time constants results in a much larger error in the calculation of VCO₂. In addition, the greater the variability of the breath size and CO2 concentrations, the longer the measurement time required for an accurate estimate of $\dot{V}CO_2$.

[0059] 6.3.2 Calculation of VCO2'. Stable transient changes in VCO₂ cannot be achieved in conscious spontaneously ventilating patients:

[0060] 6.3.2.1 Interposing a deadspace and raising their PCO₂ will stimulate spontaneously breathing conscious subjects to increase their VE and VCO₂ until the PetCO₂ is restored.

[0061] 6.3.2.2 Any change in breath size or frequency during a period of breathing, (a normal occurrence in spontaneously breathing people) changes the VCO₂ during that period. During inspiration, the deadspace gas is inhaled first followed by fresh gas. A decrease in a breath size or frequency diminishes the volume of fresh gas inhaled (and

thus the $\dot{V}CO_2$ for that breath). An increase in breath size or frequency will result in an increased volume of fresh gas delivered to the alveoli.

[0062] 6.3.2.3 Each breath is an independent event and there is no inherent method to compensate in a subsequent breath for changes in VCO₂ in the preceding breath. For the method to be implemented, therefore, measures must be taken to ensure that breath size and frequency stay absolutely constant during the test. The NICO2 method has no such built-in aspects. The method can therefore be used only in patients who have precisely uniform breathing pattern such as those that are paralysed and mechanically ventilated.

[0063] 6.3.3 Identification of PetCO₂—PaCO₂ gradient. The Gedeon and Orr methods assume, or require the establishment of, a constant gradient between the PetCO₂ and the PaCO₂. The variation in PetCO₂ is due to variations of distribution of fresh gas to various parts of the lung and any one breath does not reflect the overall state of CO₂ exchange. On the other hand, such variations are not reflected in the PaCO₂ which does reflect the overall exchange of CO₂ and remains relatively constant. Therefore, variations in PetCO₂ also confound the quantification of the PetCO₂—PaCO₂ gradient under control conditions. Although Orr provides a number of equations to correct for these limitations, these equations are empirical and do not necessarily apply to a particular patient. For example, they are applied whether or not there is irregular breathing.

[0064] The PetCO₂—PaCO₂ gradient during the test phase when rebreathing occurs is unknown. In the presence of large alveolar deadspace (as commonly occurs in many ill patients) the PetCO₂—PaCO₂ gradient will change during the rebreathing phase. Orr provides some equations to correct for this but since the volume of the alveolar deadspace is unknown, the applicability of the formula to any particular patient is unknown. This further diminishes the accuracy of calculating PaCO₂'.

[0065] The manoeuvres required to determine each of the terms required to calculate Q (VCO₂, VCO₂', PETCO₂, PETCO₂' and PaCO₂') by the Orr/Gedeon NICO2 method is awkward to implement and prone to errors in measurement in the presence of any variation in breath amplitude or breathing frequency as occurs in spontaneously breathing humans or animals.

[0066] 6.3.4 The parameter calculated by the differential Fick method as practiced by Gedeon/Orr/Respironics is pulmonary blood flow (Qp). Pulmonary blood flow may be less than the total cardiac output (Qt) when, for example, some of the Q is shunted from the right side of the circulation (superior vena cava, right atrium, right ventricle, pulmonary artery) into the left side of the circulation without passing through the lungs. This is referred to as "shunt" (Qs). About 5% of venous blood bypasses the lungs (termed shunted blood) in healthy adults. Much larger shunts occur in many medical conditions such as congenital heart disease, surgical repair of some congenital heart diseases, pneumonia, pulmonary edema, asthma, pulmonary atelectasis, adult respiratory distress syndrome, obesity, pregnancy, liver disease and others. The differential Fick method does not include shunted blood in the calculation of Q and other empiric corrections must be made to account for it.

7.0 Kim-Rahn Farhi method [0067] 7.1 Theory:

[0068] A unique maneuver was proposed by Kim, Rahn and Farhi, (J. Appl. Physiol. 21:1388-44. 1966.) as a way to calculate the oxygenated mixed venous PCO₂ (PvCO₂-oxy) as well as the true PvCO₂ and PaCO₂. It is based on a paradigm of taking a breath of O2, holding the breath, and exhaling slowly over a period equal to the recirculation time. Over this time of exhalation, the CO₂ from the mixed venous blood will diffuse into the alveoli and O2 will be absorbed. The low PO₂ in the red blood cells in the mixed venous blood maximizes the volume of CO₂ that can be carried by hemoglobin. Oxygen from the alveoli diffuses into the red blood cells, raising the PO₂ and decreasing the affinity of hemoglobin for CO₂ (Haldane effect). This releases CO₂ from the binding sites on the hemoglobin, making it available for diffusion into the alveoli. With breath holding, CO₂ will accumulate in the alveoli and the alveolar PCO₂ (PACO₂) will rise until it no longer provides a gradient for diffusion from the blood. (This PCO₂ is known as the oxygenated mixed venous PCO₂ (P vCO₂-oxy).) However, O₂ will continue to diffuse as long as the PAO₂ is greater than PvO₂. Relatively little CO₂ need diffuse into the alveoli to reach PvCO2-oxy compared to the volume of O₂ that is available for uptake before the PO₂ in the pulmonary capillary blood is in equilibrium with the PAO₂. In other words, the equilibration of CO₂ in the alveoli with the mixed venous blood will occur well before that of O_2 .

[0069] Since both $\rm O_2$ and $\rm CO_2$ are contained in the same physical volume, the changes in concentrations of each gas over a short period will reflect the rates of flux of that gas over the same period. Therefore, over a short period, the ratio of $\rm PCO_2$ to $\rm PO_2$ will reflect the respiratory quotient, RQ (defined as the rate of $\rm CO_2$ diffusion from the blood into the alveoli divided by the rate of $\rm O_2$ absorption into the blood from the alveoli). The RQ will initially be highest at the beginning of the breath when the rate of $\rm CO_2$ diffusion into the alveoli is maximal, and will approach 0 when the alveolar $\rm PCO_2$ equals $\rm P\overline{v}CO_2$ -oxy. In vitro studies have shown that $\rm PaCO_2$ equals the true $\rm P\overline{v}CO_2$ when the RQ=0.32 and equals $\rm PaCO_2$ when RQ is equal to the patient's steady state RQ (typically ~0.8).

[0071] The method suggested for performing this test would require a subject to take a maximum breath of 100% $\rm O_2$ and exhale very slowly and maximally. Over the course of this exhalation, expired gas is sampled and analyzed continuously for both $\rm PO_2$ and $\rm PCO_2$. $\rm PO_2$ is graphed vs. $\rm PCO_2$ and the RQ is calculated from the instantaneous slope of tangents to the curves at various $\rm PCO_2$ values as follows:

$$RQ = \frac{\text{slope} - (\text{FeO}_2 * \text{slope}) - \text{FeCO}_2}{1 - (\text{FeO}_2 * \text{slope}) - \text{FeCO}_2}$$

[0072] These RQ values are then plotted against their respective PCO_2 data points resulting in a linear relation as illustrated in FIGS. 4 and 5 of T. S. Kin, H. Rahn, and L. E. Farhi cited above.

[0073] 7.3 Advantages of the Method.

[0074] 7.3.1 This is the only known non-invasive method by which true $P\overline{v}CO_2$ can be calculated.

[0075] 7.3.2 The method provides an estimate of $PaCO_2$ not based on assuming a gradient between $PetCO_2$ and $PaCO_2$.

[0076] 7.3.3 Data generated by the method can be used to calculate the O₂ saturation of mixed venous blood.

[0077] 7.4 Limitations of the Kim-Rahn-Farhi Breath-Hold Method

[0078] The main limitation of this method is that it requires the subject to have a large lung capacity, hold his breath, and exhale over a prolonged duration. Patients with conditions such as pulmonary fibrosis, pneumonia, adult respiratory distress syndrome, chronic obstructive lung disease, asthma, obesity, trauma, abdominal and chest surgery, mental obtundation, confusion, pregnancy and many others have marked limitations in their ability to take a large breath. Patients are required to cooperate with their duration of breath holding and rate of exhalation. Many patients who are ill, exercising subjects, children and others are unable to perform this satisfactorily. This method is very awkward to automate or perform on ventilated patients.

8.0 Fisher Method

[0079] 8.1 Theory

[0080] In a steady state, if a subject breathes in a PCO₂ equal to PvCO2-oxy, there will be no gradient for gas exchange and the difference in PCO2 between the inspired PCO₂ (P₁CO₂) and the expired PCO₂ (P₂CO₂) will be 0. The volume of CO₂ diffusing into the alveoli will be maximal when the difference between PiCO2 and PECO2 is greatest, i.e., when the PiCO₂ is 0. Since the change in alveolar PCO₂ (PACO₂) varies directly as the volume of CO₂ diffusing into the alveoli and the volume diffusing into the alveoli varies directly as the gradient, then the difference between the PiCO₂ and PECO₂ will vary inversely as PiCO₂. In other words, graphing the difference between the PECO₂ and PiCO₂ (PECO₂-PiCO₂) vs. FICO₂ will result in a straight line. Since subjects normally breathe room air (PiCO₂ equals 0 or O₂, the control PetCO₂ provides the first point on the graph. When subjects inhale gas with any constant value of PCO₂, the PetCO₂ at the end of an equilibration period not exceeding the time for recirculation will provide a second data point which can be used to define the straight line which crosses the X axis where PiCO₂ equals PvCO₂-oxy.

[0081] 8.2 Test Method:

[0082] The subject breathes via a non-rebreathing valve. The inspiratory limb is provided with either fresh gas or test gas with any PCO₂. To perform a test, the inspired gas is switched from control gas to test gas for about one recirculation time. The PiCO₂ of the test gas, the PetCO₂ just before the test (when PiCO₂ was 0), and the PetCO₂ of the last breath before recirculation are used to calculate the $P\overline{\nu}CO_2$ -oxy.

[0083] 8.3 Advantages of the Prior Disclosed Previous Fisher Method:

- [0084] 8.3.1 Any low inspired concentration of CO₂ such as 1% is adequate to generate a data point; therefore the subject need not get a large CO₂ load.
- [0085] 8.3.2 This Fisher method extrapolates to the P $\overline{v}CO_2$ -oxy from a linear function and is therefore easier to calculate and more accurate than with the partial rebreathing test in which data points are fit to an exponential curve for extrapolation to an asymptote.
- [0086] 8.3.3 The PrCO₂ can be any value, so accurate mixtures of gases are not required.
- [0087] 8.3.4 Assuming arterial PCO₂ values (PaCO₂) can be obtained from arterial blood sample, for example, the method measures total Q, not just pulmonary blood flow.

- [0088] 8.3.5 The subject need not carry out any respiratory manoeuvre such as breath holding or hyperventilation.
- [0089] 8.3.6 The method does not entail any rebreathing. Therefore, O₂ levels remain stable throughout the test and supplemental O₂ is not needed.

8.4 Limitations of the Fisher Method.

- [0090] 8.4.1 Uniform breath size cannot be guaranteed in spontaneously breathing subjects. A change of breath size or breathing frequency during the latter parts of the test phase will affect the PetCO₂ and thus the calculation of PvCO₂-oxy. Furthermore, as the subjects are inhaling gas that contains CO₂, they may be stimulated to take larger or more frequent breaths.
- [0091] 8.4.2 The test requires an external source of CO₂. This must be supplied via a tank of CO₂ and a gas blender or via a tank of pre-mixed gas. If more than one test gas is required, then arrangements to blend additional gases must be made or more than one additional gas tank is required. This is inconvenient, costly, and adds complexity to the test method and additional bulk and weight to the test apparatus.
- [0092] 8.4.3 It is very complex to configure an automated system that works for both spontaneously breathing and mechanically ventilated patients.
- [0093] 8.4.4 There is no simple method to adapt currently available ventilators, anaesthetic machines or breathing circuits to provide a known and constant PtCO₂ for a fixed number of breaths.
- [0094] 8.4.5 The technique is difficult to adapt to anaesthetized patients breathing via a circle circuit in which both the test gas and the anaesthetic gases enter the circuit, especially in the presence of a CO₂ absorber removing CO₂ from the circuit.

OBJECT OF THE INVENTION

[0095] It is therefore a primary object of this invention to provide an improved method and apparatus for the purpose of non-invasively determining cardiac output (\dot{Q}) which may be utilized in ventilated subjects, subjects who breath spontaneously or subjects who are under controlled ventilation such as those undergoing surgical procedures under general anesthesia

[0096] It is yet a further object of this invention to provide an improved method and the apparatus related thereto for the purposes of non-invasively determining alveolar ventilation $(\dot{V}A)$ and calculating minute CO_2 production $(\dot{V}CO_2)$, oxygenated mixed venous PCO_2 ($P\overline{V}CO_2$ -oxy), true mixed venous PCO_2 (true $P\overline{V}CO_2$), pulmonary shunt, anatomical dead space, arterial PCO_2 , at a greater accuracy than prior known non-invasive methods and apparatuses would provide. [0097] It is yet another object of the invention to provide a

[0097] It is yet another object of the invention to provide a method of non-invasively calculating the oxygen saturation of mixed venous blood $(S\overline{\nu}O_2)$ which may be utilized to reveal heart failure of septic shock in a patient or the like.

[0098] It is yet a further object of this invention to provide an improved method and the apparatus related thereto for the purposes of determining \dot{Q} , \dot{V}_A and calculating $\dot{V}CO_2$, P $\overline{V}CO_2$ -oxy, true $P\overline{V}CO_2$, pulmonary shunt, anatomical dead space in a non-invasive and fully automated manner.

[0099] Further and other objects of the invention will become apparent to those skilled in the art when considering

the following summary of the invention and the more detailed description of the preferred embodiments illustrated herein.

SUMMARY OF THE INVENTION

[0100] This invention discloses a method and apparatus for calculating all of the \dot{Q} regardless of shunt, calculating the shunt, anatomical and alveolar deadspace, true mixed venous O_2 saturation, true mixed venous PCO_2 , and $PaCO_2$. Furthermore the method and apparatus can be used with ventilated subjects, subjects breathing spontaneously, even with marked variations in their tidal volume and breathing frequency, or subjects undergoing surgery under anaesthesia. Subjects need not perform any respiratory manoeuvre such as hyperventilation or breath holding.

[0101] According to one aspect of the invention there is provided an improved method and apparatus for the purposes of determining \dot{Q} and \dot{V}_A and calculating $\dot{V}CO_2$, $P\bar{v}CO_2$ -oxy, true $P\bar{v}CO_2$, PaCO₂, pulmonary shunt, and anatomical dead space which increases the accuracy of these determinations in relation to known methods and apparatus and allows the full automation of the various methods disclosed herein for these determinations and calculations

[0102] The new method:

- [0103] 1. is insensitive to changes in minute ventilation (VE) tidal volume and/or respiratory frequency so that the method can be carried out in spontaneously breathing subjects;
- [0104] 2. is simplified and less expensive to construct compared to other non-invasive automated methods of performing the differential Fick technique in that
 - [0105] a. it does not necessarily require any mechanically activated valves to be actively engaged in the patient circuit
 - [0106] b. does not require a pneumotachygraph to measure flows
 - [0107] c. does not require manual adjustment of an interposed dead space (and thus can be totally automated);
 - [0108] d. The device will be the same for all sizes of adults (one size fits all)
- [0109] 3. is compatible with a number of sequential gas delivery breathing (SGDB) circuits. A SGDB circuit provides for the sequential delivery of two gas sets to the lungs during inhalation. A gas set is composed of one or more gases and vapors. The first gas set (FGS) is provided from the beginning of inhalation and can terminate at some time during inhalation depending on the FGS flow and the VE, at which time inhalation continues with the delivery of the second gas set (SGS). For the purposes of measuring Q and the other physiologic parameters described herein, it is preferred that there is a distinct transition from FGS to SGS and there is no mixing of the gas sets. A small degree of mixing of FGS with SGS during the latter part of inhalation will reduce accuracy of the measured and calculated results. Mathematical corrections can be made to minimize effect of the mixing of FGS with SGS, but cannot completely negate the effects in all circumstances. Therefore, breathing circuits which separate the FGS from the SGS are preferred.
- [0110] 4. the generation and presentation of data will be substantially the same for controlled (mechanical) ven-

- tilation and rebreathing so that the algorithms to perform the tests and analyze the data can be substantially the same;
- [0111] 5. can institute an equilibrium steady state within one recirculation time so that the value for PetCO₂ will be a true measured value rather than one requiring multiple corrections based on unsubstantiated assumptions;
- [0112] 6. will allow the measurement of a new steady state PetCO₂ within one recirculation time and thus actualize the assumption underlying the Differential Fick approach that PvCO₂ is unchanged;
- [0113] 7. will minimize the effect of changes in tidal volume on the alveolar ventilation.
- [0114] 8. maintain the alveolar PO₂ while making pulmonary blood flow measurements;
- [0115] 9. make all calculations without a requirement to measure breath-by-breath volumes of inspired and expired CO₂ or any flows of tidal gases.
- **[0116]** According to one aspect of the invention there is provided an improved apparatus and method of identifying the alveolar ventilation (\dot{V}_A), substantially as illustrated and described herein, preferably the \dot{V}_A so determined is utilized to calculate the \dot{V}_CO_2 as $\dot{V}_A\times FetCO_2$, where $FetCO_2$ is the fractional pressure of CO_2 in end tidal gas.
- [0117] In one embodiment of the improved apparatus and method:
 - [0118] a) the Fisher approach is used to determine P $\overline{v}CO_2$ -oxy (or)
 - [0119] b) the Kim Rahn Farhi approach is used to determine
 - [0120] i) $P\overline{v}CO_2$ -oxy
 - [0121] ii) true PvCO₂
 - [0122] iii) PaCO₂
 - [0123] iv) true PvCO₂ plus the information from a pulse oximeter to determine mixed venous hemoglobin O₂ saturation (or)
 - [0124] c) the differential CO₂ Fick technique of Gedeon and Orr is utilized to determine any combination of
 - [0125] i) PvCO₂-oxy
 - [0126] ii) Q
 - [0127] iii) VCO₂
 - [0128] iv) VCO₂'
 - [0129] v) PetCO₂—PaCO₂ gradient determined using the PaCO₂ as determined by the Kim Rahn Farhi method from data collected while reducing the VCO₂ in order to perform the Differential Fick method. (or)
 - [0130] d) Q is determined via the Kim Rahn Farhi method performed during partial rebreathing using a CO₂ Fick method where the
 - [0131] i) $\dot{V}CO_2$ is calculated with or without the new method as disclosed
 - [0132] CaCO₂ and CvCO₂ are determined from the PaCO₂ and PvCO₂ respectively derived by the Kim Rahn Farhi method; (or)
 - [0133] e) calculation of the respiratory quotient (RQ) is determined as PetCO₂/(PtO₂—PeO₂); (or)
 - [0134] f) PaCO₂ is determined directly via analysis of arterial blood sample, arterialized venous sample, transcutaneous PCO₂ electrode, or other methods known to those skilled in the art.
 - [0135] wherein said apparatus or method may be utilized for very accurate non-invasive determination of Q and the other indicated parameters.

[0136] According to yet another aspect of the invention there is provided an improved method of apparatus for determining $\dot{V}A$, $\dot{V}CO_2$ and calculating \dot{Q} , $P\bar{v}CO_2$ -oxy, true $P\bar{v}CO_2$, PaCO₂, pulmonary shunt, anatomical dead space, and O_2 saturation in mixed venous blood; which increases the accuracy of these determinations and calculations in relation to known methods and apparatuses and allows for full automation thereof if necessary by using automated means well known to those skilled in the art, to:

[0137] i) induce a step change in VCO₂ by providing a step change in FGS flow to a SGDB circuit to create, with the control data at rest, two sets of data for said determination utilizing the differential Fick equations; (or)

[0138] ii) change the partial pressure of CO₂ in FGS of a SGDB circuit to create, with the control data at rest, two sets of data for said determination utilizing the Fisher or the differential Fick equations; (or)

[0139] iii) change FGS flow or change the partial pressure of CO_2 in FGS in a SGDB circuit to simulate complete or partial breath holding and utilizing the Kim-Rahn-Farhi technique, wherein the $\mathrm{P}_{ET}\mathrm{CO}_2$ of each breath is equivalent to a sequential alveolar sample;

thereby providing more relevant data to calculate desired parameters.

[0140] In yet another embodiment of the invention a ventilation circuit and method is provided for using sequential delivery of gas sets in order to identify the minute volume of gas entering the anatomical dead space and the minute volume entering the alveoli and thereby available for gas exchange (\dot{V}_A). Subsequently, setting FGS flow to substantially equal to or less than \dot{V}_A substantially controls \dot{V}_A . A step reduction in \dot{V}_A can then be induced by a step reduction in FGS flow, and resultant effects on end tidal gases such as CO_2 can be used in the to calculate \dot{Q} and other parameters as previously set out herein in the Background, disclosures and figures.

[0141] In yet another embodiment there is provided a method and apparatus of determining Q and the other parameters disclosed by utilizing any SGDB circuit for example, the circuits described and illustrated herein by reducing the FGS flow to said circuit or increasing the PCO₂ of FGS to said circuit, independent of the breathing rate thereby allowing for calculations to be made via Differential Fick equations, and/or Fisher method, and/or the Kim-Rahn-Farhi method.

[0142] Preferably the method or apparatus previously described wherein the CO_2 content as calculated from P $\overline{v}CO_2$ -oxy and true $P\overline{v}CO_2$, may be utilized to determine the O_2 saturation of mixed venous blood with known relations between CO_2 content, O_2 saturation and PCO_2 .

[0143] In one embodiment the method or apparatus disclosed may be utilized wherein the arterial O₂ hemoglobin saturation, as determined by a non-invasive pulse oximeter, which makes the measurement by shining infrared light through a finger, is utilized with the O₂ saturation value in the pulmonary artery as calculated by the Kim Rahn Farhi method, to calculate the fraction of shunted blood (assuming fully oxygenated blood in the end pulmonary capillary) thereof.

[0144] Preferably said method or apparatus is utilized to determine the fraction of shunted blood \dot{Q}_S , which in conjunction with determination of total cardiac output \dot{Q}_T (utilizing PaCO₂ as determined by the Kim Rahn Farhi method, or available from analysis or arterial blood or determined by transcutaneous PCO₂ determination or otherwise known to those skilled in the art, as a term in the Fick equation) and

pulmonary blood flow $\dot{Q}p$ (utilizing PetCO₂ in the Fick equation) may be used to determine $\dot{Q}s$ the pulmonary output via the relationship.

$$\dot{Q}_S = \dot{Q}_T - \dot{Q}_P$$

[0145] Preferably the method or apparatus disclosed wherein the O₂ saturation of haemoglobin in mixed venous blood (SaO₂), as determined therewith, is utilized to reveal a condition in a patient such as septic shock, or heart failure.

BRIEF DESCRIPTION OF THE FIGURES

[0146] FIG. 8: PCO₂ vs time tracing during a rebreathing equilibrium test for determining oxygenated mixed venous PCO₂

[014 $\tilde{7}$] FIG. 9: PCO₂ vs. time tracing during exponential method of finding oxygenated mixed venous PCO₂.

[0148] FIG. 2 is a SGDB Circuit as taught by Fisher in U.S. Pat. No. 6,622,725 referred to herein as the Fisher circuit

[0149] FIG. 3 is similar to FIG. 2 wherein the reservoir bags are remote from the patient.

[0150] FIG. 5 is a new circuit for use with spontaneous ventilation.

[0151] FIG. 3B is similar to FIG. 5 wherein bypass limb, bypass valve, and passive expiratory valve are replaced by an active expiratory valve.

[0152] FIG. 3D is similar to FIG. 2 wherein an active valve has been added to the inspiratory limb to prevent mixing of FGS with SGS during inhalation.

[0153] FIG. 5A is similar to FIG. 5 wherein an active valve has been added to the inspiratory limb to prevent mixing of FGS with SGS during inhalation.

[0154] FIG. 3E is similar to FIG. 2 wherein an active valve has replaced the passive inspiratory valve.

[0155] FIG. 5B is similar to FIG. 5 wherein an active valve has replaced the passive inspiratory valve.

[0156] FIG. 3C is similar to FIG. 3B wherein an active valve has replaced the passive inspiratory valve.

[0157] FIG. 4 shows a modification of any of the circuits shown in FIGS. 2, 3-3E, 5-5B for use with a mechanically ventilated patient.

[0158] FIG. 4B shows the preferred embodiment modified for use on ventilated patients.

[0159] FIG. 6 is a modification of the above circuits to include co-axially arranged inspiratory and expiratory limbs between the valves and the patient

[0160] FIG. 6A shows the preferred embodiment of the cardiac output circuit where inspiratory and expiratory limbs are co-axially arranged with the circuit of FIG. 5A.

[0161] FIG. 7 is a new circuit designed to allow measurement of cardiac output while delivering anesthetics or removing volatile agents from a patient.

[0162] FIG. 5C shows a detail of a circuit design where the passive valves are surrounded by the exhaled gas reservoir

[0163] FIG. 10: Apparatus for non-invasive cardiac output apparatus consisting of a breathing circuit, gas sources, gas flow controllers, gas concentration sensors, and microprocessor capable of receiving and storing analog and digital input from sensors and operators, storing and following a decision tree, and generating output signals to a computer screen and to flow controllers.

[0164] FIG. **11** Flow diagram describing automated sequence of events performed by the non-invasive cardiac output apparatus in order to automatically generate and record data non-invasively and calculate Q and other physiologic parameters.

[0165] FIG. 12 is a schematic of a standard anesthetic circle system herein provided as reference for discussion of dis-

closed system. Gas entering the anesthetic circuit consisting of oxygen, with the possible addition of air and/or nitrous oxide (N₂O), and possibly an anesthetic vapor such as isoflurane, desflurane or sevoflurane enters the fresh gas port (6) at a constant and known flow. The gas concentrations entering the circuit are set by the anesthesiologist. The patient inspires through the patient port (1) and draws fresh gas plus gas drawn from the gas reservoir bag (4) through the CO₂ absorber (5) up the inspiratory limb (8). During exhalation, the inspiratory valve (7) closes and the fresh gas passes through the CO₂ absorber (5) towards the gas reservoir bag. Expired gas flows down the expiratory limb (2) displacing gas into the gas reservoir bag (4). When the reservoir bag is full, the pressure in the circuit rises, opening the APL (airway presslure relief) valve (9), and the rest of the expired gas exits the circuit through the APL valve. Gas is sampled continuously at the patient port and is analyzed for concentrations of constituent gases. The inspiratory (2) and expiratory (8) limbs consist of tubing (T).

[0166] FIG. 13 A detail of the computer screen output of an automated analysis of test finding \dot{V}_A by progressive reduction in SGF flow method in a subject is illustrated in FIG. 13. The figure illustrates that progressive reduction of SGF (labelled "FGF" in the figure) results in a distinct inflection point when either PetCO₂ or PetO₂ is graphed as a function of SGF.

DETAILED DESCRIPTION OF THE INVENTION

Detailed Description of the Apparatus

[0167] Referring now to Figure ??, an apparatus is shown with the following components:

- [0168] 1) a breathing circuit (202), said breathing circuit preferably has the characteristic that, on exhalation, exhaled gas is kept separate from inhaled gas and on inhalation, when VE is greater than the flow of a first gas set (FGS) into the circuit, the subject inhales FGS gas first and then inhales a second gas set (SGS) gas, preferably said SGS containing CO₂ and where SGS may be mostly previously exhaled gas. Any SGDB circuit can be used to greater or lesser benefit, according to its characteristics. We provide below detailed descriptions of several alternate configurations and outline their particular advantages and drawbacks with respect to measuring Q and related parameters outlined above.
- [0169] 2) a gas sample line (204.1) leading to a gas analyzer (204) that monitors the concentration of gases, for example CO₂, O₂, at the patient-circuit interface and outputs preferably an electric signal corresponding to the concentrations (204.2) (for example if the gases of interest are O₂ and CO₂, the "#17500 O₂ and CO₂ analyzer set" (Vacumed, Ventura Calif., USA))
- [0170] 3) a precise gas flow controller (200), preferably one that can control the flow of one or more pressurized gases (such as oxygen, air, CO₂) singly or in combination, and that can be set manually or via an automated system such as via machine intelligence (for example, the Voltek gas flow controller by Voltek Enterprises, Toronto, Canada);
- [0171] 4) a source of FGS (201), preferably containing O₂ and/or air with or without CO₂;
- [0172] 5) means (205) to identify phase of breathing, for example using electronic pressure sensors with tubing to sample pressures at the patient-circuit interface (205.1) or in other locations in the circuit and generating electrical signal corresponding to the sensed pressures. Such means will provide electrical signal (205.2). Phase of

- breathing can also be determined from analysis of gas sensor output by machine intelligence.
- [0173] 6) a computer or machine intelligence (207) which records, stores, analyzes signals from gas analyzer (204) and pressure transducer (if present), contains a predetermined set of instructions regarding the analysis of data such as calculation of Q and physiologic parameters, determination of phase of respiration, display of information on a computer screen, and control of gas flow controller (200) including the timing, sequence and flow of gas.
- [0174] 7) wherein said device may be utilized for non-invasive measurement and determination of Q and other parameters such as VA, VCO₂, PvCO₂-oxy, true PvCO₂, PaCO₂, pulmonary shunt, and anatomical dead space

Detailed Description of Breathing Circuits

[0175] FIG. 5 shows a breathing circuit which provides sequential delivery of the FGS followed by the SGS when \dot{V}_E exceeds FGSF, with the manifold containing the valves and the FGS reservoir bag and the expiratory gas reservoir bag remote from the patient. This improvement reduces the bulk of the patient manifold, and eliminates the possibility of the SGS mixing with the FGS due to vigorous exhalation.

[0176] Referring to FIG. 5, Patient (38) breathes via a Y connector (40). Valve (31) is an inspiratory valve and valve (33) is an expiratory valve. Valve (35) is a bypass valve in the bypass limb (34) that bypasses the expiratory valve (33) and has an opening pressure greater than inspiratory valve (31). Valves (35, 33) may be close to or distal from the patient manifold as desired, as long as they are on the expiratory limb (39). However, in the preferred embodiment, they are distal to the patient to reduce the bulk of the patient manifold. Inspiratory valve (31) may be close to, or distal from, the patient manifold as desired, as long as it is on the inspiratory limb (32). In the preferred embodiment, it is distal to the patient as well. FGS enters the circuit via port (30).

Function:

[0177] During exhalation, increased pressure in the circuit closes inspiratory valve (31) and bypass valve (35). Gas is directed into the exhalation limb (39), past one-way valve (33) into the expiratory gas reservoir bag (36). Excess gas is vented via port (41) in expiratory gas reservoir bag (36). FGS enters via port (30) and fills FGS reservoir (37). During inhalation, inhalation valve (31) opens and FGS from the FGS reservoir (37) and FGS port (30) enter the inspiratory limb (32) and are delivered to the patient. If FGSF is less than VE, the FGS reservoir (37) empties before the end of the breath, and continued respiratory effort results in a further reduction in pressure in the circuit. When the opening pressure of the bypass valve (35) is reached, it opens and gas from the expiratory gas reservoir (36) passes into the expiratory limb (39) and makes up the balance of the breath with SGS.

[0178] Thus when FGSF is less than \dot{V}_E , the subject inhales FGS, then SGS, and no contamination of FGS occurs.

[0179] FIG. 3B shows an alternate embodiment of the circuit illustrated in FIG. 5 where the passive expiratory valve (33) and expiratory bypass limb (34), and expiratory limb bypass valve (35) are replaced with a control valve that is triggered by the collapse of the inspiratory reservoir. Referring to FIG. 3B, a control valve (401) is placed in the expiratory limb (16) anywhere along its length between the patient port (10) and the expiratory reservoir bag (18). When the patient's V_E exceeds the FGSF during inspiration the reservoir bag (20) collapses. This is detected by pressure sensing

means (405) through port (406) as an acute reduction in pressure. Pressure sensing means (405) could be an electronic pressure transducer capable of detecting changes 2 cm H₂O pressure, for example. Immediately afterwards, valve (401) is then opened by control means (403), which could be an electronic signal for activating a solenoid valve, for example, leading to depressurization and collapse of a balloon valve, as is known to those skilled in the art, resulting in SGS is being inhaled for the balance of inhalation. During exhalation, patient exhales through expiratory tube (16) past valve (401) into the SGS reservoir (18). At end of exhalation, as detected by pressure sensing means (405) as a reduction of pressure, valve (401) is closed by control means (403), which could be an electronic signal for toggling a solenoid valve, for example, leading to pressurization and inflation of a balloon valve, as is known to those skilled in the art.

[0180] While the circuits of FIG. 5 and FIG. 3B present the advantages over the Fisher circuit of reducing the bulk of the patient manifold, and eliminating the possibility of the SGS mixing with the FGS due to vigorous exhalation, they still have the following drawback: When FGS reservoir (20, 37) is emptied and the patient is breathing SGS for the balance of an inspiration, the circuit does not deliver SGS alone but a mixture of SGS and FGS. The FGS continues to flow into the circuit and is drawn by inhalation past one-way inspiratory valve (31,3) and allows FGS gas to be inhaled from the inspiratory limb (32,14). To optimize the generation of data required to measure of cardiac output, it is necessary to redirect the FGS into the FGS reservoir (37,20) for the balance of inhalation after the initial collapse of the FGS reservoir. This would prevent mixing of FGS with SGS during the period of inhalation where the patient breathes SGS. This limitation of circuits illustrated in FIGS. 5 and 3B with respect to measuring cardiac output are shared with the Fisher circuit.

[0181] FIG. 3D shows an improved circuit that prevents contamination of the SGS by FGS when SGS is being delivered to the patient. Referring to FIG. 3D, FGS control valve (400) is added to the inspiratory limb (14) at some point between the FGS port (12) and the inspiratory valve (11). Pop-off valve (425) is connected to the inspiratory limb on the side of the FGS control valve (400) that is proximal to the inspiratory reservoir bag (425). During exhalation, gas passes from the patient port (10), through the expiratory one-way check valve (15) down the expiratory limb (16) into the expiratory reservoir bag (18). Excess gas exits the expiratory reservoir bag (18) at the opening (19) remote from the entrance. FGS enters the circuit at a constant flow via a fresh gas port (12). As the inspiratory one-way check valve (11) is closed during exhalation, the fresh gas accumulates in the fresh gas reservoir bag (20). During inhalation, FGS entering from the port (12) and the FOS reservoir (20) passes through the inspiratory valve (11) and enters the patient. If the FGSF is less than VE, the FGS reservoir bag (20) collapses, as detected by pressure sensing means (405) connected to pressure sensing port (406). FGS control valve (400) is closed via valve control means (403), and valve (17) in the bypass limb (13) opens, directing previously exhaled gas to the patient. When the FGS control valve (400) is closed, any FGSF entering the circuit during the balance of inspiration is directed only to the FGS reservoir bag (20) and not to the patient, who is receiving SGS for the balance of inspiration. FGS control valve (400) may be re-opened any time from the beginning of expiration to just before the next inspiration. FGS control valve (400) may be any type of valve, and is preferably an active valve such as a balloon valve, known to those skilled in the art, that can be controlled by automated means. The popoff valve (425) opens when the reservoir bag (20) is full to prevent the reservoir bag (20) from overfilling.

[0182] The circuit illustrated in FIG. 5A is similar to that in FIG. 5 but has the addition of a FGS control valve (400), together with pressure sensing means (405) and port (406), and valve control means (403), added to the inspiratory limb of the circuit (32) distal to the one-way inspiratory valve (31) and proximal to the FGS inflow port (30). Similarly, a FGS control valve, together with pressure sensing means and port, and valve control means, may be added to the inspiratory limb (14) of the circuit illustrated in FIG. 3B positioned distal to the one-way inspiratory valve (31) and proximal to the FOS inflow port (12) to achieve the same result, namely, prevention of contamination of SGS by FGS when VE exceeds FGSF and the FGSF reservoir bag is emptied.

[0183] We present two additional circuits that are configured by adding FGS control valve (400) together with pressure sensing means (405) and port (406), and valve control means (403), to the Fisher circuit and the circuit illustrated in FIG. 5 and removing the passive one way inspiratory valve (11, 31), as shown in FIGS. 3E and 5B respectively. These circuits function identically to those illustrated in FIGS. 3D and 5A with respect to complete separation of FGS and SGS during inhalation. In such a circuit, during inspiration, FGS control valve (400) is open until FGSF reservoir bag (20,37) is emptied, then it is closed so that any additional FGSF entering the circuit during the balance of inspiration is directed only to the reservoir bag (20) and not to the patient. As the patient continues to inspire, bypass valve (17,35) opens allowing the patient to inhale SGS for the balance of inspiration.

[0184] Another embodiment of each of the circuits whereby the valves can be remote from the patient without loss of sequential delivery of FGS and SGS, such as those illustrated in FIGS. 5, 3B, 5A, 5B, 3C, 4B, is the replacement of separate inspiratory limbs and expiratory limbs with coaxially arranged inspiratory and expiratory limbs as shown in FIG. 6. FIG. 6A shows the preferred embodiment of the invention: The circuit valves are configured as in the circuit illustrated in FIG. 5A with the improvement of co-axially arranged inspiratory (59) and expiratory (51) limbs. The limbs (51, 59) are co-axial so that one limb is contained within the other for some length of tubing, with the limbs separating at some point along its length, such that the expiratory limb (51) leads to the exhaled gas reservoir (54) and the inspiratory limb (59) leads to the FGS reservoir (56). This has two important advantages over the circuit of FIG. 5:

- [0185] 1. A single tube is connected to the patient interface making it easier to manage sick patients
- [0186] 2. The heat contained in the expiratory limb (51) warms the FGS entering through the inspiratory limb (50)
- [0187] 3. If the inner tube is of a material that allows moisture to pass through it but not gas, such as Nafion, will promote moisture exchange as well, so that FGS will become slightly moisturized and more comfortable for the patient to breathe if the SGS is moist.

[0188] It should be understood that co-axial tubing may be used with any of the SGDB circuits described herein.

DESCRIPTION OF A PREFERRED EMBODIMENT

[0189] Referring to FIG. 6A, Patient port (50) opens directly to the inspiratory limb (59) and expiratory limb (51) without a Y connector, where the limbs are arranged coaxially. Valve (31) is an inspiratory valve and valve (33) is an expiratory valve. Valve (35) is a bypass valve in the bypass

limb (34) that bypasses the expiratory valve (33) and has an opening pressure greater than inspiratory valve (31). Valves (35, 33) are preferably distal from the patient on the expiratory limb (51) to reduce the bulk of the patient interface. Inspiratory valve (31) is also preferably distal from, the patient on the inspiratory limb (59). FGS enters the circuit via port (30). FGS control valve (400) is on the inspiratory limb (59) between port (30) and inspiratory valve (31). FGS reservoir bag (37) is connected to inspiratory limb (59) distal to the patient, past port (37). SGS reservoir bag (36) is distal to the patient on the expiratory limb (51) past expiratory valve (33) and bypass valve (35). Excess expiratory gas vents to the atmosphere via port (41). Pressure sensing means (405) is connected to pressure sensing port (406) which is connected to the patient port (50), and valve control means (403). Pressure sensing port (406) may be connected to the co-axial inspiratory (59) and expiratory limb arrangement (51) anywhere along its length between the inspiratory valve (31) and the patient port (50) or between the expiratory valve (33) and the patient. Pop-off valve (425) is connected to the inspiratory limb on the side of the FGS control valve (400) that is proximal to the inspiratory reservoir bag (425).

Function:

[0190] During exhalation, increased pressure in the circuit closes inspiratory valve (31) and bypass valve (35). Gas is directed into the exhalation limb (51), past one-way valve (33) into the expiratory gas reservoir bag (36). Excess gas is vented via port (41) in expiratory gas reservoir bag (36). FGS enters via port (30) and fills FGS reservoir (37). During inhalation, inhalation valve (31) opens and FGS from the FGS reservoir (37) and FGS port (30) enter the inspiratory limb (59) and are delivered to the patient. If FGSF is less than VE, the FGS reservoir (37) empties before the end of the breath, and continued respiratory effort results in a further reduction in pressure in the circuit. When the opening pressure of the bypass valve (35) is reached, it opens and gas from the expiratory gas reservoir (36) passes into the expiratory limb (39) and makes up the balance of the breath with SGS. The emptying of FGS reservoir bag (37) is detected by pressure sensing means (405) such as an electronic pressure transducer, known to those skilled in the art, connected to pressure sensing port (406), and FGS control valve (400) such as a balloon valve known to those skilled in the art, is closed via valve control means (403) such as access to gas pressure controlled by an electronically toggled solenoid valve known to those skilled in the art. When the FGS control valve (400) is closed, any additional FGSF entering the circuit during the balance of inspiration is directed only to the FGS reservoir bag (20) and not to the patient, who is inhaling only SGS for the balance of inspiration. FGS control valve (400) may be re-opened any time from the beginning of expiration, as sensed by the reverse of pressure by the pressure sensing means (405), to just before the next inspiration, also sensed by pressure changes in the breathing circuit. Pop-off valve (425) prevents the FGS reservoir bag (20) from overfilling when FGS

[0191] Thus when FGSF is less than \dot{V}_E , the subject inhales FOS, then SGS, and no contamination of SGS with FGS occurs.

Use of Circuits for Ventilated Patients

[0192] Any of the SGDB circuits disclosed herein as well as the Fisher circuit can be used for a patient under controlled ventilation by enclosing the FGS reservoir (20) and exhaled gas reservoir (18) within a rigid container (21) with exit ports for the inspiratory limb of the circuit (24) and expiratory limb of the circuit (25) and port for attachment to a patient interface of a ventilator (22) as illustrated in FIG. 4. In FIG. 4, the inspiratory limb (500) represents the inspiratory limb of any of the SGDB circuits herein described, and expiratory limb (501) corresponds to the expiratory limb of any of the SGDB circuits herein described. The FGS reservoir bag (20) and expiratory gas reservoir bag (18) are enclosed in a rigid airtight container such that the inspiratory limb (500) enters the container via port (24) and expiratory limb (501) enters the container via port (25) such that the junctions of the outside of the limbs form an air-tight seal with the inside surface of the ports. A further port (22) is provided for attachment of the Y piece of any ventilator (23). Detachment from the ventilator allows the circuit to be used with a spontaneously breathing patient. During the inspiratory phase of the ventilator, the pressure inside the container (21) rises putting the contents of the FGS reservoir bag (20) and the expiratory gas reservoir bag (18) under the same pressure. Since the opening pressure of the inspiratory valve is less than that of the bypass valve for circuits using passive bypass valves (for example those shown in FIGS. 2, 3, 5, 5B, 5A, 3E, and 3D), the FOS reservoir (20) will be emptied preferentially. When the FGS reservoir (20) is empty, the pressure in the container (21) and inside the expiratory gas reservoir (18) will open the bypass valve (35, 17, 206) and begin emptying exhaled gas reservoir (18) delivering SGS to the patient. For circuits using an actively engaged control valve (400) in the inspiratory limb of the circuit, a valve opening detection means (407) such as an electronic circuit that is broken by the opening of the valve when the valve is part of a closed electronic circuit, not shown, detects opening of the one way valve (35, 17, 206) in the exhalation bypass limb. The FGS control valve (400) is then closed, directing FGS into the FGS reservoir bag until the collapse of the FGS reservoir during the next inspiratory phase.

[0193] During the exhalation phase of the ventilator, the ventilator's expiratory valve is opened and contents of the container (21) are opened to atmospheric pressure, allowing the patient to exhale into the expiratory gas reservoir (18) and the FGS to flow into the FGS reservoir bag (20). Thus, the FGS and SGS are inhaled sequentially during inhalation with controlled ventilation without mixing of FGS with SGS at any time.

[0194] FIG. 4B shows the ventilator configuration described above as used with the preferred circuit shown in FIG. 6A. This is the preferred embodiment for ventilated patients.

[0195] The primary difference between the standard anesthetic circle circuit of the prior art (FIG. 12) and the circuits disclosed herein is that with the circuits disclosed herein, both a SGS reservoir (18) and a FGS reservoir (20) are in the rigid box. With the valve configurations disclosed herein, there will be sequential delivery of the FGS, then the SGS, when $\dot{\rm VE}$ exceeds the FGSF. This does not occur with the standard anesthetic circle circuit, even if the $\rm CO_2$ absorber is removed from the circuit.

Circuit for Calculation of Q and Related Physiologic Parameters while Modifying Second Gas Set

[0196] FIG. 7 shows the preferred circuit for measuring cardiac output while maintaining the ability to modify the SGS. The circuit consists of the following components:

[0197] 200 patient port

[0198] 201 three-port connector

[0199] 202 expiratory limb

[0200] 203 expiratory valve

[0201] 204 canister on bypass conduit that may be switched to be empty, contain CO₂ absorbing crystals, zeolyte, charcoal or similar substance that filters anesthetic agents, or hopcalite for filtering carbon monoxide

[0202] 205 bypass conduit.

[0203] 206 one-way bypass valve with opening pressure slightly greater than that of the inspiratory valve (219)

[0204] 207 SGS reservoir bag [0205] 208 port in rigid container for entrance of expiratory limb of circuit in an air-tight manner

[0206] 209 exit port for expired gas from expired gas reservoir

[0207] 210 a 2-way manual valve that can be turned so that the gas in the rigid box (216) is continuous with either the ventilator Y piece (211) or the manual ventilation assembly consisting of ventilating bag (212) and APL valve (213)

[0208] 211 the ventilator Y piece

[0209] 212 the ventilation bag

[0210] 213 APL valve

[0211]214 ventilation port in rigid box (216)

[0212] 215 FGS reservoir

[0213] 216 rigid box

[0214] 217 port in rigid container for entrance of inspiratory limb of circuit (220) in an air-tight manner

[0215]218 FGS inlet port

[0216] 219 inspiratory valve

[0217]220 inspiratory limb

[0218]221 bypass limb proximal to canister (204)

[0219] 400 active FGS Control valve

[0220] 403 valve control means

[0221]407 bypass valve opening sensing means

Function of the Circuit as an Anesthetic Circuit:

[0222] For spontaneous ventilation, 3-way valve (210) is open between rigid container (216) and manual ventilation assembly consisting of ventilation bag (212) and API, valve (213). When the patient exhales, increased pressure in the circuit closes inspiratory valve (219) and bypass valve (206). Exhaled gas is directed into the exhalation limb (202), past one-way valve (203) into the expiratory reservoir bag (207). FGS enters via port (218) and fills the FGS reservoir (215). During inhalation, inhalation valve (219) opens and FGS from the FGS reservoir (215) and FGS port (218) enter the inspiratory limb (220) and are delivered to patient. If FGSF is less than VE, the FGS reservoir (215) empties before the end of the breath; continued respiratory effort results in a further reduction in pressure in the circuit. When the opening pressure of the bypass valve (206) is exceeded, it opens and gas from the expiratory gas reservoir (207) passes through the canister (204) into the rebreathing limb (221) and makes up the balance of the breath with SGS. The opening of bypass valve (206) is detected by valve opening sensing means (407) signals are sent to close FGS control valve (400) by activating valve control means (403). When the FGS control valve (400) is closed, any additional FGSF entering the circuit during the balance of inspiration is directed only to the FGS reservoir bag (215) and not to the patient. When valve (400) is closed patient receives only SGS for the balance of inspiration. FGS control valve (400) may be re-opened any time from the beginning of expiration to just before the next inspiration. Phase of ventilation is sensed by sensor (407).

[0223] For the purposes of functioning as an anesthetic delivery circuit, part of the FGS entering the circuit would be the anesthetic vapor, for example Desflurane, and the canister (204) would contain CO₂ absorbent material. The SGS passes through the canister (204) but still contains expired O₂ and anesthetic, which can both be safely rebreathed by the patient. In this respect, the circuit in FIG. 7 functions like a circle anesthetic circuit in which the FGSF containing O2 and anesthetic can be reduced to match the consumption or absorption by the patient. However, by bypassing the canister (204), the circuit can be used for measuring cardiac output.

[0224] If the canister (204) is filled with hopcalite it can be used to remove carbon monoxide from the patient, since the SGS still contains expired O₂ and CO₂. If the canister (204) is filled with zeolite it can be used to remove volatile agents such as anesthetics from the patient.

Advantages of Circuit Over Previous Art.

[0225] 1. It is comparable to the circle anesthesia circuit with respect to efficiency of delivery of anesthesia, and ability to conduct anesthesia with spontaneous ventilation as well as controlled ventilation.

[0226] 2) It is often important to measure tidal volume and VE during anesthesia. With a circle circuit, a pneumotach with attached tubing and cables must be placed at the patient interface, increasing the dead-space, bulk and clutter at the head of the patient. With our circuit, the pneumotach (or a spirometer if the patient is breathing spontaneously) can be placed at port (214) and thus remote from the

[0227] 3) Sasano (Anesth Analg 2001; 93:1188-1191) taught a circuit that can be used to accelerate the elimination of anesthesia However that circuit required additional devices such as an external source of gas (reserve gas), a demand regulator, self-inflating bag or other manual ventilating device, 3-way stopcock and additional tubing. Furthermore, Sasano did not disclose a method whereby mechanical ventilation can be used. In fact it appears that it cannot be used—patients must be ventilated by hand for that method. With the apparatus and method disclosed herein, there is no requirement for an additional external source of gas or demand regulator;

[0228] 4) the patient can be ventilated with the ventilation bag (212) already on the circuit or the circuit ventilator, or any ventilator; no other tubing or devices are required.

[0229] 5) Circle circuits cannot deliver FGS and then SGS sequentially. Such control is required to make physiological measurements such as cardiac output during anesthe-

[0230] With the circuit of FIG. 7, if the canister (204) is bypassed, the circuit becomes the equivalent of the one described in FIG. 5 with the addition of the ventilator apparatus shown in FIG. 4. With the circuit of FIG. 7, box (216) could be opened to atmosphere instead of connected to a ventilator, and the circuit could be used with spontaneously breathing patients for measuring cardiac output while modifying SGS.

[0231] It should be recognized to those skilled in the art that various embodiments of the invention disclosed in this patent application are possible without departing from the scope including, but not limited to:

[0232] a) using multiple inspiratory and expiratory limbs in combination provided that

[0233] i) the inspiratory and expiratory limbs are kept separate except at a single point prior to reaching the patient where they are joined

[0234] ii) each limb has the corresponding valves as in the arrangement above, and

[0235] iii) the valves have the same relative pressures so as to keep the inspired gas delivery sequential as discussed above.

- [0236] b) using active valves, for example electronic, solenoid, or balloon valves, instead of passive valves, provided said valves are capable of occluding the limbs, and means is provided for triggering and controlling said active valves. The advantage of active valves is more precise control. The disadvantage is that they are more costly.
- [0237] c) replacing reservoir bags with extended tubes or other means for holding gases
- [0238] d) surrounding valves in exhalation limb and/or in the inspiratory limb of circuit with the exhaled gas reservoir causing them to be surrounded by warm exhaled air and prevent freezing and sticking of valves in cold environments.
- [0239] e) Changing the composition of FGS and SGS to change alveolar concentrations of gases other than CO₂, for example O₂. By analogy to CO₂, with respect to O₂: alveolar PO₂ is determined by FGS flow and the PO₂ of FGS. When PO₂ of SGS is the same as the PO₂ in the alveoli, inhaling SGS does not change flux of O₂ in the alveoli. Therefore, those skilled in the art can arrange the partial pressure of component gases in FGS and SGS and the flows of FGS such that they can achieve any alveolar concentration of component gases independent of Ve, as long as Ve exceeds sufficiently flow of FGS.
- [0240] As many changes can be made to the various embodiments of the invention without departing from the scope thereof; it is intended that all matter contained herein be interpreted as illustrative of the invention but not in a limiting sense.
- [0241] To clarify the function of the automated cardiac output device, we will contrast it to a standard anaesthetic machine which has the same configuration of listed components
- [0242] 1) The preferred SGDB circuits we describe differ from any anaesthetic circuit. The SGDB circuit first provides the FGS, then the SGS. This allows the circuit to compensate for changes in CO₂ elimination on any particular breath. For example, during a small breath, the unused FGS remains in the FGS reservoir and is available to provide the exact additional VA for each gas in the set when a larger breath is taken or frequency of breathing increases subsequently. As a result, changes in VCO₂ can be instituted independent of breathing pattern.
- [0243] 2) Anesthetic machines do not automatically alter the fresh gas flows. Fresh gas flows are manually controlled by the anesthesiologist
- [0244] 3) Anesthetic machines do not calculate \dot{V} A and cannot calculate \dot{V} CO₂, and \dot{Q} .
- [0245] 4) Anesthetic machines cannot generate the data required to make the calculations for Q and its associated parameters because the circuit is inappropriate and the gas flows are not configured to be controlled by a computer.
- [0246] 5) The flowmeters on commonly used anesthetic machines are too imprecise and inaccurate to perform these tests and calculations. There is no need for such precision and accuracy of flow for routine clinical anesthetic care.
- 9.0 Method of Generating Data Required to Make Calculations of Q and Related Physiologic Parameters (See FIG. 11):
 - [0247] Cardiac Output can be measured in several ways according to the methods and apparatus disclosed herein. These include:
- [0248] 9.1 Set-Up Phase
 - [0249] 9.1.1 Set Flow of FGS>VE
 - [0250] 9.1.2 Access default values
 - [0251] 9.1.3 Check pressure sensor or PCO₂ sensor during inhalation. If fresh gas reservoir collapsed or CO₂ is

- detected during inhalation, increase FGS flow until the reservoir until reservoir does not collapse fully and no CO_2 is detected during inhalation
- [0252] 9.1.4 Identify $P_{ET}CO_2$ from the CO_2 gas analyzer [0253] 9.2 Find \dot{V}_A via one of two methods:
 - [0254] 9.2.1 Calculate VA by inducing two reductions in FGS flow below VA without first identifying VA by following the following steps:
 - [0255] 9.2.1.1 Calculate a preliminary minimum VA for the subject based on body weight, temperature, sex and other parameters known to those skilled in the art.
 - [0256] 9.2.1.2 Provide luxuriant FGS flow greater than the patient's resting $\dot{V}_{\rm E}$ until steady state $P_{ET}CO_2$ is reached
 - [0257] 9.2.1.3 Impose a Va by setting FGS Flow below assumed Va, to VA^x preferably just below the calculated preliminary Va, for a time less than or equal to a recirculation time, and measure PetCO₂^x, the end tidal CO₂ concentration during equilibrium if an equilibrium end tidal value is reached within a recirculation time, otherwise it is the equilibrium value of end tidal CO₂ as extrapolated from the exponential rise in end tidal CO₂ values within the recirculation time.
 - [0258] 9.2.1.4 Set FGS flow above V_E until steady state $P_{ET}CO_2$ is reached as identified by a less than a threshold change in $P_{ET}CO_2$ over a designated time period. The actual thresholds and time periods are user defined according to the circumstances of the test and can be determined by those skilled in the art.
 - [0259] 9.2.1.5 Impose a VA by setting FGS Flow below assumed VA, to VA* where VA* is less than calculated preliminary minimum VA and not equal to VA*, for a time approximately equal to a recirculation time, about 30 s at rest. Measure PetCO₂*, the end tidal CO₂ concentration during equilibrium if an equilibrium end tidal value is reached within a recirculation time, otherwise it is the equilibrium value of end tidal CO₂ as extrapolated from the exponential rise in end tidal CO₂ values within the recirculation time.
 - [0260] 9.2.1.6 On a graph of $P_{ET}CO_2$ vs FGS flow, plot the points $(P_{ET}CO_2^y, \dot{V}A^y)$ and $(P_{ET}CO_2^x, \dot{V}A^x)$. Extrapolate the line formed by connecting these two point to intersect a horizontal line at $P_{ET}CO_2$ -resting $P_{ET}CO_2$. The FGS flow at the intersection point is determined to be $\dot{V}A$.
 - [0261] 9.2.2 Progressive Reduction of FGS flow method of finding V_A:
 - [0262] 9.2.2.1 Use FGS that preferably has no CO₂
 - [0263] 9.2.2.2 Wait for steady state as indicated by less than a threshold change in PetCO₂ over a designated time period. The actual thresholds and time periods are user defined according to the circumstances of the test and can be determined by those skilled in the art.
 - [0264] 9.2.2.3 When in steady state, reduce FGS flow by a small fixed flow, for example 0.1 L/min, preferably at regular intervals of time or after each breath. Alternate flow reduction rates could be used, and the reduction need not be linear in time.
 - [0265] 9.2.2.4 When P_{ET}CO₂ begins to rise above a threshold value which is approximately the mean steady state P_{ET}CO₂, continue the reduction in the FGS flow for a time approximately equal to one recirculation time.
 - [0266] 9.2.2.5 After approximately one recirculation time, usually about 30 s, raise FGS flow above resting

 $\dot{\text{VE}}$. A relation of $P_{ET}\text{CO}_2$ vs FGS flow is calculated and two lines of best fit are calculated, one for the set of steady state $P_{ET}\text{CO}_2$ values, and one for the set of raised $P_{ET}\text{CO}_2$ values above the mean of the steady state values. The FGS flow corresponding to the intersection of said lines corresponds to $\dot{\text{VA}}$. FIG. 13 illustrates that progressive reduction of SGF (labelled "FGF" in the figure) results in a distinct inflection point when either $P_{\text{ET}}\text{CO}_2$ or $P_{\text{ET}}\text{O}_2$ is graphed as a function of SGF. We define the SGF corresponding to this inflection point as equal to $\dot{\text{VA}}$.

[0267] 9.2.2.6 These two methods of finding \dot{V}_A are physiologically equivalent and one may have some advantages over the other in particular clinical or research circumstances. The Progressive Reduction method should be contrasted with the method for calculating VA taught by Preiss et al. (Canadian Patent Application 2346517). In that method, while fresh gas flow into a sequential gas delivery circuit was reduced stepwise, after each reduction, the subject was observed for several breaths looking for an exponential rise in P_{ET}CO₂. The Preiss method requires continued breathing at each fresh gas flow looking for development of a new steady when fresh gas flow falls below VA. This process is very time consuming and is unlikely to be tolerated by most patients. If, in the attempt to shorten the time for finding the fresh gas flow below V_A the fresh gas flow reduction are large, resolution of critical fresh gas flow is lost. If the steps are small, when the fresh gas flow is just barely less than VA, it will be difficult to discern the small rise in PetCO₂ from the normal variation in PetCO₂. The progressive breath-by-breath reduction in FGS flow disclosed herein results in a rapid linear rise in PetCO₂ and fall in PetO₂, both of which can be used to identify the FGS flow corresponding to VA as illustrated in FIG. 13.

[0268] 9.3 Calculations with the Differential Fick Equation [0269] There are two methods of calculating cardiac output with the Differential Fick equation. (It is understood that the general methods are disclosed without the details well known to those skilled in the art of the multiple standard corrections for temperature, moisture, barometric pressure and the like):

[0270] 9.3.1 Find \dot{V}_A by the Progressive Reduction of FGS flow method of finding \dot{V}_A :

[0271] 9.3.1.1 Find VA

[0272] 9.3.1.2 Set FGS Flow=VA and calculate VCO₂ using the equation VCO₂=VA×FETCO₂.

[0273] 9.3.1.3 Impose a transient step change in VA to VA' for a time approximately equal to a recirculation time, about 30 s at rest, by changing FGS flow to a value below VA. To fully automate the process, select a VA' that will be below the VA. Calculate VCO₂'=VA'× F_{ET}CO₂'. Where F_{ET}CO₂' is the fractional end tidal CO₂ concentration during equilibrium if an equilibrium end tidal value is reached within a recirculation time, otherwise it is the equilibrium value of end tidal CO₂ as extrapolated from the exponential rise in end tidal CO₂ values within the recirculation time.

[0274] 9.3.1.4 Calculate Q according to the differential Fick equation using VCO₂ ν_{CO2} , and CCO₂ and CCO₂ where CCO₂ and CCO₂ are the contents of CO₂ of end capillary blood as calculated from P_{E7}CO₂ and P_{E7}CO₂ using known relationships between PerCO₂, and other characteristics related to the blood such as hemoglobin concentration, tem-

perature oxygen partial pressure and other parameters that are accessible or can be used as default values by those skilled in the art.

[0275] 9.3.1.5 Calculate Q according to the differential Fick equation using VCO₂ and P_{ET}CO₂ data from steady state phase and step change phase and the PaCO₂ from the Kim Rahn Farhi method. This allows the identification of the P_{ET}CO₂—PaCO₂ gradient without an arterial blood sample.

[0276] 9.3.2 Generate required data by inducing two reductions in FGS flow below \dot{V}_A without first identifying \dot{V}_A by following the following steps:

[0277] 9.3.2.1 Calculate a preliminary minimum VA for the subject based on body weight, temperature, sex and other parameters known to those skilled in the art.

[0278] 9.3.2.2 Provide luxuriant FGS flow greater than the patient's resting \dot{V}_E until steady state $P_{ET}CO_2$ is reached

[0279] 9.3.2.3 Impose a VA and hence a VCO₂ by setting FGS Flow below preliminary calculated VA, to VA^x, preferably just below the preliminarily calculated VA, for a time less than or equal to a recirculation time, and calculate VCO₂^x using the equation VCO₂^{x=VA^x}×F_{ET}CO₂^x where F_{ET}CO₂^x is the fractional end tidal CO₂ concentration during equilibrium if an equilibrium end tidal value is reached within a recirculation time, otherwise it is the equilibrium value of end tidal CO₂ as extrapolated from the exponential rise in end tidal CO₂ values within the recirculation time.

[0280] 9.3.2.4 Set FGS flow above V_E until steady state $P_{ET}CO_2$ is reached as identified by a less than a threshold change in $PetCO_2$ over a designated time period. The actual thresholds and time periods are user defined according to the circumstances of the test and can be determined by those skilled in the art.

[0281] 9.3.2.5 Impose a transient step change in VA to VA^y where VA^y is less than calculated VA and not equal to VA^x, for a time approximately equal to a recirculation time, about 30 s at rest. Calculate VCO₂^y=VA^y× FETCO₂^y. FETCO₂^y is the end tidal CO₂ concentration during equilibrium if an equilibrium end tidal value is reached within a recirculation time, otherwise it is the equilibrium value of end tidal CO₂ as extrapolated from the exponential rise in end tidal CO₂ values within the recirculation time.

[0282] 9.3.2.6 Calculate Q according to the differential Fick equation using VCO₂^x, VCO₂^y, and CCO₂^x and CCO₂^x where CCO₂^x and CCO₂^y are the contents of CO₂ of end capillary blood as calculated from P_{ET}CO₂^x, and P_{ET}CO₂^y using known relationships between PETCO₂, and other characteristics related to the blood such as hemoglobin concentration, temperature oxygen partial pressure and other parameters that are accessible or can be used as default values by those skilled in the art.

[0283] 9.3.2.7 Calculate Q according to the differential Fick equation using VCO₂ and P_{ET}CO₂ data from steady state phase and step change phase and the PaCO₂ from the Kim Rahn Farhi method to identify the PetCO₂—PaCO₂ gradient. This allows the identification of the PetCO₂—PaCO₂ gradient without an arterial blood sample.

[0284] Difference between this method and previous methods to perform the differential Fick:

[0285] (a) With the new method, the decrease in VCO₂ is performed by reducing the FOP to a SGDB circuit as opposed to insertion of a dead-space at the patient-circuit interface. As a result, if the subject increases his breathing rate or breath size, there is no change in VCO₂ and the calculations via the differential Fick equation are not affected.

[0286] (b) The VCO₂ is known using the VA (identified by one of the new or the previously disclosed method) and the PetCO₂, two robust and highly reliable measures. This is unlike the need for a pneumotachymeter and the error-prone breath-bybreath analysis of VCO₂ required by previous art.

[0287] (c) Va is not identified with the previous differential Fick methods.

[0288] (d) The PetCO₂ to PaCO₂ gradient is calculated from two independently derived values in the same subject. In the previous art, this gradient is calculated from empirical formulae derived from averaged values and do not necessarily apply to the subject.

[0289] Therefore our method provides more accurate values for VCO₂, V CO₂ and PaCO₂ than the previous art.

[0290] 9.4 Kim-Rahn-Farhi

[0291] 9.4.1 A period of reduced FGS flow simulates complete or partial breath holding. The PetCO₂ of each breath is equivalent to a sequential alveolar sample in the KU prolonged exhalation method. The substitution of sequential PetCO₂ values for sequential samples from a single exhalation is used to calculate true PvCO₂, PvCO₂-oxy, PaCO₂ and hemoglobin O₂ saturation in mixed venous blood SvO₂ using the Kim Rahn Farhi method

[0292] 9.4.2 \dot{Q} can be calculated using the Fick approach where the $P\bar{\nu}CO_2$ -oxy and $PaCO_2$ as calculated by the Kim Rahn Farhi method are used to calculate the respective CO_2 contents using methods well known to those skilled in the art, and the $\dot{\nu}CO_2$ is as calculated from $\dot{\nu}A$ and $FetCO_2$ as derived in the sequence of steps described above.

[0293] 9.4.3 Mixed venous O₂ hemoglobin saturation are calculated as follows. $\dot{V}O_2$ is calculated from $\dot{V}O_2=\dot{V}_{AX}$ (FiO₂-FetO₂) where FiO₂ and FetO₂ are the fractional concentration of inspired and end tidal O₂ respectively. Using $\dot{V}O_2$, \dot{Q} as calculated by Differential Fick or Kim Rahn Farhi or Fisher Method, end capillary O₂ oxygen content (assuming end capillary blood is fully saturated with oxygen), Mixed venous O₂ saturation can be calculated from the standard Fick equation.

[0294] 9.4.4 Information regarding the arterial O_2 hemoglobin saturation (Sa O_2) (as read from a non-invasive commonly available pulse oximeter that makes the measurement by shining an infrared light through a finger), and the $S\overline{V}O_2$) can be used to calculate the fraction of shunted blood (\dot{Q}_S) (assuming fully oxygenated blood in the end pulmonary capillary) by using the following equation

$$\dot{Q}s = \frac{(SpO_2)\dot{Q}t - (SaO_2)\dot{Q}p}{S\overline{\nu}O_2}$$

[0295] Our method of performing the Kim Rahn Farhi is an improvement over the previous art in that

[0296] (a) Test is performed simultaneously with a test for differential Fick in spontaneously breathing subject.

[0297] (b) Data are pooled with the test as outlined above so calculation of VCO₂, is simultaneous to the other calculations. In the previous art, the VCO₂, calculation cannot be done during a breath hold or simulated breath hold by rebreathing.

[0298] (c) VCO₂, measurement does not require a pneumotachymeter which is expensive, cumbersome and error-prone. In the previous art, VCO₂, required for the calculation of Q required additional apparatus such as pneumatchymeter or gas collection and volume measuring apparatus.

[**0299**] 9.5 Fisher E-I Test

[0300] 9.5.1 Calculate VA from the calibration phase, set FGS flow=VA.

[0301] 9.5.2 With FGS Flow at VA, the PCO₂ in the FGS is changed to any value and held at that value for a time approximately equal to a recirculation time, about 30 s at rest

[0302] 9.5.3 PvCO₂-oxy is calculated using the PetCO₂-PtCO₂ method described by Fisher.

[0303] Our method of the Fisher E-I test is an improvement over the previous art in that the effect of change in breath size on the equilibrium value of PetCO₂ is minimized by the SGDB circuit such that a larger breath delivers physiologically neutral previously expired gas instead of additional test gas.

10.0 Method of Finding \dot{V}_{E} Using Progressive Reduction of FGS Flow:

[0304] 10.1 Use FGS that preferably has no CO₂

[0305] 10.2 Wait for steady state as indicated by less than a threshold change in PetCO₂ over a designated time period. The actual thresholds and time periods are user defined according to the circumstances of the test and can be determined by those skilled in the art.

[0306] 10.3 When in steady state, reduce FGS flow by a small fixed flow, for example 0.1 L/min, preferably at regular intervals of time or after each breath. Alternate flow reduction rates could be used, and the reduction need not be linear in time.

[0307] 10.4 Using a means for measuring pressure within the FGS reservoir in the breathing circuit, for example a pressure transducer, monitor when the FGS reservoir bag first collapses. VE is the FGS flow rate when the reservoir bag first collapses.

11.0 Method for Measuring Anatomical Dead Space

[0308] 11.1 Measure \dot{V}_E and \dot{V}_A using any of the methods disclosed above

[0309] 11.2 Measure the respiratory rate, preferrably using the apparatus for cardiac output disclosed herein.

[0310] 11.3 Calculate Anatomical Dead Space $\dot{V}_{DAN}=(\dot{V}_{E}-\dot{V}_{A})/respiratory$ rate

[0311] As many changes can be made to the various embodiments of the invention without departing from the scope thereof; it is intended that all matter contained herein be interpreted as illustrative of the invention but not in a limiting sense.

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- **62**. A method of identifying the alveolar ventilation (\dot{V}_A) , the method comprising:
 - (1) using a breathing circuit characterized in that:
 - i. on exhalation, exhaled gas is kept substantially sepa-
 - rate from inhaled gas; ii. on inhalation, when \dot{V}_E is less than FGS flow, the subject inhales FGS first and the balance of the minute ventilation is substantially SGS,
 - (2) setting the FGS flow into the circuit at a rate greater than the subjects minute ventilation (\dot{V}_E) ;
 - (3) measuring the end tidal CO_2 concentration ($P_{ET}CO_2$) in a steady state
 - (4) progressively lowering the FGS flow into the circuit, either breath by breath or continuously, until after a time equal to the time of recirculation of CO₂ within the patient after a rise in P_{ET}CO₂ above a threshold value is
 - (5) deriving \dot{V}_A as the rate of FGS flow at a point of the intersection between two lines comprising:
 - a. the average $P_{ET}CO_2$ in steady state; and
 - b. a line fit to the $P_{ET}CO_2$ values after the rise begins until the recirculation time.



专利名称(译)	在自发和受控通气期间通过肺非侵入性地测量心脏相关参数的方法		
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[标]发明人	FISHER JOSEPH PREISS DAVID ASAMI TAKAFUMI VESELY ALEX PRISMAN ELTAN SOMOGYI RON ISCOE STEVE		
发明人	FISHER, JOSEPH PREISS, DAVID ASAMI, TAKAFUMI VESELY, ALEX PRISMAN, ELTAN SOMOGYI, RON ISCOE, STEVE		
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摘要(译)

一种测量肺血流量和心输出量(Q)的装置,包括:a)呼气回路,当呼气时,当呼吸气体大于第一气体流量(FGS)流量时,呼出气体将呼出气体与吸入气体分开并吸入在吸入FGS的受试者中然后吸入第二组气体 201、(SGS),以实现吸入的平衡; b)用于监测患者 - 回路界面处的气体浓度的气体传感器; c)气流控制装置,用于控制进入呼吸回路的FGS流量; d)由能够控制气流控制装置的计算机或逻辑电路组成的机器智能,其接收气体传感器装置的输出并输出肺血流。

