



US009113801B2

(12) **United States Patent**
DiLorenzo

(10) **Patent No.:** **US 9,113,801 B2**
(45) **Date of Patent:** **Aug. 25, 2015**

(54) **METHODS AND SYSTEMS FOR
CONTINUOUS EEG MONITORING**

(75) Inventor: **Daniel John DiLorenzo**, Seattle, WA
(US)

(73) Assignee: **CYBERONICS, INC.**, Houston, TX
(US)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 1463 days.

(21) Appl. No.: **11/647,785**

(22) Filed: **Dec. 29, 2006**

(65) **Prior Publication Data**

US 2007/0161919 A1 Jul. 12, 2007

Related U.S. Application Data

(63) Continuation of application No. 10/889,844, filed on
Jul. 12, 2004, now Pat. No. 7,231,254, and a
continuation-in-part of application No. 10/818,333,
filed on Apr. 5, 2004, now Pat. No. 7,277,758, and a

(Continued)

(51) **Int. Cl.**
A61B 5/04 (2006.01)
A61B 5/0476 (2006.01)

(Continued)

(52) **U.S. Cl.**
CPC **A61B 5/04001** (2013.01); **A61B 5/04012**
(2013.01); **A61B 5/0476** (2013.01); **A61B**
5/0488 (2013.01); **A61B 5/1101** (2013.01);
A61B 5/4094 (2013.01); **A61B 5/4839**
(2013.01); **A61N 1/36082** (2013.01); **A61N**
1/372 (2013.01); **A61B 5/6852** (2013.01); **A61B**
5/6864 (2013.01); **A61B 5/6868** (2013.01);
A61B 5/6882 (2013.01); **A61B 5/7203**
(2013.01);

(Continued)

(58) **Field of Classification Search**

CPC ... A61B 5/0476; A61B 5/0478; A61B 5/0482
USPC 600/544-545
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

3,218,638 A 11/1965 Honig
3,498,287 A 3/1970 Ertl

(Continued)

FOREIGN PATENT DOCUMENTS

CA 2251852 4/1999
CA 2423840 2/2002

(Continued)

OTHER PUBLICATIONS

DiLorenzo, Daniel; U.S. Appl. No. 12/177,060 entitled "Closed-loop
feedback-driven neuromodulation," filed Jul. 21, 2008.

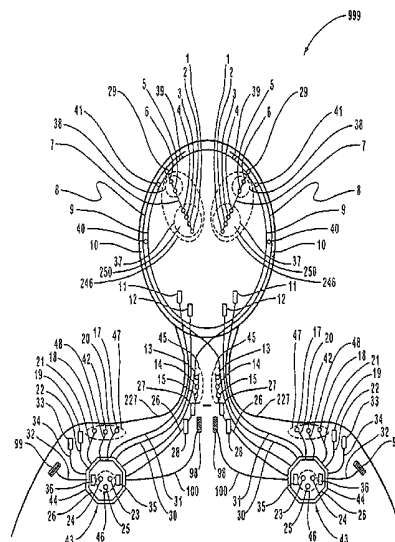
(Continued)

Primary Examiner — Michael D'Angelo

(57) **ABSTRACT**

A neurological control system for modulating activity of any component or structure comprising the entirety or portion of the nervous system, or any structure interfaced thereto, generally referred to herein as a "nervous system component." The neurological control system generates neural modulation signals delivered to a nervous system component through one or more neuromodulators to control neurological state and prevent neurological signs and symptoms. Such treatment parameters may be derived from a neural response to previously delivered neural modulation signals sensed by one or more sensors, each configured to sense a particular characteristic indicative of a neurological or psychiatric condition.

12 Claims, 66 Drawing Sheets



Related U.S. Application Data

continuation-in-part of application No. 10/753,205, filed on Jan. 6, 2004, now Pat. No. 7,242,984, and a continuation-in-part of application No. 10/718,248, filed on Nov. 20, 2003, now Pat. No. 7,209,787, which is a continuation-in-part of application No. 10/008,576, filed on Nov. 11, 2001, now Pat. No. 6,819,956, which is a continuation-in-part of application No. 09/340,326, filed on Jun. 25, 1999, now Pat. No. 6,366,813.

- (60) Provisional application No. 60/562,487, filed on Apr. 14, 2004, provisional application No. 60/460,140, filed on Apr. 3, 2003, provisional application No. 60/438,286, filed on Jan. 6, 2003, provisional application No. 60/095,413, filed on Aug. 5, 1998, provisional application No. 60/427,699, filed on Nov. 20, 2002, provisional application No. 60/436,792, filed on Dec. 27, 2002.

(51) **Int. Cl.**

A61B 5/0488 (2006.01)

A61B 5/11 (2006.01)

A61B 5/00 (2006.01)

A61N 1/36 (2006.01)

A61N 1/372 (2006.01)

A61N 1/05 (2006.01)

A61N 1/378 (2006.01)

(52) **U.S. Cl.**

CPC *A61N1/0529* (2013.01); *A61N 1/0531* (2013.01); *A61N 1/0539* (2013.01); *A61N 1/36114* (2013.01); *A61N 1/3787* (2013.01)

(56) **References Cited**

U.S. PATENT DOCUMENTS

3,522,811	A	8/1970	Schwartz
3,575,162	A	4/1971	Gaarder
3,837,331	A	9/1974	Ross
3,850,161	A	11/1974	Liss
3,863,625	A *	2/1975	Viglione et al. 600/545
3,882,850	A	5/1975	Bailin et al.
3,918,461	A	11/1975	Cooper
3,967,616	A	7/1976	Ross
3,993,046	A	11/1976	Fernandez
4,201,224	A	5/1980	John
4,214,591	A	7/1980	Sato et al.
4,279,258	A	7/1981	John
4,305,402	A	12/1981	Katims
4,334,545	A	6/1982	Shiga
4,407,299	A	10/1983	Culver
4,408,616	A	10/1983	Duffy et al.
4,417,592	A *	11/1983	John 600/544
4,421,122	A	12/1983	Duffy
4,471,786	A	9/1984	Inagaki
4,494,950	A	1/1985	Fischell
4,505,275	A	3/1985	Chen
4,545,388	A	10/1985	John
4,556,061	A	12/1985	Barreras et al.
4,566,464	A	1/1986	Piccone et al.
4,573,481	A	3/1986	Bullara
4,579,125	A	4/1986	Strobl et al.
4,590,946	A	5/1986	Loeb
4,612,934	A	9/1986	Borkan
4,679,144	A	7/1987	Cox et al.
4,686,999	A	8/1987	Snyder et al.
4,702,254	A	10/1987	Zabara
4,735,208	A	4/1988	Wylar et al.
4,768,176	A	8/1988	Kehr et al.
4,768,177	A	8/1988	Kehr et al.
4,785,827	A	11/1988	Fischer

4,793,353	A	12/1988	Borkam
4,817,628	A	4/1989	Zealear
4,838,272	A	6/1989	Lieber
4,844,075	A	7/1989	Liss et al.
4,852,573	A	8/1989	Kennedy
4,867,164	A	9/1989	Zabara
4,873,981	A	10/1989	Abrams et al.
4,878,498	A	11/1989	Abrams et al.
4,903,702	A	2/1990	Putz
4,920,979	A	5/1990	Bullara
4,926,865	A	5/1990	Oman
4,955,380	A	9/1990	Edell
4,978,680	A	12/1990	Sofia
4,979,511	A	12/1990	Terry
4,991,582	A	2/1991	Byers et al.
5,010,891	A	4/1991	Chamoun
5,016,635	A	5/1991	Graupe
5,025,807	A	6/1991	Zabara
5,029,590	A	7/1991	Allain et al.
5,031,618	A	7/1991	Mullett
5,070,873	A	12/1991	Graupe et al.
5,082,861	A	1/1992	Sofia
5,097,835	A	3/1992	Putz
RE34,015	E	8/1992	Duffy
5,154,172	A	10/1992	Terry
5,167,229	A	12/1992	Peckham et al.
5,179,950	A	1/1993	Stanislaw
5,181,520	A	1/1993	Wertheim et al.
5,186,170	A	2/1993	Varichio
5,188,104	A	2/1993	Wernicke
5,190,029	A	3/1993	Byron et al.
5,193,539	A	3/1993	Schulman et al.
5,193,540	A	3/1993	Schulman et al.
5,205,285	A	4/1993	Baker, Jr.
5,215,086	A	6/1993	Terry, Jr. et al.
5,215,088	A	6/1993	Normann
5,215,089	A	6/1993	Baker, Jr.
5,222,494	A	6/1993	Baker, Jr.
5,222,503	A	6/1993	Ives
5,231,988	A	8/1993	Wernicke et al.
5,235,980	A	8/1993	Varichio et al.
5,237,991	A	8/1993	Baker, Jr.
5,251,634	A	10/1993	Weinberg
5,263,480	A	11/1993	Wernicke et al.
5,265,619	A	11/1993	Comby et al.
5,269,302	A	12/1993	Swartz et al.
5,269,303	A	12/1993	Wernicke et al.
5,269,315	A	12/1993	Leuchter et al.
5,292,772	A	3/1994	Sofia
5,293,879	A	3/1994	Vonk
5,299,118	A	3/1994	Martens et al.
5,299,569	A	4/1994	Wernicke et al.
5,300,094	A	4/1994	Kallok et al.
5,304,206	A	4/1994	Baker, Jr. et al.
5,311,876	A	5/1994	Olsen et al.
5,312,439	A	5/1994	Loeb
5,314,458	A	5/1994	Najafi et al.
5,324,316	A	6/1994	Schulman et al.
5,330,515	A	7/1994	Rutecki et al.
5,335,657	A	8/1994	Terry et al.
5,342,408	A	8/1994	deColriolis et al.
5,342,409	A	8/1994	Mullett
5,343,064	A	8/1994	Spangler et al.
5,349,962	A	9/1994	Lockard et al.
5,351,394	A	10/1994	Weinberg
5,361,760	A	11/1994	Normann
5,365,939	A	11/1994	Ochs
5,376,359	A	12/1994	Johnson
5,392,788	A	2/1995	Hudspeth
5,405,365	A	4/1995	Hoegnelid et al.
5,405,367	A	4/1995	Schulman et al.
5,411,540	A	5/1995	Edell et al.
5,458,117	A	10/1995	Chamoun
5,474,547	A	12/1995	Aebischer et al.
5,476,494	A	12/1995	Edell et al.
5,486,999	A	1/1996	Mebane
5,513,649	A	5/1996	Gevens
5,531,778	A	7/1996	Maschino et al.
5,540,730	A	7/1996	Terry

(56)

References Cited

U.S. PATENT DOCUMENTS

5,540,734 A	7/1996	Zabara	6,094,598 A	7/2000	Elsberry et al.
5,549,656 A	8/1996	Reiss	6,109,269 A	8/2000	Rise et al.
5,555,191 A	9/1996	Hripcsak	6,117,066 A	9/2000	Abrams et al.
5,571,148 A	11/1996	Loeb et al.	6,128,537 A	10/2000	Rise et al.
5,571,150 A	11/1996	Wernicke	6,128,538 A	10/2000	Fischell et al.
5,575,813 A	11/1996	Edell et al.	6,134,474 A	10/2000	Fischell et al.
5,578,036 A	11/1996	Stone et al.	6,161,045 A	12/2000	Fischell et al.
5,611,350 A	3/1997	John	6,167,304 A	12/2000	Loos
5,626,145 A	5/1997	Clapp et al.	6,171,239 B1	1/2001	Humphrey
5,626,627 A	5/1997	Krystall et al.	6,176,242 B1	1/2001	Rise
5,638,826 A	6/1997	Wolpaw	6,205,359 B1	3/2001	Boveja
5,649,068 A	7/1997	Boser et al.	6,208,893 B1	3/2001	Hofmann
5,672,154 A	9/1997	Sillen et al.	6,221,011 B1	4/2001	Bardy
5,683,422 A	11/1997	Rise	6,227,203 B1	5/2001	Rise et al.
5,683,432 A	11/1997	Goedeke et al.	6,230,049 B1	5/2001	Fischell et al.
5,690,681 A	11/1997	Geddes et al.	6,248,126 B1	6/2001	Lesser et al.
5,690,691 A	11/1997	Chen et al.	6,249,703 B1	6/2001	Stanton
5,697,369 A	12/1997	Long	6,259,889 B1 *	7/2001	LaDue 434/308
5,700,282 A	12/1997	Zabara	6,263,237 B1	7/2001	Rise
5,704,352 A	1/1998	Tremblay et al.	6,280,198 B1	8/2001	Calhoun et al.
5,707,400 A	1/1998	Terry et al.	6,304,775 B1	10/2001	Iasemidis et al.
5,711,316 A	1/1998	Elsberry et al.	6,309,406 B1	10/2001	Jones et al.
5,713,923 A *	2/1998	Ward et al. 607/3	6,328,699 B1	12/2001	Eigler
5,715,821 A	2/1998	Faupel	6,337,997 B1	1/2002	Rise
5,716,377 A	2/1998	Rise et al.	6,339,725 B1	1/2002	Naritoku
5,720,294 A	2/1998	Skinner	6,341,236 B1	1/2002	Osorio et al.
5,730,146 A	3/1998	Itil et al.	6,343,226 B1	1/2002	Sunde et al.
5,735,814 A	4/1998	Elsberry et al.	6,353,754 B1	3/2002	Fischell et al.
5,743,860 A	4/1998	Hively et al.	6,354,299 B1	3/2002	Fischell et al.
5,749,908 A	5/1998	Snell	6,356,784 B1	3/2002	Lozano et al.
5,752,979 A	5/1998	Benabid	6,356,788 B2	3/2002	Boveja
5,759,199 A	6/1998	Snell et al.	6,358,203 B2	3/2002	Bardy
5,769,778 A	6/1998	Abrams et al.	6,358,281 B1	3/2002	Berrang et al.
5,776,434 A	7/1998	Purewal et al.	6,360,122 B1	3/2002	Fischell
5,782,798 A	7/1998	Rise	6,366,813 B1	4/2002	DiLorenzo
5,782,874 A	7/1998	Loos	6,366,814 B1	4/2002	Boveja
5,782,891 A	7/1998	Hassler et al.	6,374,140 B1	4/2002	Rise
5,792,186 A	8/1998	Rise	6,386,882 B1	5/2002	Linberg
5,800,474 A	9/1998	Benabid et al.	6,402,678 B1	6/2002	Fischell et al.
5,813,993 A	9/1998	Kaplan	6,411,854 B1	6/2002	Tziviskos et al.
5,814,014 A	9/1998	Elsberry et al.	6,427,086 B1	7/2002	Fischell et al.
5,815,413 A	9/1998	Hively et al.	6,434,419 B1	8/2002	Gevens et al.
5,816,247 A	10/1998	Maynard	6,442,421 B1	8/2002	Quyen et al.
5,824,021 A	10/1998	Rise	6,443,891 B1	9/2002	Grevious
5,832,932 A	11/1998	Elsberry et al.	6,453,198 B1	9/2002	Torgerson
5,833,709 A	11/1998	Rise et al.	6,463,328 B1	10/2002	John
5,857,978 A	1/1999	Hively et al.	6,466,822 B1	10/2002	Pless
5,862,803 A	1/1999	Besson et al.	6,471,645 B1	10/2002	Warkentin et al.
5,876,424 A	3/1999	O'Phelan et al.	6,473,639 B1	10/2002	Fischell et al.
5,899,922 A	5/1999	Loos	6,473,644 B1	10/2002	Terry et al.
5,913,881 A	6/1999	Benz et al.	6,480,743 B1	11/2002	Kirkpatrick
5,916,239 A	6/1999	Geddes et al.	6,484,132 B1	11/2002	Hively et al.
5,917,429 A	6/1999	Otis, Jr. et al.	6,488,617 B1	12/2002	Katz
5,928,272 A	7/1999	Adkins	6,496,724 B1	12/2002	Levendowski et al.
5,931,791 A	8/1999	Saltzstein et al.	6,505,077 B1	1/2003	Kast et al.
5,938,689 A	8/1999	Fischell et al.	6,510,340 B1	1/2003	Jordan
5,941,906 A	8/1999	Barreras et al.	6,511,424 B1	1/2003	Moore-Ede
5,950,632 A	9/1999	Reber et al.	6,529,774 B1	3/2003	Greene
5,957,861 A	9/1999	Combs et al.	6,534,693 B2	3/2003	Fischell et al.
5,971,594 A	10/1999	Sahai et al.	6,547,746 B1	4/2003	Marino
5,975,085 A	11/1999	Rise	6,549,804 B1	4/2003	Osorio et al.
5,978,702 A	11/1999	Ward et al.	6,553,262 B1	4/2003	Lang et al.
5,978,710 A	11/1999	Prutchi et al.	6,560,486 B1	5/2003	Osorio et al.
5,995,868 A	11/1999	Dorfmeister et al.	6,571,123 B2	5/2003	Ives et al.
6,006,124 A	12/1999	Fischell et al.	6,571,125 B2	5/2003	Thompson
6,016,449 A	1/2000	Fischell et al.	6,572,528 B2	6/2003	Rohan et al.
6,018,682 A	1/2000	Rise	6,587,719 B1	7/2003	Barrett et al.
6,033,073 A *	3/2000	Potapova et al. 351/211	6,591,132 B2	7/2003	Gotman et al.
6,042,548 A	3/2000	Giuffre	6,591,137 B1	7/2003	Fischell et al.
6,042,579 A	3/2000	Elsberry et al.	6,591,138 B1	7/2003	Fischell et al.
6,051,017 A	4/2000	Loeb et al.	6,597,954 B1	7/2003	Pless et al.
6,052,619 A	4/2000	John	6,600,956 B2	7/2003	Maschino
6,061,593 A *	5/2000	Fischell et al. 600/544	6,609,025 B2	8/2003	Barrett et al.
6,066,163 A	5/2000	John	6,618,623 B1	9/2003	Pless et al.
6,081,744 A	6/2000	Loos	6,620,415 B2	9/2003	Donovan
			6,622,036 B1	9/2003	Suffin
			6,622,038 B2	9/2003	Barrett et al.
			6,622,041 B2	9/2003	Terry et al.
			6,622,047 B2	9/2003	Barrett et al.

(56)

References Cited

U.S. PATENT DOCUMENTS

6,658,287	B1	12/2003	Litt et al.	2003/0167078	A1	9/2003	Weisner et al.
6,665,562	B2	12/2003	Gluckman et al.	2003/0174554	A1	9/2003	Dunstone et al.
6,668,191	B1	12/2003	Boveja	2003/0176806	A1	9/2003	Pineda et al.
6,671,555	B2	12/2003	Gielen	2003/0181955	A1	9/2003	Gielen
6,678,548	B1	1/2004	Echaz et al.	2003/0187621	A1	10/2003	Nikitin et al.
6,684,105	B2	1/2004	Cohen et al.	2003/0195574	A1	10/2003	Osorio et al.
6,687,538	B1	2/2004	Hrdlicka et al.	2003/0195588	A1	10/2003	Fischell et al.
6,735,467	B2	5/2004	Wilson	2003/0195602	A1	10/2003	Boling
6,760,626	B1	7/2004	Boveja	2004/0034368	A1	2/2004	Pless et al.
6,768,969	B1	7/2004	Nikitin et al.	2004/0039427	A1	2/2004	Barrett et al.
6,778,854	B2	8/2004	Puskas	2004/0039981	A1	2/2004	Riedl et al.
6,782,292	B2	8/2004	Whitehurst	2004/0054297	A1	3/2004	Wingeier et al.
6,879,859	B1	4/2005	Boveja	2004/0059761	A1	3/2004	Hively
6,893,395	B1	5/2005	Kraus et al.	2004/0068199	A1	4/2004	Echaz et al.
6,901,294	B1	5/2005	Whitehurst et al.	2004/0073273	A1	4/2004	Gluckman et al.
6,901,296	B1	5/2005	Whitehurst	2004/0077995	A1	4/2004	Ferek-Petric
6,912,419	B2	6/2005	Hill	2004/0078160	A1	4/2004	Frei et al.
6,921,538	B2	7/2005	Donovan	2004/0082984	A1	4/2004	Osorio et al.
6,921,541	B2	7/2005	Chasin et al.	2004/0087835	A1	5/2004	Hively
6,923,784	B2	8/2005	Stein	2004/0097802	A1	5/2004	Cohen
6,931,274	B2	8/2005	Williams	2004/0122281	A1	6/2004	Fischell et al.
6,934,580	B1	8/2005	Osorio	2004/0122335	A1	6/2004	Sackellares et al.
6,937,891	B2	8/2005	Leinders et al.	2004/0127810	A1	7/2004	Sackellares et al.
6,944,501	B1	9/2005	Pless	2004/0133119	A1	7/2004	Osorio et al.
6,950,706	B2	9/2005	Rodriquez	2004/0133120	A1 *	7/2004	Frei et al. 600/544
6,973,342	B1	12/2005	Swanson	2004/0133248	A1	7/2004	Frei et al.
6,990,372	B2	1/2006	Perron et al.	2004/0133390	A1	7/2004	Osorio et al.
7,010,351	B2	3/2006	Firlik et al.	2004/0138516	A1	7/2004	Osorio et al.
7,089,059	B1	8/2006	Pless	2004/0138517	A1	7/2004	Osorio et al.
7,117,108	B2	10/2006	Rapp et al.	2004/0138536	A1	7/2004	Frei et al.
7,174,212	B1	2/2007	Klehn et al.	2004/0138578	A1	7/2004	Pineda et al.
7,177,701	B1	2/2007	Pianca	2004/0138579	A1	7/2004	Deadwyler et al.
7,212,851	B2	5/2007	Donoghue et al.	2004/0138580	A1	7/2004	Frei et al.
7,231,254	B2	6/2007	DiLorenzo	2004/0138581	A1	7/2004	Frei et al.
7,373,198	B2	5/2008	Bibian et al.	2004/0138647	A1	7/2004	Osorio et al.
7,463,917	B2	12/2008	Martinez	2004/0138711	A1	7/2004	Osorio et al.
7,631,015	B2	12/2009	Gupta et al.	2004/0138721	A1	7/2004	Osorio et al.
7,805,196	B2	9/2010	Miesel et al.	2004/0147969	A1	7/2004	Mann et al.
7,881,798	B2	2/2011	Miesel et al.	2004/0152958	A1	8/2004	Frei et al.
8,055,348	B2	11/2011	Heruth et al.	2004/0153129	A1	8/2004	Pless et al.
2001/0051819	A1	12/2001	Fischell et al.	2004/0158119	A1	8/2004	Osorio et al.
2001/0056290	A1	12/2001	Fischell et al.	2004/0172089	A1	9/2004	Whitehurst et al.
2002/0002390	A1	1/2002	Fischell et al.	2004/0176359	A1	9/2004	Wermeling
2002/0035338	A1	3/2002	Dear et al.	2004/0181263	A1	9/2004	Balzer et al.
2002/0054694	A1	5/2002	Vachtsevanos et al.	2004/0199212	A1	10/2004	Fischell
2002/0072770	A1	6/2002	Pless	2004/0210269	A1	10/2004	Shalev et al.
2002/0072776	A1	6/2002	Osorio et al.	2004/0243146	A1	12/2004	Chesbrough et al.
2002/0072782	A1	6/2002	Osorio et al.	2004/0267152	A1	12/2004	Pineda et al.
2002/0077670	A1	6/2002	Archer et al.	2005/0004621	A1	1/2005	Boveja et al.
2002/0095099	A1	7/2002	Quyen et al.	2005/0010261	A1	1/2005	Luders et al.
2002/0099412	A1	7/2002	Fischell et al.	2005/0015128	A1	1/2005	Rezai et al.
2002/0103512	A1	8/2002	Echaz et al.	2005/0015129	A1	1/2005	Mische
2002/0109621	A1 *	8/2002	Khair et al. 341/174	2005/0021103	A1	1/2005	DiLorenzo
2002/0111542	A1	8/2002	Warkentin et al.	2005/0021104	A1	1/2005	DiLorenzo
2002/0116042	A1	8/2002	Boling	2005/0021105	A1	1/2005	Firlik et al.
2002/0126036	A1	9/2002	Flaherty et al.	2005/0021108	A1	1/2005	Klosterman et al.
2002/0147388	A1	10/2002	Mass et al.	2005/0021313	A1	1/2005	Nikitin et al.
2002/0169485	A1	11/2002	Pless et al.	2005/0027328	A1	2/2005	Greenstein
2002/0177882	A1	11/2002	DiLorenzo	2005/0033369	A1	2/2005	Badelt
2002/0188330	A1	12/2002	Gielen et al.	2005/0043772	A1	2/2005	Stahmann et al.
2003/0004428	A1	1/2003	Pless	2005/0043774	A1	2/2005	Devlin et al.
2003/0009207	A1	1/2003	Paspa et al.	2005/0049649	A1	3/2005	Luders et al.
2003/0013981	A1	1/2003	Gevens et al.	2005/0059867	A1	3/2005	Cheng
2003/0018367	A1	1/2003	DiLorenzo	2005/0070970	A1	3/2005	Knudson et al.
2003/0028072	A1	2/2003	Fischell et al.	2005/0075067	A1	4/2005	Lawson et al.
2003/0050549	A1	3/2003	Sochor	2005/0096710	A1	5/2005	Kieval
2003/0073917	A1	4/2003	Echaz et al.	2005/0113885	A1	5/2005	Haubrich et al.
2003/0074033	A1	4/2003	Pless et al.	2005/0119703	A1	6/2005	DiLorenzo
2003/0083716	A1	5/2003	Nicolelis et al.	2005/0124863	A1	6/2005	Cook
2003/0114886	A1	6/2003	Gluckman et al.	2005/0131493	A1	6/2005	Boveja et al.
2003/0144709	A1	7/2003	Zabara et al.	2005/0137640	A1	6/2005	Freeberg et al.
2003/0144711	A1	7/2003	Pless et al.	2005/0143786	A1	6/2005	Boveja
2003/0144829	A1	7/2003	Geatz et al.	2005/0143787	A1	6/2005	Boveja et al.
2003/0149457	A1	8/2003	Tcheng et al.	2005/0149123	A1	7/2005	Lesser et al.
2003/0158587	A1	8/2003	Esteller et al.	2005/0182308	A1	8/2005	Bardy
				2005/0182464	A1	8/2005	Schulte et al.
				2005/0187789	A1	8/2005	Hatlestad
				2005/0197590	A1	9/2005	Osorio et al.
				2005/0203366	A1	9/2005	Donoghue et al.

(56)

References Cited**U.S. PATENT DOCUMENTS**

2005/0203584 A1 9/2005 Twetan et al.
 2005/0209218 A1 9/2005 Meyerson et al.
 2005/0222626 A1 10/2005 DiLorenzo
 2005/0222641 A1 10/2005 Pless
 2005/0228461 A1 10/2005 Osorio et al.
 2005/0245971 A1 11/2005 Brockway et al.
 2006/0015034 A1 1/2006 Martinerie et al.
 2006/0111644 A1 5/2006 Gutttag et al.
 2006/0142822 A1 6/2006 Tulgar
 2006/0173510 A1 8/2006 Besio et al.
 2006/0200038 A1 9/2006 Savit et al.
 2006/0212092 A1 9/2006 Pless et al.
 2006/0212093 A1 9/2006 Pless et al.
 2006/0224191 A1 10/2006 Dilorenzo
 2006/0293720 A1 12/2006 DiLorenzo et al.
 2007/0073355 A1 3/2007 DiLorenzo
 2007/0142862 A1 6/2007 DiLorenzo
 2007/0149952 A1 6/2007 Bland et al.
 2007/0150024 A1 6/2007 Leyde et al.
 2007/0150025 A1 6/2007 DiLorenzo et al.
 2007/0162086 A1 7/2007 DiLorenzo
 2007/0167991 A1 7/2007 DiLorenzo
 2007/0250077 A1 10/2007 Skakoon et al.
 2008/0114417 A1 5/2008 Leyde
 2008/0161712 A1 7/2008 Leyde
 2008/0161713 A1 7/2008 Leyde et al.
 2010/0023089 A1 1/2010 DiLorenzo
 2011/0319785 A1 12/2011 Snyder et al.

FOREIGN PATENT DOCUMENTS

CA 2428116 5/2002
 CA 2428383 5/2002
 CA 2425122 6/2002
 CA 2425004 8/2002
 CA 2456443 1/2003
 CA 2491987 1/2004
 DE 69832022 12/2005
 EP 0124663 A1 11/1984
 EP 0898460 3/1999
 EP 1017313 7/2000
 EP 1107693 6/2001
 EP 1145735 A2 10/2001
 EP 1145736 A2 10/2001
 EP 1292900 3/2003
 EP 1307260 5/2003
 EP 1331967 8/2003
 EP 1335668 8/2003
 EP 1341580 9/2003
 EP 1404216 4/2004
 EP 1333753 9/2004
 EP 1525551 4/2005
 EP 1558121 8/2005
 EP 1558128 8/2005
 EP 1558130 8/2005
 EP 1558131 8/2005
 EP 1558132 8/2005
 EP 1558330 8/2005
 EP 1558334 8/2005
 EP 1562674 8/2005
 EP 0911061 B1 10/2005
 EP 1609414 A2 12/2005
 JP 24033673 A2 2/2004
 SU 1074484 2/1984
 WO WO 85/01213 A1 3/1985
 WO WO 92/00119 A1 1/1992
 WO WO 97/26823 A1 7/1997
 WO WO 97/34522 A1 9/1997
 WO WO 97/34524 A1 9/1997
 WO WO 97/34525 A1 9/1997
 WO WO 97/39797 A1 10/1997
 WO WO 97/42990 A1 11/1997
 WO WO 97/45160 A1 12/1997
 WO WO 98/49935 A1 11/1998

WO WO 99/20342 A1 4/1999
 WO WO 99/56821 A1 11/1999
 WO WO 99/56822 A1 11/1999
 WO WO 00/07494 A2 2/2000
 WO WO 00/10455 3/2000
 WO WO 01/41867 A1 6/2001
 WO WO 01/48676 A1 7/2001
 WO WO 01/49364 A2 7/2001
 WO WO 01/67288 A2 9/2001
 WO WO 01/75660 A1 10/2001
 WO WO 02/09610 A1 2/2002
 WO WO 02/09811 A1 2/2002
 WO WO 02/36003 A1 5/2002
 WO WO 02/38031 A2 5/2002
 WO WO 02/38217 A2 5/2002
 WO WO 02/49500 A2 6/2002
 WO WO 02/058536 A2 8/2002
 WO WO 02/067122 A1 8/2002
 WO WO 03/001996 A2 1/2003
 WO WO 03/009207 A1 1/2003
 WO WO 03/030734 A2 4/2003
 WO WO 03/035165 A1 5/2003
 WO WO 03/084605 A1 10/2003
 WO WO 2004/008373 A2 1/2004
 WO WO 2004/032720 A2 4/2004
 WO WO 2004/034231 A2 4/2004
 WO WO 2004/034879 A2 4/2004
 WO WO 2004/034880 A2 4/2004
 WO WO 2004/034881 A2 4/2004
 WO WO 2004/034882 A2 4/2004
 WO WO 2004/034883 A2 4/2004
 WO WO 2004/034885 A2 4/2004
 WO WO 2004/034982 A2 4/2004
 WO WO 2004/034997 A2 4/2004
 WO WO 2004/034998 A2 4/2004
 WO WO 2004/035130 A2 4/2004
 WO WO 2004/036370 A2 4/2004
 WO WO 2004/036372 A2 4/2004
 WO WO 2004/036376 A2 4/2004
 WO WO 2004/036377 A2 4/2004
 WO WO 2004/037342 A2 5/2004
 WO WO 2004/043536 A1 5/2004
 WO WO 2004/091718 A1 10/2004
 WO WO 2005/007236 A2 1/2005
 WO WO 2005/028026 A1 3/2005
 WO WO 2005/028028 A1 3/2005
 WO WO 2005/031630 A2 4/2005
 WO WO 2005/051167 A1 6/2005
 WO WO 2005/051306 A2 6/2005

OTHER PUBLICATIONS

Bland et al.; U.S. Appl. No. 12/180,996 entitled "Patient advisory device," filed Jul. 28, 2008.

Adjouadi, et al. A new mathematical approach based on orthogonal operators for the detection of interictal spikes in epileptogenic data. Biomed. Sci. Instrum. 2004; 40: 175-80.

Adjouadi, et al. Interictal spike detection using the Walsh transform. IEEE Trans. Biomed. Eng. 2004; 51(5): 868-72.

Aksenova, et al. Nonparametric on-line detection of changes in signal spectral characteristics for early prediction of epilepsy seizure onset. J. Automation and Information Sciences. 2004; 36(8): 35-45.

Aksenova, et al. On-line disharmony detection for early prediction of epilepsy seizure onset. 5th International Workshop Neural Coding 2003. Aulla (Italy) Sep. 20-25, 2003. (Abstract).

Andrzejak, et al. Bivariate surrogate techniques: necessity, strengths, and caveats. Physical Review E. 2003; 68: 066202-1-066202-15.

Andrzejak, et al. Testing the null hypothesis of the nonexistence of a preseizure state. Physical Review E. 2003; 67: 010901-1-010901-4.

Aschenbrenner-Scheibe, et al. How well can epileptic seizures be predicted? An evaluation of a nonlinear method. Brain. 2003; 126: 2616-26.

Baruchi, et al. Functional holography of recorded neuronal networks activity. Neuroinformatics. 2004; 2(3): 333-51.

Ben-Hur, et al. Detecting stable clusters using principal component analysis. Methods Mol. Biol. 2003; 224: 159-82.

(56)

References Cited

OTHER PUBLICATIONS

- Bergey, et al. Epileptic seizures are characterized by changing signal complexity. *Clin. Neurophysiol.* 2001; 112(2): 241-9.
- Betterton, et al. Determining State of Consciousness from the Intracranial Electroencephalogram (IEEG) for Seizure Prediction. From Proceeding (377) Modeling, Identification, and Control. 2003; 377-201: 313-317.
- Bhattacharya, et al. Enhanced phase synchrony in the electroencephalograph gamma band for musicians while listening to music. *Phys. Rev. E.* 2001; 64:012902-1-4.
- Boley, et al. Training Support Vector Machine using Adaptive Clustering. 2004 SIAM International Conference on Data Mining, Apr. 22-Apr. 24, 2004. Lake Buena Vista, FL, USA. 12 pages.
- Cao, et al. Detecting dynamical changes in time series using the permutation entropy. *Physical Review E.* 2004; 70:046217-1-046217-7.
- Carretero-Gonzalez, et al. Scaling and interleaving of subsystem Lyapunov exponents for spatio-temporal systems. *Chaos.* 1999; 9(2): 466-482.
- Cerf, et al. Criticality and synchrony of fluctuations in rhythmical brain activity: pretransitional effects in epileptic patients. *Biol. Cybern.* 2004; 90(4): 239-55.
- Chaovalitwongse, et al. EEG Classification in Epilepsy. *Annals.* 2004; 2 (37): 1-31.
- Chavez, et al. Spatio-temporal dynamics prior to neocortical seizures: amplitude versphase couplings. *IEEE Trans. Biomed. Eng.* 2003; 50(5):571-83.
- D'Alessandro, et al. Epileptic seizure prediction using hybrid feature selection over multiple intracranial EEG electrode contacts: a report of four patients. *IEEE Trans. Biomed. Eng.* 2003; 50(5): 603-15.
- Drury, et al. Seizure prediction using scalp electroencephalogram. *Exp. Neurol.* 2003; 184 Suppl 1: S9-18.
- Elger, et al. Nonlinear EEG analysis and its potential role in epileptology. *Epilepsia.* 2000; 41 Suppl 3: S34-8.
- Esteller, et al. A Comparison of Waveform Fractal Dimension Algorithms. *IEEE Transactions on Circuits and Systems.* 2001; vol. 48(2): 177-183.
- Esteller, et al. Feature Parameter Optimization for Seizure Detection/prediction. Proceedings of the 23rd Annual International Conference of the IEEE Engineering in Medicine and Biology Society, Istanbul, Turkey. Oct. 2001.
- Fell, et al. Linear inverse filtering improves spatial separation of nonlinear brain dynamics: a simulation study. *J. Neurosci. Methods.* 2000; 98(1): 49-56.
- Fisher et al. 1999. Reassessment: Vag nerve stimulation for epilepsy, A report of the therapeutics and technology assessment subcommittee of the American Academy of Neurology. *Neurology.* 53: 666-669.
- Franszczuk et al.; An autoregressive method for the measurement of synchronization of interictal and ictal EEG signals; *Biological Cybernetics*, vol. 81; No. 1; pp. 3-9; 1999.
- Gardner, A. B. A Novelty Detection Approach to Seizure Analysis from Intracranial EEG. Georgia Institute of Technology. Apr. 2004. A dissertation available at http://etd.gatech.edu/theses/available/etd-04122004-132404/unrestricted/gardner_andrew_b_200405_phd.pdf. Accessed Feb. 28, 2006.
- Gigola, et al. Prediction of epileptic seizures using accumulated energy in a multiresolution framework. *J. Neurosci. Methods.* 2004; 138(1-2): 107-111.
- Guyon, I. An introduction to variable and feature selection. *Journal of Machine Learning Research.* 2003; 3:1157-1182.
- Guyon, I. Multivariate Non-Linear Feature Selection with Kernel Multiplicative Updates and Gram-Schmidt Relief. BISC FLINT-CIBI 2003 Workshop. Berkeley. 2003; p. 1-11.
- Hively, et al. Channel-consistent forewarning of epileptic events from scalp EEG. *IEEE Trans. Biomed. Eng.* 2003; 50(5): 584-93.
- Hively, et al. Detecting dynamical changes in nonlinear time series. *Physics Letters A.* 1999; 258: 103-114.
- Hively, et al. Epileptic Seizure Forewarning by Nonlinear Techniques. ORNL/TM-2000/333 Oak Ridge National Laboratory. Nov. 2000. Available at http://computing.ornl.gov/cse_home/staff/hively/NBICradaAnnualRpt FY00.pdf. Accessed Feb. 28, 2006.
- Hsu, et al. A practical guide to support vector classification. Technical report, Department of Computer Science and Information Technology, National Taiwan University, 2003. Available at <http://www.csie.ntu.edu.tw/~cjlin/papers/guide/guide.pdf>. Accessed Feb. 28, 2006.
- Iasemidis, et al. Adaptive epileptic seizure prediction system. *IEEE Trans. Biomed. Eng.* 2003; 50(5):616-27.
- Iasemidis, et al. Measurement and Quantification of Spatio-Temporal Dynamics of Human Epileptic Seizures. In: *Nonlinear Signal Processing in Medicine*, Ed. M. Akay, IEEE Press. 1999; pp. 1-27.
- Iasemidis, et al. Quadratic binary programming and dynamic system approach to determine the predictability of epileptic seizures. *Journal of Combinatorial Optimization.* 2001; 5: 9-26.
- Iasemidis, L. D. Epileptic seizure prediction and control. *IEEE Trans. Biomed. Eng.* 2003; 50(5):549-58.
- Jerger, et al. Early seizure detection. *Journal of Clin. Neurophysiol.* 2001; 18(3):259-68.
- Jouny, et al. Characterization of epileptic seizure dynamics using Gabor atom density. *Clin. Neurophysiol.* 2003; 114(3):426-37.
- Korn, et al. Is there chaos in the brain? II. Experimental evidence and related models. *C. R. Biol.* 2003; 326(9):787-840.
- Kreuz, et al. Measure profile surrogates: a method to validate the performance of epileptic seizure prediction algorithms. *Phys. Rev. E.* 2004; 69(6 Pt 1):061915-1-9.
- Lachaux, et al. Measuring phase synchrony in brain signals. *Hum. Brain Mapp.* 1999; 8(4):194-208.
- Lai, et al. Controlled test for predictive power of Lyapunov exponents: their inability to predict epileptic seizures. *Chaos.* 2004; 14(3):630-42.
- Lai, et al. Inability of Lyapunov exponents to predict epileptic seizures. *Phys. Rev. Lett.* 2003; 91(6):068102-1-4.
- Latka, et al. Wavelet analysis of epileptic spikes. *Phys. Rev. E.* 2003; 67 (5 Pt 1):052902 (6 pages).
- Le Van Quyen, et al. Anticipating epileptic seizures in real time by a non-linear analysis of similarity between EEG recordings. *Neuroreport.* 1999; 10(10):2149-55.
- Le Van Quyen, et al. Author's second reply. *The Lancet.* 2003; 361:971.
- Le Van Quyen, et al. Comparison of Hilbert transform and wavelet methods for the analysis of neuronal synchrony. *J. Neurosci. Methods.* 2001; 111(2):83-98.
- Le Van Quyen, et al. Nonlinear analyses of interictal EEG map the brain interdependences in human focal epilepsy. *Physica D.* 1999; 127:250-266.
- Lehnertz, et al. Nonlinear EEG analysis in epilepsy: its possible use for interictal focus localization, seizure anticipation, and prevention. *J. Clin. Neurophysiol.* 2001; 18(3):209-22.
- Lehnertz, et al. Seizure prediction by nonlinear EEG analysis. *IEEE Eng. Med. Biol. Mag.* 2003; 22(1):57-63.
- Lehnertz, K. Non-linear time series analysis of intracranial EEG recordings in patients with epilepsy—an overview. *Int. J. Psychophysiol.* 1999; 34(1):45-52.
- Li, et al. Linear and nonlinear measures and seizure anticipation in temporal lobe epilepsy. *J. Comput. Neurosci.* 2003; 15(3):335-45.
- Li, et al. Non-linear, non-invasive method for seizure anticipation in focal epilepsy. *Math. Biosci.* 2003; 186(1):63-77.
- Litt, et al. Prediction of epileptic seizures. *Lancet Neurol.* 2002; 1(1):22-30.
- Litt, et al. Seizure prediction and the pre seizure period. *Curr. Opin. Neurol.* 2002; 15(2):173-7.
- Maiwald, et al. Comparison of three nonlinear seizure prediction methods by means of the seizure prediction characteristic. *Physica D.* 2004; 194:357-368.
- Mangasarian, et al. Lagrangian Support Vector Machines. *Journal of Machine Learning Research.* 2001; 1:161-177.
- McSharry, et al. Comparison of predictability of epileptic seizures by a linear and a nonlinear method. *IEEE Trans. Biomed. Eng.* 2003; 50(5):628-33.
- McSharry, et al. Linear and non-linear methods for automatic seizure detection in scalp electro-encephalogram recordings. *Med. Biol. Eng. Comput.* 2002; 40(4):447-61.

(56)

References Cited

OTHER PUBLICATIONS

- McSharry, P. E. Detection of dynamical transitions in biomedical signals using nonlinear methods. *Lecture Notes in Computer Science* 2004; 3215:483-490.
- Meng, et al. Gaussian mixture models of ECoG signal features for improved detection of epileptic seizures. *Med. Eng. Phys.* 2004; 26(5):379-93.
- Mizuno-Matsumoto, et al. Wavelet-crosscorrelation analysis can help predict whether bursts of pulse stimulation will terminate after discharges. *Clin. Neurophysiol.* 2002; 113(1):33-42.
- Mormann, et al. Automated detection of a pre-seizure state based on a decrease in synchronization in intracranial electroencephalogram recordings from epilepsy patients. *Phys. Rev. E.* 2003; 67(2 Pt 1):021912-1-10.
- Mormann, et al. Epileptic seizures are preceded by a decrease in synchronization. *Epilepsy Res.* 2003; 53(3):173-85.
- Mormann, et al. Mean phase coherence as a measure for phase synchronization and its application to the EEG of epilepsy patients. *Physica D.* 2000; 144:358-369.
- Navarro, et al. Seizure anticipation in human neocortical partial epilepsy. *Brain.* 2002; 125:640-55.
- Niederhauser, et al. Detection of seizure precursors from depth-EEG using a sign periodogram transform. *IEEE Trans. Biomed. Eng.* 2003; 50(4):449-58.
- Nigam, et al. A neural-network-based detection of epilepsy. *Neurological Research.* 2004; 26(1):55-60.
- Osorio, et al. Performance reassessment of a real-time seizure-detection algorithm on long ECoG series. *Epilepsia.* 2002; 43(12):1522-35.
- Pflieger, et al. A noninvasive method for analysis of epileptogenic brain connectivity. Presented at the American Epilepsy Society 2004 Annual Meeting, New Orleans. Dec. 6, 2004. *Epilepsia.* 2004; 45(Suppl. 7):70-71.
- Platt, et al. Large Margin DAGs for Multiclass Classification. S.A. Solla. T.K. Leen and K. R. Muller (eds.). 2000; pp. 547-553.
- Platt, J. C. Using Analytic QP and Sparseness to Speed Training of Support Vector Machines. *Advances in Neural Information Processing Systems.* 1999; 11:557-563.
- Protopopescu, et al. Epileptic event forewarning from scalp EEG. *J. Clin. Neurophysiol.* 2001; 18(3):223-45.
- Sackellares, J. C. Epilepsy—when chaos fails. In: *chaos in the brain?* Eds. K. Lehnertz & C.E. Elger. World Scientific. 2000 (22 pages).
- Schindler, et al. EEG analysis with simulated neuronal cell models helps to detect pre-seizure changes. *Clin. Neurophysiol.* 2002; 113(4):604-14.
- Sheridan, T. *Humans and Automation.* NY: John Wiley. 2002.
- Shoeb et al. Patient-specific seizure detection. MIT Computer Science and Artificial Intelligence Laboratory. 2004; pp. 193-194.
- Staba, et al. Quantitative analysis of high-frequency oscillations (80-500 Hz) recorded in human epileptic hippocampal and entorhinal cortex. *J. Neurophysiol.* 2002; 88(4):1743-52.
- Stefanski, et al. Using chaos synchronization to estimate the largest Lyapunov exponent of nonsmooth systems. *Discrete Dynamics in Nature and Society.* 2000; 4:207-215.
- Terry, et al. An improved algorithm for the detection of dynamical interdependence in bivariate time-series. *Biol. Cybern.* 2003; 88(2):129-36.
- Tetzlaff, et al. Cellular neural networks (CNN) with linear weight functions for a prediction of epileptic seizures. *Int'l. J. of Neural Systems.* 2003; 13(6):489-498.
- Van Drongelen, et al. Seizure anticipation in pediatric epilepsy: use of Kolmogorov entropy. *Pediatr. Neurol.* 2003; 29(3): 207-13.
- Van Putten, M. Nearest neighbor phase synchronization as a measure to detect seizure activity from scalp EEG recordings. *J. Clin. Neurophysiol.* 2003; 20(5):320-5.
- Venugopal, et al. A new approach towards predictability of epileptic seizures: KLT dimension. *Biomed Sci. Instrum.* 2003; 39:123-8.
- Vonck, et al. Long-term amygdalohippocampal stimulation for refractory temporal lobe epilepsy. *Ann. Neurol.* 2002; 52(5):556-65.
- Vonck, et al. Neurostimulation for refractory epilepsy. *Acta. Neurol. Belg.* 2003; 103(4):213-7.
- Widman, et al. Reduced signal complexity of intracellular recordings: a precursor for epileptiform activity? *Brain Res.* 1999; 836(1-2):156-63.
- Winterhalter, et al. The seizure prediction characteristic: a general framework to assess and compare seizure prediction methods. *Epilepsy Behav.* 2003; 4(3):318-25.
- Yang et al.; Testing whether a prediction scheme is better than guess; Ch. 14 in *Quantitative Neuroscience: Models, Algorithms, Diagnostics, and Therapeutic Applications*; pp. 252-262; 2004.
- Yang, et al. Relation between Responsiveness to Neurotransmitters and Complexity of Epileptiform Activity in Rat Hippocampal CA1 Neurons. *Epilepsia.* 2002; 43(11):1330-1336.
- Zhang, et al. High-resolution EEG: cortical potential imaging of interictal spikes. *Clin. Neurophysiol.* 2003; 114(10):1963-73.
- Leyde et al.; U.S. Appl. No. 12/020,507 entitled "Methods and systems for measuring a subject's susceptibility to a seizure," filed Jan. 25, 2008.
- Snyder et al.; U.S. Appl. No. 12/020,450 entitled "Systems and methods for identifying a contra-ictal condition in a subject," filed Jan. 25, 2008.
- Snyder et al.; U.S. Appl. No. 12/035,335 entitled "Methods and systems for characterizing and generating a patient-specific seizure prediction system," filed Feb. 21, 2008.
- Snyder et al.; U.S. Appl. No. 12/053,312 entitled "Implantable systems and methods for identifying a contra-ictal condition in a subject," filed Mar. 21, 2008.
- Leyde et al.; U.S. Appl. No. 12/343,376 entitled "Systems and method for recording clinical manifestations of a seizure," filed Dec. 23, 2008.
- Brown et al.; U.S. Appl. No. 12/343,386 entitled "Housing for an implantable medical device," filed Dec. 23, 2008.
- Himes, David M.; U.S. Appl. No. 12/630,300 entitled "Universal Electrode Array for Monitoring Brain Activity," filed Dec. 3, 2009.
- Himes et al.; U.S. Appl. No. 12/646,783 entitled "Brain State Analysis Based on Select Seizure Onset Characteristics and Clinical Manifestations," filed Dec. 23, 2009.
- Echaz et al.; U.S. Appl. No. 12/649,098 entitled "Processing for Multi-Channel Signals," filed Dec. 29, 2009.
- Floyd et al.; U.S. Appl. No. 12/685,543 entitled "Medical Lead Termination Sleeve for Implantable Medical Devices," filed Jan. 11, 2010.
- Harris et al.; U.S. Appl. No. 12/691,650 entitled "Minimally invasive system for selecting patient-specific therapy parameters," filed Jan. 21, 2010.
- Rimes et al.; U.S. Appl. No. 12/716,132 entitled "Displaying and Manipulating Brain Function Data Including Enhanced Data Scrolling Functionality," filed Mar. 2, 2010.
- Himes et al.; U.S. Appl. No. 12/716,147 entitled "Displaying and Manipulating Brain Function Data Including Filtering of Annotations," filed Mar. 2, 2010.
- Rothman et al.; *Local Cooling: a therapy for intractable neocortical epilepsy*; *Epilepsy Currents*; vol. 3; No. 5; pp. 153-156; Sep./Oct. 2003.
- DiLorenzo, Daniel, U.S. Appl. No. 10/858,899, entitled "Closed-loop feedback-driven neuromodulation," filed Jun. 1, 2004.
- DiLorenzo, Daniel, U.S. Appl. No. 11/706,630, entitled "Methods and systems for administering an appropriate pharmacological treatment to a patient for managing epilepsy and other neurological disorders," filed Feb. 14, 2007.
- DiLorenzo, Daniel, U.S. Appl. No. 11/743,607, entitled "Controlling a Subject's Susceptibility to a Seizure," filed May 2, 2007.
- DiLorenzo, Daniel, U.S. Appl. No. 11/743,611, entitled "Providing Output Indicative of Subject's Disease State," filed May 2, 2007.
- DiLorenzo, Daniel, U.S. Appl. No. 11/282,317 entitled "Closed-loop vagus nerve stimulation," filed Nov. 17, 2005.
- Harris et al.; U.S. Appl. No. 11/766,742, entitled "Minimally Invasive Monitoring Systems," filed Jun. 21, 2007.
- Harris et al.; U.S. Appl. No. 11/766,751, entitled "Minimally Invasive Monitoring Methods," filed Jun. 21, 2007.
- Harris et al.; U.S. Appl. No. 11/766,756, entitled "Methods and Systems for Facilitating Clinical Trials," filed Jun. 21, 2007.

(56)

References Cited

OTHER PUBLICATIONS

- Harris et al; U.S. Appl. No. 11/766,760, entitled "Minimally Invasive System for Selecting Patient-Specific Therapy Parameters," filed Jun. 21, 2007.
- Harris et al; U.S. Appl. No. 11/766,761, entitled "Minimally Invasive Monitoring Systems for Monitoring a Patient's Propensity for a Neurological Event," filed Jun. 21, 2007.
- Harris, John, U.S. Appl. No. 11/734,190, entitled "Methods and Template Assembly for Implanting an Electrode Array in a Patient," filed Apr. 11, 2007.
- Bangham et al. Diffusion of univalent ions across the lamellae of swollen phospholipids. 1965. *J. Mol. Biol.* 13: 238-252.
- Burges, C. A Tutorial on Support Vector Machines for Pattern Recognition. *Data Mining and Knowledge Discovery*. 1998; 2: 121-167.
- Casdagli, et al. Characterizing nonlinearity in invasive EEG recordings from temporal lobe epilepsy. *Physica D*. 1996; 99 (2/3): 381-399.
- Casdagli, et al. Nonlinear Analysis of Mesial Temporal Lobe Seizures Using a Surrogate Data Technique. *Epilepsia*. 1995; 36, suppl. 4, pp. 142.
- Casdagli, et al. Non-linearity in invasive EEG recordings from patients with temporal lobe epilepsy. *Electroencephalogr. Clin. Neurophysiol.* 1997; 102(2): 98-105.
- Crichton, Michael, "Terminal Man", 1972, Ballantine Books, NY, NY, pp. 21-24, 32-33, 70-71, and 74-81.
- Ebersole, J. S. Functional neuroimaging with EEG source models to localize epileptogenic foci noninvasively. *Neurology*. Available at http://www.uchospitals.edu/pdf/uch_001471.pdf. Accessed Feb. 28, 2006.
- Elbert et al. Chaos and Physiology: Deterministic Chaos in Excitable Cell Assemblies. *Physiological Reviews*. 1994; 74(1):1-47.
- Elger, et al. Seizure prediction by non-linear time series analysis of brain electrical activity. *Eur. J. Neurosci.* 1998; 10(2): 786-789.
- Fein, et al. Common reference coherence data are confounded by power and phase effects. *Electroencephalogr. Clin. Neurophysiol.* 1988; 69:581-584.
- Geva, et al. Forecasting generalized epileptic seizures from the EEG signal by wavelet analysis and dynamic unsupervised fuzzy clustering. *IEEE Trans. Biomed. Eng.* 1998; 45(10): 1205-16.
- Hearst M. Trends & Controversies: Support Vector Machines. *IEEE Intelligent Systems*. 1998; 13: 18-28.
- Hjorth, B. Source derivation simplifies topographical EEG interpretation. *Am. J. EEG Technol.* 1980; 20: 121-132.
- Iasemidis, et al. Automated Seizure Prediction Paradigm. *Epilepsia*. 1998; vol. 39, pp. 56.
- Iasemidis, et al. Chaos Theory and Epilepsy. *The Neuroscientist*. 1996; 2:118-126.
- Iasemidis, et al. Detection of the Preictal Transition State in Scalp-Sphenoidal EEG Recordings. *American Clinical Neurophysiology Society Annual Meeting*, Sep. 1996. pp. C206.
- Iasemidis, et al. Dynamical Interaction of the Epileptogenic Focus with Extrafocal Sites in Temporal Lobe Epilepsy (TLE). *Ann. Neurol.* 1997; 42, pp. 429. pp. M146.
- Iasemidis, et al. Epileptogenic Focus Localization by Dynamical Analysis of Interictal Periods of EEG in Patients with Temporal Lobe Epilepsy. *Epilepsia*. 1997; 38, suppl. 8, pp. 213.
- Iasemidis, et al. Localizing Preictal Temporal Lobe Spike Foci Using Phase Space Analysis. *Electroencephalography and Clinical Neurophysiology*. 1990; 75, pp. S63-64.
- Iasemidis, et al. Long-Time-Scale Temporo-spatial Patterns of Entrainment of Preictal Electroencephalographic Data in Human Temporal Lobe Epilepsy. *Epilepsia*. 1990; 31(5):621.
- Iasemidis, et al. Modelling of ECoG in temporal lobe epilepsy. *Biomed. Sci. Instrum.* 1988; 24: 187-93.
- Iasemidis, et al. Nonlinear Dynamics of ECoG Data in Temporal Lobe Epilepsy. *Electroencephalography and Clinical Neurophysiology*. 1998; 5, pp. 339.
- Iasemidis, et al. Phase space topography and the Lyapunov exponent of electrocorticograms in partial seizures. *Brain Topogr.* 1990; 2(3): 187-201.
- Iasemidis, et al. Preictal Entrainment of a Critical Cortical Mass is a Necessary Condition for Seizure Occurrence. *Epilepsia*. 1996; 37, suppl. 5, pp. 90.
- Iasemidis, et al. Preictal-Postictal Versus Postictal Analysis for Epileptogenic Focus Localization. *J. Clin. Neurophysiol.* 1997; 14, pp. 144.
- Iasemidis, et al. Quantification of Hidden Time Dependencies in the EEG within the Framework of Non-Linear Dynamics. *World Scientific*. 1993; pp. 30-47.
- Iasemidis, et al. Spatiotemporal dynamics of human epileptic seizures. *World Scientific*. 1996; pp. 26-30.
- Iasemidis, et al. Spatiotemporal Evolution of Dynamical Measures Precedes Onset of Mesial Temporal Lobe Seizures. *Epilepsia*. 1994; 358, pp. 133.
- Iasemidis, et al. Spatiotemporal Transition to Epileptic Seizures: A Nonlinear Dynamical Analysis of Scalp and Intracranial EEG Recordings. (In SILVA, F.L. *Spatiotemporal Models in Biological and Artificial Systems*. Ohmsha IOS Press. 1997; 37, pp. 81-88.).
- Iasemidis, et al. The evolution with time of the spatial distribution of the largest Lyapunov exponent on the human epileptic cortex. *World Scientific*. 1991; pp. 49-82.
- Iasemidis, et al. The Use of Dynamical Analysis of EEG Frequency Content in Seizure Prediction. *American Electroencephalographic Society Annual Meeting*, Oct. 1993.
- Iasemidis, et al. Time Dependencies in Partial Epilepsy. 1993; 34, pp. 130-131.
- Iasemidis, et al. Time dependencies in the occurrences of epileptic seizures. *Epilepsy Res.* 1994; 17(1): 81-94.
- Katz, et al. Does interictal spiking change prior to seizures? *Electroencephalogr. Clin. Neurophysiol.* 1991; 79 (2):153-6.
- Kraskov, A. Synchronization and Interdependence Measures and Their Application to the Electroencephalogram of Epilepsy Patients and Clustering of Data. Available at <http://www.kfa-juelich.de/nic-series/volume24/nic-series-band24.pdf>. Accessed Apr. 17, 2006 (106 pp).
- Lemos, et al. The weighted average reference montage. *Electroencephalogr. Clin. Neurophysiol.* 1991; 79 (5):361-70.
- Martinerie, et al. Epileptic seizures can be anticipated by non-linear analysis. *Nat. Med.* 1998; 4(10):1173-6.
- Osorio, et al. Real-time automated detection and quantitative analysis of seizures and short-term prediction of clinical onset. *Epilepsia*. 1998; 39(6):615-27.
- Rahimi, et al. On the Effectiveness of Aluminum Foil Helmets: An Empirical Study. Available at <http://people.csail.mit.edu/rahimi/helmet/>. Accessed Mar. 2, 2006.
- Remington's Pharmaceutical Sciences, latest edition, Mack Publishing Co., Easton P.
- Robinson, et al. Steady States and Global Dynamics of Electrical Activity in the Cerebral Cortex. *Phys. Rev. E*. 1998; (58):3557-3571.
- Sackellares et al. Computer-Assisted Seizure Detection Based on Quantitative Dynamical Measures. *American Electroencephalographic Society Annual Meeting*, Sep. 1994.
- Sackellares et al. Dynamical Studies of Human Hippocampal Limbic Epilepsy. *Neurology*. 1995; 45, Suppl. 4, pp. A 404.
- Sackellares et al. Epileptic Seizures as Neural Resetting Mechanisms. *Epilepsia*. 1997; vol. 38, Sup. 3.
- Sackellares et al. Measurement of Chaos to Localize Seizure Onset. *Epilepsia*. 1989; 30 (5):663.
- Sackellares et al. Relationship Between Hippocampal Atrophy and Dynamical Measures of EEG in Depth Electrode Recordings. *American Electroencephalographic Society Annual Meeting*, Sep. 1995. pp. A105.
- Salant, et al. Prediction of epileptic seizures from two-channel EEG. *Med. Biol. Eng. Comput.* 1998; 36(5):549-56.
- Schwartzkroin, P. Origins of the Epileptic State. *Epilepsia*. 1997; 38, supply. 8, pp. 853-858.
- Szoka et al. Procedure for preparation of liposomes with large internal aqueous space and high capture volume by reverse phase evaporation. 1978. *Proc. Natl Acad. Sci. USA*. 75: 4194-4198.
- Tass, et al. Detection of n: m Phase Locking from Noisy Data: Application to Magnetoencephalography. *Physical Review Letters*. 1998; 81(15):3291-3294.

(56)

References Cited

OTHER PUBLICATIONS

Theiler, et al. Testing for non-linearity in time series: the method of surrogate data. *Physica D*. 1992; 58:77-94.

Tsakalis, K. S. Prediction and control of epileptic seizures: Coupled oscillator models. Arizona State University. (Slide: 53 pages) (No date).

Weiss, P. Seizure prelude found by chaos calculation. *Science News*. 1998; 153(20):326.

Yang, et al. A supervised feature subset selection technique for multivariate time series. Available at <http://infolab.usc.edu/DocsDemos/fsdm05.pdf>. Accessed Mar. 2, 2006.

Yang, et al. CLe Ver: A feature subset selection technique for multivariate time series. T. B. Ho, D. Cheung, and H. Liu (Eds.): *PAKDD*. 2005; *LNAI* 3518: 516-522.

Yatsenko, et al. Geometric Models, Fiber Bundles, and Biomedical Applications. *Proceedings of Institute of Mathematics of NAS of Ukraine*. 2004; 50 (Part 3):1518R1525.

Zaveri et al. Time-Frequency Analyses of Nonstationary Brain Signals. *Electroencephalography and Clinical Neurophysiology*. 1991; 79, pp. 28P-29P.

DiLorezo, Daniel; U.S. Appl. No. 12/774,550 entitled "Systems for Monitoring a Patient's Neurological Disease State," filed May 5, 2010.

Echaz et al.; U.S. Appl. No. 12/792,582 entitled "Processing for Multi-Channel Signals," filed Jun. 2, 2010.

Higgins et al.; U.S. Appl. No. 13/026,961 entitled "Neurological monitoring and alerts," filed Feb. 14, 2011.

Harris et al.; U.S. Appl. No. 13/050,839 entitled "System and methods for analyzing seizure activity," filed Mar. 17, 2011.

Leyde et al.; U.S. Appl. No. 13/070,333 entitled "Communication Error Alerting in an Epilepsy Monitoring System," filed Mar. 23, 2011.

Leyde et al.; U.S. Appl. No. 13/070,357 entitled "Patient Entry Recording in an Epilepsy Monitoring System," filed Mar. 23, 2011.

Chen et al.; Clinical utility of video-EEG monitoring; *Pediatric Neurology*; vol. 12; No. 3; pp. 220-224; 1995.

Spector et al.; High and Low Perceived Self-Control of Epileptic Seizures; *Epilepsia*, vol. 42(4), Apr. 2001; pp. 556-564.

* cited by examiner

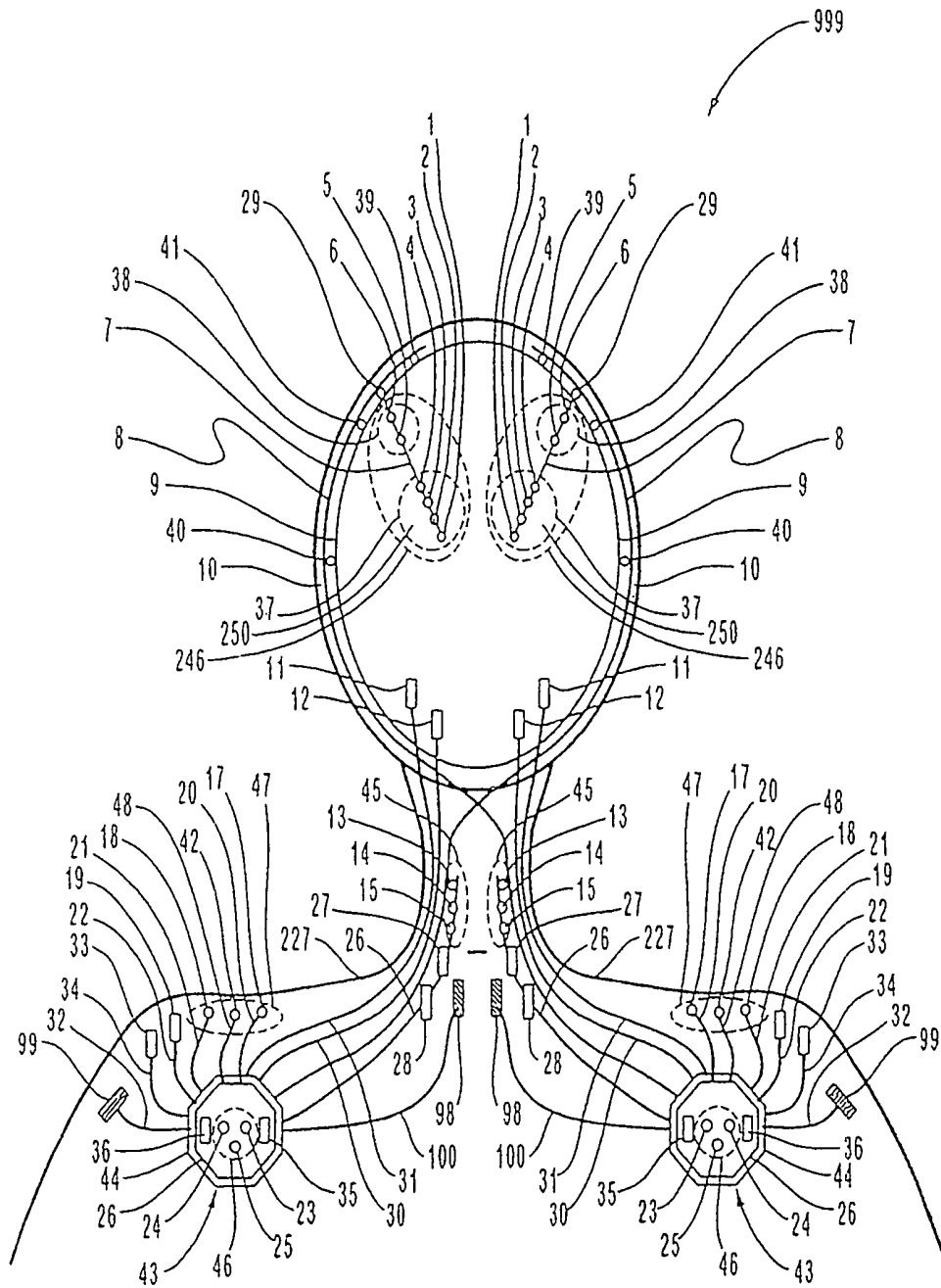


FIG. 1

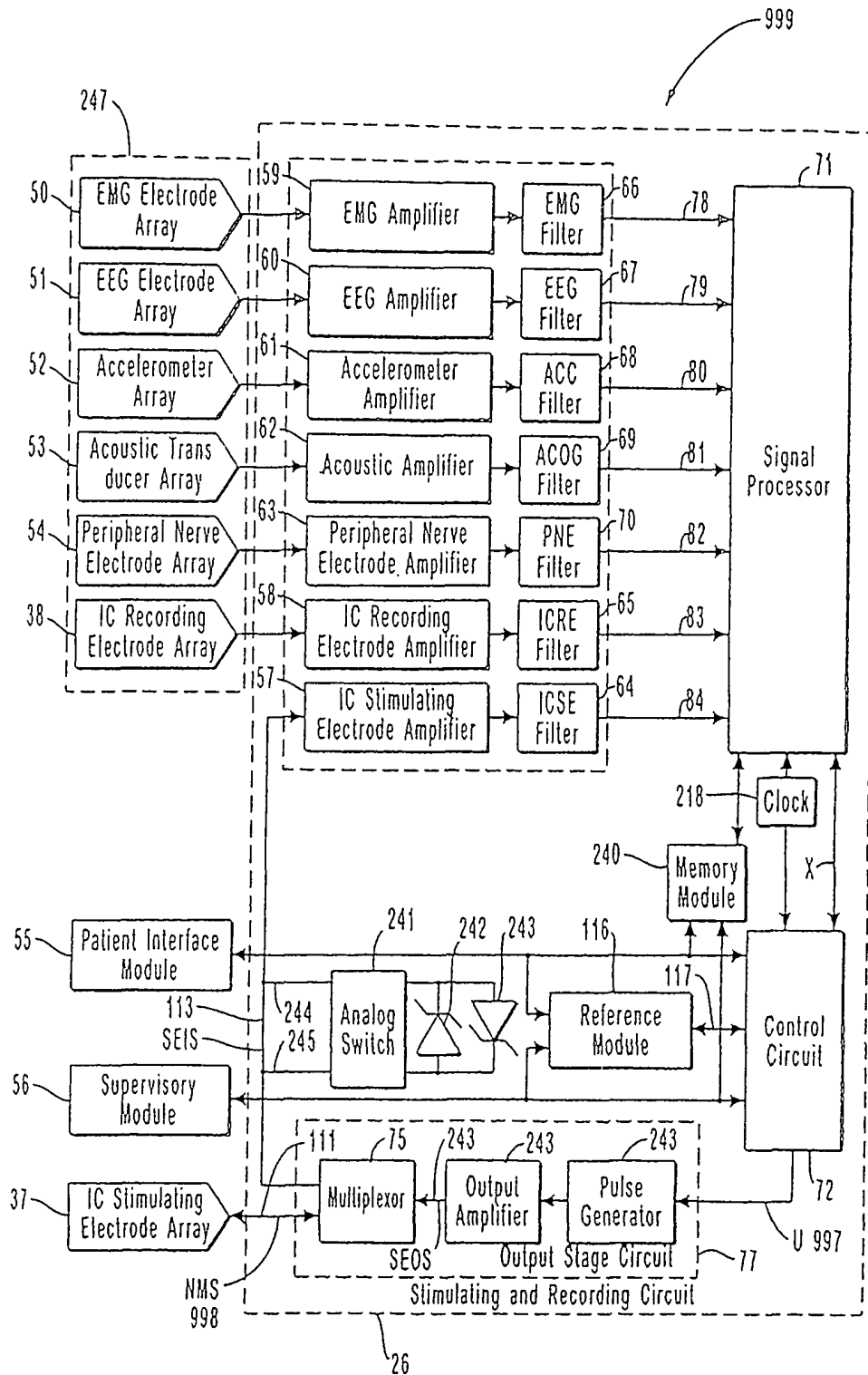


FIG. 2

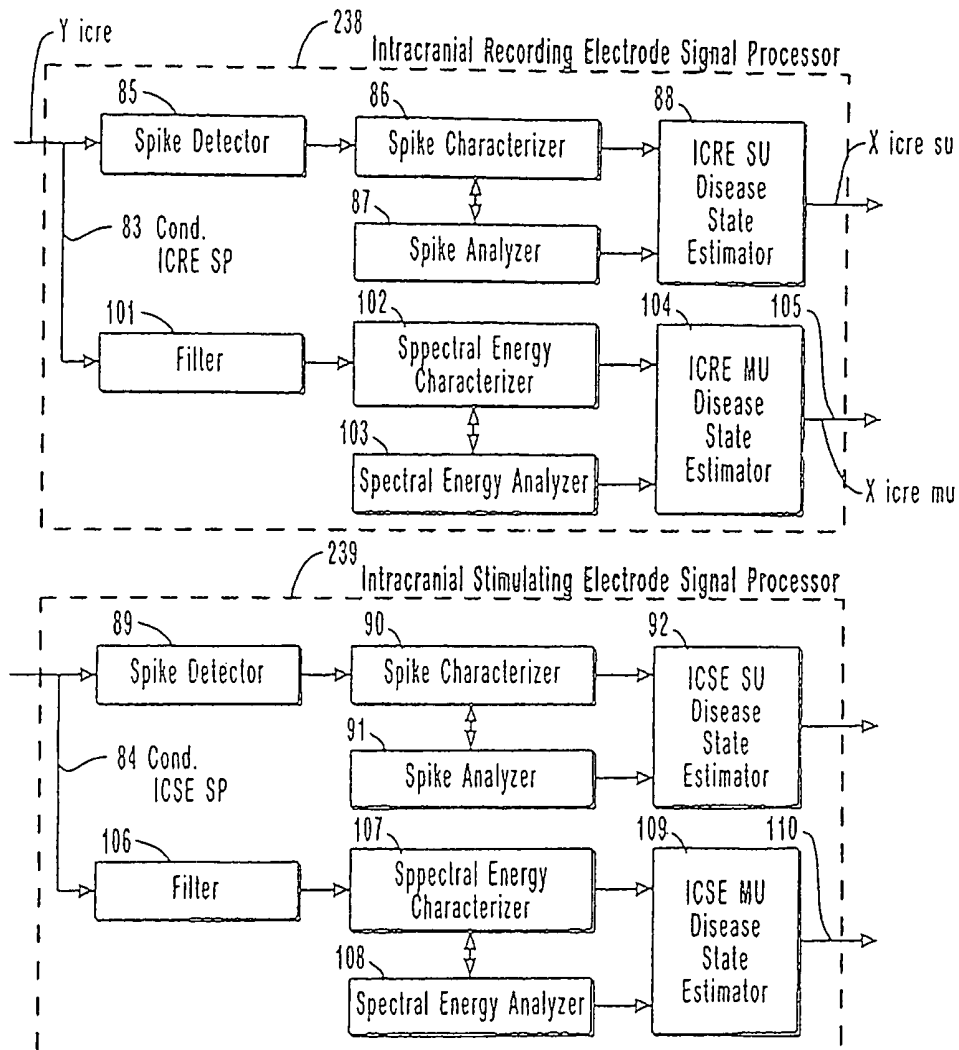


FIG. 3

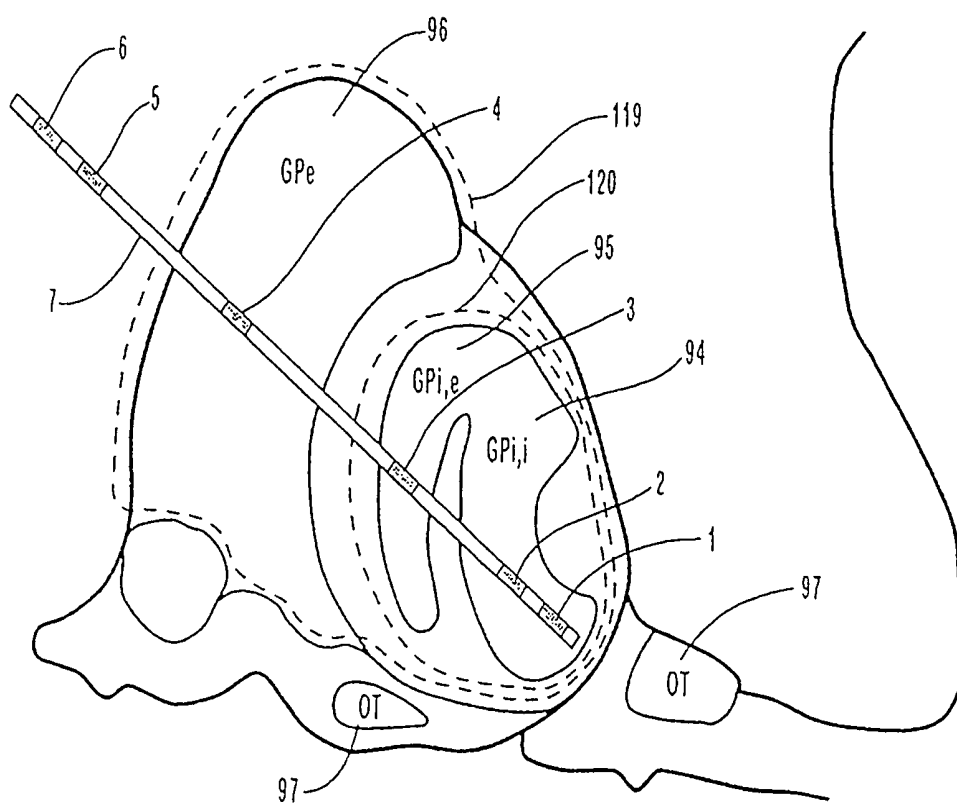


FIG. 4

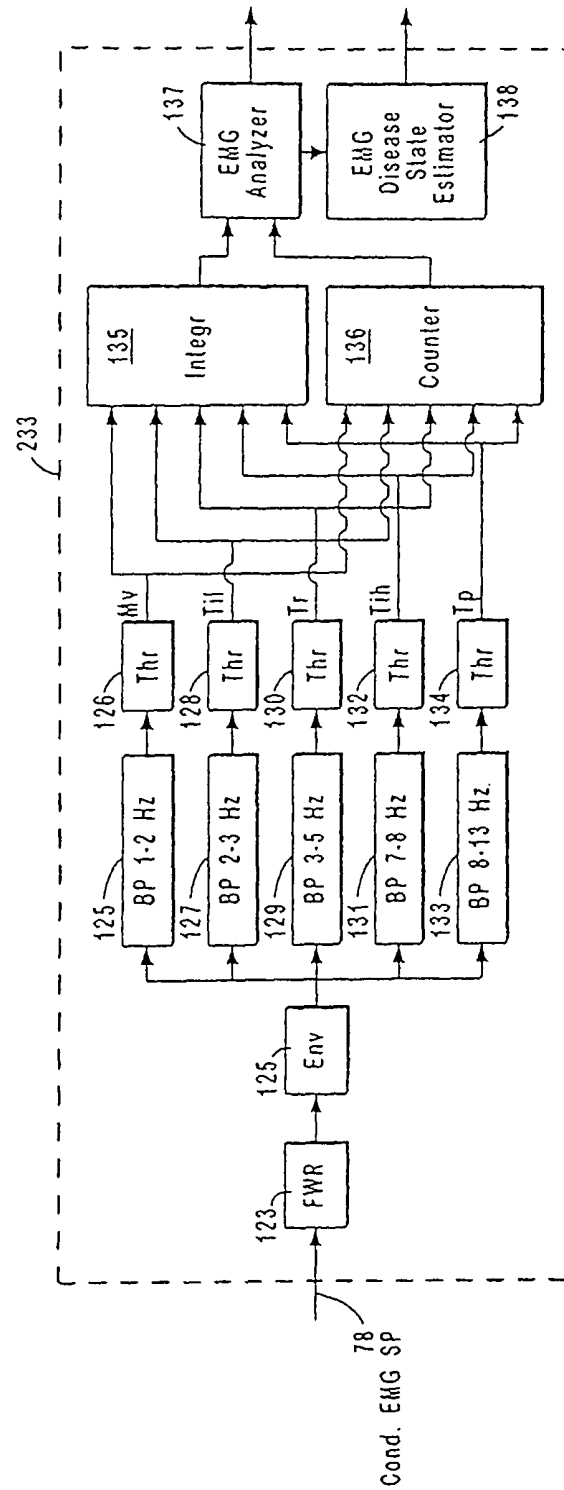


FIG. 5

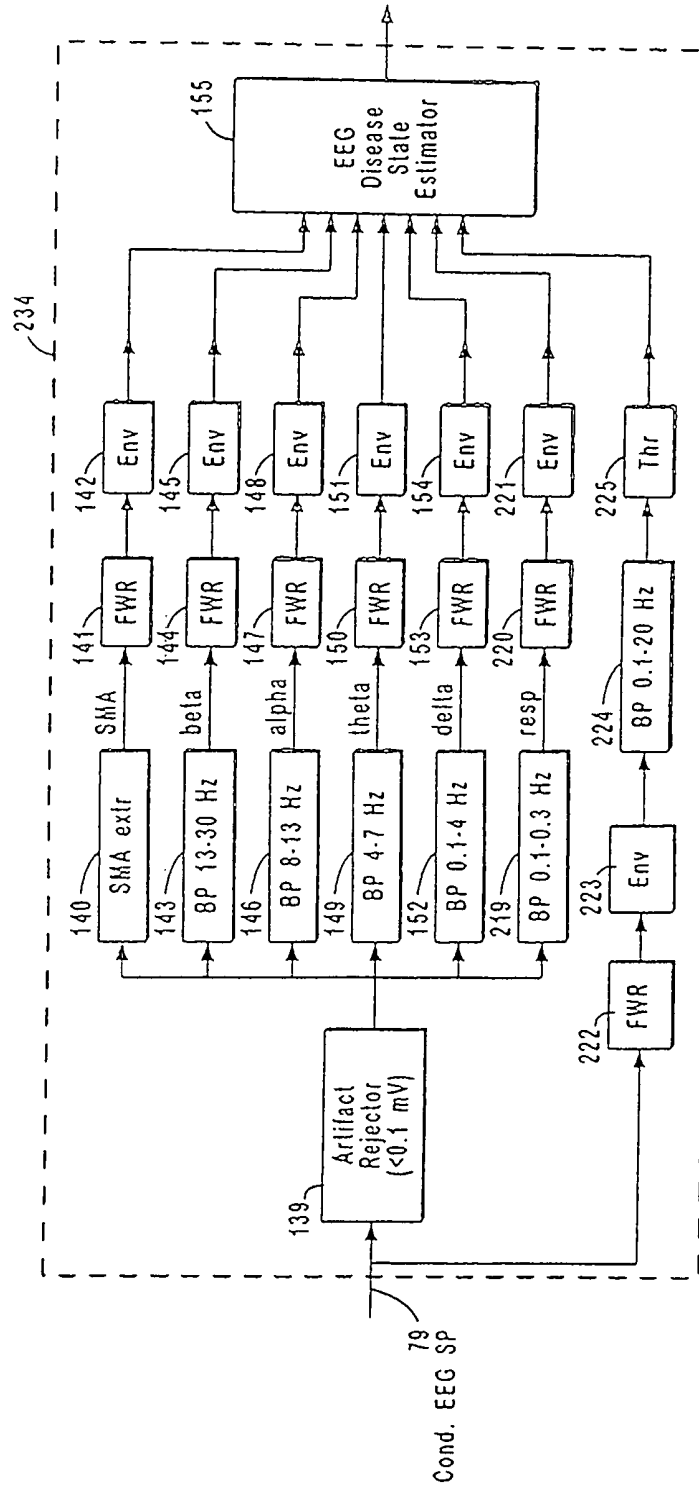


FIG. 6

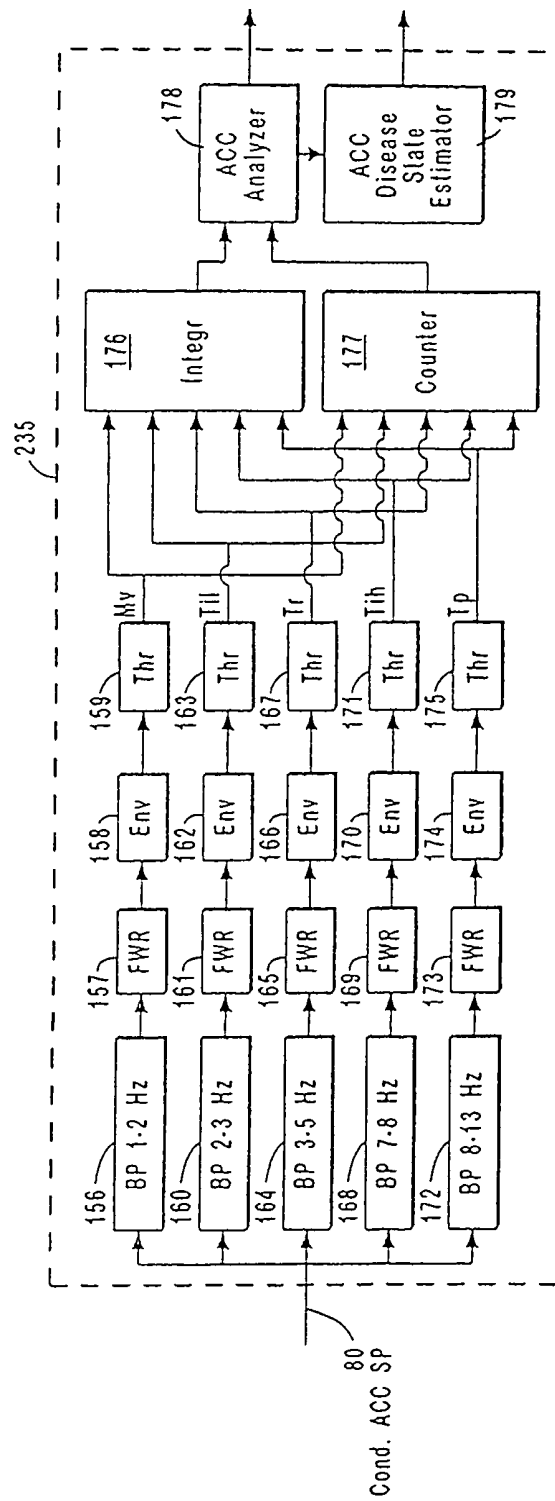


FIG. 7

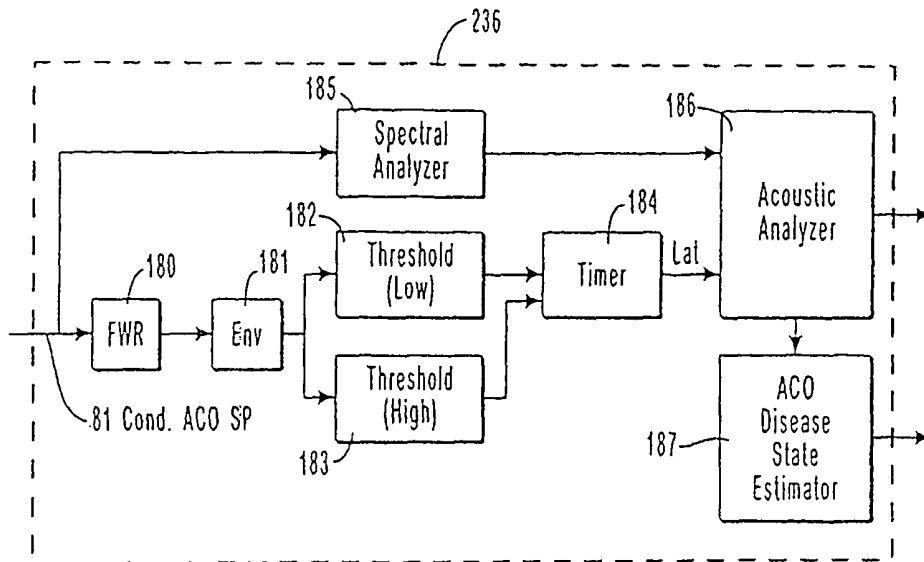


FIG. 8

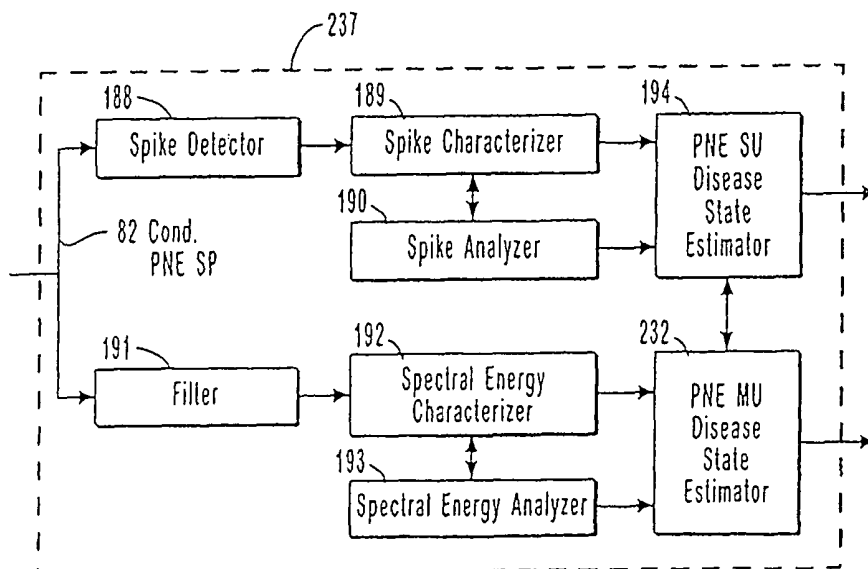


FIG. 9

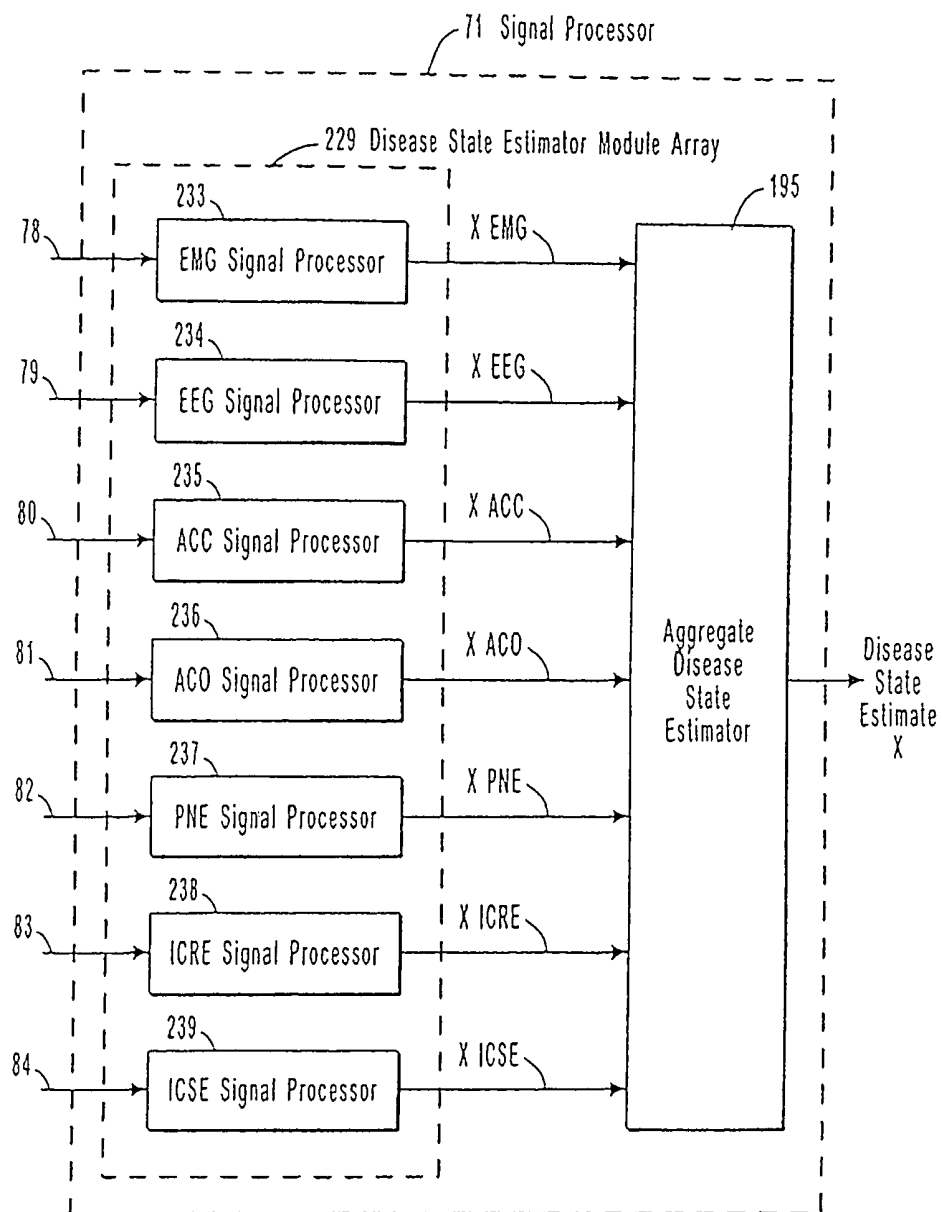


FIG. 10

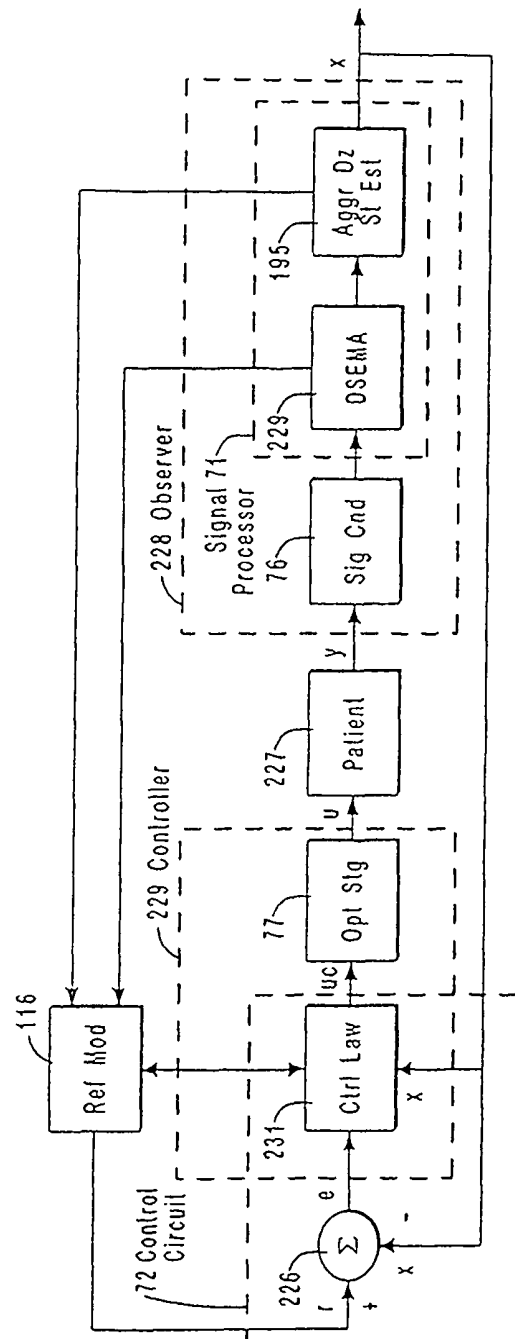


FIG. 11

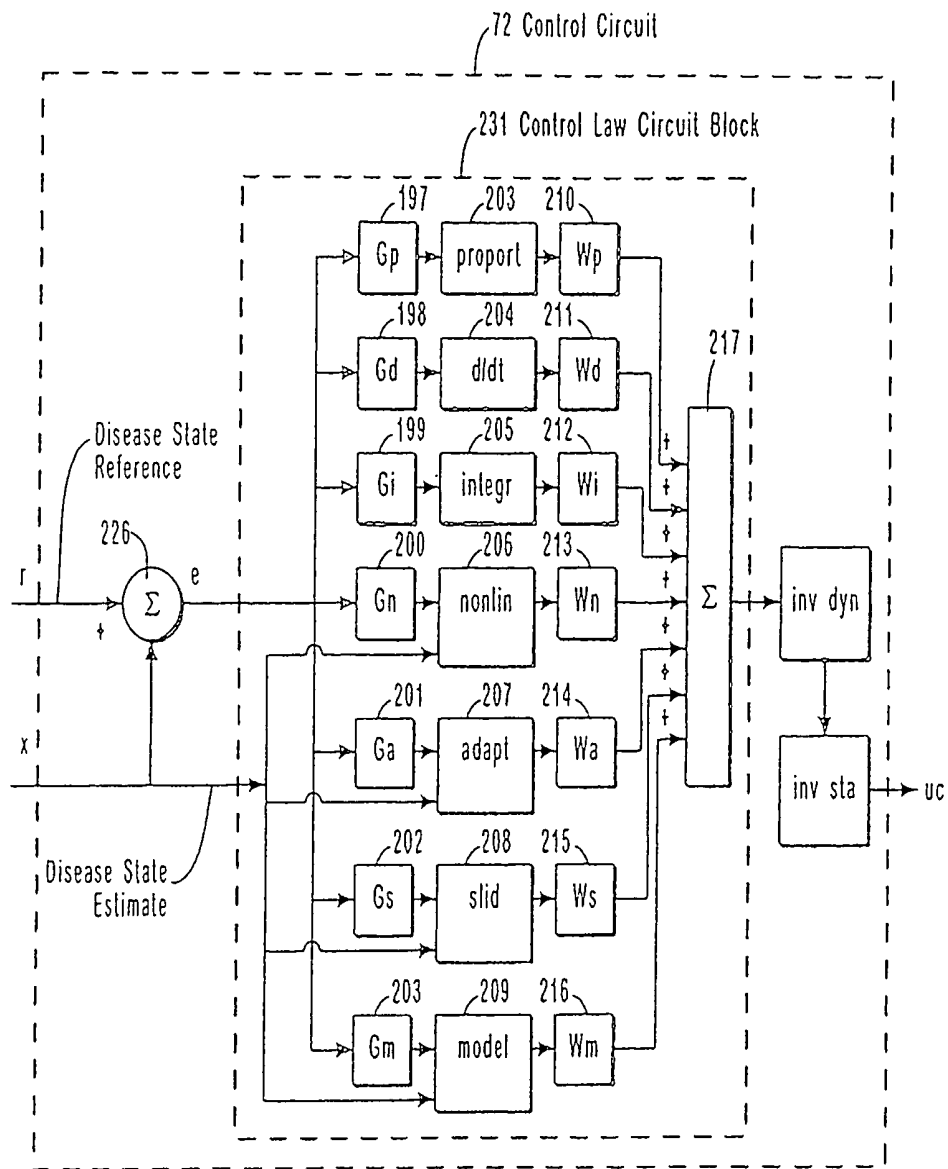


FIG. 12

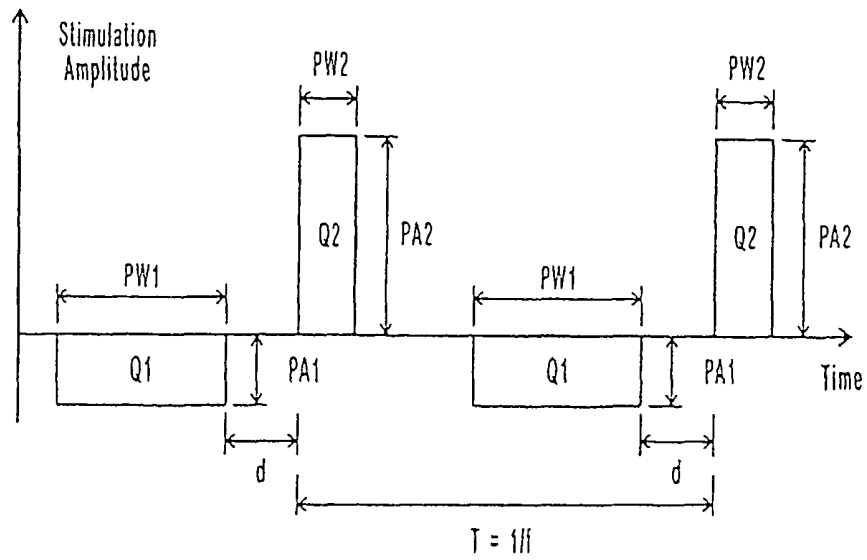


FIG. 13

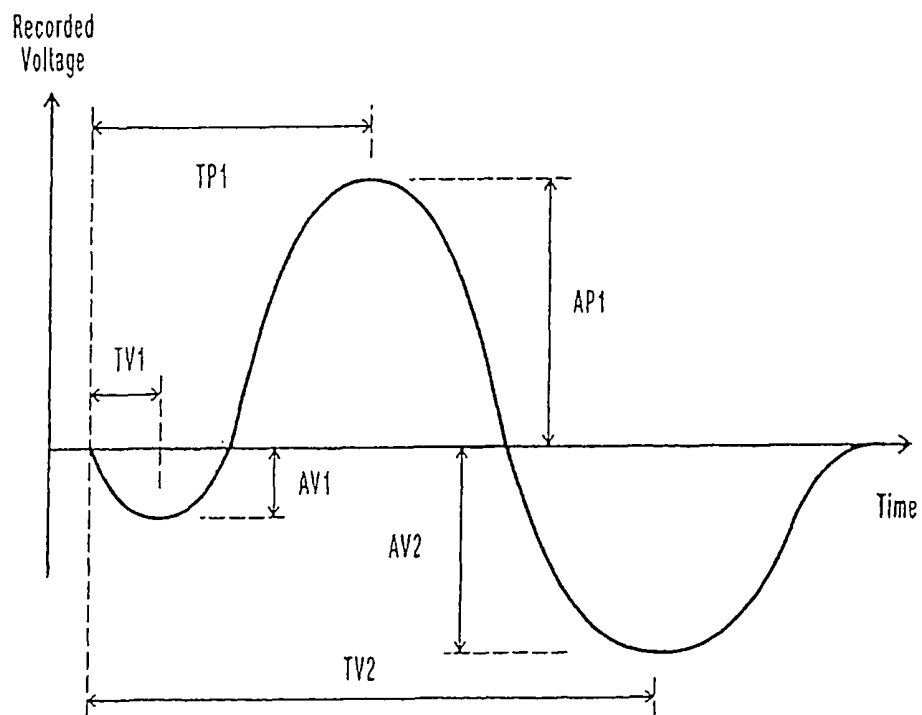


FIG. 14

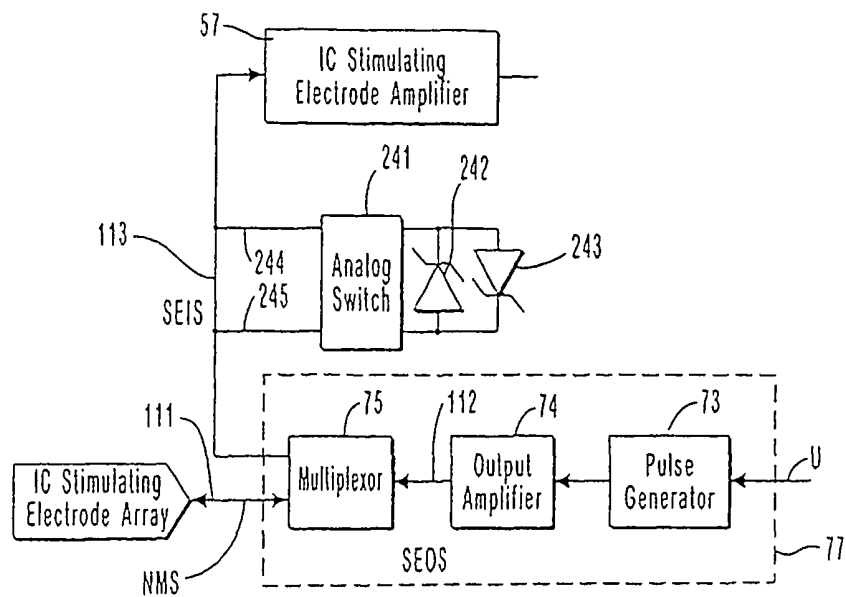


FIG. 15

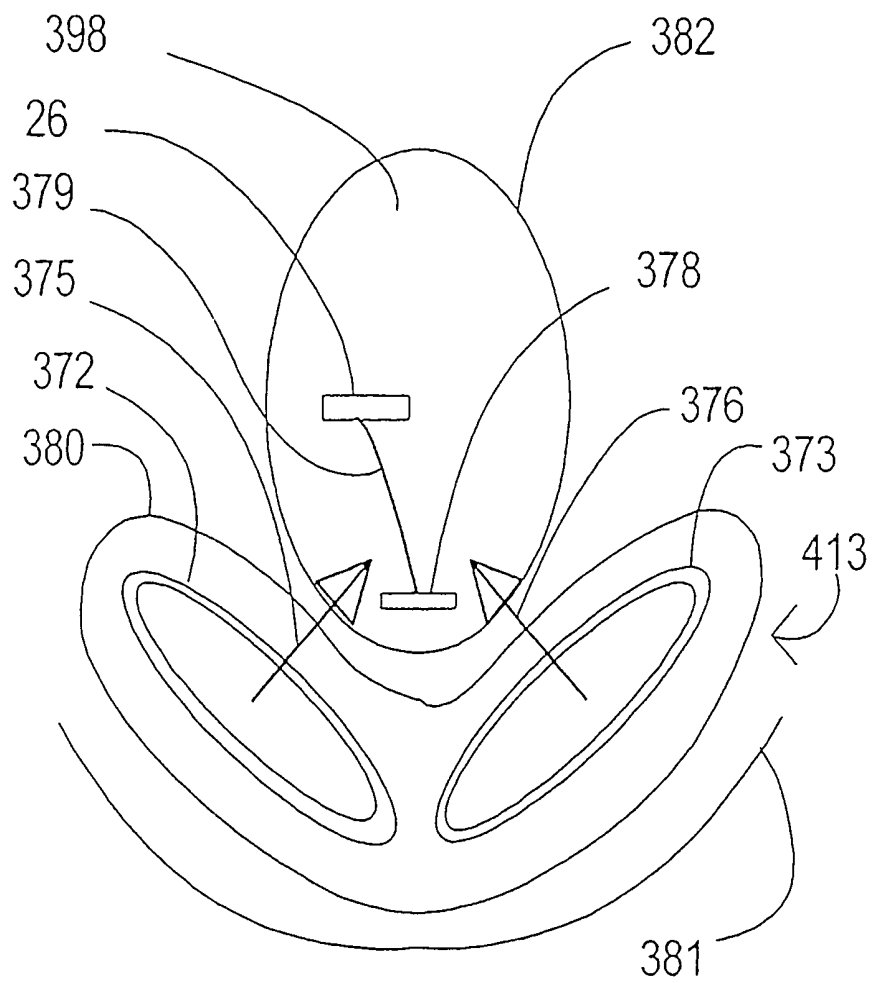


Figure 16

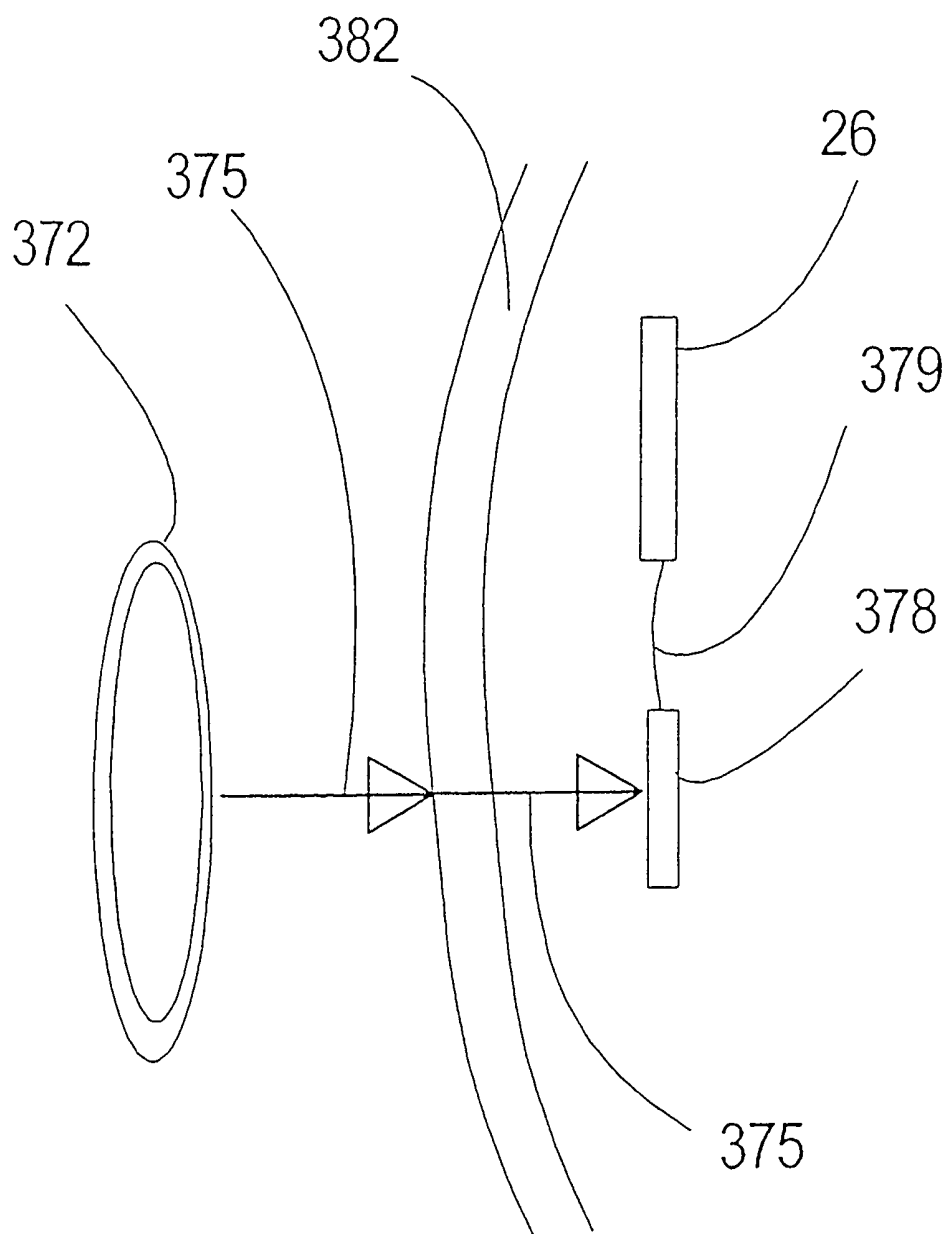


Figure 17

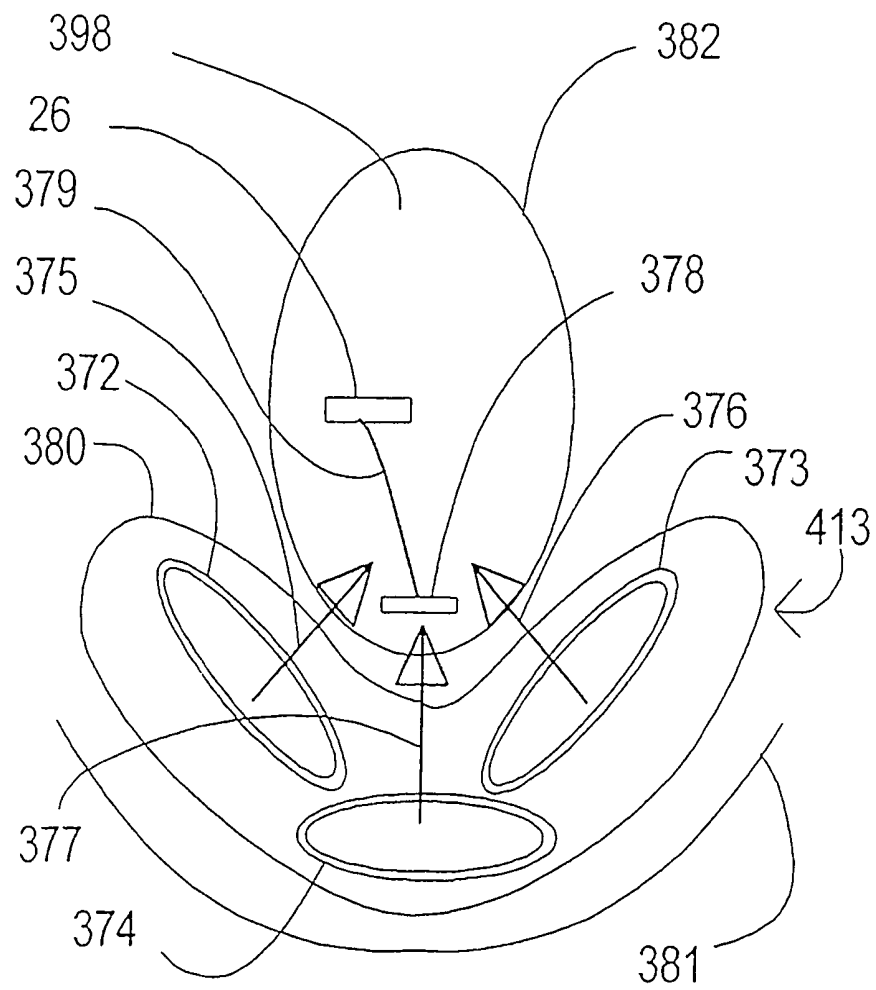


Figure 18

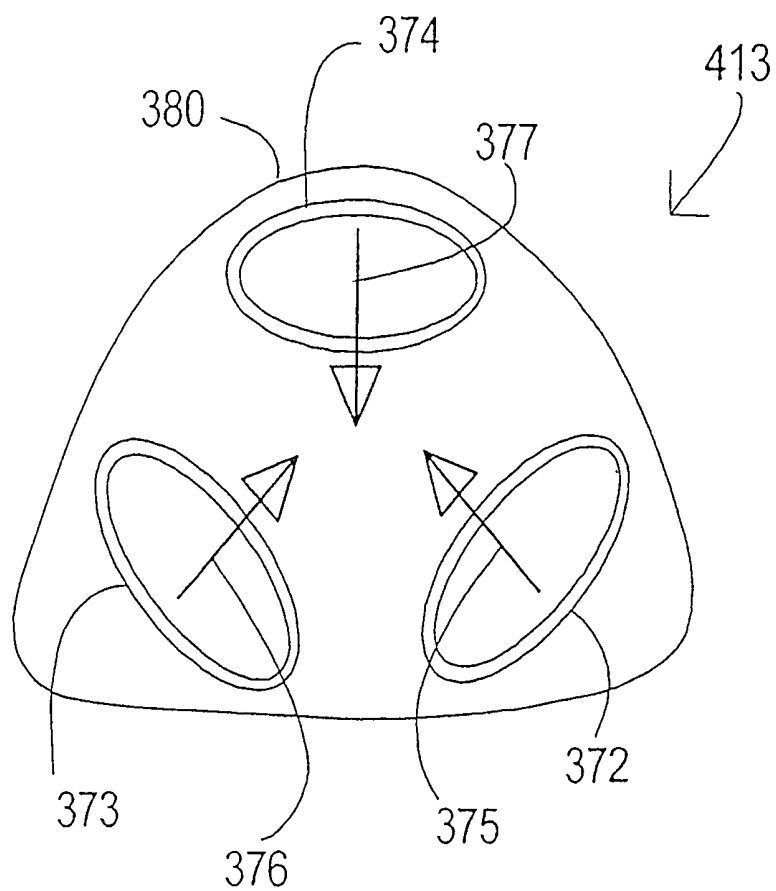


Figure 19

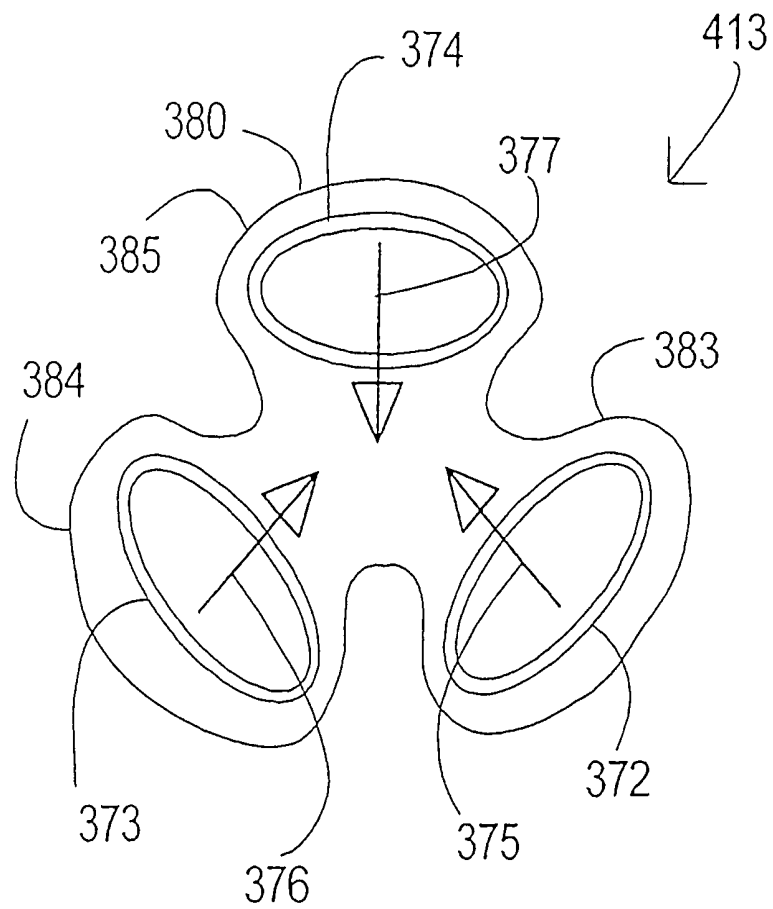


Figure 20

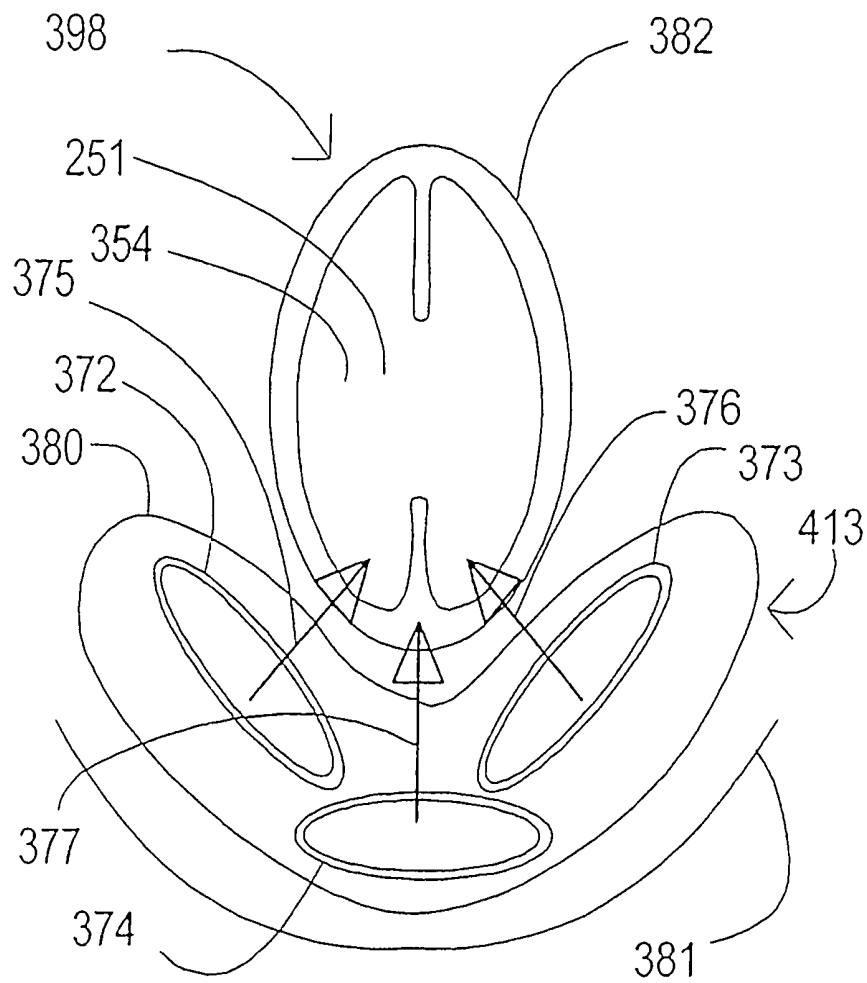


Figure 21

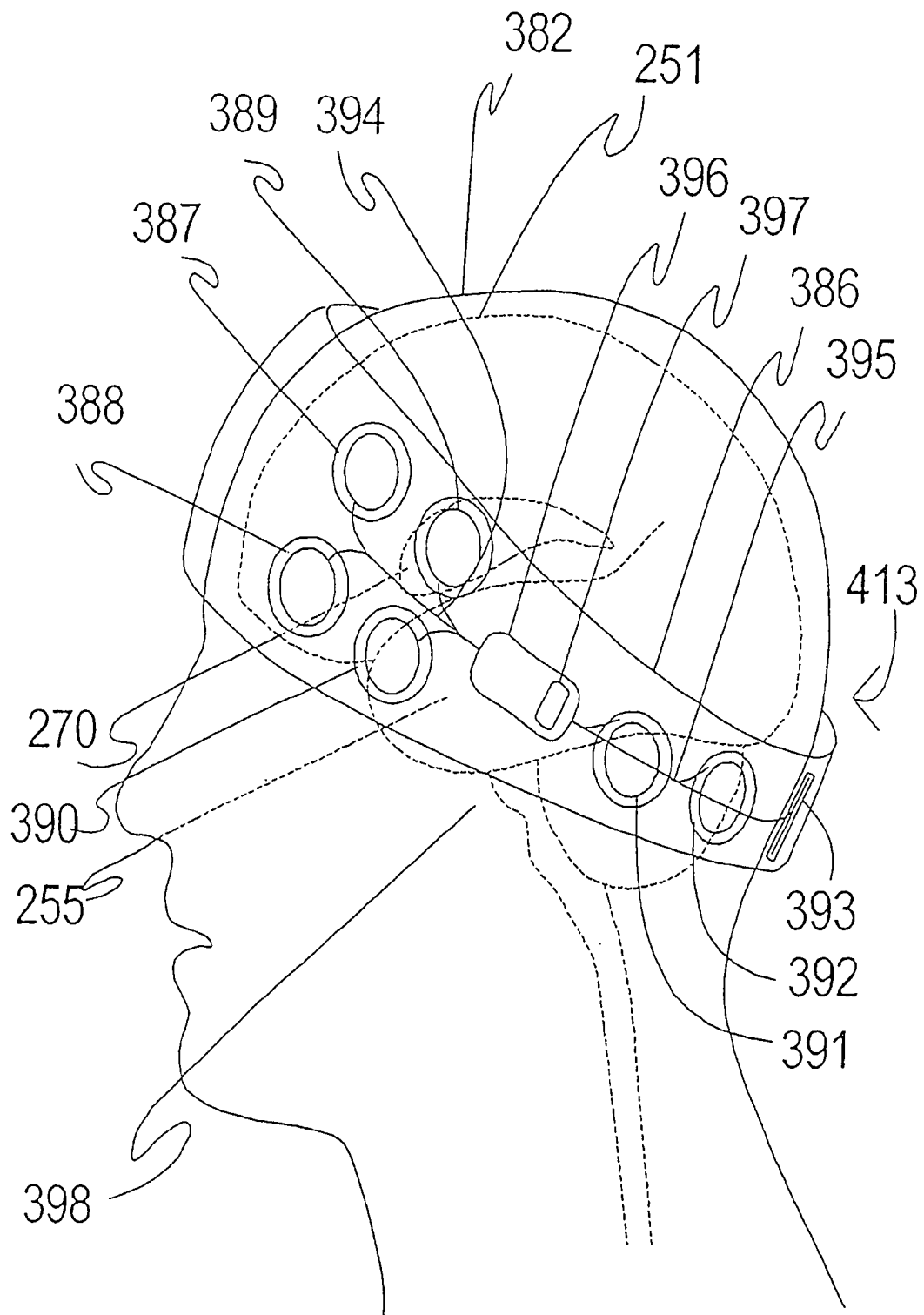


Figure 22

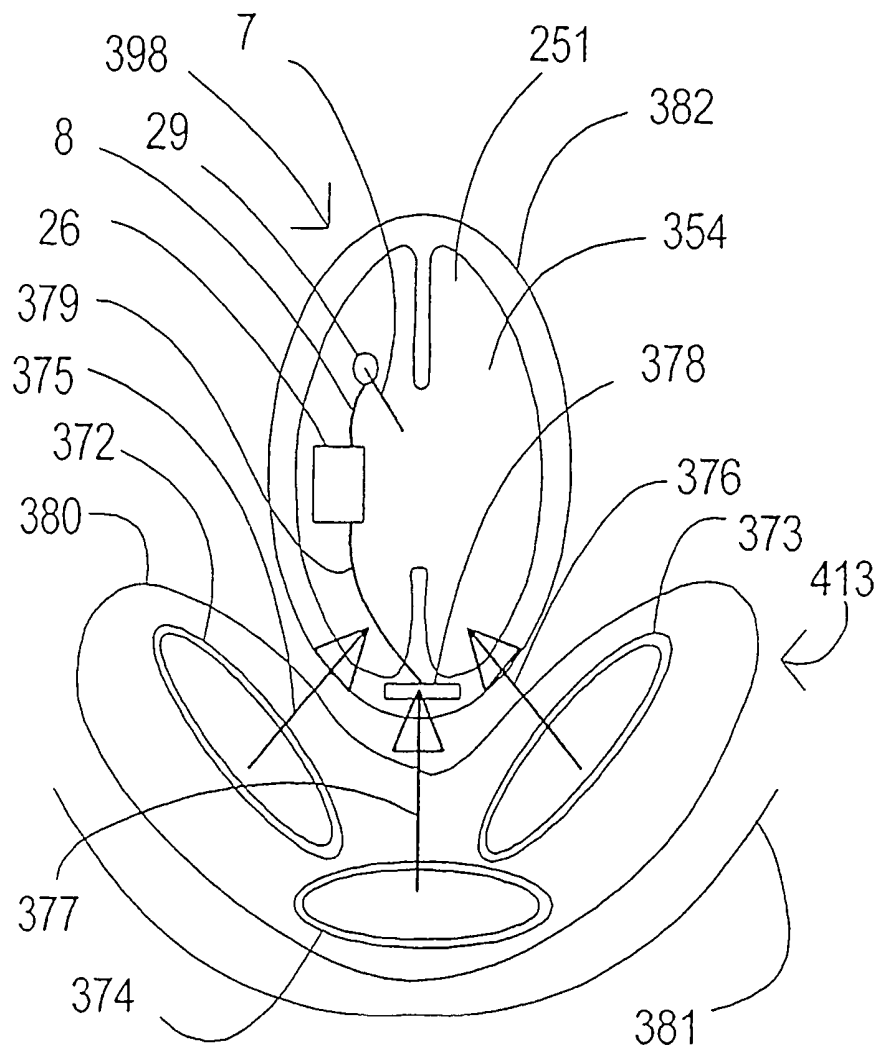


Figure 23

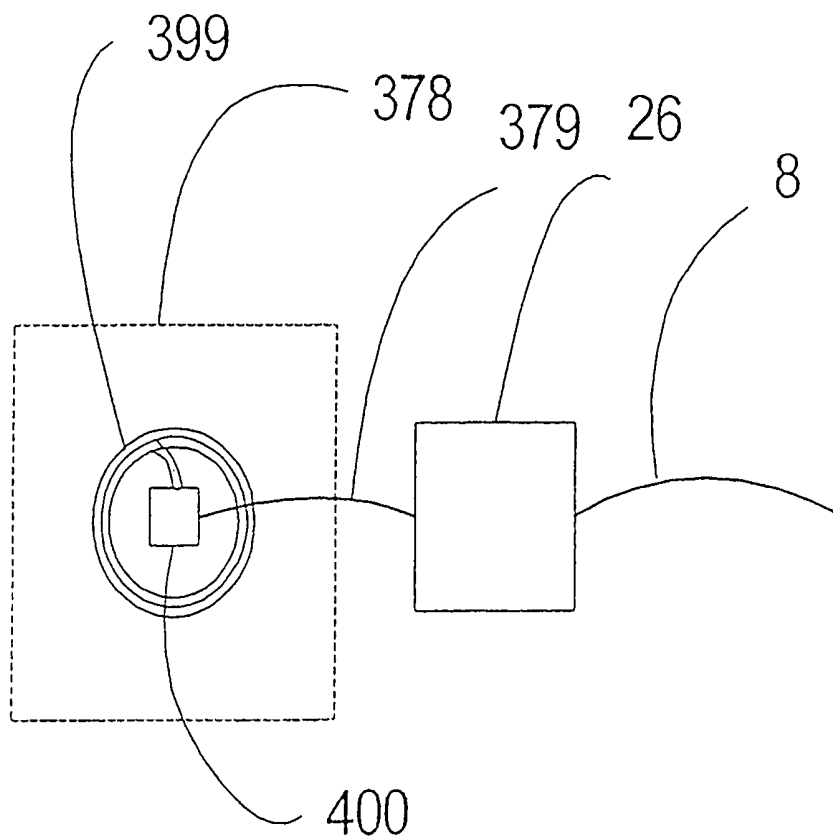


Figure 24

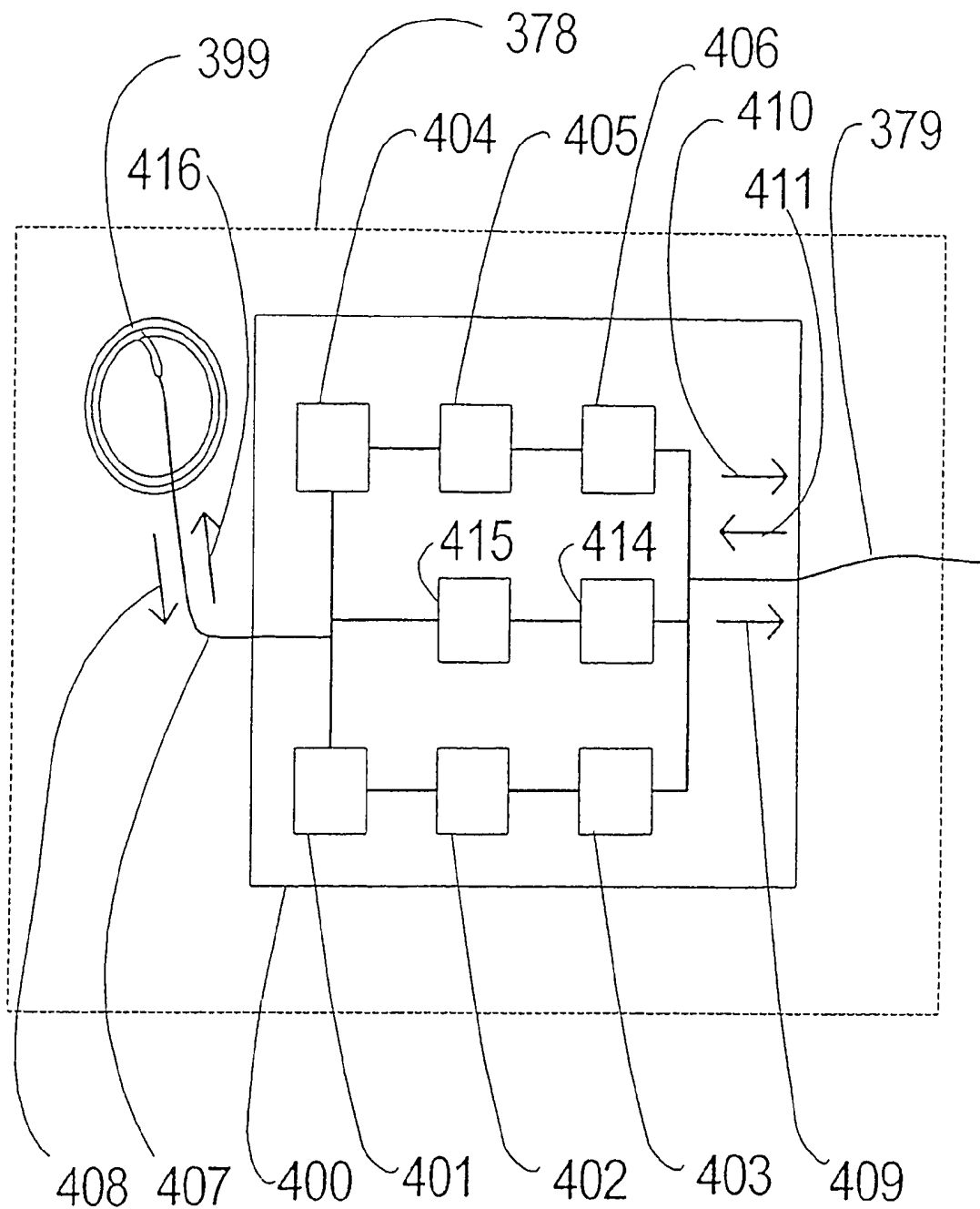


Figure 25

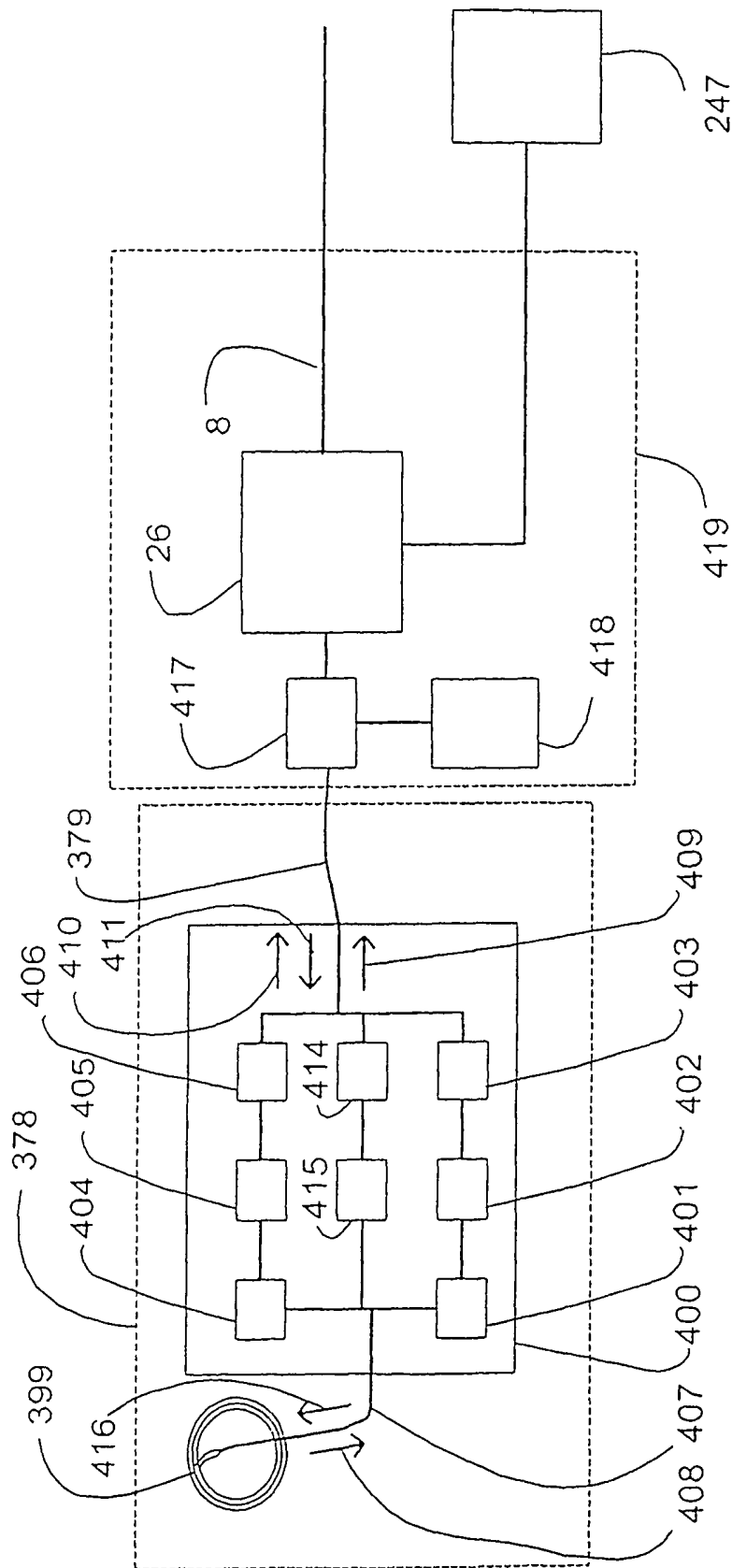


Figure 26

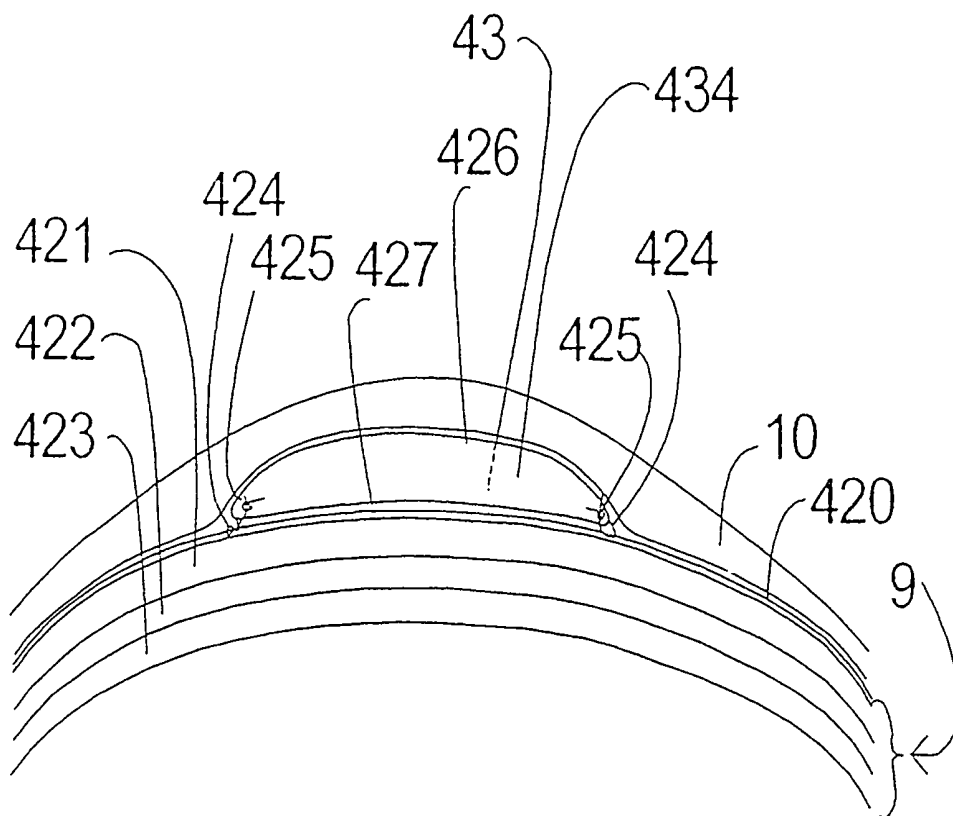


Figure 27

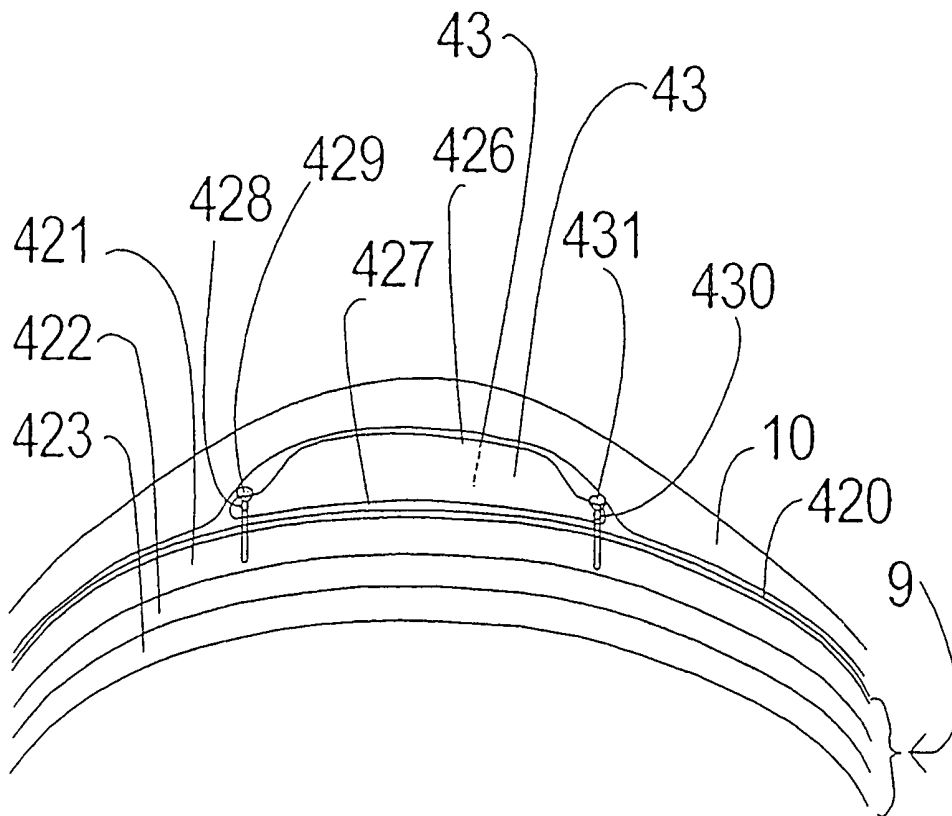


Figure 28

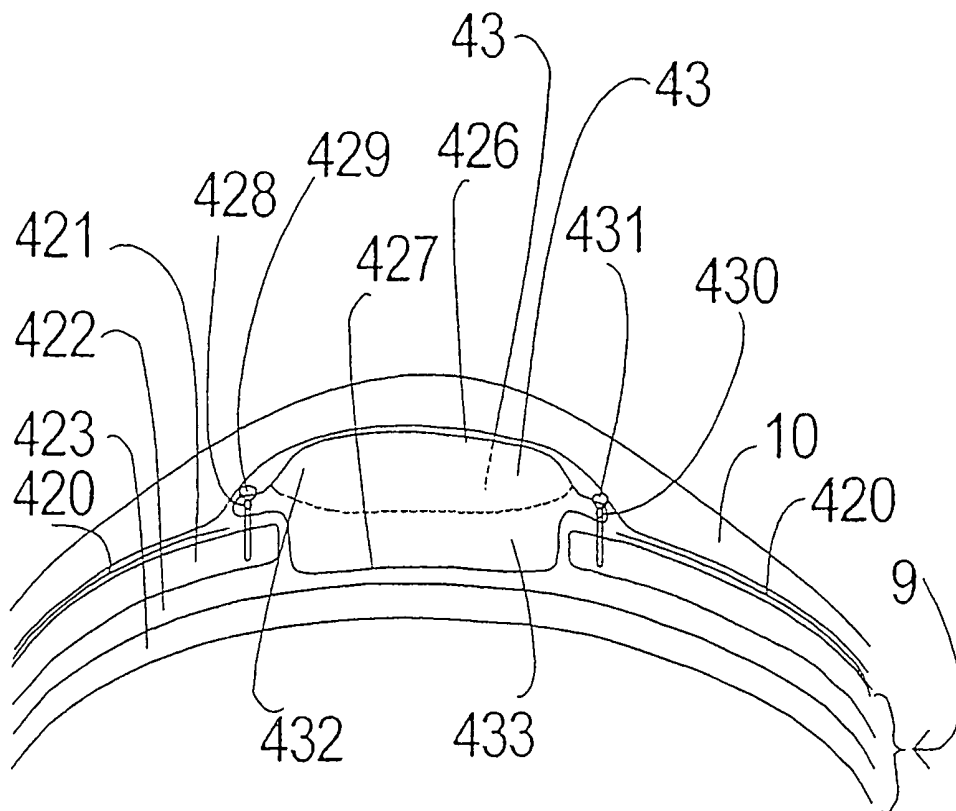


Figure 29

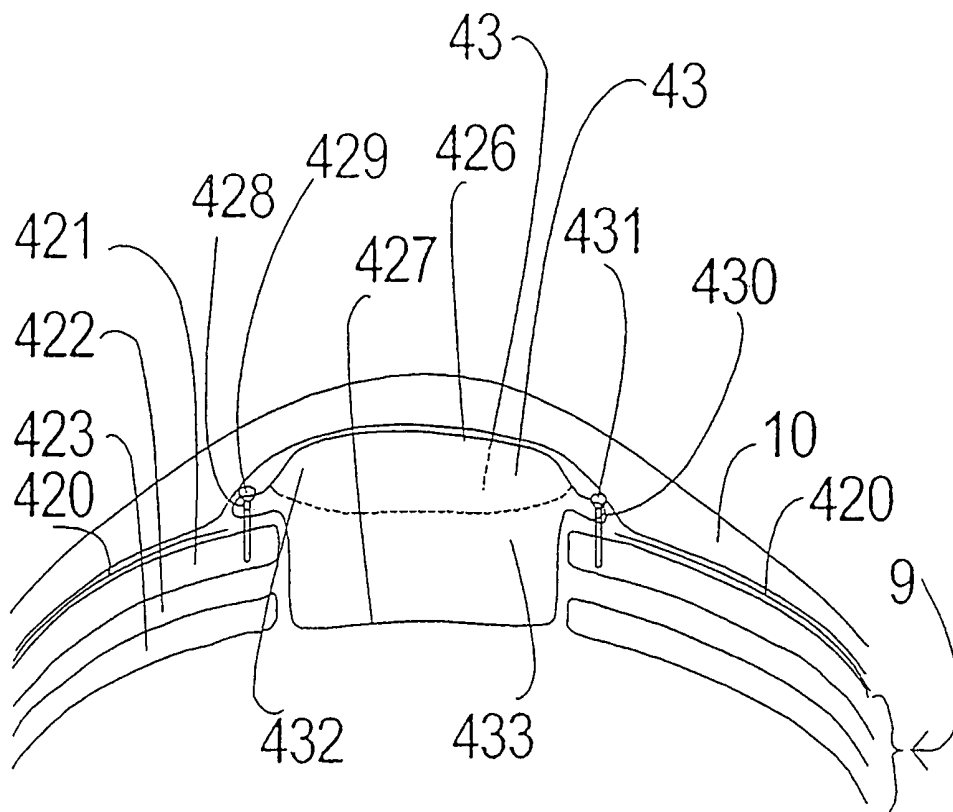


Figure 30

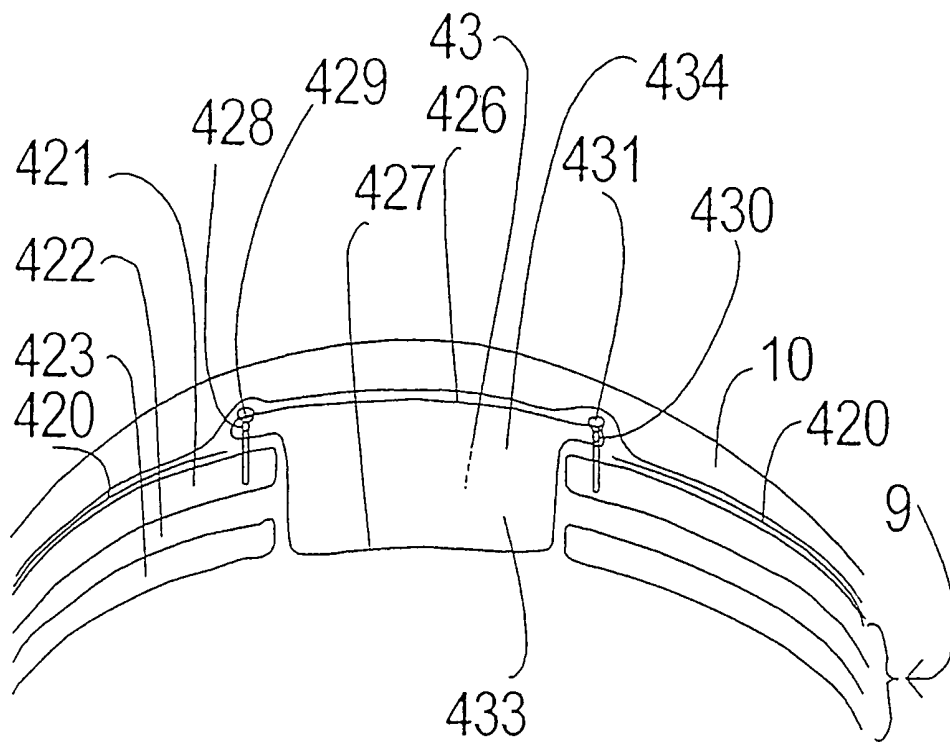


Figure 31

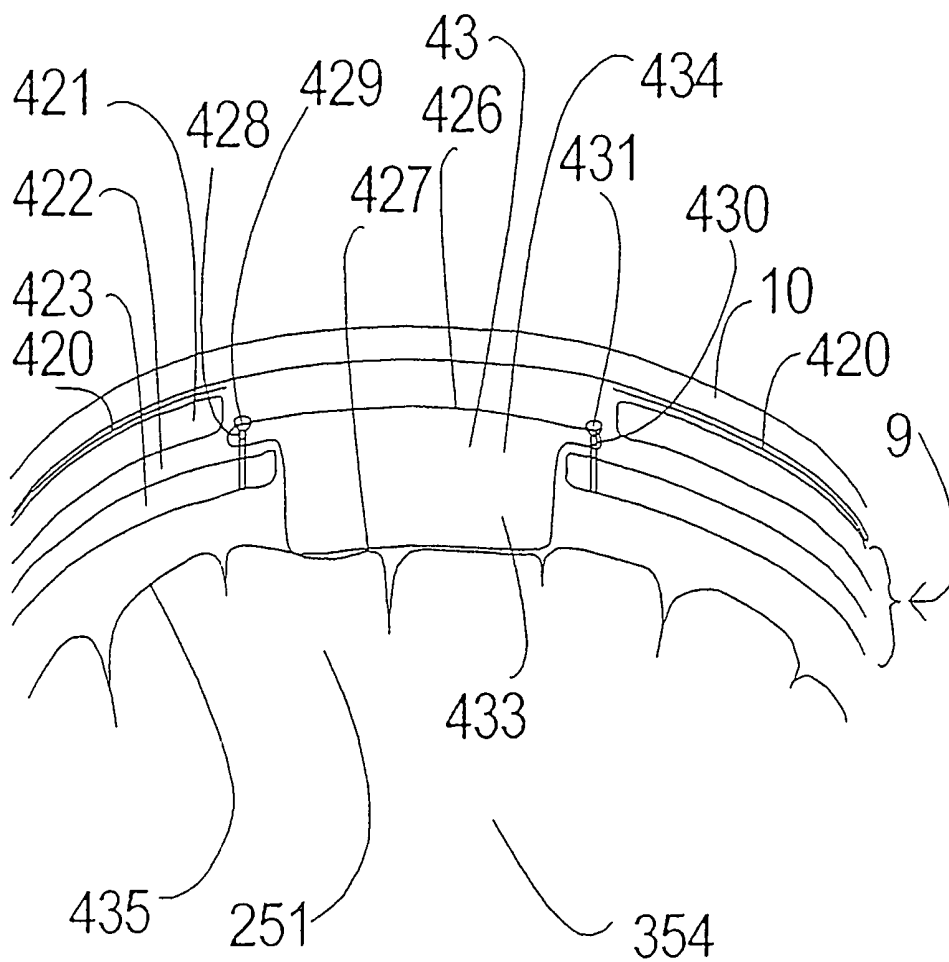


Figure 32

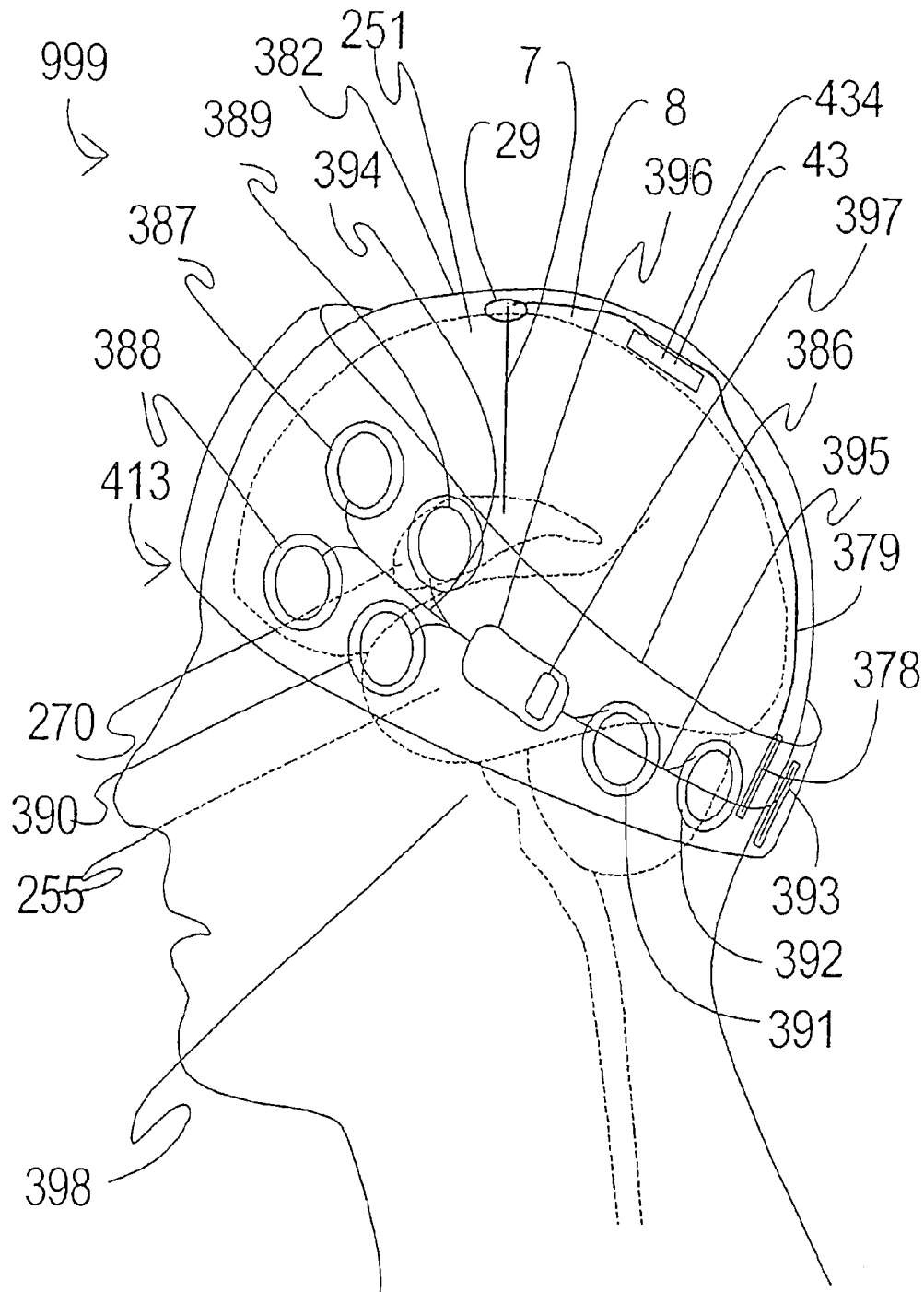


Figure 33

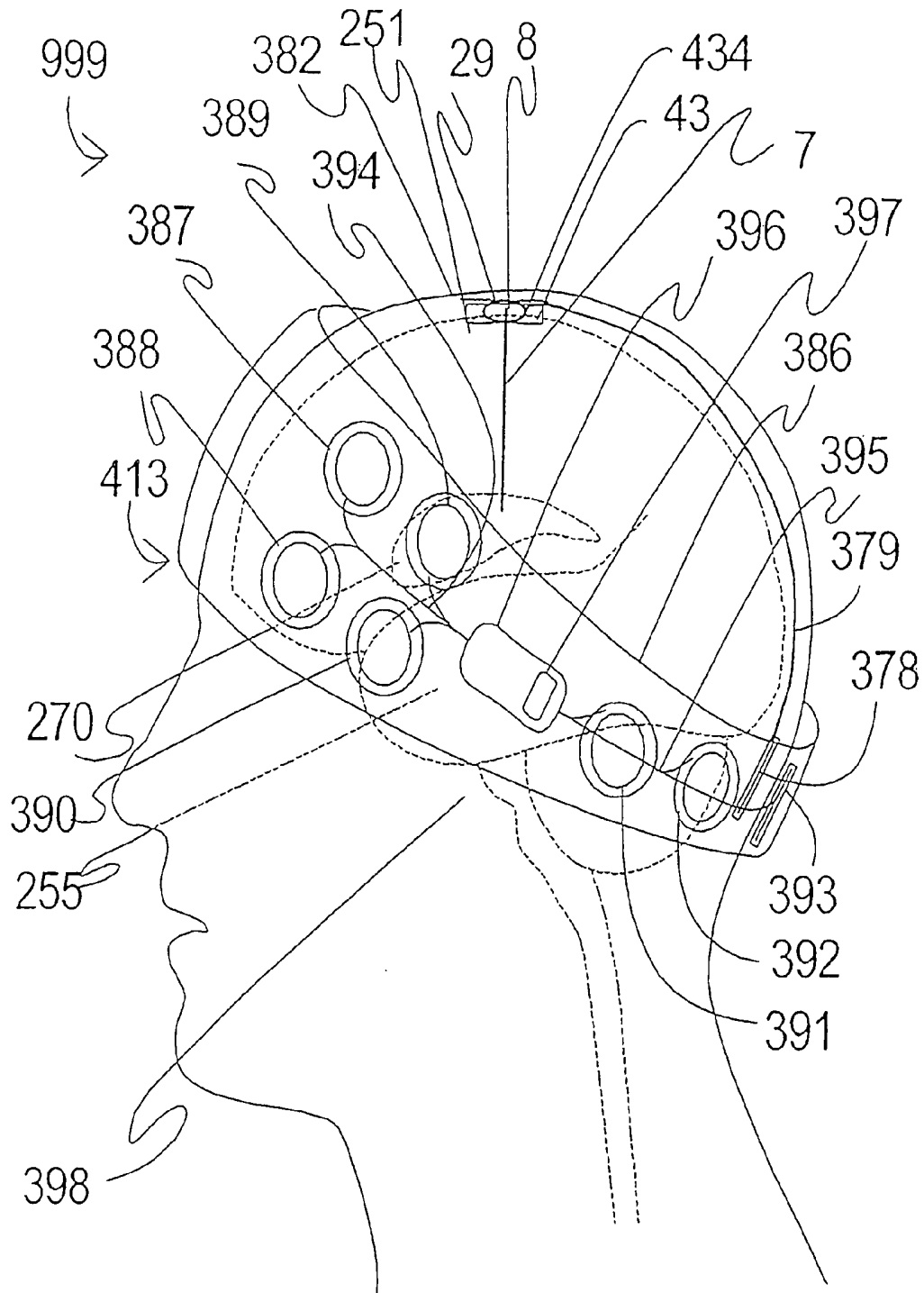


Figure 34

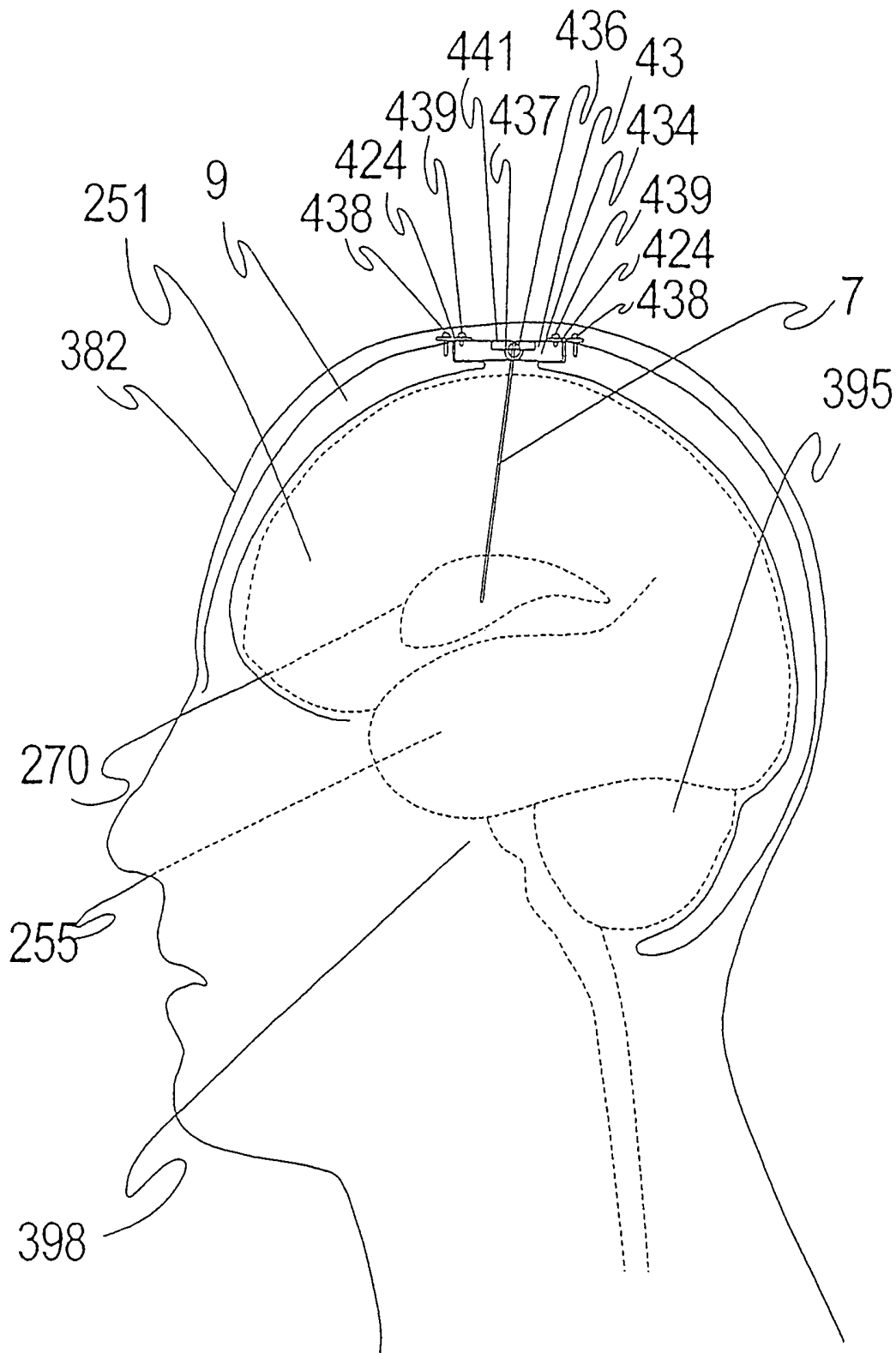


Figure 35

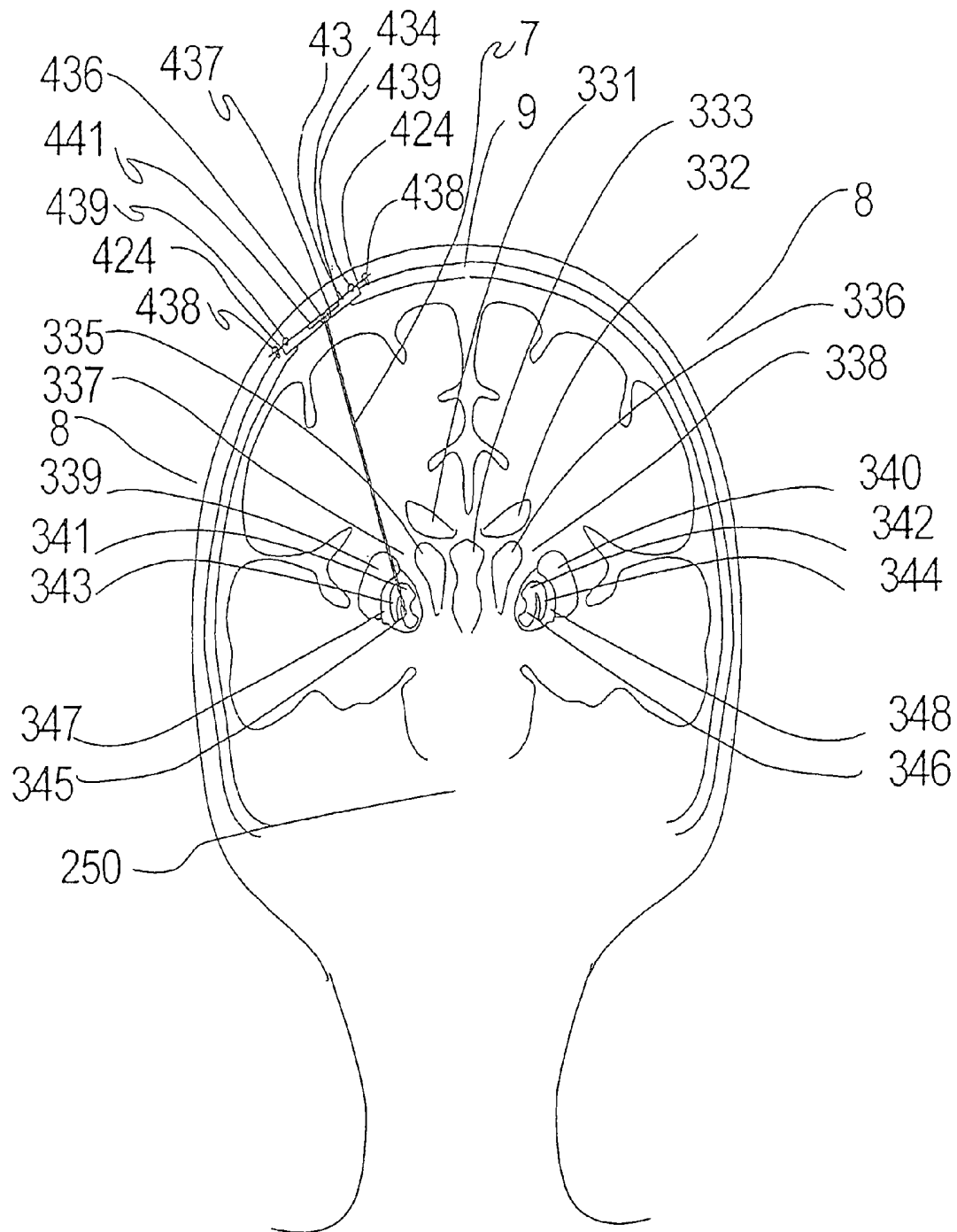


Figure 36

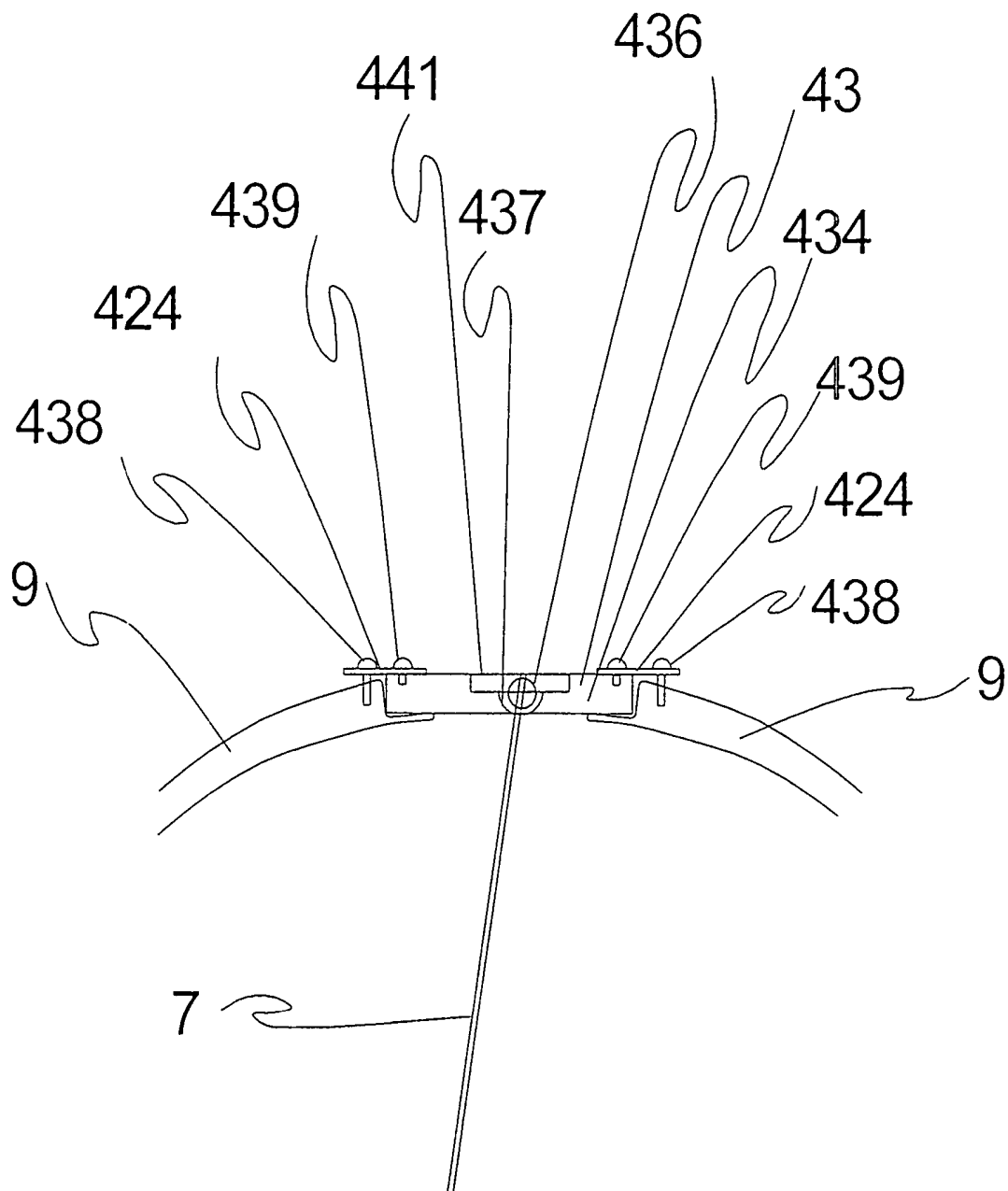


Figure 37

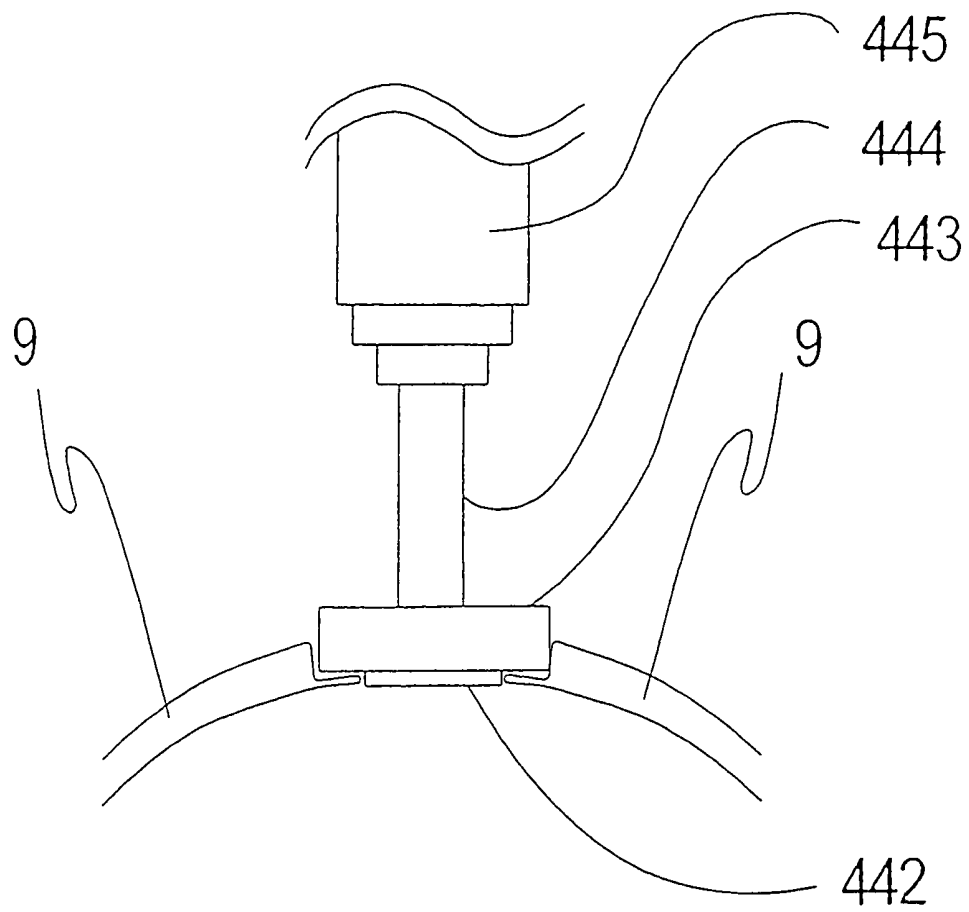


Figure 38

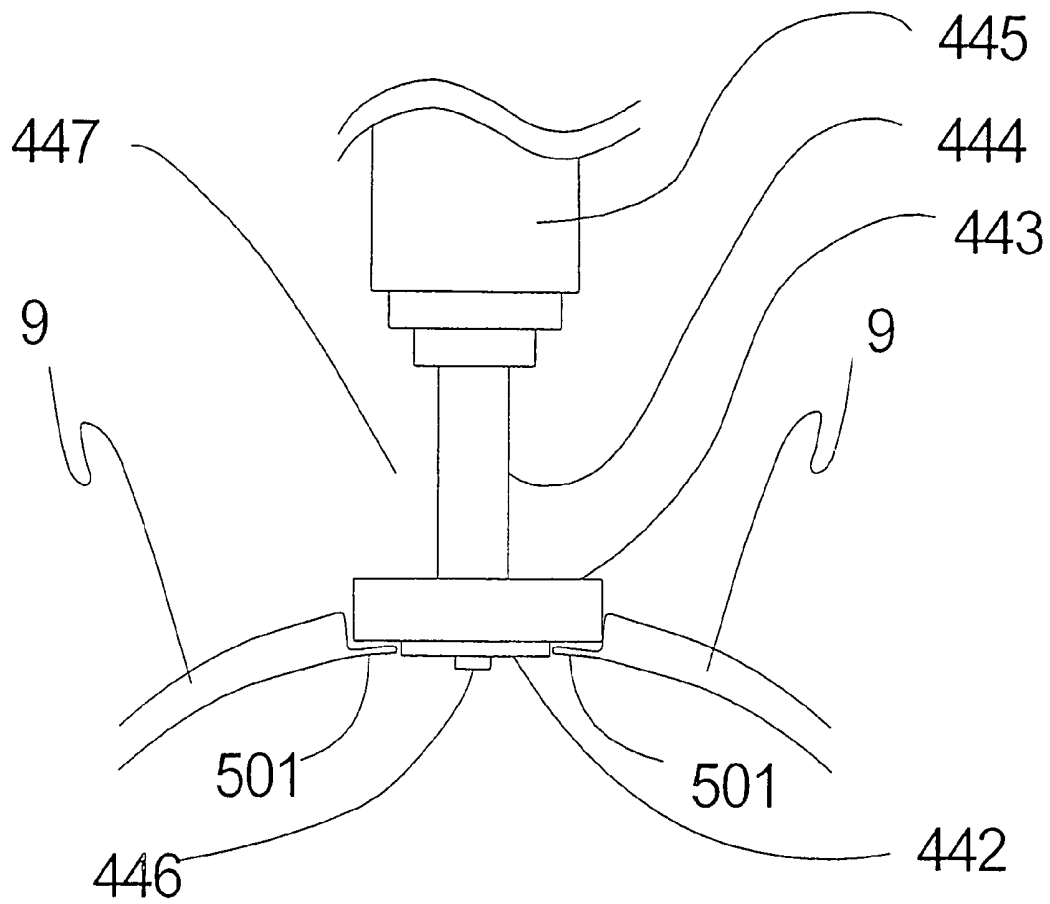


Figure 39

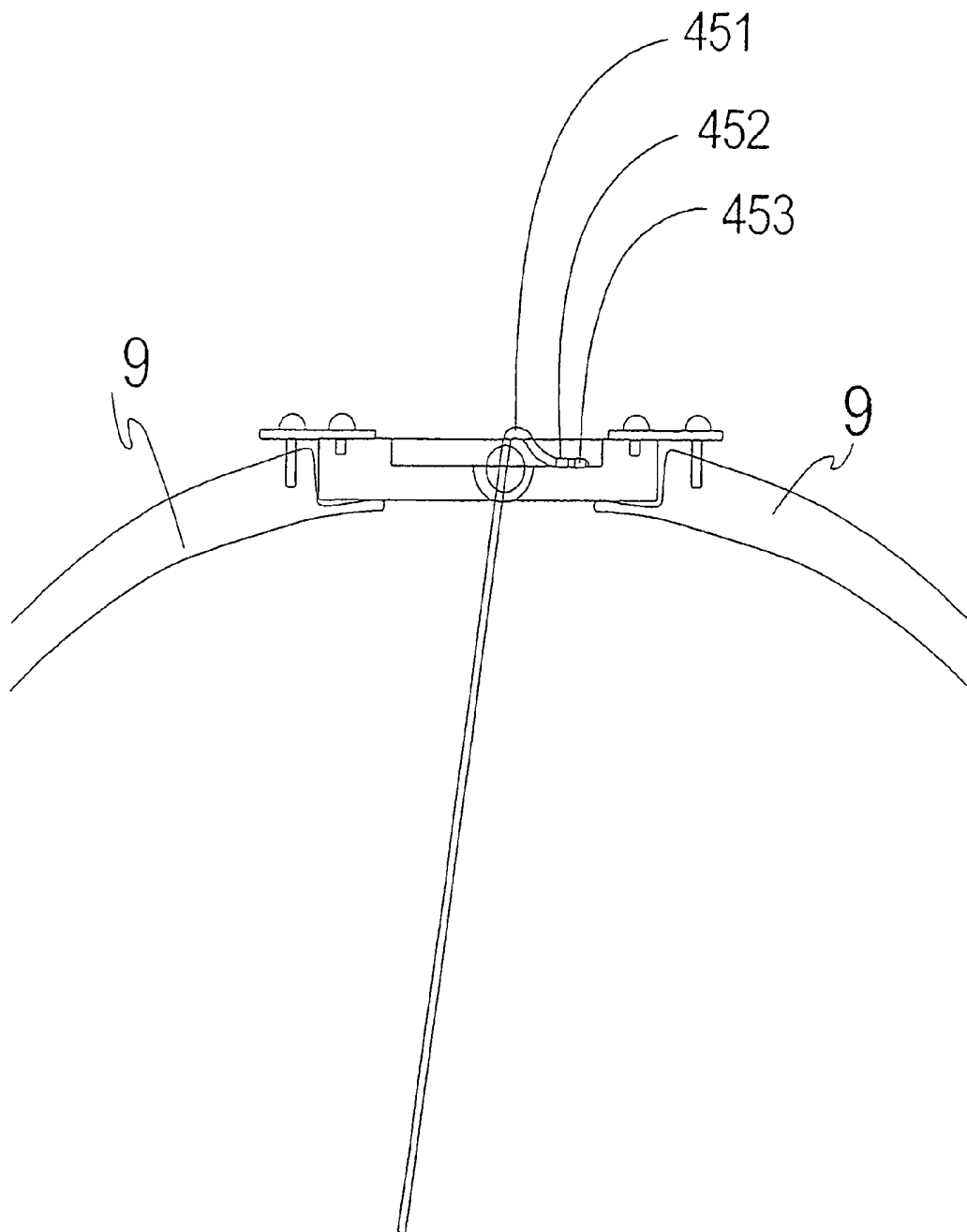


Figure 40

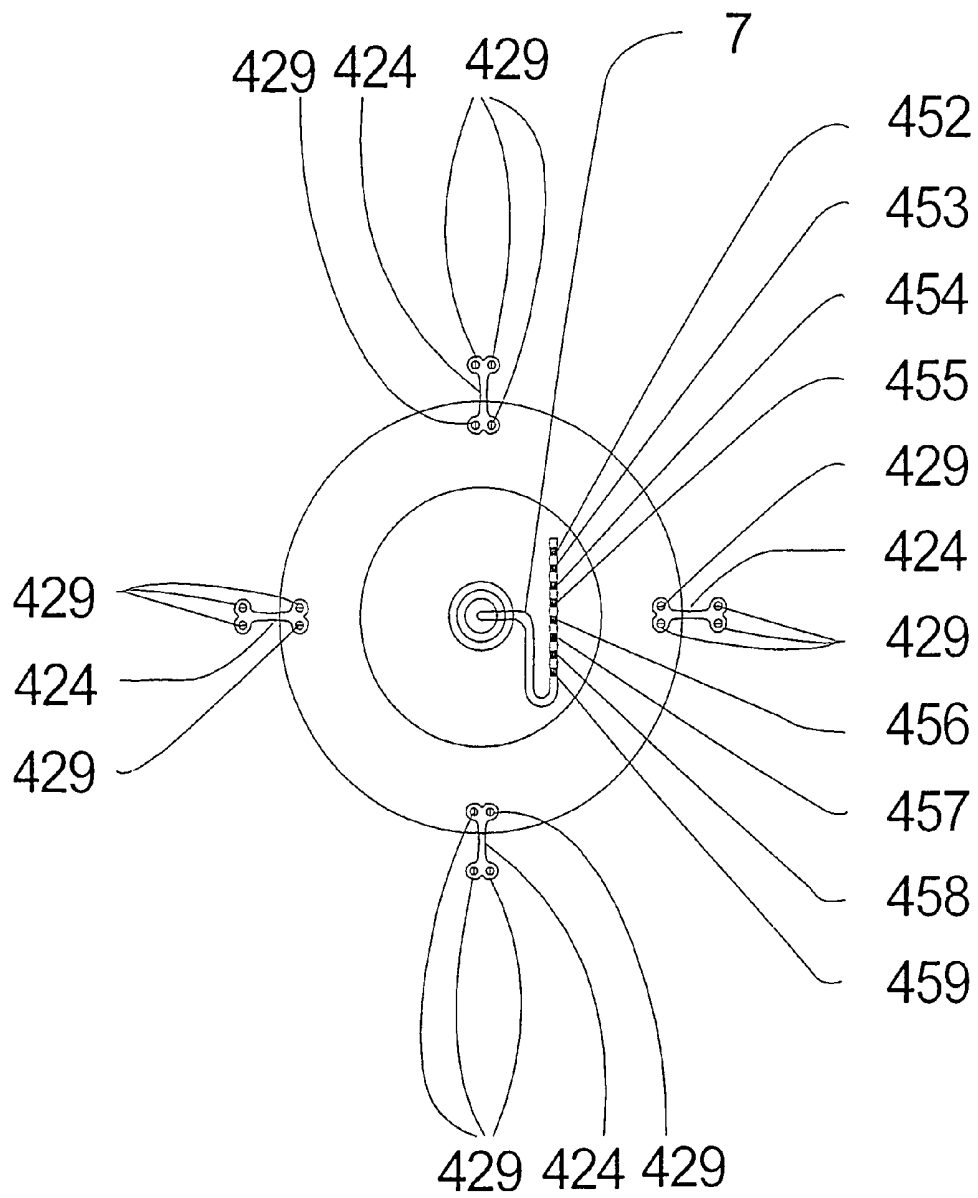


Figure 41

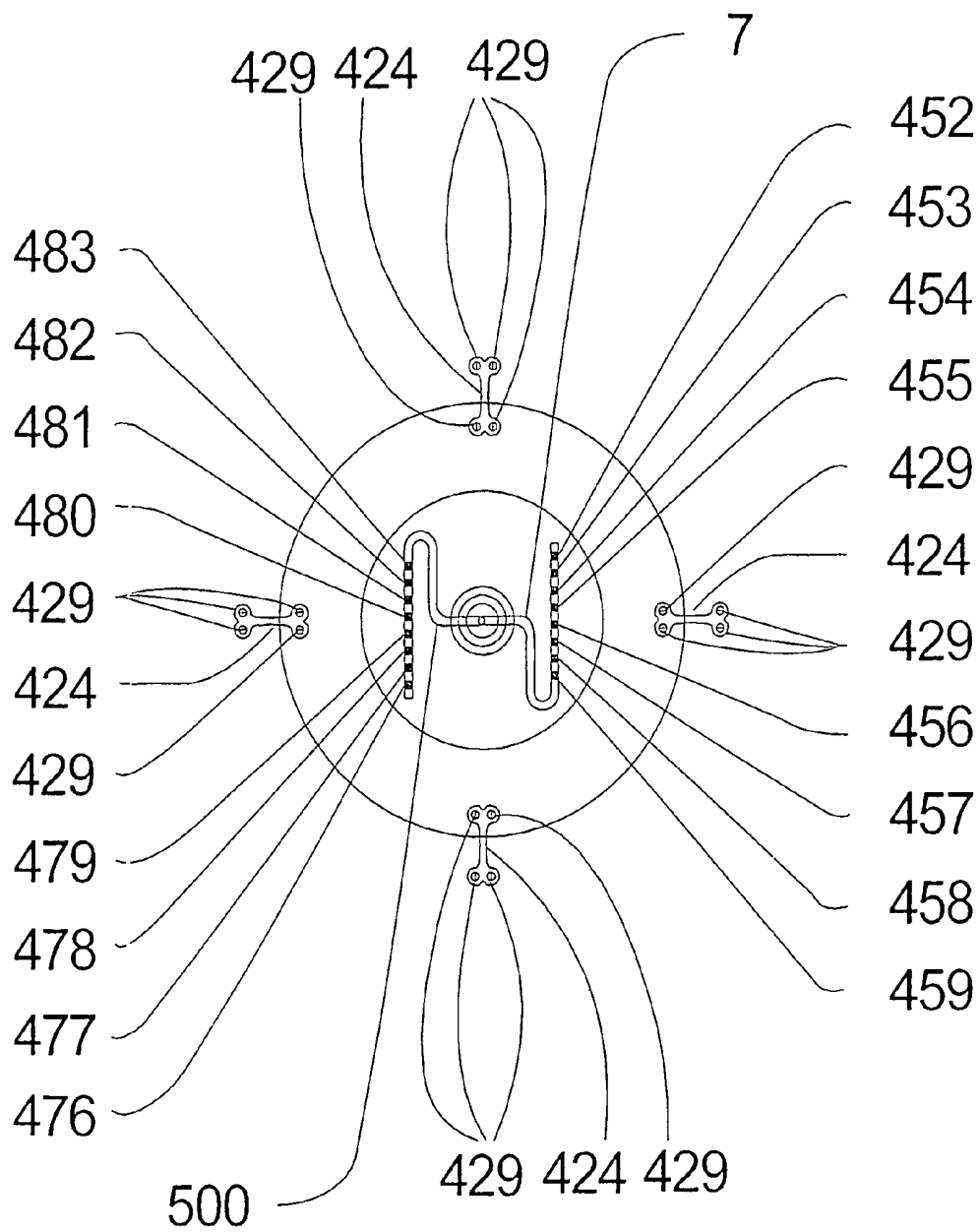


Figure 42

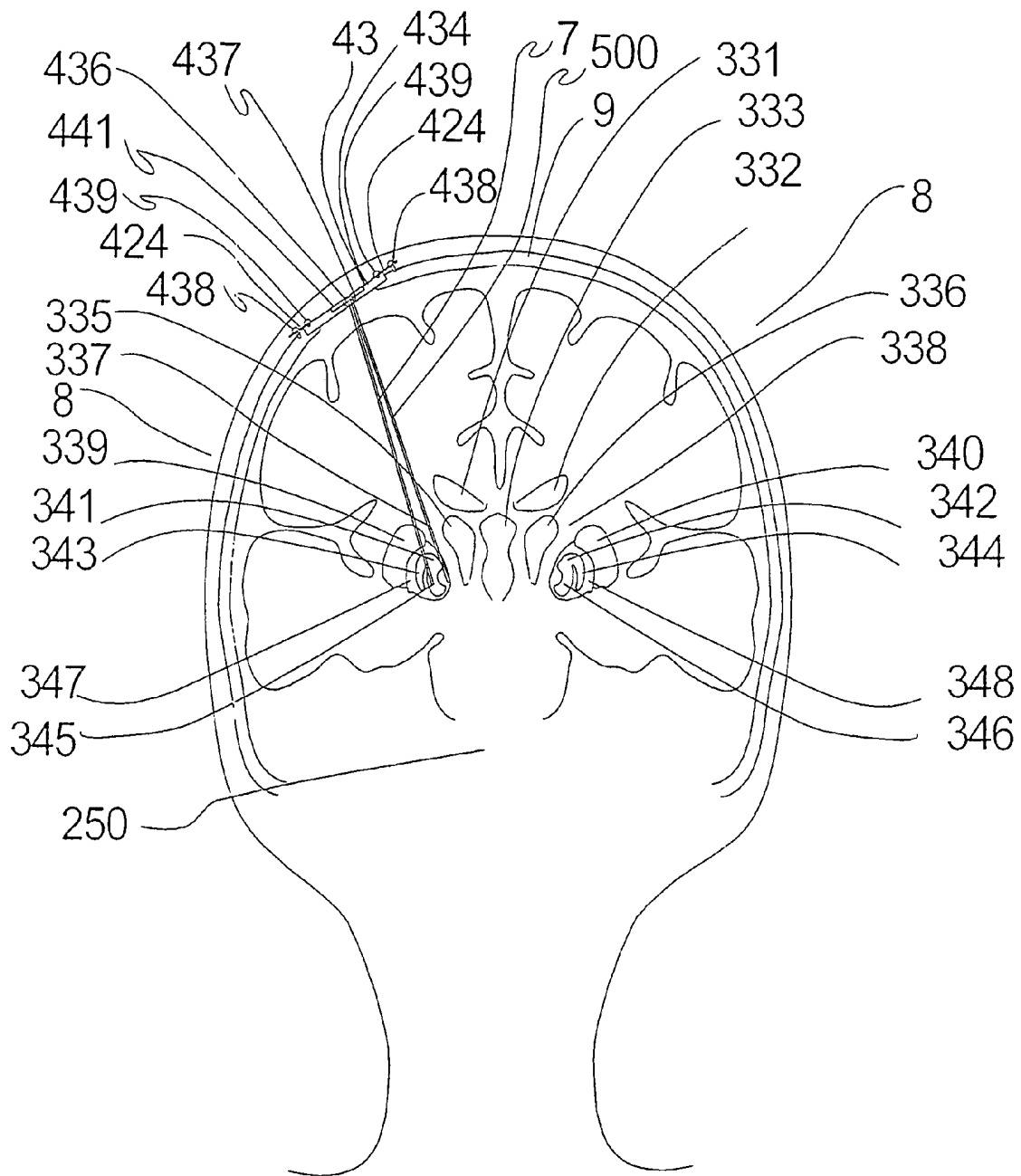


Figure 43

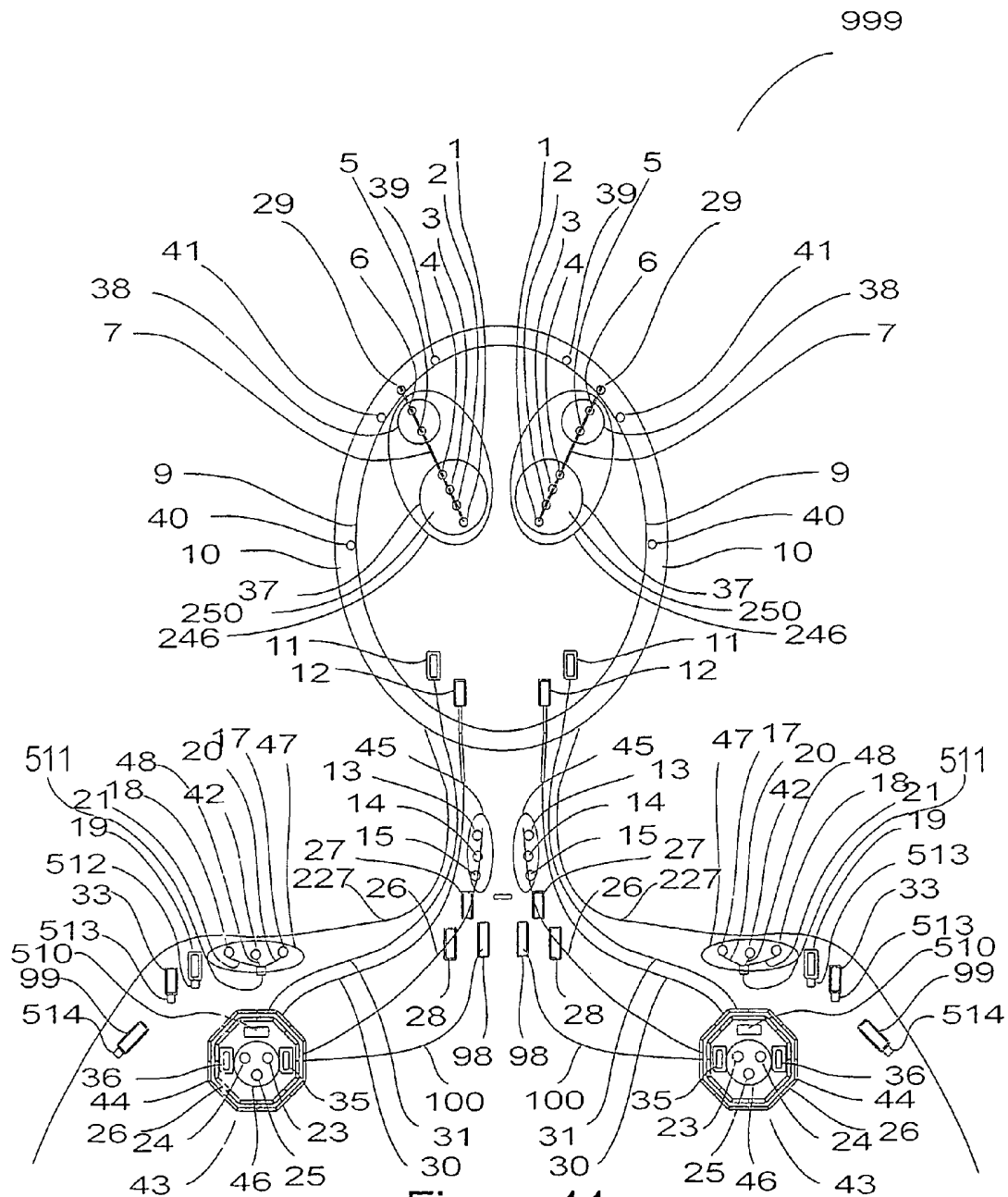


Figure 44

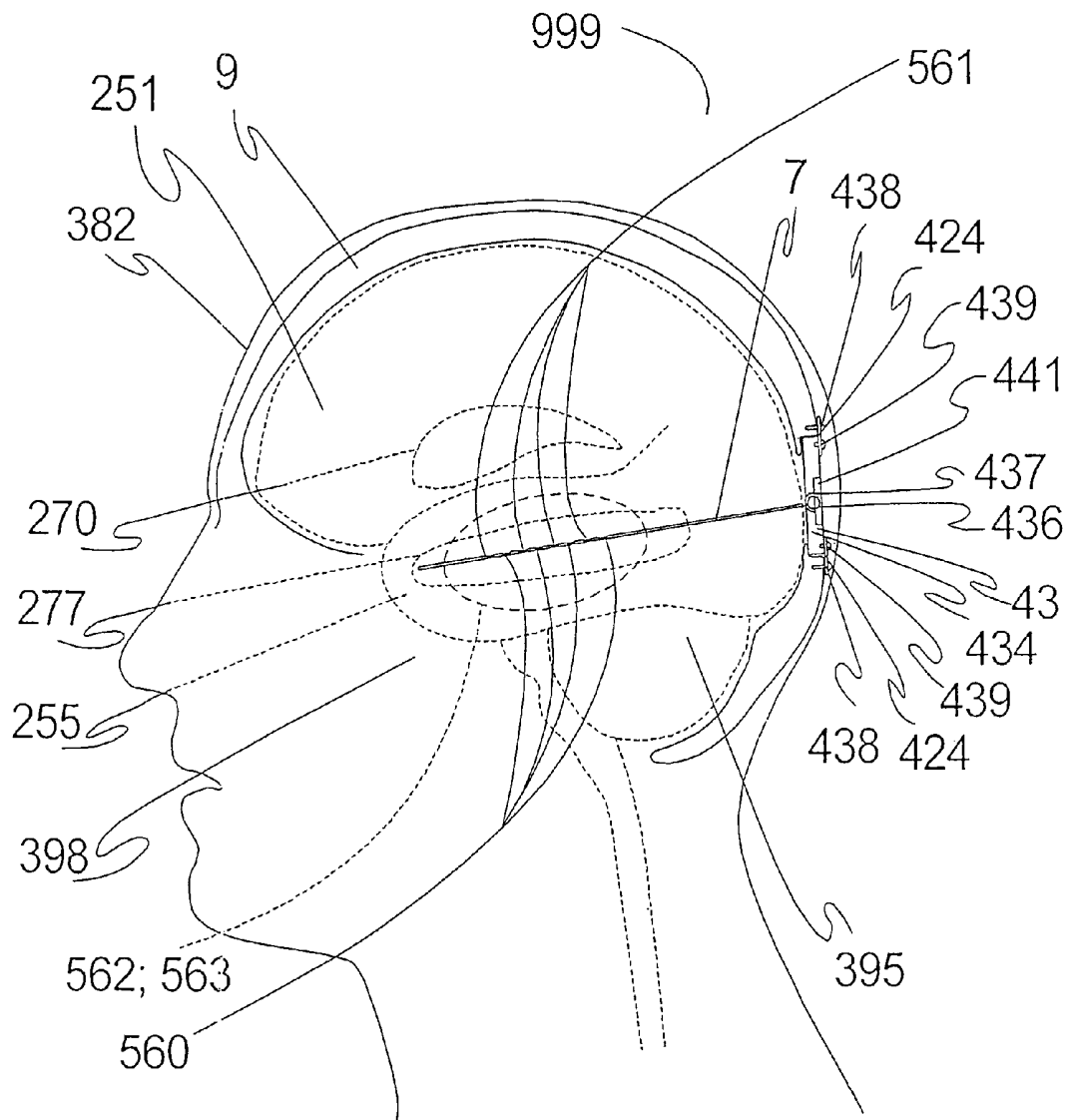


Figure 45

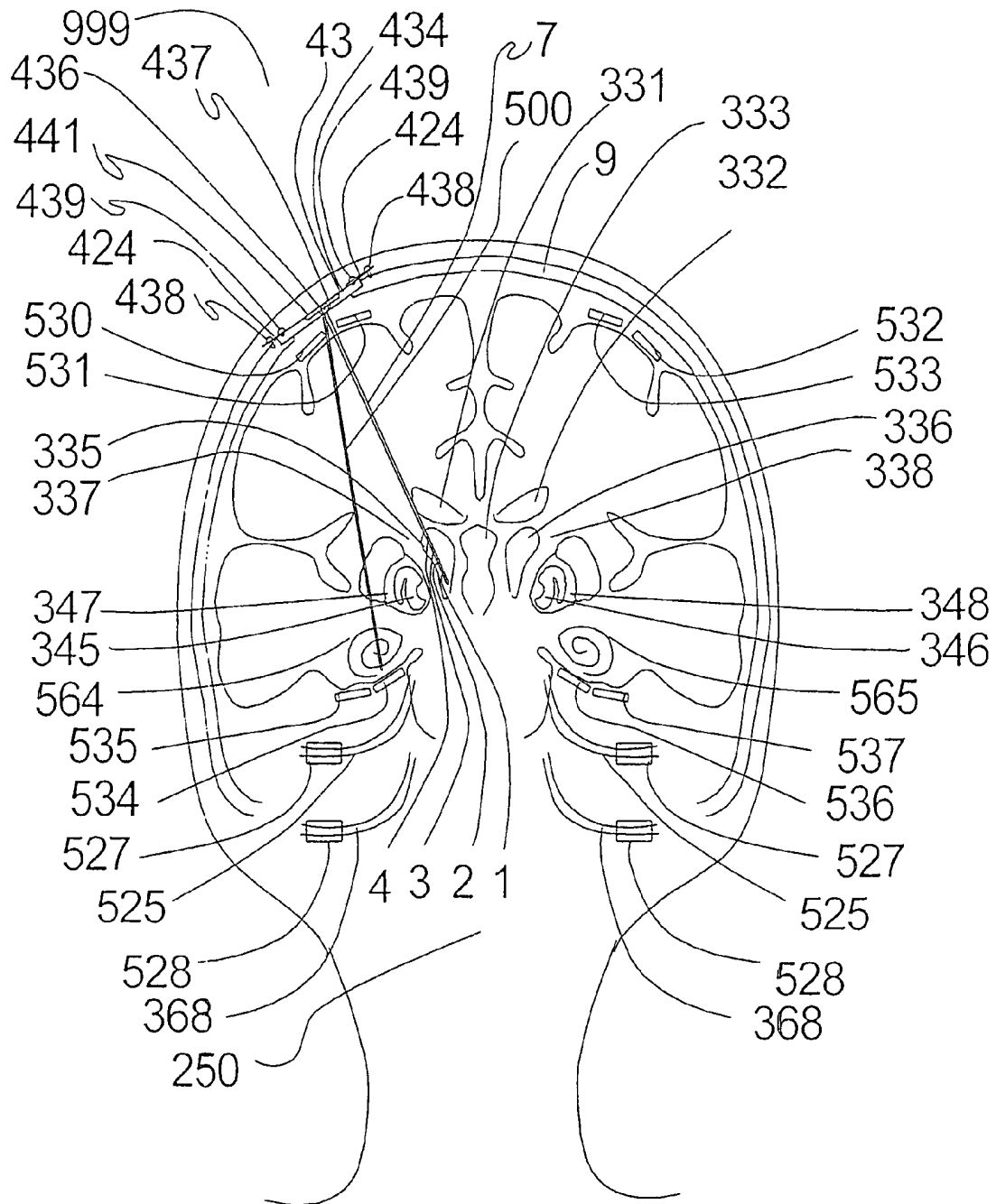
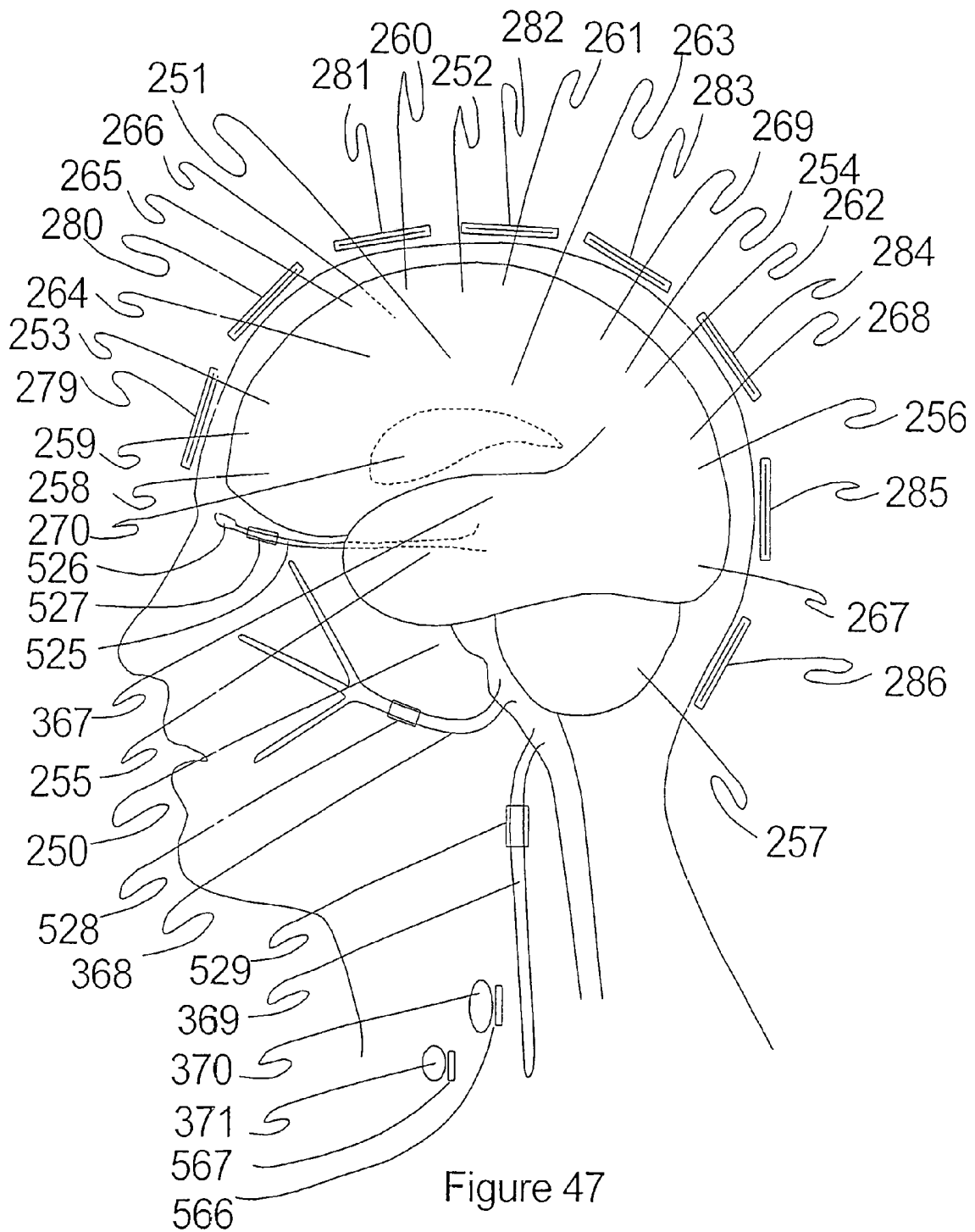


Figure 46



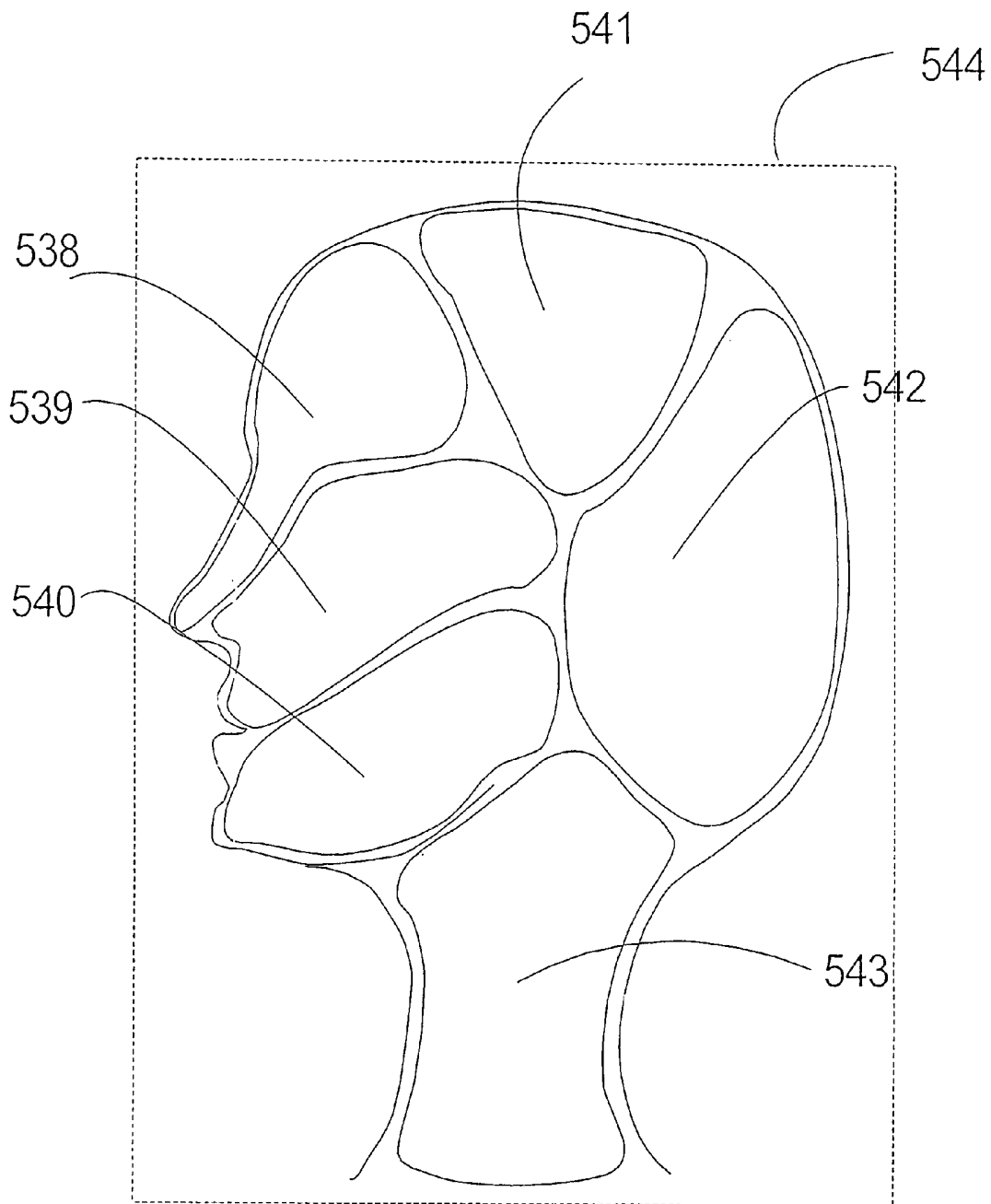


Figure 48

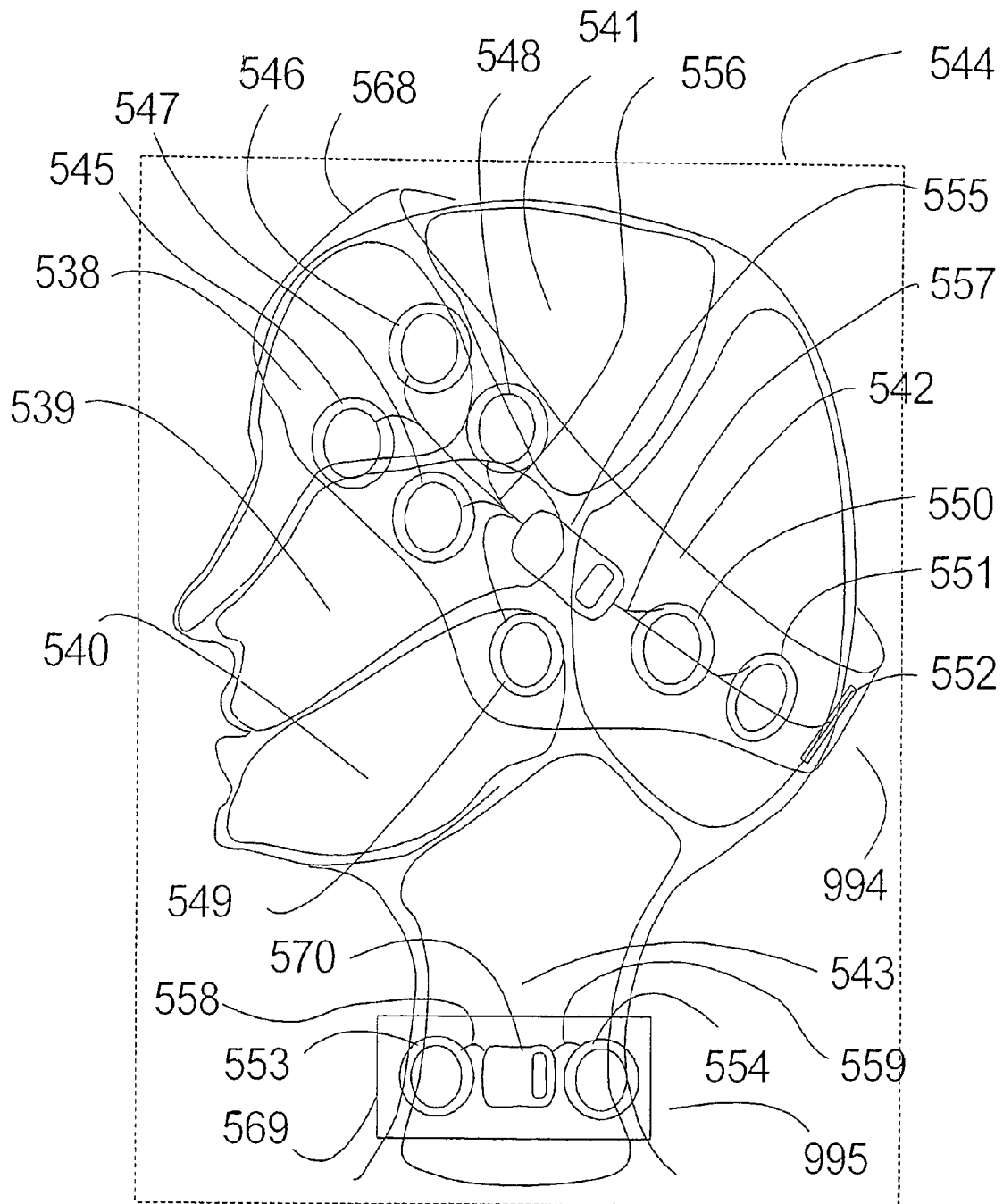


Figure 49

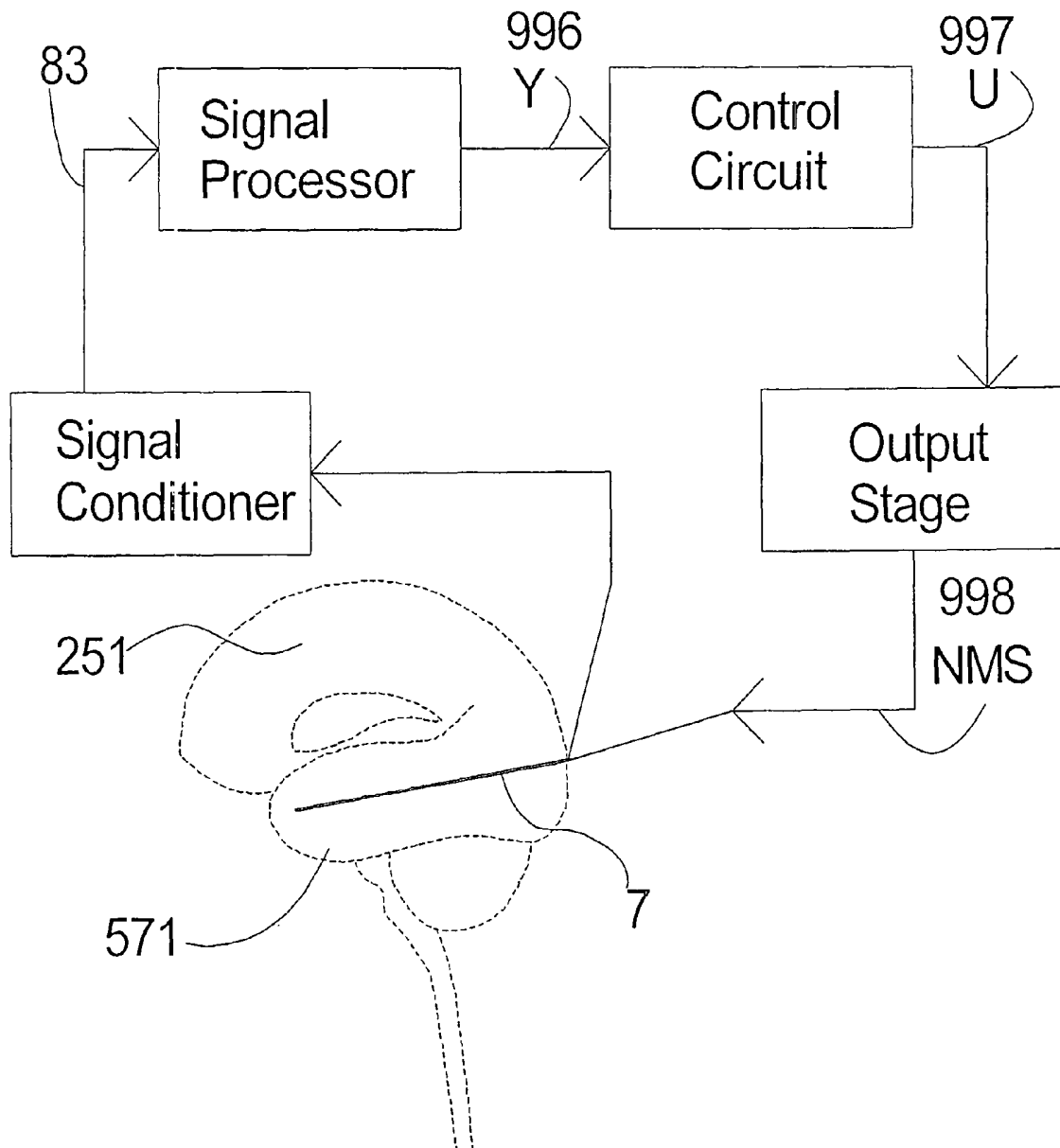


Figure 50

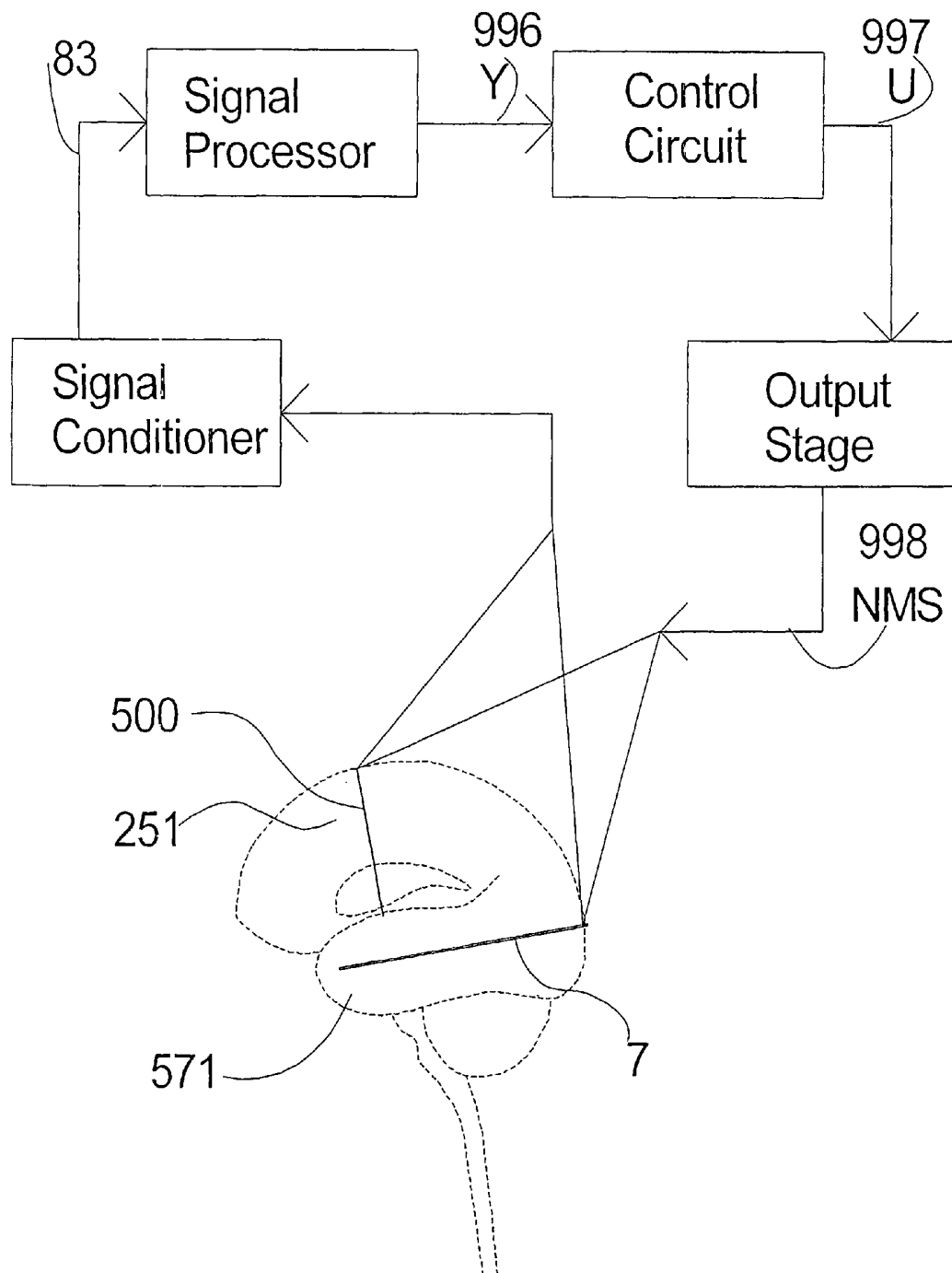


Figure 51

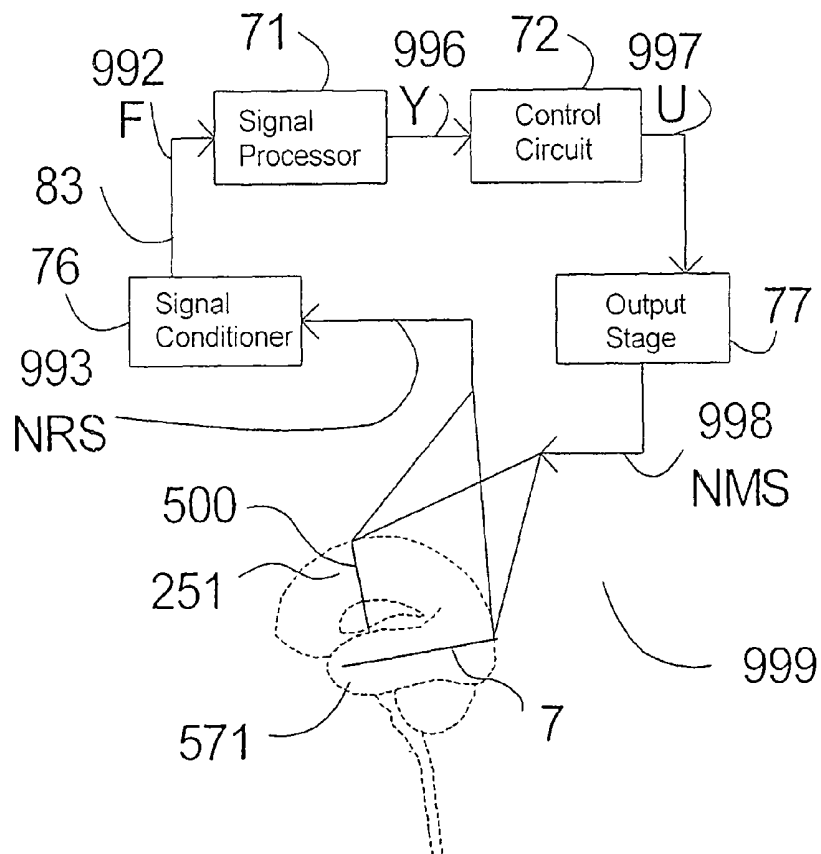
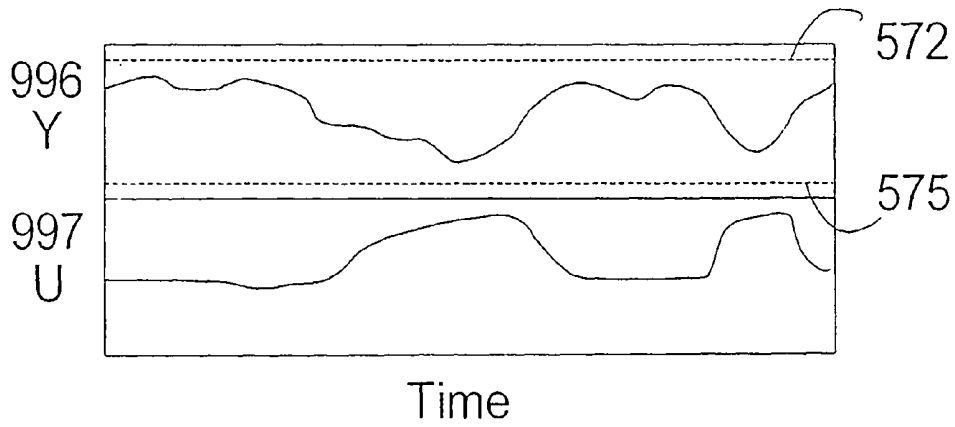


Figure 52

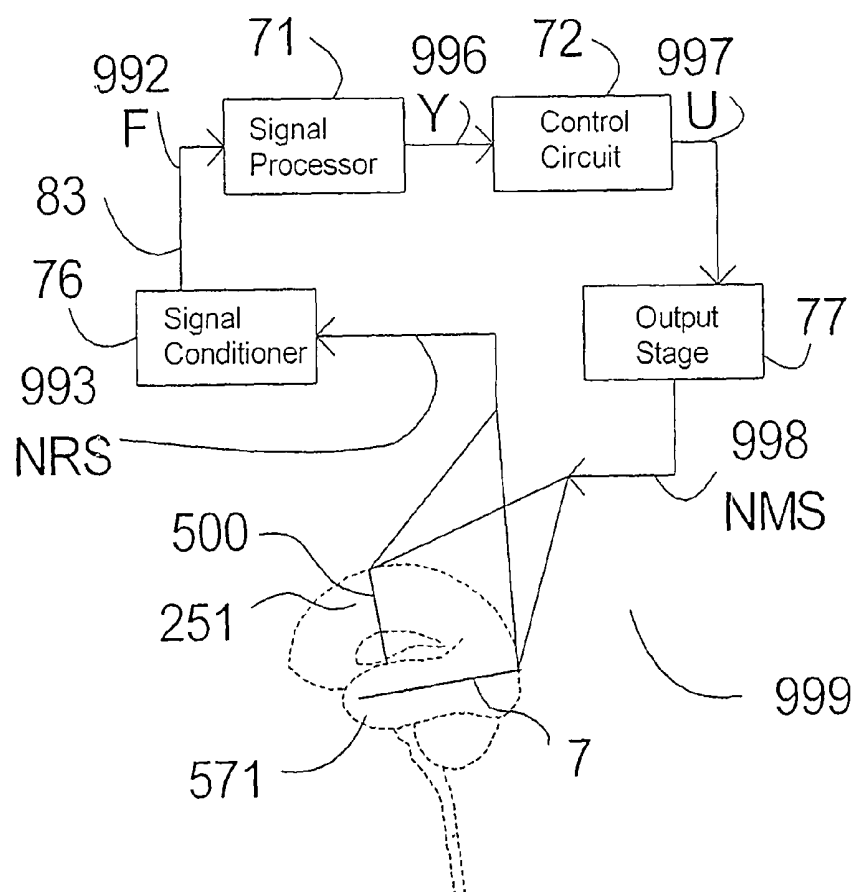
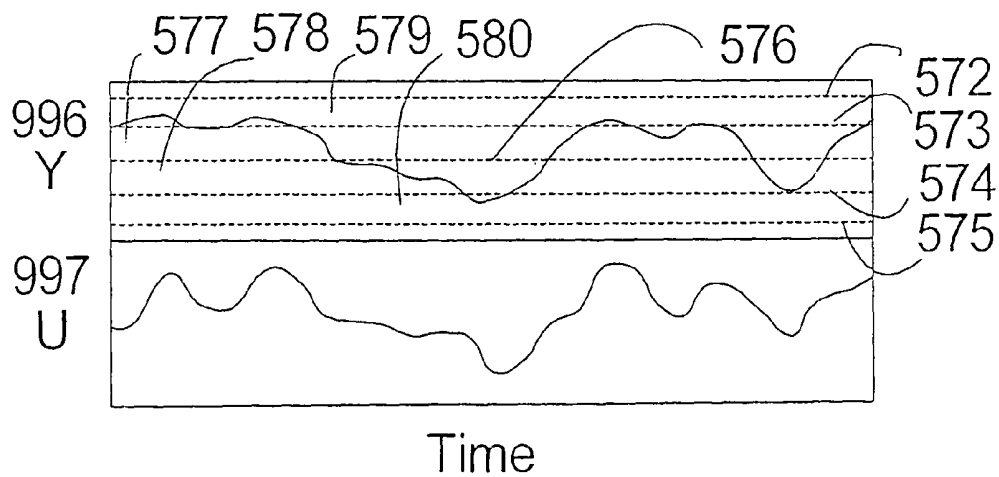


Figure 53

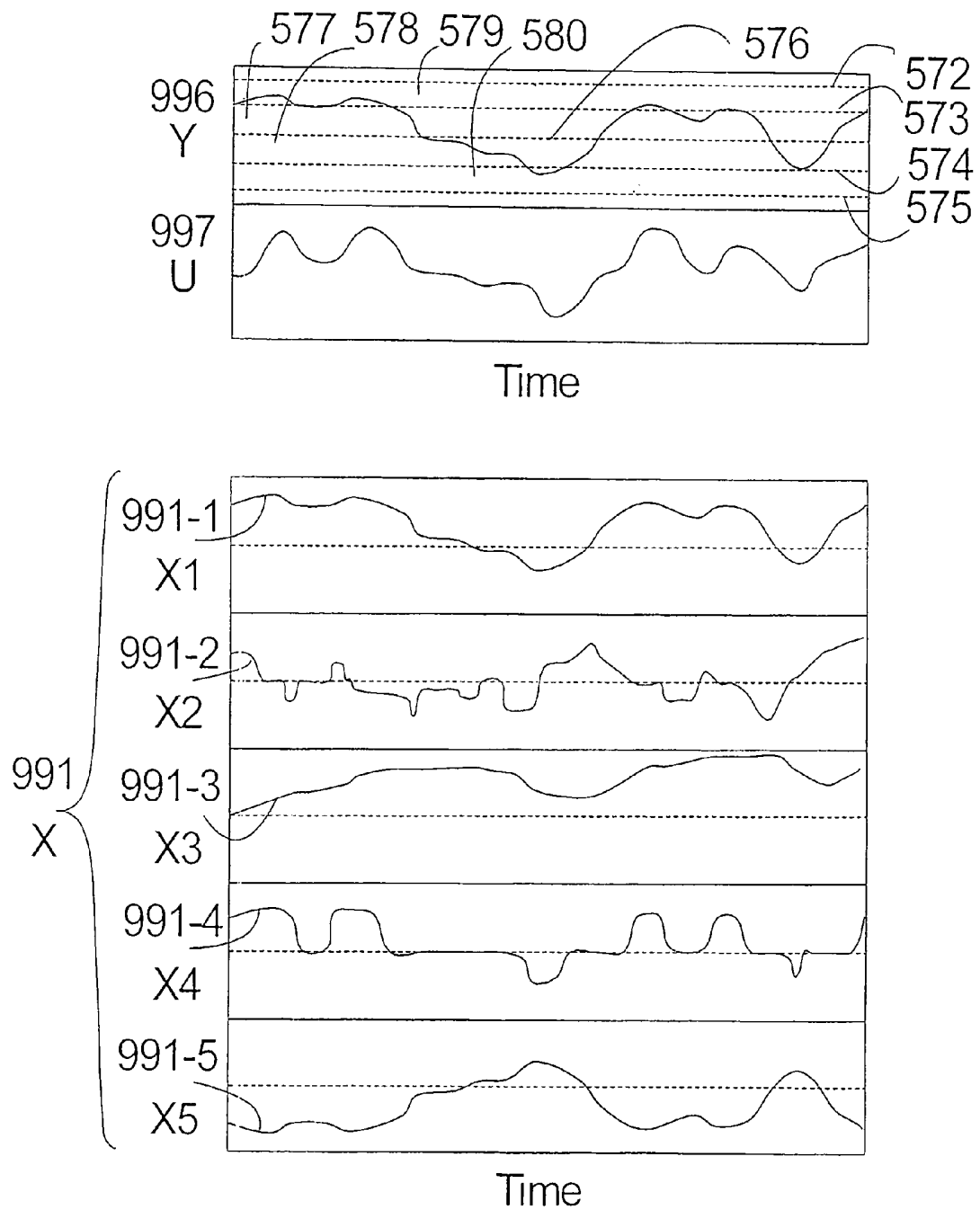


Figure 54

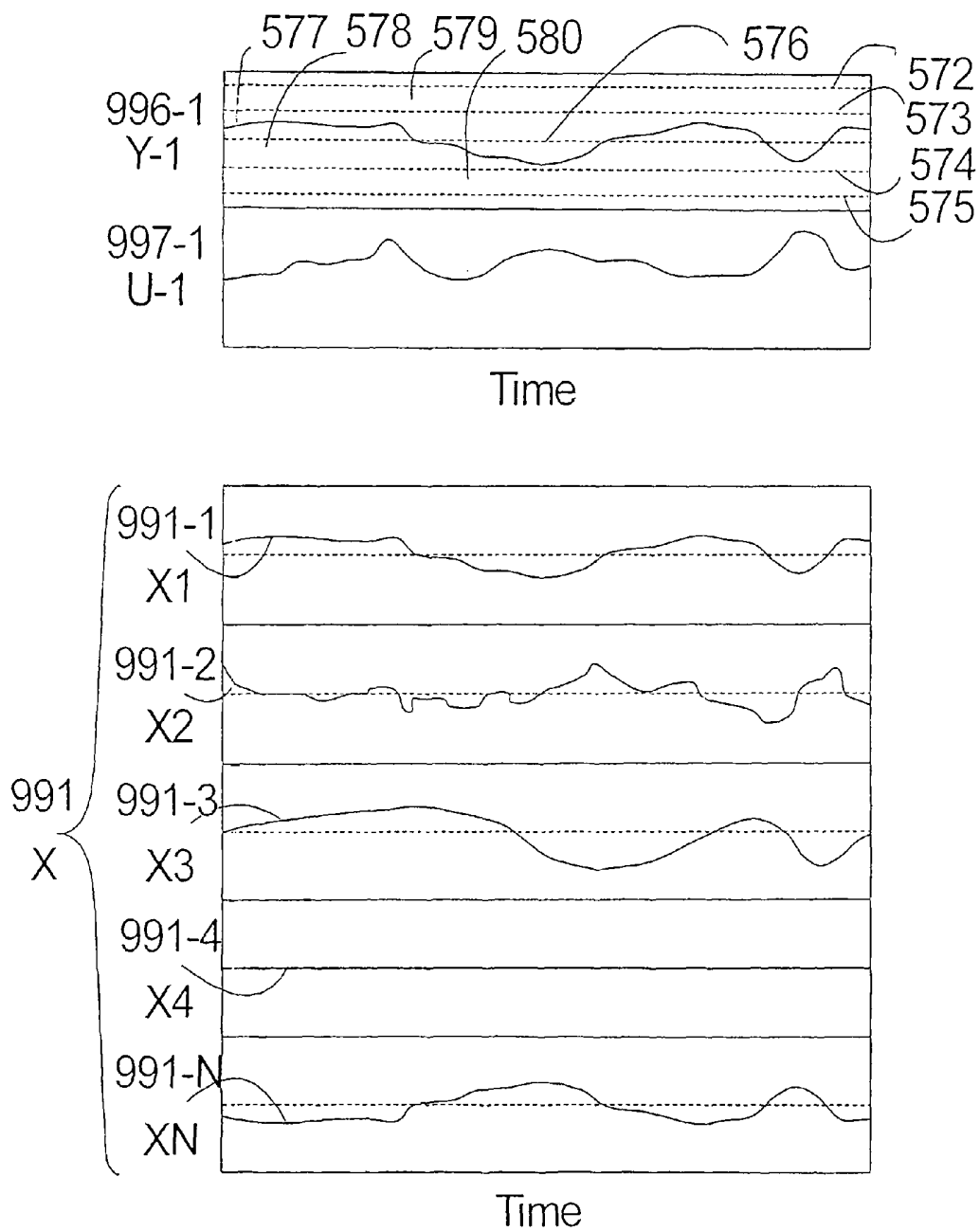


Figure 55

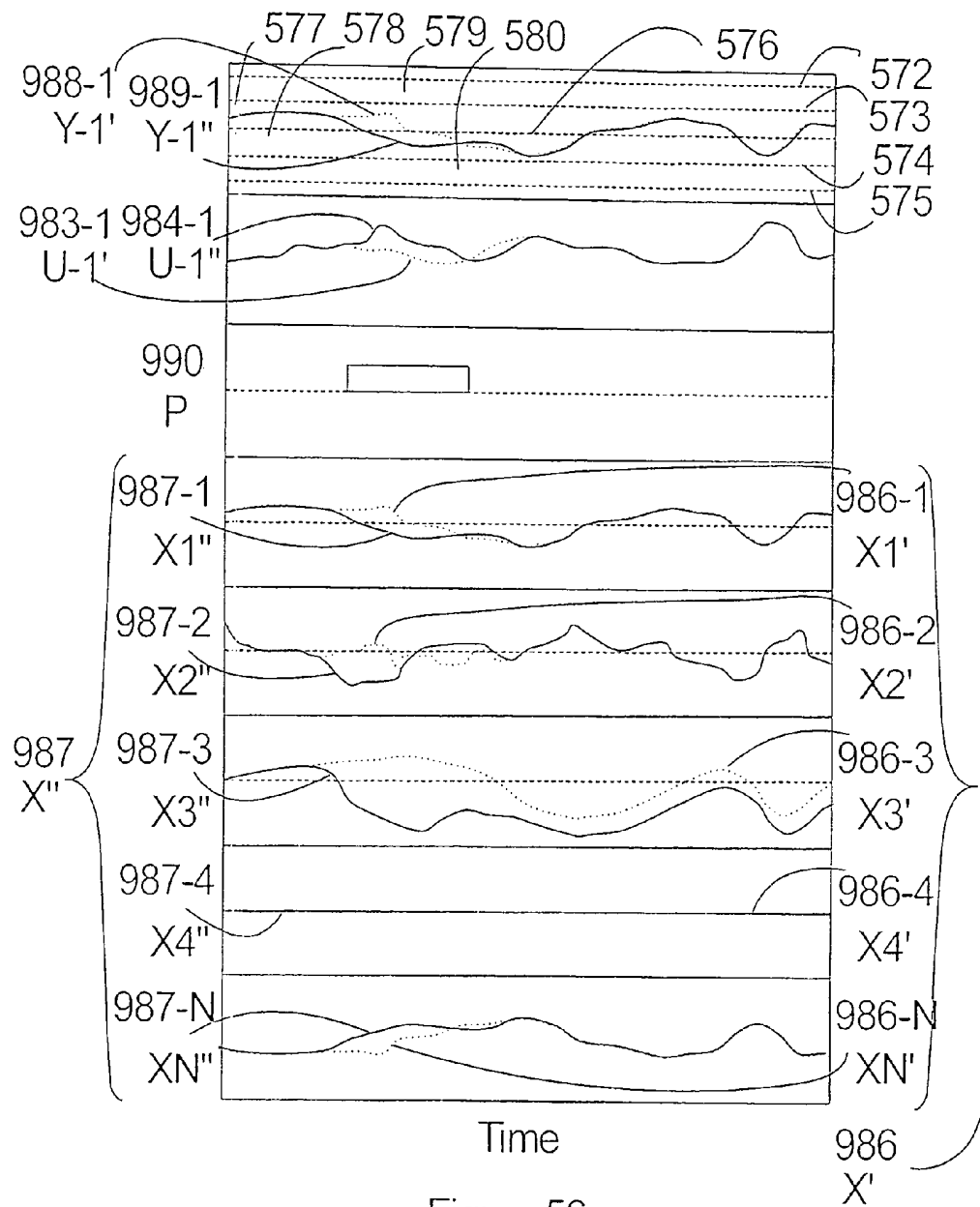
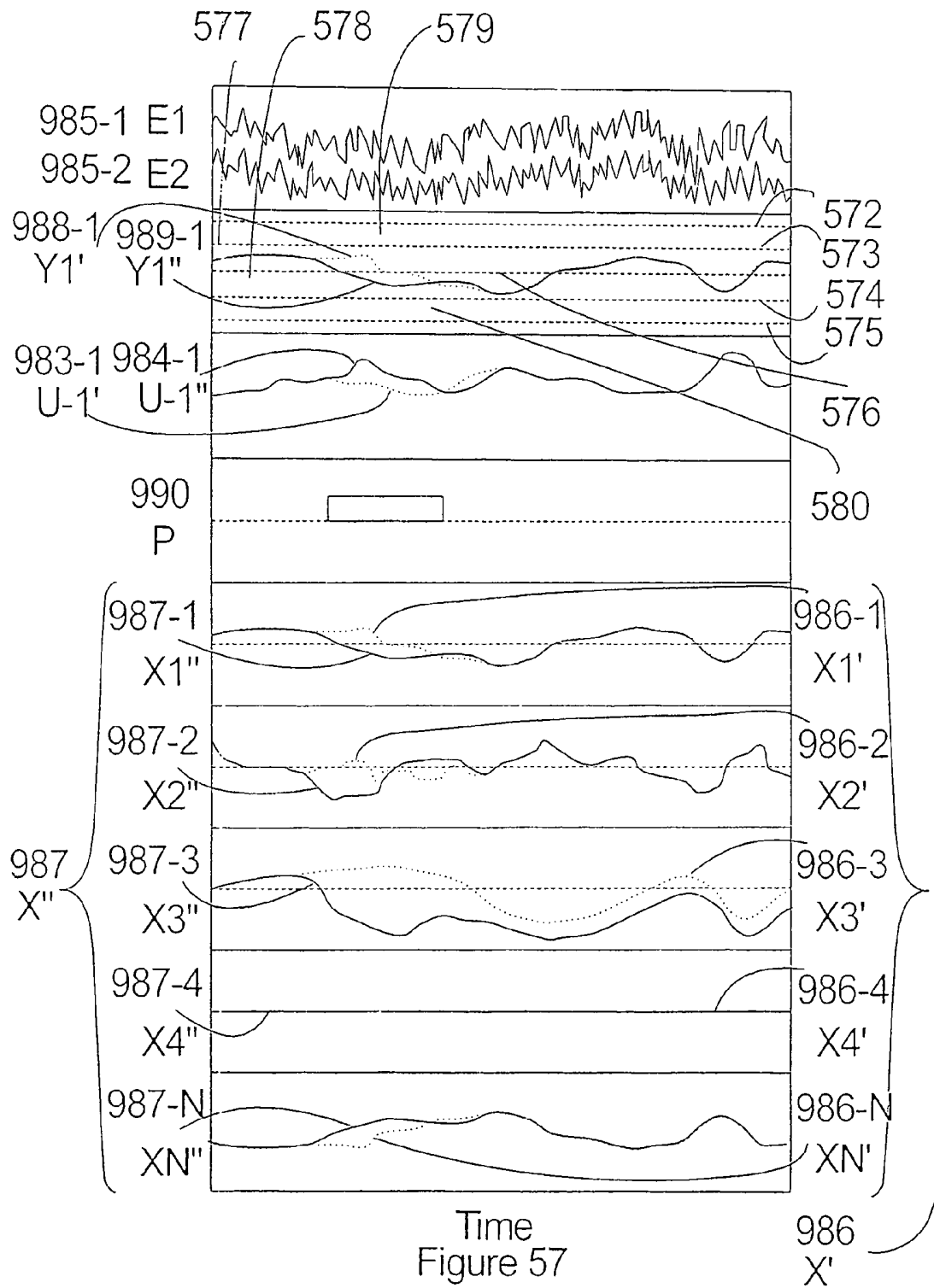
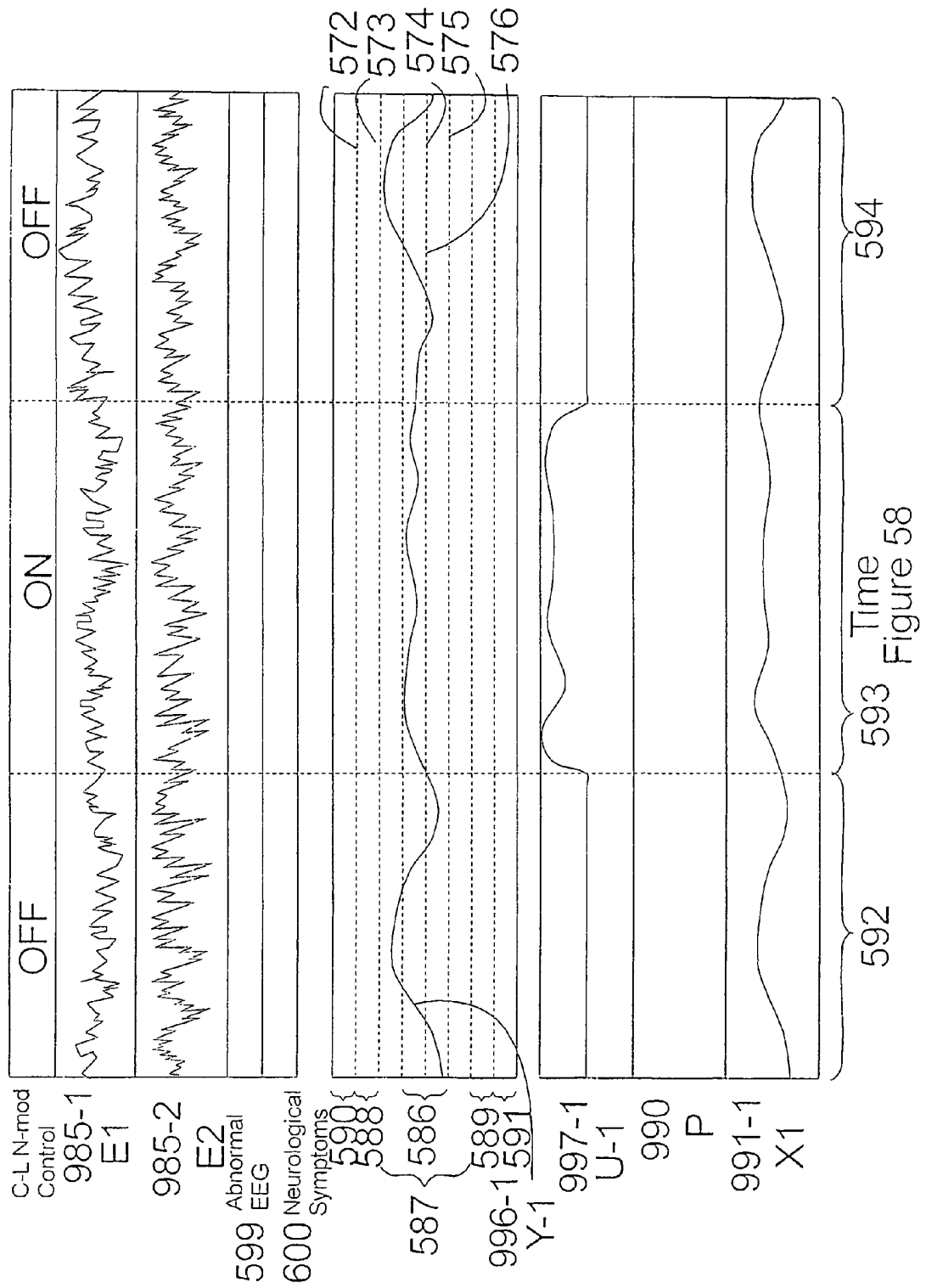
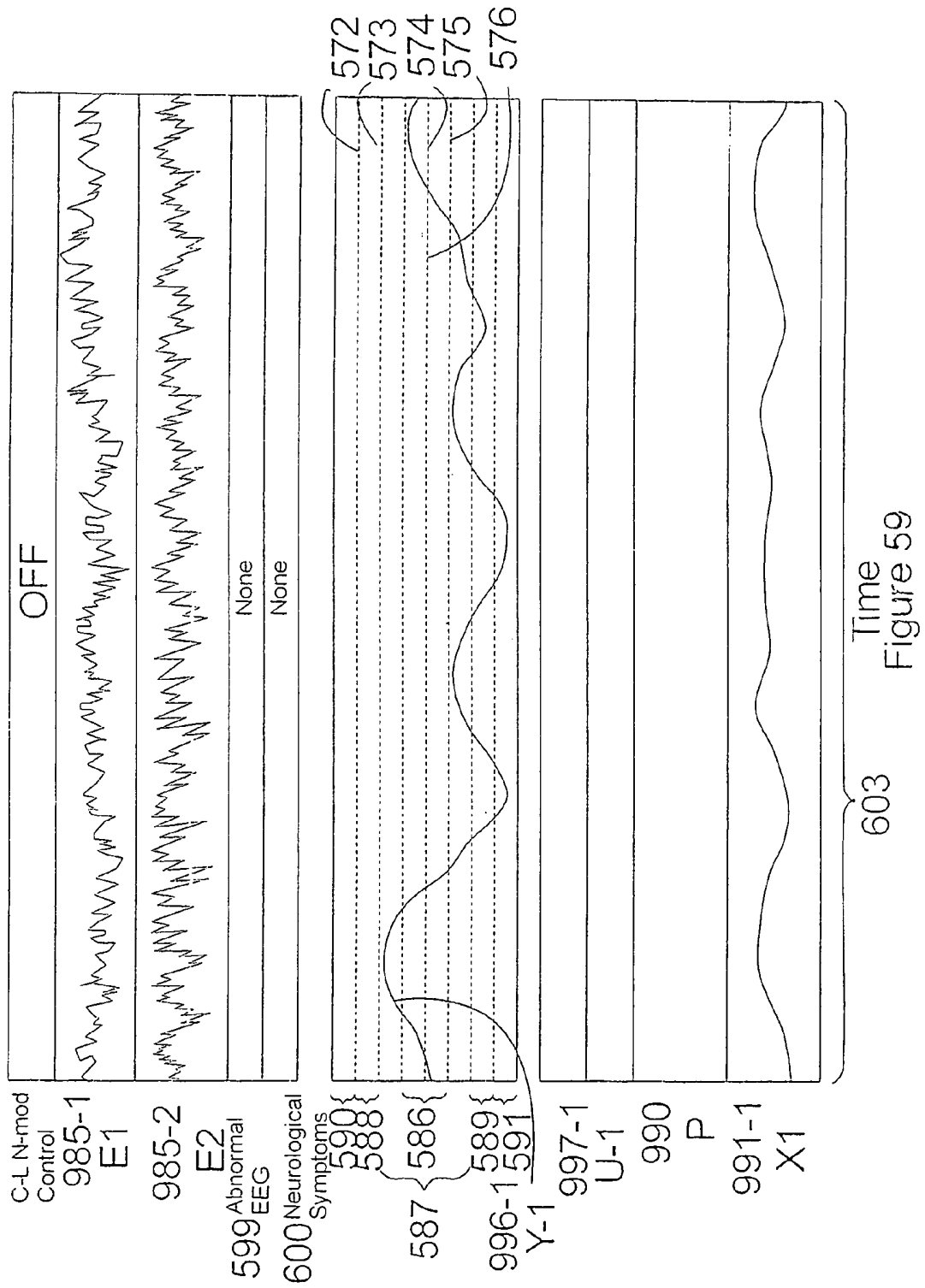
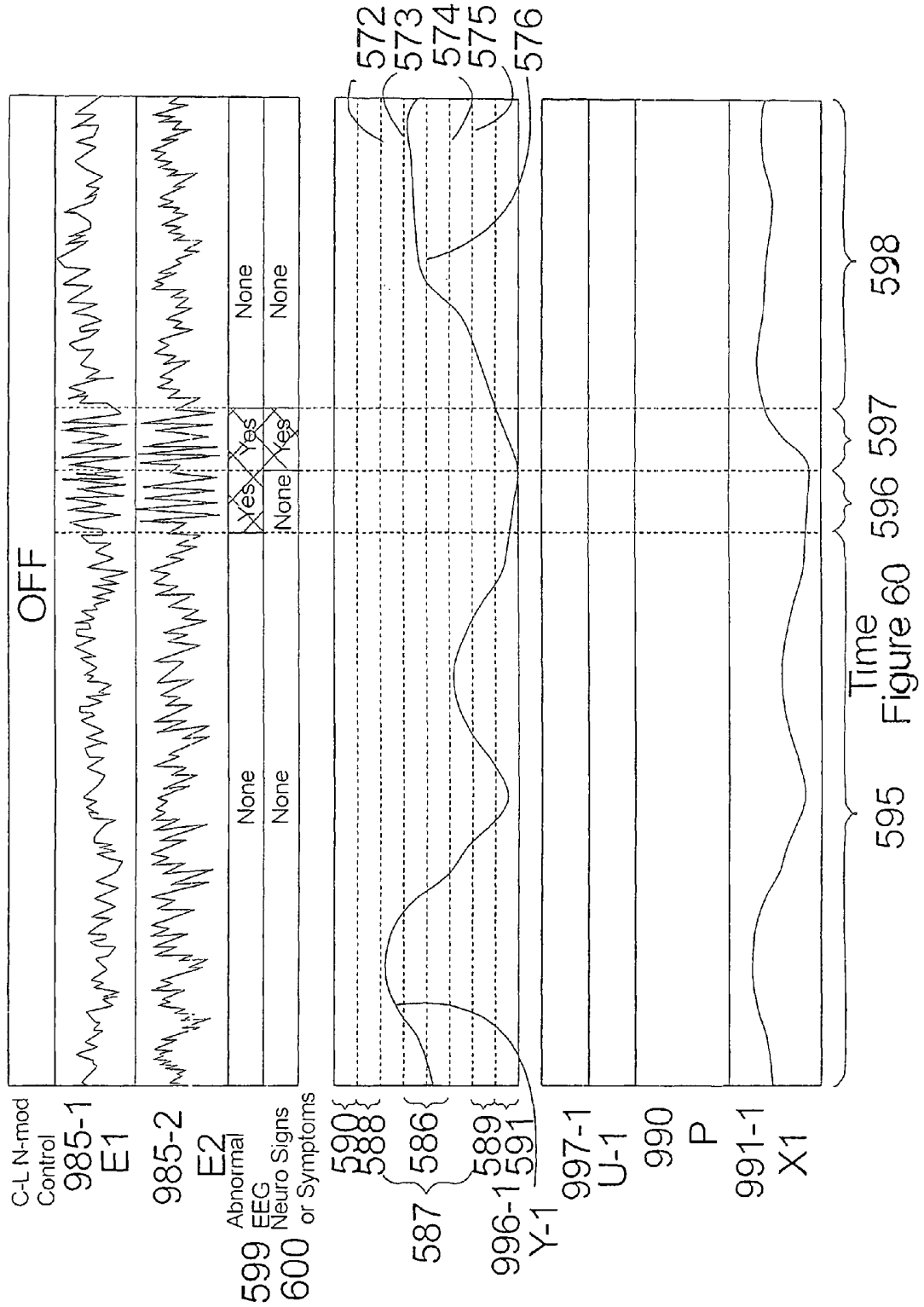


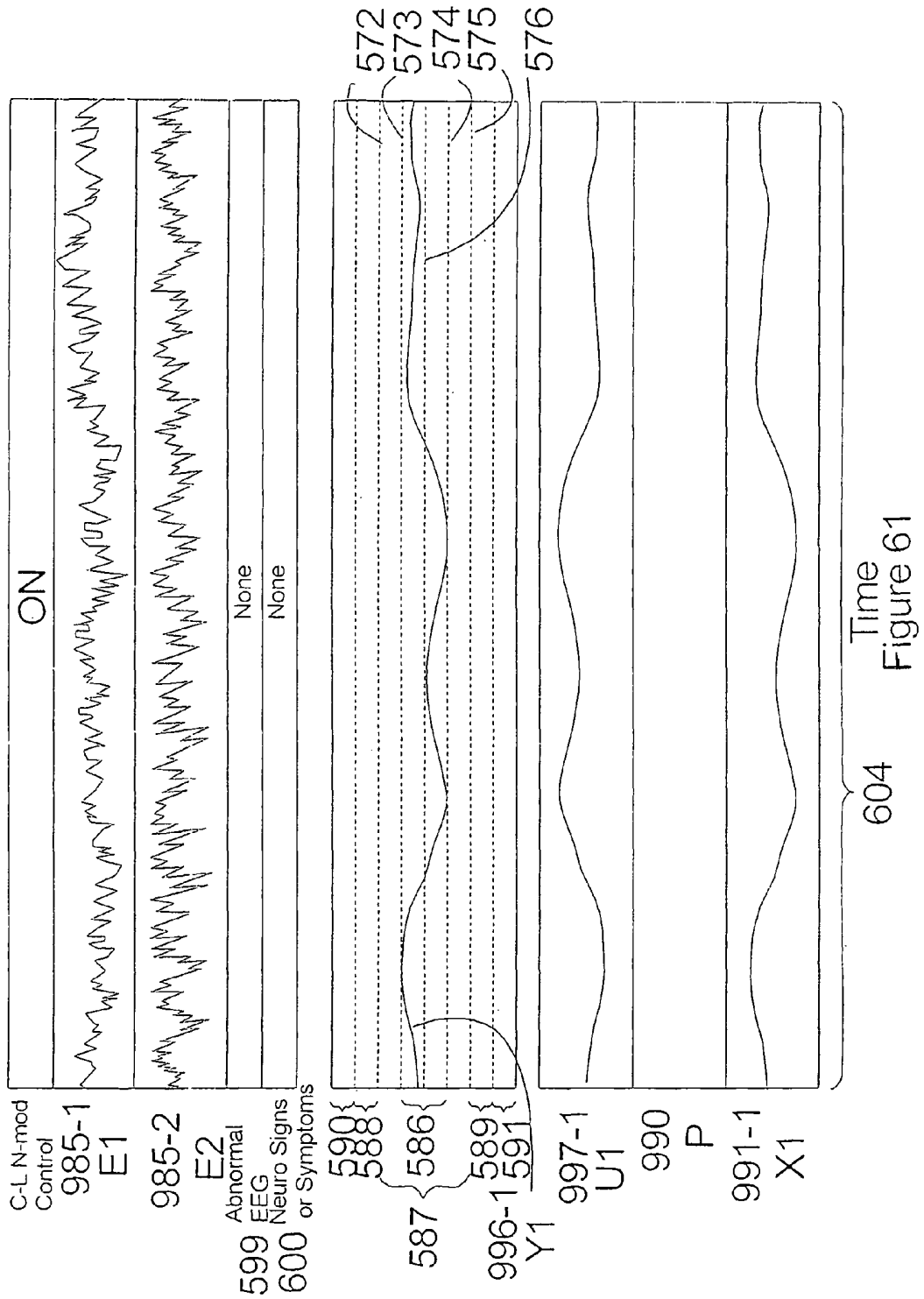
Figure 56

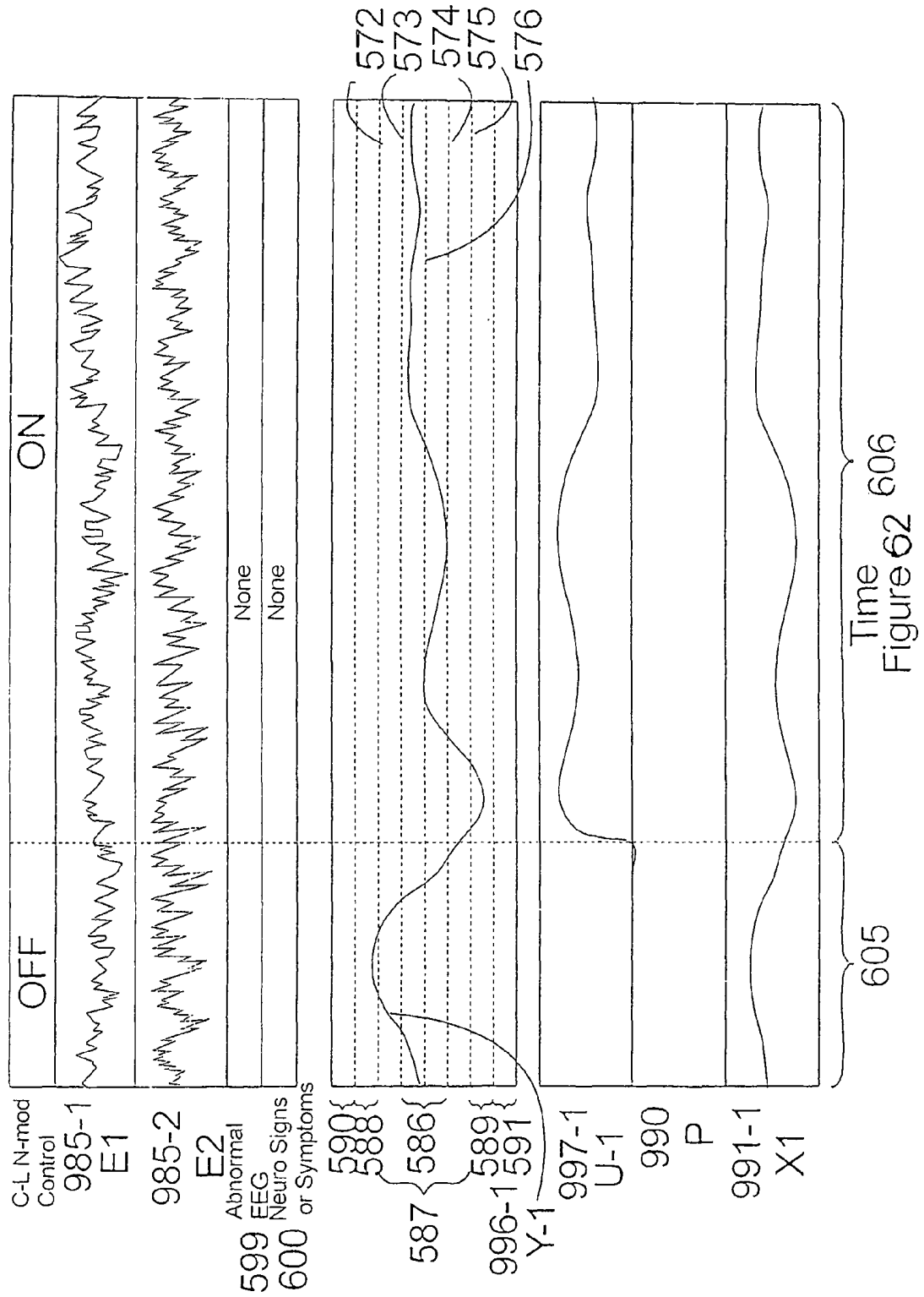


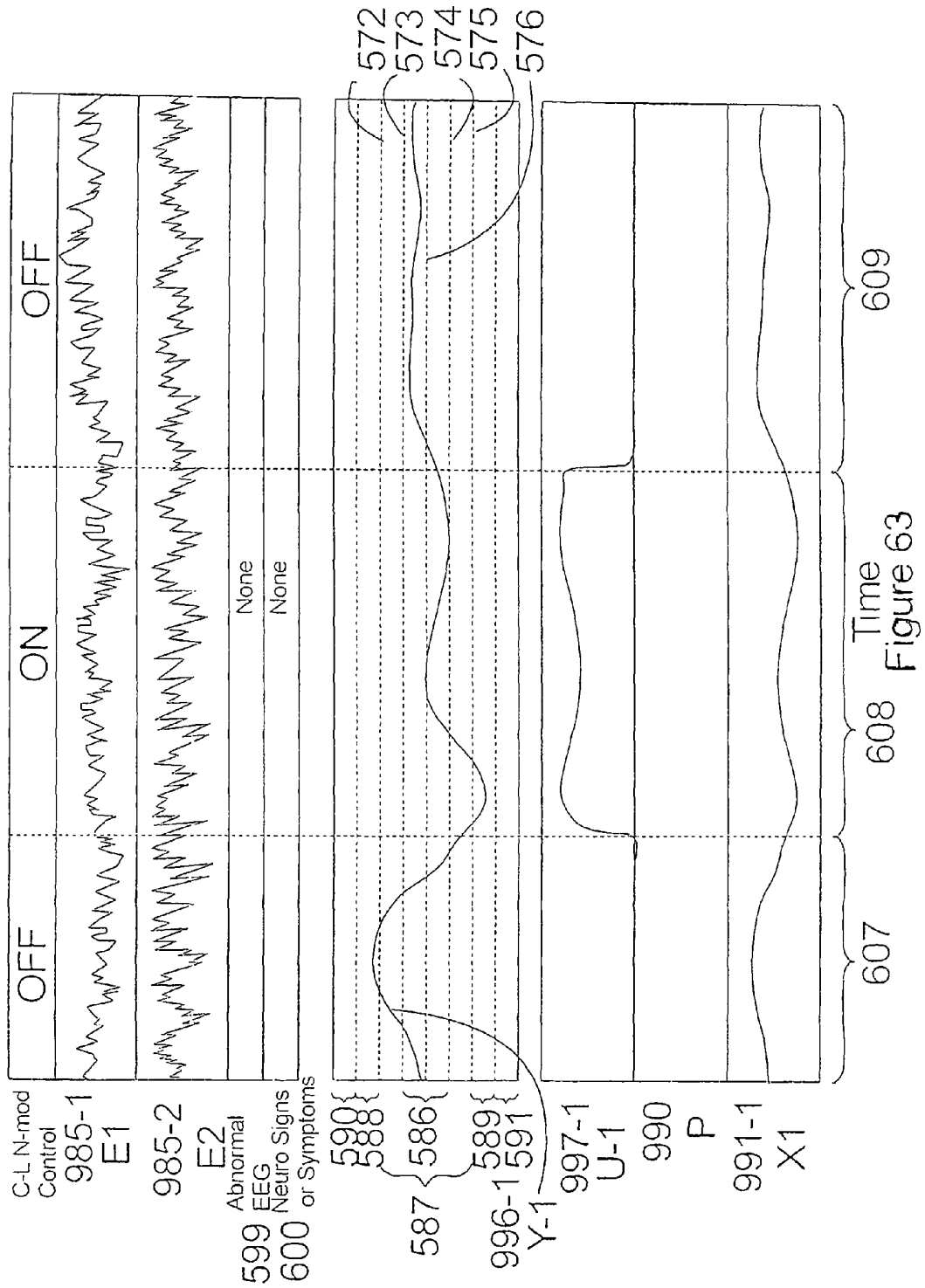


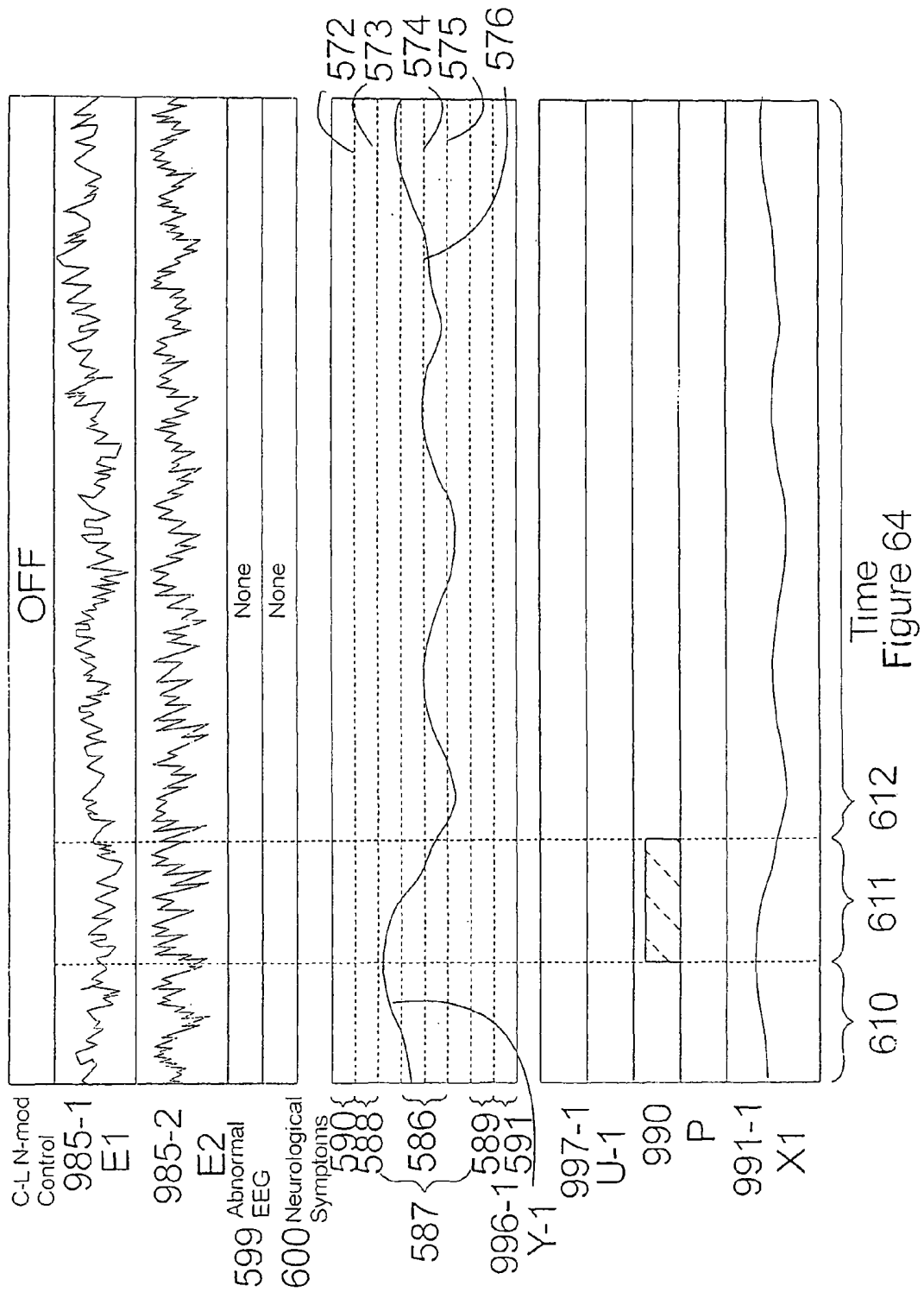


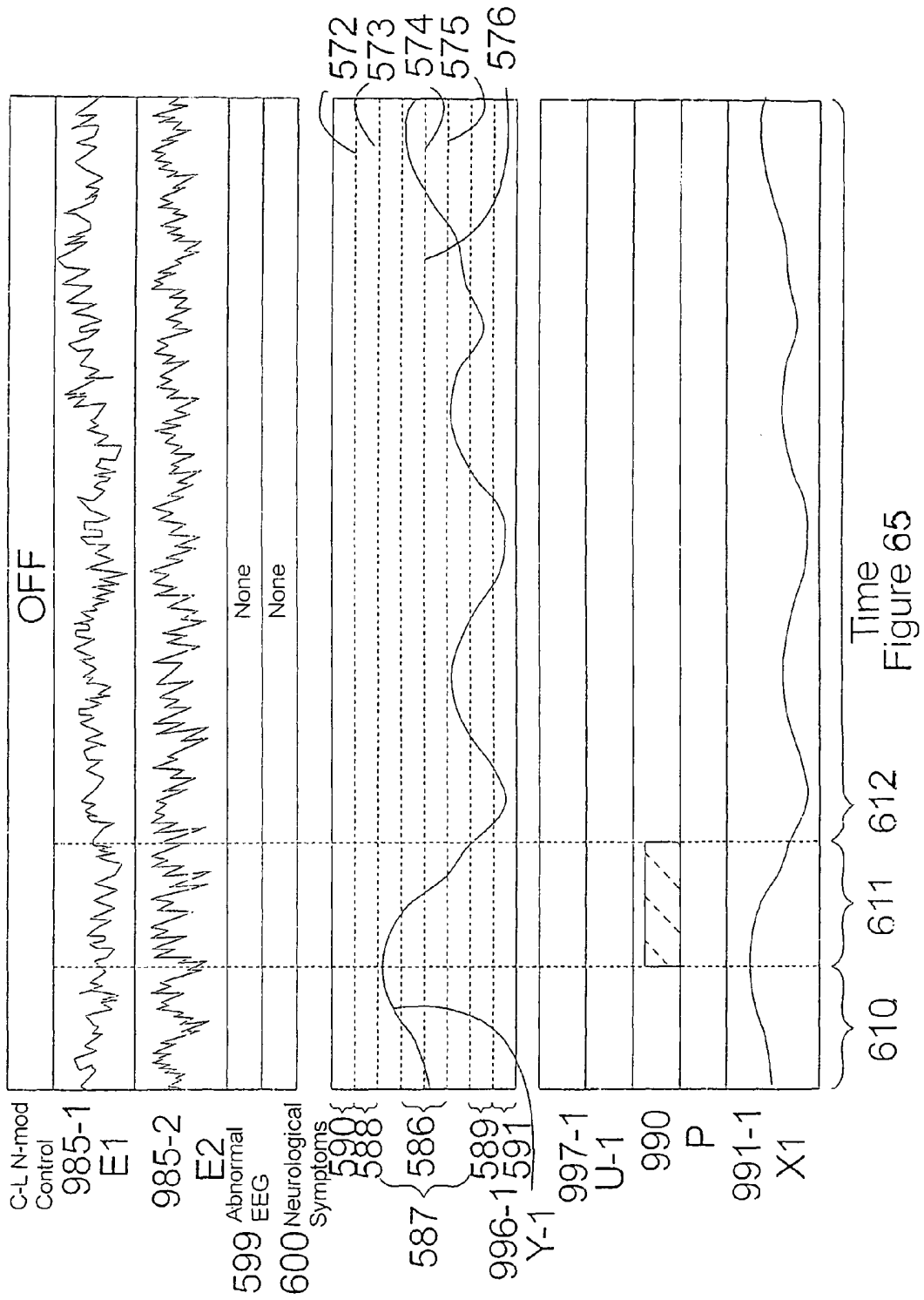


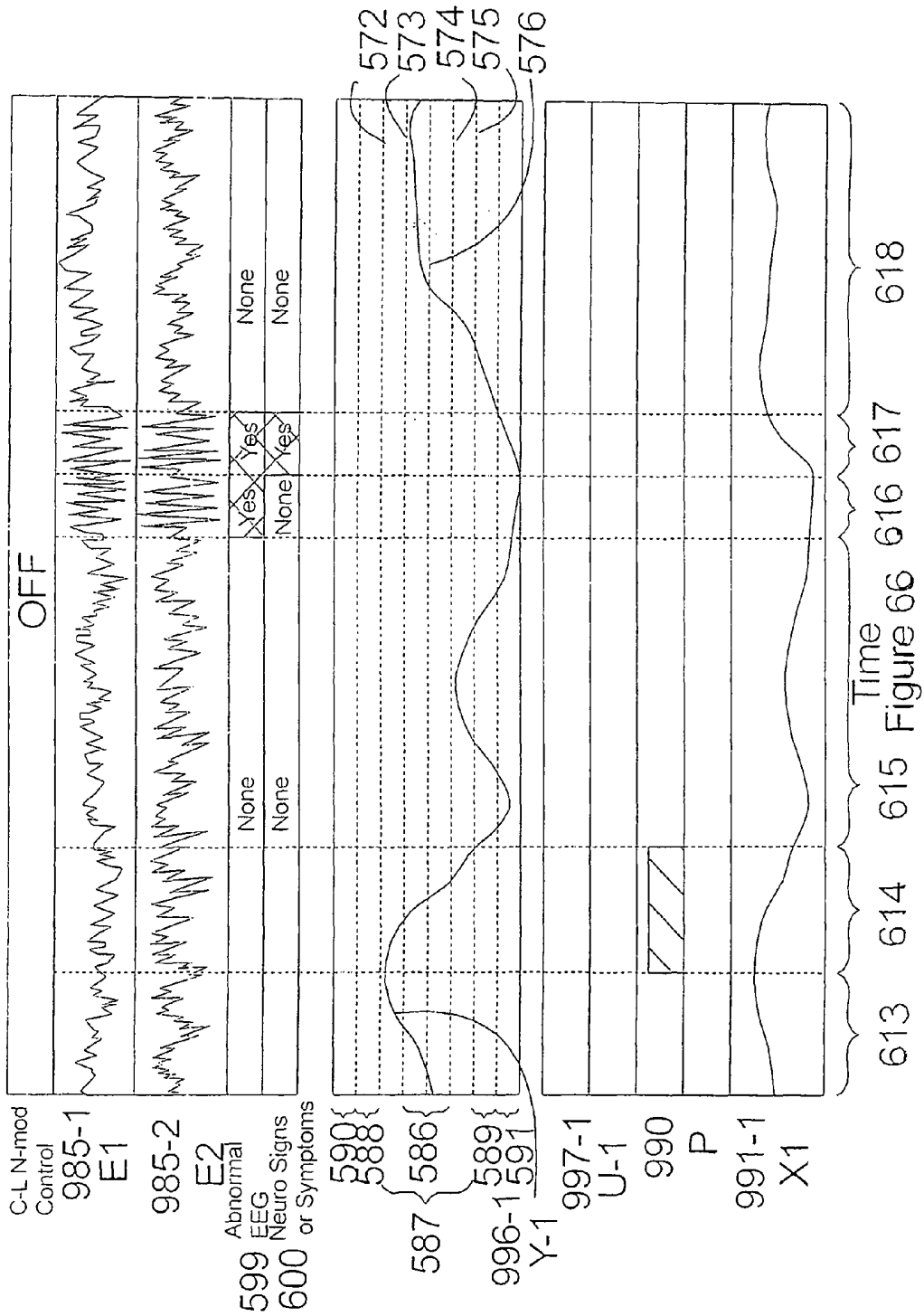


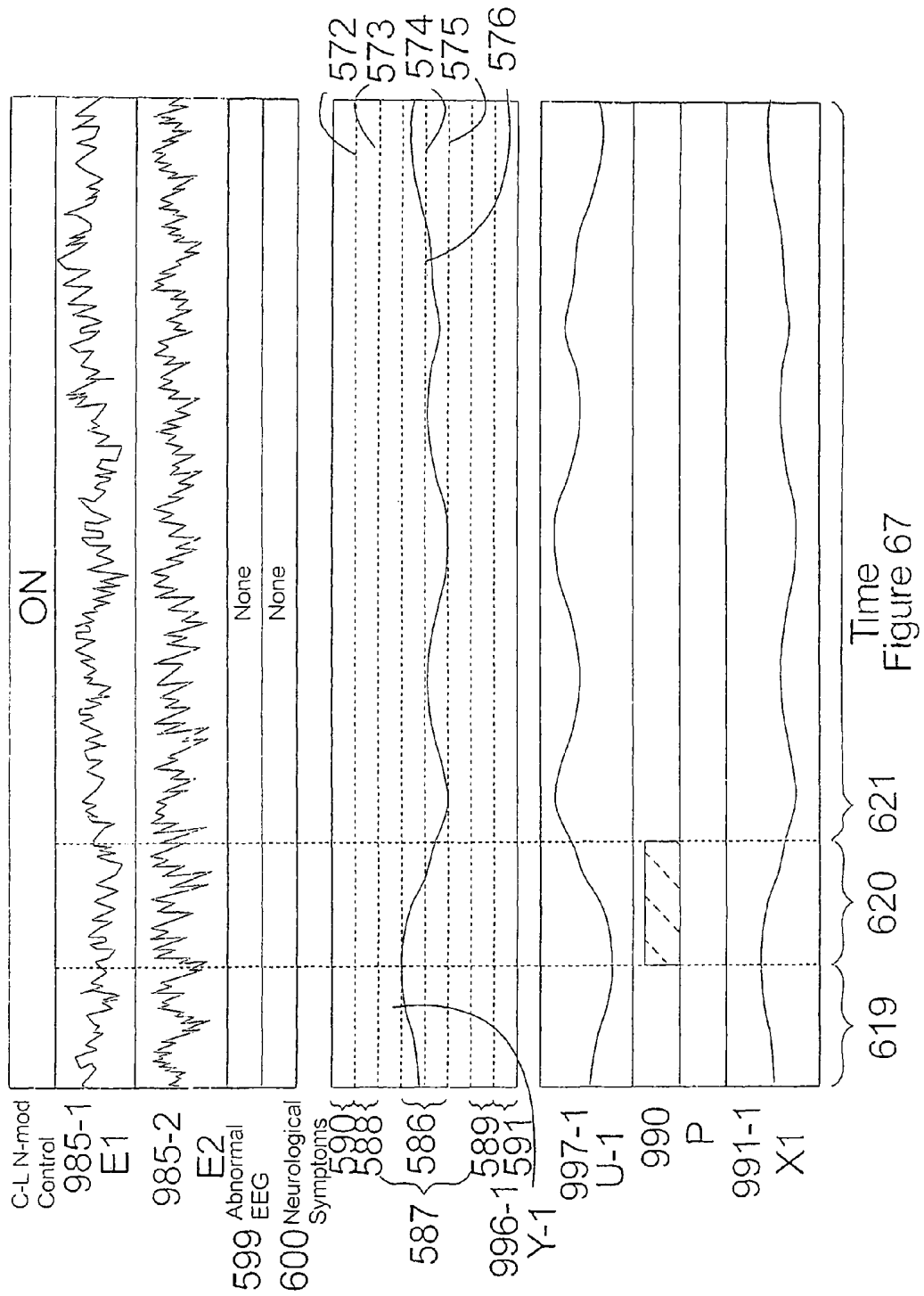


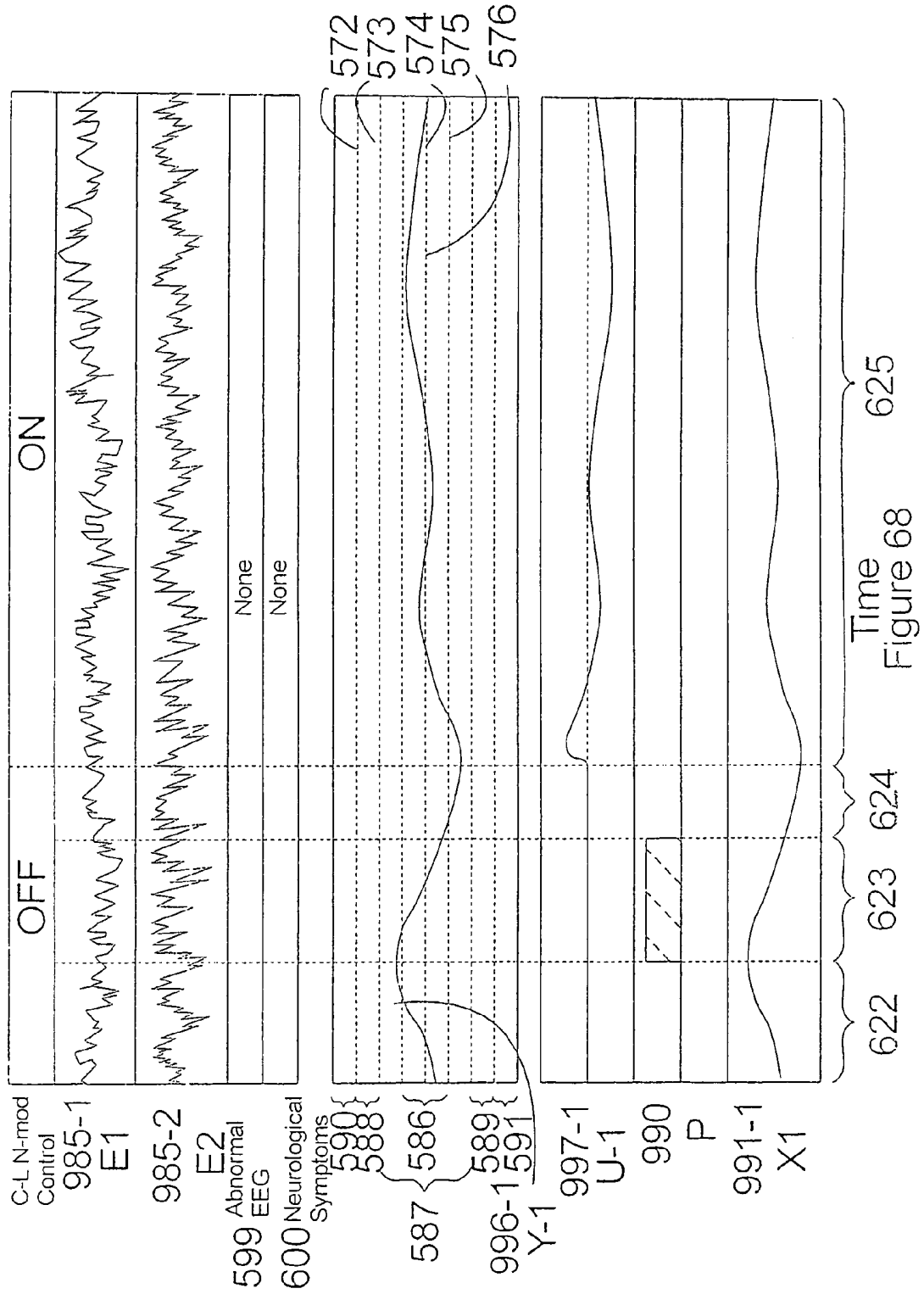












METHODS AND SYSTEMS FOR CONTINUOUS EEG MONITORING

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a continuation of application Ser. No. 10/889,844, filed Jul. 12, 2004, now U.S. Pat. No. 7,231,254, which claims benefit of U.S. Provisional Application No. 60/562,487, filed Apr. 14, 2004, and is a continuation-in-part of U.S. application Ser. No. 10/818,333, filed Apr. 5, 2004, now U.S. Pat. No. 7,277,758, which, in turn, claims benefit of U.S. Provisional Application No. 60/460,140, filed Apr. 3, 2003; U.S. application Ser. No. 10/818,333 is also a continuation-in-part of U.S. application Ser. No. 10/753,205, filed Jan. 6, 2004, now U.S. Pat. No. 7,242,984, which claims the benefit of U.S. Provisional Application No. 60/438,286, filed Jan. 6, 2003; U.S. application Ser. No. 10/753,205 is also a continuation-in-part of U.S. application Ser. No. 10/718,248, filed Nov. 20, 2003; now U.S. Pat. No. 7,209,787, which is a continuation-in-part of U.S. application Ser. No. 10/008,576, filed Nov. 11, 2001, now U.S. Pat. No. 6,819,956; which is a continuation-in-part of U.S. application Ser. No. 09/340,326, filed Jun. 25, 1999, now U.S. Pat. No. 6,366,813; which claims the benefit of U.S. Provisional Application No. 60/095,413, filed Aug. 5, 1998; U.S. application Ser. No. 10/718,248 also claims benefit of U.S. Provisional Application Nos. 60/427,699, filed Nov. 20, 2002, and 60/436,792, filed Dec. 27, 2002.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates generally to neurological disease and, more particularly, to intracranial stimulation for optimal control of movement disorders and other neurological disease.

2. Related Art

There are a wide variety of treatment modalities for neurological disease including movement disorders such as Parkinson's disease, Huntington's disease, and Restless Leg Syndrome, as well as psychiatric disease including depression, bipolar disorder and borderline personality disorders. These treatment modalities are moderately efficacious; however, they suffer from several severe drawbacks. Each of these traditional treatment modalities and their associated limitations are described below.

One common conventional technique for controlling neurological disease includes the use of dopaminergic agonists or anticholinergic agents. Medical management using these techniques requires considerable iteration in dosing adjustments before an "optimal" balance between efficacy and side effect minimization is achieved. Variation, including both circadian and postprandial variations, causes wide fluctuation in symptomatology. This commonly results in alternation between "on" and "off" periods during which the patient possesses and loses motor functionality, respectively.

Another traditional approach for controlling movement disorders is tissue ablation. Tissue ablation is most commonly accomplished through stereotactic neurosurgical procedures, including pallidotomy, thalamotomy, subthalamotomy, and other lesioning procedures. These procedures have been found to be moderately efficacious. However, in addition to posing risks that are inherent to neurosurgical operations, these procedures suffer from a number of fundamental limitations. One such limitation is that tissue removal or destruc-

tion is irreversible. As a result, excessive or inadvertent removal of tissue cannot be remedied.

Furthermore, undesirable side effects, including compromise of vision and motor or sensory functions, are likely to be permanent conditions. In particular, bilateral interventions place the patient at considerable risk for developing permanent neurologic side effects, including incontinence, aphasia, and grave psychic disorders. An additional drawback to this approach is that the "magnitude" of treatment is constant. That is, it is not possible to vary treatment intensity over time, as may be required to match circadian, postprandial, and other fluctuations in symptomatology and consequent therapeutic needs. Thus, decrease in treatment "magnitude" is not possible while an increase in treatment "magnitude" necessitates reoperation. Some adjustment is possible through augmentation with pharmacologic treatment; however, these additional treatments are subject to the above-noted limitations related to drug therapy.

Another traditional approach for controlling movement disorders and other neurological disease includes tissue transplantation, typically from animal or human mesencephalic cells. Although tissue transplantation in humans has been performed for many years, it remains experimental and is limited by ethical concerns when performed using a human source. Furthermore, graft survival, as well as subsequent functional connection with intracranial nuclei, are problematic. The yield, or percentage of surviving cells, is relatively small and is not always predictable, posing difficulties with respect to the control of treatment "magnitude."

Another traditional approach for controlling neurological disease is the continuous electrical stimulation of a predetermined neurological region. Chronic high frequency intracranial electrical stimulation is typically used to inhibit cellular activity in an attempt to functionally replicate the effect of tissue ablation, such as pallidotomy and thalamotomy. Acute electrical stimulation and electrical recording and impedance measuring of neural tissue have been used for several decades in the identification of brain structures for both research purposes as well as for target localization during neurosurgical operations for a variety of neurological diseases. During intraoperative electrical stimulation, reduction in tremor has been achieved using frequencies typically on the order of 75 to 330 Hz. Based on these findings, chronically implanted constant-amplitude electrical stimulators have been implanted in such sites as the thalamus, subthalamic nucleus and globus pallidus.

Chronic constant-amplitude stimulation has been shown to be moderately efficacious. However, it has also been found to be limited by the lack of responsiveness to change in patient system symptomatology and neuromotor function. Following implantation, a protracted phase of parameter adjustment, typically lasting several weeks to months, is endured by the patient while stimulation parameters are interactively adjusted during a series of patient appointments. Once determined, an "acceptable" treatment magnitude is maintained as a constant stimulation level. A drawback to this approach is that the system is not responsive to changes in patient need for treatment. Stimulation is typically augmented with pharmacological treatment to accommodate such changes, causing fluctuation of the net magnitude of treatment with the plasma levels of the pharmacologic agent.

As noted, while the above and other convention treatment modalities offer some benefit to patients with movement disorders, their efficacy is limited. For the above-noted reasons, with such treatment modalities it is difficult and often impossible to arrive at an optimal treatment "magnitude," that is, an optimal dose or intensity of treatment. Furthermore, patients

are subjected to periods of overtreatment and undertreatment due to variations in disease state. Such disease state variations include, for example, circadian fluctuations, postprandial (after meal) and nutrition variations, transients accompanying variations in plasma concentrations of pharmacological agents, chronic progression of disease, and others.

Moreover, a particularly significant drawback to the above and other traditional treatment modalities is that they suffer from inconsistencies in treatment magnitude. For example, with respect to drug therapy, a decrease in responsiveness to pharmacologic agents eventually progresses to eventually preclude effective pharmacologic treatment. With respect to tissue ablation, progression of disease often necessitates reoperation to extend pallidotomy and thalamotomy lesion dimensions. Regarding tissue transplantation, imbalances between cell transplant formation rates and cell death rates cause unanticipated fluctuations in treatment magnitude. For continuous electrical stimulation, changes in electrode position, electrode impedance, as well as patient responsiveness to stimulation and augmentative pharmacologic agents, cause a change in response to a constant magnitude of therapy.

Currently, magnets commonly serve as input devices used by patients with implantable stimulators, including deep brain stimulators, pacemakers, and spinal cord stimulators. Current systems require the patient to manually turn the system off at night time to conserve battery power and use such magnets to maintain system power. This presents considerable difficulty to many patients whose tremor significantly impairs arm function, as they are unable to hold a magnet in a stable manner over the implanted electronics module. Consequently, many patients are unable to turn their stimulators on in the morning without assistance.

What is needed, therefore, is an apparatus and method for treatment of patients with neurological disease in general and movement disorders in particular that is capable of determining and providing an optimal dose or intensity of treatment. Furthermore, the apparatus and method should be responsive to unpredictable changes in symptomatology and minimize alternations between states of overtreatment and undertreatment. The system should also be capable of anticipating future changes in symptomatology and neuromotor functionality, and being responsive to such changes when they occur.

SUMMARY OF THE INVENTION

One aspect of the invention is a method of monitoring and storing EEG signals from a patient. The method includes monitoring EEG signals from a patient with an implanted unit, wirelessly transmitting a substantially real-time monitored EEG signal from the implanted unit to a communication module that is external to the patient's body, and storing the substantially-real time EEG signal in a memory that is external to the patient's body.

In some embodiments the communication module is a portable patient interface module, and the memory is within the portable patient interface module. The communication module may also be a supervisory module that is used by a healthcare provider wherein the memory is within the supervisory module.

In some embodiments the method may further comprise establishing a wireless communication link between the implanted unit and the communication module, where the monitoring and transmitting the EEG signal is performed substantially continuously while the wireless communication link is established between the implanted unit and the communication module. Storing the transmitted EEG signal may create a time history record of the EEG signal for a time

period when the wireless communication link is established between the implanted unit and the communication module.

In some embodiments the method can further comprise wirelessly transmitting a stored time history of the EEG signals that was previously stored in a memory of the implanted unit.

In some embodiments wirelessly transmitting the monitored EEG signal facilitates a real-time monitoring of the EEG signal with the communication module.

In some embodiments the method further comprises analyzing the EEG signal to estimate the patient's condition. Analyzing the EEG signal from the patient to estimate the patient's condition may include predicting future symptomatology of the patient's condition. The condition may comprise epilepsy and the future symptomatology may comprise a seizure. The embodiment may further comprise initiating delivery of a therapy to the patient when the future symptomatology is predicted.

In some embodiments the method further comprises using the communication module to provide an output to the patient that is indicative of the patient's estimated condition.

The condition can also comprise Parkinson's disease and the future symptomatology can comprise a tremor. The condition can also comprise depression or bipolar disorder.

Another aspect of the invention is a method of monitoring a patient's neurological or psychiatric condition. The method includes transmitting substantially real-time EEG signals that are sampled with an implanted device to an external device, and analyzing the substantially real-time EEG signals to estimate the patient's condition and/or predict future symptomatology of the patient's condition.

In some embodiments the method includes facilitating delivery of a therapy to manage the patient's condition and/or predicted future symptomatology. The therapy can comprise at least one of electrical stimulation of a peripheral nerve, electrical stimulation of brain tissue, and drug delivery.

In some embodiments the method also includes using the external device to provide an output to the patient that is indicative of the patient's estimated condition.

In some embodiments transmitting is performed continuously during a time period in which there is a wireless communication link established between the implanted device and the external device.

In some embodiments the patient's condition is epilepsy and the future symptomatology comprises a seizure. The patient's condition can comprise Parkinson's disease and the future symptomatology can comprise a tremor. The patient's condition can also comprise depression or can comprise bipolar disorder.

Another aspect of the invention is a method of monitoring a patient's epilepsy condition. The method includes establishing a wireless communication link between an implanted device and an external device, sampling an EEG signal from a patient with the implanted device, continuously transmitting a substantially real-time EEG signal to the external device while the communication link is established, extracting parameters from the EEG signal to estimate the patient's epilepsy condition, wherein the parameters are indicative or predictive of a seizure, and providing a substantially real-time output to the patient with the external device that is indicative of the patient's epilepsy condition.

In some embodiments the method includes initiating delivery of a therapy to manage the patient's epilepsy condition. In some embodiments the therapy comprises at least one of electrical stimulation of a peripheral nerve, electrical stimulation of brain tissue, and drug delivery.

In some embodiments continuously transmitting is carried out using a wireless radiofrequency communication link.

In some embodiments the method also includes storing the sampled EEG signal in a memory of the external device.

Further features and advantages of the present invention, as well as the structure and operation of various embodiments of the present invention, are described in detail below with reference to the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

The present invention is described with reference to the accompanying drawings. In the drawings, like reference numerals indicate identical or functionally similar elements.

FIG. 1 is a schematic diagram of one embodiment of the present invention implanted bilaterally in a human patient.

FIG. 2 is an architectural block diagram of one embodiment of the neurological control system of the present invention.

FIG. 3 is a block diagram of one embodiment of an intracranial recording electrode (ICRE) signal processor and an intracranial stimulating electrode (ICSE) signal processor each of which are included within the signal processor illustrated in FIG. 2.

FIG. 4 is a schematic diagram of a globus pallidus implanted with stimulating and recording electrodes in accordance with one embodiment of the present invention.

FIG. 5 is a block diagram of one embodiment of an EMG signal processor that is included in one embodiment of the signal processor illustrated in FIG. 2.

FIG. 6 is a block diagram of one embodiment of an EEG signal processor module that is included in one embodiment of the signal processor illustrated in FIG. 2.

FIG. 7 is a block diagram of one embodiment of an accelerometer signal processor that is incorporated into certain embodiments of the signal processor illustrated in FIG. 2.

FIG. 8 is a block diagram of one embodiment of an acoustic signal processor that is included in certain embodiments of the signal processor illustrated in FIG. 2.

FIG. 9 is block diagram of one embodiment of a peripheral nerve electrode (PNE) signal processor 237 that is implemented in certain embodiments of signal processor 71. PNE signal

FIG. 10 is a schematic diagram of one embodiment of the signal processor illustrated in FIG. 2.

FIG. 11 is a schematic diagram of the patient-neural modulator system illustrated in FIG. 2 illustrated to show its controller and observer components.

FIG. 12 is a schematic diagram of one embodiment of the control circuit illustrated in FIG. 2.

FIG. 13 is a schematic diagram of electrical stimulation waveforms for neural modulation.

FIG. 14 is a schematic diagram of one example of the recorded waveforms.

FIG. 15 is a schematic block diagram of an analog switch used to connect one or an opposing polarity pair of Zener diodes across the noninverting and inverting inputs of an intracranial recording electrode amplifier.

FIG. 16 is a diagram of a two coil embodiment of the power delivery unit.

FIG. 17 is a magnification of the configurations of FIGS. 16 and 18 showing the magnetic flux penetrating the skin.

FIG. 18 is a diagram of a three coil embodiment of the power delivery unit.

FIG. 19 is a diagram of the power delivery unit with coil holder.

FIG. 20 is a diagram of the coil holder.

FIG. 21 is a diagram of the electromagnetic copies in proximity to the head of a patient.

FIG. 22 is a diagram of multiple coil embodiments.

FIG. 23 is a diagram of the paracranial design, with implanted components in close proximity to the patient's head.

FIG. 24 is a diagram of the power conversion unit that includes the electromagnetic coupling element.

FIG. 25 is a diagram of the power conversion circuit.

FIG. 26 is a diagram of the overall system.

FIG. 27 is a diagram of the stimulating and recording unit.

FIG. 28 is a diagram of the system enclosure secured to the calvarium.

FIG. 29 is a diagram of a lower profile design.

FIG. 30 is a diagram of a lower profile design with the system enclosure partially recessed into the calvarium.

FIG. 31 is a diagram of a lower profile design with the system enclosure fully recessed into the calvarium.

FIG. 32 is a diagram of a second lower profile design with the system enclosure fully recessed into the calvarium.

FIG. 33 is a diagram of a neurological control system.

FIG. 34 is a diagram of a second neurological control system.

FIG. 35 is a schematic diagram of one embodiment of the present invention implanted unilaterally in a human patient, with the system enclosure recessed in the calvarium, shown as a lateral view.

FIG. 36 is a schematic diagram of one embodiment of the present invention implanted unilaterally in a human patient, with the system enclosure recessed in the calvarium, shown as an anteroposterior view.

FIG. 37 is a schematic diagram of a cross section of calvarium with system enclosure shown implanted recessed within the calvarium.

FIG. 38 is a schematic diagram of a cross section of calvarium with drill bit shown in place after completion of process of drilling hole in calvarium.

FIG. 39 is a schematic diagram of a cross section of calvarium with drill bit, with a penetration detection release mechanism, shown in place after completion of process of drilling hole in calvarium.

FIG. 40 is a diagram depicting the path of the intracranial catheter and its connection to the electrical elements.

FIG. 41 depicts one design for the system enclosure for implantation in the calvarium.

FIGS. 42 and 43 depict a dual intracranial catheter design.

FIG. 44 is a schematic diagram of one embodiment of the present invention implanted bilaterally in a human patient.

FIG. 45 is a schematic diagram, lateral view, of one embodiment of the present invention implanted unilaterally in a human patient.

FIG. 46 is a schematic diagram, anteroposterior view, of one embodiment of the present invention implanted unilaterally in a human patient, with multiple catheters and neuromodulators.

FIG. 47 is a schematic diagram of one embodiment of the present invention, depicting a set of noninvasive and implanted sensors and neuromodulators in a human patient.

FIG. 48 is a schematic diagram of the dermatomal distributions recruited by neuromodulators.

FIG. 49 is a schematic diagram of a noninvasive version of neurological control system, with sensors and neuromodulators overlying dermatomal distributions recruited by neuromodulators.

FIG. 50 is a functional block diagram of neurological control system, with sensors and neuromodulators implanted in the temporal lobe.

FIG. 51 is a functional block diagram of neurological control system, with sensors and neuromodulators implanted a multiplicity of locations, including the temporal lobe and deep brain regions.

FIG. 52 is a functional block diagram of neurological control system, with sensors and neuromodulators implanted a multiplicity of locations, including the temporal lobe and deep brain regions, showing control input and control output waveforms versus time.

FIG. 53 is a functional block diagram of neurological control system, with sensors and neuromodulators implanted a multiplicity of locations, including the temporal lobe and deep brain regions, showing control input and control output waveforms versus time.

FIG. 54 is a timing diagram showing control input and control output waveforms versus time along with five representative elements of neural state vector X over time, during which time control input deviates outside target range.

FIG. 55 is a timing diagram showing control input and control output waveforms versus time along with five representative elements of neural state vector X over time, during which time control input remains within target range.

FIG. 56 is a timing diagram showing control input and control output waveforms versus time along with five representative elements of neural state vector X over time, during which time control input remains within target range, under conditions with and without the application of a perturbation.

FIG. 57 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with five representative elements of neural state vector X over time, during which time control input remains within target range, under conditions with and without the application of a perturbation.

FIG. 58 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input remains within normal range, under conditions without the application of a perturbation. Closed-loop neuromodulation control is turned on for a duration during which time control input is more tightly maintained in control range, a subset of normal range.

FIG. 59 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input deviates outside normal range into borderline range and into critical range, under conditions without the application of a perturbation. Closed-loop neuromodulation control remains off, and there are no EEG abnormalities nor neurological sings or symptoms.

FIG. 60 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input deviates outside normal range into borderline range and into critical range, under conditions without the application of a perturbation. Closed-loop neuromodulation control remains off, and EEG abnormalities develop and are followed by neurological sings or symptoms.

FIG. 61 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input remains within normal range, under conditions without the application of a perturbation. Closed-loop neuromodulation control remains on, and there are no EEG abnormalities nor neurological sings or symptoms.

FIG. 62 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input deviates outside normal range and closed-loop neuromodulation control is turned on, bringing control input back into control range, which is a subset of normal range. There are no EEG abnormalities nor neurological sings or symptoms during this time.

FIG. 63 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time control input deviates outside normal range and closed-loop neuromodulation control is turned on, bringing control input back into control range, which is a subset of normal range, following which time closed-loop neuromodulation control is turned back off. There are no EEG abnormalities nor neurological sings or symptoms during this time.

FIG. 64 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time a perturbation is applied and control input decreases but remains inside normal range. Closed-loop neuromodulation control remains off, and there are no EEG abnormalities nor neurological sings or symptoms during this time.

FIG. 65 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time a perturbation is applied and control input deviates outside normal range into borderline range and critical range and then returns spontaneously to normal range. Closed-loop neuromodulation control remains off, and there are no EEG abnormalities nor neurological sings or symptoms during this time.

FIG. 66 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time a perturbation is applied and control input deviates outside normal range into borderline range and critical range, then back into borderline range and normal range and again into borderline range and critical range, following which EEG abnormalities develop and which are followed by neurological sings or symptoms. Closed-loop neuromodulation control remains off during this time.

FIG. 67 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time a perturbation is applied and control input decreases but remains inside control range, which is a subset of normal range. Closed-loop neuromodulation control remains on, and there are no EEG abnormalities nor neurological sings or symptoms during this time.

FIG. 68 is a timing diagram showing EEG waveforms, control input and control output waveforms versus time along with one representative element of neural state vector X over time, during which time a perturbation is applied and control input deviates outside control range yet remains within normal range. Closed-loop neuromodulation control is turned on, and control input is brought back within control range. Control input remains within normal range and there are no EEG abnormalities nor neurological signs or symptoms during this time.

DETAILED DESCRIPTION

FIG. 1 is a schematic diagram of one embodiment of the intracranial stimulator of the present invention implanted

bilaterally in a human patient. In the embodiment illustrated in FIG. 1, two neurological control systems 999 are shown implanted bilaterally. Each system 999 includes a stimulating and recording unit 26 and one or more intracranial components described below. As described in this illustrative embodiment, the intracranial components preferably include a stimulating electrode array 37. However, it should become apparent to those of ordinary skill in the relevant art after reading the present disclosure that the stimulating electrodes may also be extracranial; that is, attached to a peripheral nerve in addition to or in place of being located within the cranium. As shown in FIG. 1, stimulating and recording unit 26 of each neurological control system 999 is preferably implanted contralateral to the intracranial components of the device.

As one skilled in the relevant art would find apparent from the following description, the configuration illustrated in FIG. 1 is just one example of the present invention. Many other configurations are contemplated. For example, in alternative embodiments of the present invention, the stimulating and recording unit 26 is implanted ipsilateral or bilateral to the intracranial components. It should also be understood that the stimulating and recording unit 26 can receive ipsilateral, contralateral or bilateral inputs from sensors and deliver ipsilateral, contralateral, or bilateral outputs to a single or a plurality of intracranial stimulating electrode arrays 37. Preferably, these inputs are direct or preamplified signals from at least one of EMG electrode array 50, EEG electrode array 51, Accelerometer Array 52, Acoustic Transducer Array 53, Peripheral Nerve Electrode Array 54, and Intracranial Recording Electrode Array 38. The signals input from these sensors will be referred to herein as "sensory input modalities" 247. The outputs include but are not limited to one or more stimulating current signals or stimulating voltage signals to Intracranial Stimulating Electrode Array 37.

In the embodiment illustrated in FIG. 1, the two unilateral systems 26 are shown to receive sensory inputs from the side contralateral as well as the intracranial stimulating electrode arrays 37. In the illustrative embodiment, systems 26 also receive sensory inputs from intracranial recording electrode arrays 38. As will become apparent from the following description, intracranial recording electrode arrays 38 may provide valuable feedback information.

It should be understood that this depiction is for simplicity only, and that any combination of ipsilateral, contralateral or bilateral combination of each of the multiple sensory input modalities and multiple stimulation output channels may be employed. In addition, stimulating and recording units 26 may be a single device, two communicating devices, or two independent devices. Accordingly, these and other configurations are considered to be within the scope of the present invention. It is anticipated that stimulating and recording units 26, if implemented as distinct units, would likely be implanted in separate procedures (soon after clinical introduction) to minimize the likelihood of drastic neurological complications.

In the exemplary embodiment illustrated in FIG. 1, the intracranial stimulating electrode array 37 includes a plurality of intracranial stimulating electrodes 1, 2, 3 and 4. Array 37 may, of course, have more or fewer electrodes than that depicted in FIG. 1. These intracranial stimulating electrodes 1-4 may be used to provide stimulation to a predetermined nervous system component. The electrical stimulation provided by the intracranial stimulating electrodes 1-4 may be excitatory or inhibitory, and this may vary in a manner which is preprogrammed, varied in real-time, computed in advance using a predictive algorithm, or determined using another technique now or latter developed.

The intracranial recording electrode arrays 38 includes intracranial recording electrodes 5 and 6. In accordance with one embodiment of the present invention, the intracranial recording electrodes 5, 6 are used to record cortical activity as a measure of response to treatment and as a predictor of impending treatment magnitude requirements. In the illustrative embodiment, intracranial recording electrodes 5 and 6 are depicted in a location superficial to the intracranial stimulating electrodes 1-4. However, this positioning may be reversed or the intracranial stimulating electrodes 1-4 and intracranial recording electrodes 5 and 6 may be interspersed in alternative embodiments. For example, these electrodes may be placed in at least one of motor cortex, premotor cortex, supplementary motor cortex, other motor cortical areas, somatosensory cortex, other sensory cortical areas, Wernicke's area, Broca's area, other cortical region, other intracranial region, and other extracranial region.

In the illustrative embodiment, an intracranial catheter 7 is provided to mechanically support and facilitate electrical connection between intracranial and extracranial structures. In this embodiment, intracranial catheter 7 contains one or more wires connecting extracranial stimulating and recording circuit 26 to the intracranial electrodes, including but not limited to, intracranial stimulating electrodes 1-4 and intracranial recording electrodes 5, 6. The wires contained within intracranial catheter 7 transmit stimulating electrode output signal (SEOS) to intracranial stimulating electrode array 37. Such wires additionally transmit stimulating electrode input signal (SEIS) and recording electrode input signal (REIS), from intracranial stimulating electrode array 37 and intracranial recording electrode array 38 respectively, to stimulating and recording circuit 26.

Stimulating and recording circuit 26 is protected within a circuit enclosure 44. Circuit enclosure 44 and contained components, including stimulating and recording circuit 26 comprise stimulating and recording unit 43. It should be understood that more or fewer of either type of electrode as well as additional electrode types and locations may be incorporated or substituted without departing from the spirit of the present invention. Furthermore, stimulating and recording circuit 26 can be placed extra cranially in a subdavian pocket as shown in FIG. 1, or it may be placed in other extracranial or intracranial locations.

Connecting cable 8 generally provides electrical connection between intracranial or intracranial locations. A set of electrical wires provides the means for communication between the intracranial and extracranial components; however, it should be understood that alternate systems and techniques such as radiofrequency links, optical (including infrared) links with transcranial optical windows, magnetic links, and electrical links using the body components as conductors, may be used without departing from the present invention. Specifically, in the illustrative embodiment, connecting cable 8 provides electrical connection between intracranial components 246 and stimulating and recording circuit 26. In embodiments wherein stimulating and recording circuit 26 has an intracranial location, connecting cable 8 would likely be entirely intracranial. Alternatively, connecting in embodiments wherein stimulating and recording circuit 26 is implanted under scalp 10 or within or attached to calvarium 9, connecting cable 8 may be confined entirely to subcutaneous region under the scalp 10.

A catheter anchor 29 provides mechanical connection between intracranial catheter 7 and calvarium 9. Catheter anchor 29 is preferably deep to the overlying scalp 10. Such a subcutaneous connecting cable 8 provides electrical connection between intracranial electrodes 246 and stimulating and

recording circuit 26. Cable 8 may also connect any other sensors, including but not limited to any of sensory input modalities 247, or other stimulating electrodes, medication dispensers, or actuators with stimulating and recording circuit 26.

Sensory feedback is provided to recording and stimulating unit 26 from a multiplicity of sensors, collectively referred to as sensory input modalities 247. Intracranial recording electrode array 38, previously described, is intracranial in location. Additional sensors, most of which are located extracranially in the preferred embodiment, comprise the remainder of sensory input modalities 247. Sensory input modalities 247 provide information to stimulating and recording unit 26. As will be described in greater detail below, such information is processed by stimulating and recording unit 26 to deduce the disease state and progression and its response to therapy.

In one embodiment of the invention, a head-mounted acoustic sensor 11 is used to monitor any number of vibratory characteristics such as high frequency head vibration, muscle vibration, and/or speech production. Head-mounted acoustic sensor 11 is connected to stimulating and recording circuit 26 with an acoustic sensor connecting cable 30.

A head-mounted accelerometer 12 is implemented in certain embodiments of the present invention to monitor head movement and position with respect to gravity. Head-mounted accelerometer 12 may be mounted to any structure or structures that enables it to accurately sense a desired movement. Such structures include, for example, the skull base, calvarium, clavicle, mandible, extraocular structures, soft tissues and vertebrae. Head-mounted accelerometer 12 is connected to stimulating and recording circuit 26 with an accelerometer connecting cable 31.

A proximal electromyography (EMG) electrode array 45 is also included in certain preferred embodiments of the invention. Proximal EMG electrode array 45 includes a positive proximal EMG electrode 13, a reference proximal EMG electrode 14, and a negative proximal EMG electrode 15. As one skilled in the relevant art would find apparent, proximal EMG electrode array 45 may include any number of type of electrodes. Proximal EMG electrode array 45 is implanted in or adjacent to muscle tissue. In the embodiment illustrated in FIG. 1, proximal EMG electrode array 45 is shown implanted within the neck of the human patient. However, it should be understood that this location is illustrative only and that proximal EMG electrode array 45 may be implanted in or adjacent to any muscle without departing from the spirit of the present invention.

A proximal acoustic sensor 27 may also be implemented in the present invention. Proximal acoustic sensor 27 senses muscle vibration and may be used to augment, supplement or replace EMG recording. Also, a proximal accelerometer 28 may be used to sense movement, including tremor and voluntary activity, and orientation with respect to gravity. Proximal connecting cable 16 provides electrical connection from the proximal EMG electrodes 14 and 15, proximal acoustic sensor 27, and proximal accelerometer 28 to stimulating and recording circuit 26. In the illustrative embodiment, these sensors are shown connected to a common proximal connecting cable 16. However, in alternative embodiments, this configuration may include the use of multiple connecting cables or implement other types of communication media without departing from the present invention. It should also be understood from the preceding description that the number of each type of sensor may also be increased or decreased, some sensor types may be eliminated, and other sensor types may be included without departing from the spirit of the present invention.

A distal EMG electrode array 47 may also be included in certain embodiments of the present invention. In such embodiments, distal EMG electrode array 47 typically includes a positive distal EMG electrode 17, a reference distal EMG electrode 42, and a negative distal EMG electrode 18. Positive distal EMG electrode 17 is connected to stimulating and recording circuit 26 by positive distal EMG connecting cable 20. Negative distal EMG electrode 18 is connected to stimulating and recording circuit 26 by negative distal EMG connecting cable 21. Reference distal EMG electrode 42 is connected to stimulating and recording circuit 26 by reference distal EMG connecting cable 48.

In other embodiments, a distal acoustic sensor 19 is connected to stimulating and recording circuit 26 by distal acoustic connecting cable 22. Distal accelerometer 33 is connected to stimulating and recording circuit 26 by distal accelerometer connecting cable 34. Distal accelerometer 33 is connected to stimulating and recording circuit 26 by distal accelerometer connecting cable 34.

In the embodiment illustrated in FIG. 1, distal EMG electrode array 47, distal acoustic sensor 19, and distal accelerometer 33 are shown located in the shoulder region. However, the distal EMG electrode array 47 may be located in other locations, including, for example, the masseter, temporalis, sternocleidomastoid, other portion of the head and neck, pectoralis, torso, abdomen, upper extremities, lower extremities, and other locations. The number of each type of sensor may be increased or decreased, some sensor types may be eliminated, and other sensor types may be included without departing from the spirit of the present invention.

An enclosure-mounted EMG electrode array 46 is illustrated in FIG. 1. Enclosure-mounted EMG electrode array 46 includes enclosure-mounted positive EMG electrode 23, enclosure-mounted negative EMG electrode 24 and enclosure-mounted reference EMG electrode 25, all of which are attached to the circuit enclosure 44 that encloses stimulating and recording unit 26. The circuit enclosure 44 is preferably included to provide robustness against potential lead entanglement and fracture. In one particular embodiment, circuit enclosure 44 is constructed of titanium and epoxy, or other single or combination of bio-compatible materials. Enclosure-mounted acoustic sensor 35 and enclosure-mounted accelerometer 36 are mounted to stimulating and recording unit 43. The number of each type of sensor may be increased or decreased, their locations changed, some sensor types eliminated, and other sensor types included without departing from the spirit of the present invention.

In the embodiment illustrated in FIG. 1, EEG electrodes 39, 40, 41 are provided. The EEG electrodes may be mounted directly to connecting cable 8 or may be connected via intermediate cables. Any one of the numerous standard and new electrode configurations, or montages, may be employed in EEG electrodes 39-41 without departing from the present invention.

In one embodiment, a proximal peripheral nerve electrode array 98 is connected to stimulating and recording circuit 26 by proximal peripheral nerve electrode array connecting cable 100. Proximal peripheral nerve electrode array 98 is shown located in the neck region. In this location proximal peripheral nerve electrode array 98 can interface with the vagus nerve, spinal accessory nerve, or nerve arising from cervical roots.

A distal peripheral nerve electrode array 99 is connected to stimulating and recording circuit 26 by distal peripheral nerve electrode array connecting cable 32. Distal peripheral nerve electrode array 99 is shown located by the proximal arm, in position to interface with the brachial plexus or proximal arm

nerve. One or more of these peripheral nerve electrode arrays may be implanted in these or other locations, including but not limited to the head, cranial nerves, neck, torso, abdomen, upper extremities, and lower extremities, without departing from the present invention.

In one preferred embodiment, the peripheral nerve electrode arrays are each comprised of three epineural platinum-iridium ring electrodes, each in with an internal diameter approximately 30% larger than that of the epineurium, longitudinally spaced along the nerve. Electrodes of differing dimensions and geometries and constructed from different materials may alternatively be used without departing from the present invention. Alternative electrode configurations include but are not limited to epineural, intrafascicular, or other intraneural electrodes; and materials include but are not limited to platinum, gold, stainless steel, carbon, and other element or alloy.

FIG. 2 is an architectural block diagram of one embodiment of the neurological control system 999 of the present invention for modulating the activity of at least one nervous system component in a patient. As used herein, a nervous system component includes any component or structure comprising an entirety or portion of the nervous system, or any structure interfaced thereto. In one preferred embodiment, the nervous system component that is controlled by the present invention includes the globus pallidus internus. In another preferred embodiment, the controlled nervous system component is the subthalamic nucleus.

The neurological control system 999 includes one or more implantable components 249 including a plurality of sensors each configured to sense a particular characteristic indicative of a neurological or psychiatric condition. One or more intracranial (IC) stimulating electrodes in an IC stimulating electrode array 37 delivers a neural modulation signal to the same or other nervous system component as that being monitored by the system 26. One or more sensors 38, 51, 52, 53, and 54 sense the occurrence of neural responses to the neural modulation signals. Stimulating and recording unit 26 generates the neural modulation signal based on the neural response sensed by the sensors.

The neurological control system 999 preferably also includes a patient interface module 55 and a supervisory module 56. A control circuit 72 (described below) is communicably coupled to the patient interface module 55 and receives signal inputs from and provides signal outputs to patient interface module 55 and supervisory module 56. In one preferred embodiment, patient interface module 55 and supervisory module 56 remain external to the body of the patient. However either of these devices may be connected via percutaneous leads or be partially or totally implanted without departing from the present invention.

Patient interface module 55 and supervisory module 56 facilitate adjustment of control parameters, monitoring of disease state, monitoring of response to therapy, monitoring of stimulating and recording circuit 26, monitoring of impedance and other characteristics of intracranial stimulating electrode array 37, monitoring of physiologic parameters, monitoring of vital signs, monitoring of any other characteristic or function of components of the present invention, including but not limited to the stimulating and recording circuit 26, stimulating and recording unit 43, circuit enclosure 44, EMG electrode array 50, EEG electrode array 51, accelerometer array 52, acoustic transducer array 53, peripheral nerve electrode array 54, and intracranial recording electrode array 38. Such monitoring and adjustment is accomplished through the use of any well known bi-directional communication between control circuit 72 and supervisory module 56. In one pre-

ferred embodiment, a radiofrequency link is employed. In alternative embodiments, other communication technologies, including but not limited to optical, percutaneous, or electromagnetic, may be used.

In one preferred embodiment, patient interface module 55 and supervisory module 56 are placed adjacent to the patients garments overlying the implanted stimulating and recording unit 43. When neurological control system 999 is turned on in this position, a communications handshaking protocol is executed. Communication handshaking routines are known to those of ordinary skill in the art, and they enable establishment of a communication rate and protocol and facilitate mutual identification of devices. Patient interface module 55 automatically downloads parameters from stimulating and recording circuit 26 and stores values of such parameters in a memory. When the transfer of these parameter values is complete, patient interface module 55 emits an audible signal such as a series of beeps, and the patient turns off patient interface module 55 and removes it from its position overlying the implanted stimulating and recording unit 43. Parameter values may then be retrieved by the patient by a routine including but not limited to a menu driven interface, and the values may be transmitted via telephone conversation or other communication method to a health care professional. Supervisory module 56 operates in the same manner with one addition; a step is provided during which the health care professional may upload parameters to stimulating and recording circuit 26 to alter its function including by means of changing parameters including but not limited to control laws gains and thresholds, filter parameters, signal processing parameters, stimulation waveform modes (including at least one of current regulated, voltage regulated, frequency regulated, or pulse width regulated), and stimulation waveform parameters.

Control laws, well known to those of ordinary skill in the field of control theory, are defined by a set of parameters specific to the particular control law. Common parameters include preset gains, threshold levels, saturation amplitudes, sampling rates, and others. Adaptive controllers change in response to the behavior of the system being controlled; as such, in addition to preset parameters, adaptive controllers possess a set of varying parameters. These varying parameters contain information indicative of the behavior of the system being controlled; downloading of these parameters provides one set of measures of the disease state and its response to therapy.

Such monitoring includes observation of time history of disease state, stimulation parameters, response to therapy, and control law parameters, including time-varying adaptive controller parameters. Such adjustments includes modification of actual stimulation parameters and allowable ranges thereof, including but not limited to pulse width, pulse amplitude, interpulse interval, pulse frequency, number of pulses per burst frequency. Adjustments can further include modification of actual control law parameters and allowable ranges thereof, including but not limited to gains, thresholds and sampling rates of said stimulation waveforms. Signal processor 71 contains signal processor modules for each of the sensory input modalities 247. Signal processing algorithms for each of the said sensory input modalities 247 may be independent. Additionally, signal processing algorithms the said sensory input modalities 247 may be coupled, such that the processing of one of the sensory input modalities 247 is dependent on another of the sensory input modalities 247. Adjustments may additionally include modification of actual signal processor parameters and allowable ranges thereof, including but not limited to gains, filter cutoff frequencies,

filter time constants, thresholds, and sampling rates. In a preferred embodiment, the stimulation and control law parameters are stored in at least one of random access memory and central processing unit registers (not shown).

It is anticipated that patient interface module 55 is to be used by the patient, a family member or associate, or home health care personnel to monitor the functions and performance of neurological control system 999. In such an embodiment, the use of the patient interface module 55 is restricted to monitoring operations; adjustment of stimulation and control parameters is not enabled. However, adjustment of all or a subset of stimulation and control parameters (described below) may be facilitated by patient interface module 55 without departing from the present invention. Supervisory module 56, on the other hand, is used by a physician or other health care personnel to monitor function and performance of neurological control system 999 and to adjust stimulation and control parameters. Control parameters controlled by patient interface module 55 and supervisory module 56 include allowable stimulation magnitude range, such as maximum combination of stimulation voltage, current, pulse width, pulse frequency, train frequency, pulse train count, pulse train duration. Control parameters may also include variables and constants used to define control laws implemented in control circuit 72. Such control parameters include, but are not limited to, control law gains 197-203, and other parameters for control laws, including but not limited to proportional controller 230, differential controller 204, integral controller 205, nonlinear controller 206, adaptive controller 207, sliding controller 208, model reference controller 209, and other controllers. In addition, amplitudes for other controller parameters, including but not limited to amplitudes for controller weights 210-216 may be set by supervisory module 56. Additionally, the parameters specifying the maximum amplitudes, or saturation values, may be set by supervisory module 56. Control circuit 72 (FIG. 12) will be described in detail below.

The majority of the computation accomplished by stimulating and recording circuit 26 is performed in signal conditioning circuit 76, signal processor 71, and control circuit 72; the algorithms and behavior of which are determined by corresponding sets of control parameters, of which some may be set by the supervisory module 56 and a typically more restricted set by patient interface module 55. In one embodiment, control parameters further includes signal conditioning parameters. Signal conditioning parameters may include, for example, amplifier gains, filter gains and bandwidths, threshold values, and other parameters. In certain embodiments, control parameters additionally include signal processing parameters, including envelope determinator gains and time constants, filter passbands, filter gains, threshold values, integrator gains, analyzer parameters, disease state estimator parameters, artifact rejecter thresholds, envelope determinator time constants, rectifier parameters, spectral analyzer parameters and timer parameters.

In the illustrative embodiment described herein, control parameters further include spike detector 188 (FIG. 9) parameters, spike characterizer 189 (FIG. 9) parameters, spike analyzer 190 (FIG. 9) parameters, spectral energy characterizer 192 (FIG. 9) parameters, spectral energy analyzer 193 (FIG. 9) parameters, aggregate disease state estimator 195 (FIG. 10) parameters.

In accordance with the present invention, tremor are quantified and monitored by any sensors over time as indicators of disease state. Such sensors include but are not limited to EMG electrode array 50, EEG electrode array 51, accelerometer array 52, acoustic transducer array 53, peripheral nerve elec-

trode array 54, intracranial recording electrode array 38, and intracranial stimulating electrode array 37. In one particular embodiment, the sensed tremor characteristics include, but are not limited to, magnitude, frequency, duration and frequency of occurrence of tremors. Changes in these and other parameters are compared to current levels of, and changes in, treatment parameters. These changes are then used by aggregate disease state estimator 195 to estimate the response to therapy as functions of various electrical stimulation treatment parameters. Electrical stimulation treatment parameters are adjusted by control circuit 72 in real-time to provide optimal control of disease state.

Modulation parameters are optimized to achieve at least one of minimization of disease state, minimization of symptoms of disease, minimization of stimulation magnitude, minimization of side effects, and any constant or time-varying weighted combination of these. Patient interface module 55 and supervisory module 56 also preferably monitor the function and operation of other components of neurological control system 999, including stimulating and recording unit 26 and implanted components 249.

Stimulating and recording unit 26 receives and processes signals generated by implanted components 249 to provide conditioned signals 78-84 to a signal processor 71. For each type of implanted components 249 coupled to stimulating and recording unit 26, signal conditioning circuit 76 preferably includes an associated amplifier and filter. Each amplifier and associated filter is configured to receive and process the signal generated by the associated one of the set of sensors 38, 51, 52, 53, and 54.

In the illustrative embodiment, implanted components 249 include an electromyography (EMG) electrode array 50 which generate EMG signals. Preferably, EMC electrode array 50 comprises of all EMG electrodes implemented in the particular embodiment of the present invention. These include, in the exemplary embodiment illustrated in FIG. 1, proximal EMG electrode array 45, enclosure-mounted EMG electrode array 46 and distal EMG electrode array 47. Array 50 may also include, for example, EMG electrodes implanted in the head or other location, and surface EMG electrodes.

Implanted components 249 also include an electroencephalography (EEG) electrode array 51 which generate EEG signals and accelerometer array 52 which generates acceleration signals. EEG electrodes 39, 40, 41 illustrated in FIG. 1 are representative of EEG electrode array 51. EEG electrodes 39-41 may be mounted directly to connecting cable 8 or connected via intermediate cables. EEG electrode array 51 may include more or fewer elements than EEG electrodes 39-41 depicted; and any of numerous standard and new electrode configurations, or montages, may be employed without departing from the present invention.

Accelerometer array 52, which produces well-known acceleration signals, preferably includes all accelerometers implemented in the patient associated with the present invention. For example, in the embodiment illustrated in FIG. 1, accelerometer array 52 includes head-mounted accelerometer 12, proximal accelerometer 28, enclosure-mounted accelerometer 36 and distal accelerometer 33. Accelerometer array 52 may include more or fewer accelerometers than these accelerometers, and accelerometers of any types and locations may be employed without departing from the present invention.

Acoustic transducer array 53 includes all acoustic sensors utilized by the present invention. In the exemplary embodiment illustrated in FIG. 1, acoustic transducer array 53, includes head-mounted acoustic sensor 11, proximal acoustic sensor 27, enclosure-mounted acoustic sensor 35 and distal

acoustic sensor 19. It should be understood that acoustic transducer array 53 may include more or fewer elements than said acoustic sensors listed above; and any of numerous acoustic sensor types and locations may be employed without departing from the present invention.

Peripheral nerve electrode array 54 generates peripheral neural signals, including but not limited to efferent and afferent axonal signals. Preferably, peripheral nerve electrode array 54 includes all peripheral nerve electrodes implemented in present invention. For example, in the illustrative embodiment illustrated in FIG. 1, peripheral nerve electrode array 54 includes proximal peripheral nerve electrode array 98 and distal peripheral nerve electrode array 99. The single or plurality of individual peripheral nerve electrode arrays which comprise peripheral nerve electrode array 54 may be implanted in the illustrated or other locations, as noted above.

Intracranial (IC) recording electrode array 38 generates central neural signals, including but not limited to cortical, white matter, and deep brain nuclear signals. Neural activity to be sensed includes but is not limited to that found in the primary motor cortex, premotor cortex, supplementary motor cortex, somatosensory cortex, white matter tracts associated with these cortical areas, the globus pallidus internal segment, the globus pallidus external segment, the caudate, the putamen, and other cortical and subcortical areas. As one of ordinary skill in the relevant art will find apparent, the present invention may include additional or different types of sensors that sense neural responses for the type and particular patient. Such sensors generate sensed signals that may be conditioned to generate conditioned signals, as described below. One example of the placement of these electrodes is described above with reference to the embodiment illustrated in FIG. 1. Many others are contemplated by the present invention.

As noted, for each of the different types of sensors included in implanted components 249, signal conditioning circuit 76 includes an associated amplifier and filter in the illustrative embodiment. Accordingly, signal conditioning circuit 76 includes an EMG amplifier 59 and filter 66, each constructed and arranged to amplify and filter, respectively, the EMG signals received from EMG electrode array 50. Similarly, signal conditioning circuit 76 also includes an EEG amplifier 60 and filter 67, accelerometer (ACC) amplifier 61 and filter 68, acoustic (ACO) amplifier 62 and filter 69, peripheral nerve electrode (PNE) amplifier 63 and filter 70 and intracranial (IC) recording electrode (ICRE) amplifier 58 and filter 65.

Simplifiers 57-63 may be single or multi-channel amplifiers depending upon the number of electrodes with which it interfaces. In one preferred embodiment, amplifiers 57-63 are physically located in the same enclosure as filters 64-70; that is, in a single signal conditioning circuit 76. Preferably, signal conditioning circuit 76 is physically contained within stimulating and recording unit 102. However, amplifiers 57-63 may be located separately from stimulating recording unit 102. For example, amplifiers 57-63 may be affixed to or situated proximate to their associated electrode arrays 38, 50-54. This arrangement facilitates the preamplification of the associated signals generated by the associated electrode arrays 38, 50-54, increasing the signal-to-noise ratio of the signals. Amplifiers 57-63 may be any known voltage amplifier now or later developed suitable for amplifying the particular signals generated by their associated electrodes.

As noted, the amplified signals are passed to their associated filters 64-70 as shown in FIG. 2. As with amplifiers 57-59, filters 64-70 may be physically separate from or incorporated into signal conditioning circuit 76 and stimulating and recording unit 26. In one preferred embodiment, filters

64-70 are low pass filters having a cut-off frequency of, for example, 3,000 Hz. In alternative embodiments, filters 64-70 may include a notch filter to remove, for example, 60 Hz noise, or other types of filters appropriate for the type of signals generated by the associated sensors 38, 51, 52, 53, and 54. Selection of the appropriate frequencies for the cut-off and notch filter frequencies is considered to be well known in the relevant art and within the scope of the present invention. Filters 66-70, 65 and 64 generate conditioned sensed signals 84, 83 and 78-82, respectively.

Signal processor 71 processes the conditioned sensed neural response signals 78-84 generated by signal conditioning circuit 76 in accordance with the present invention to determine neural system states. Signal processor 71 generally performs well known filtering operations in the time and frequency domains. In one preferred embodiment, the neural system states include one or more physiologic or disease states. Signal processor 71, which can be implemented in a fast microprocessor, a DSP (digital signal processor) chip, or as analog circuitry, for example, is described in detail below.

Control circuit 72, responsive to the signal processor 71, patient interface module 55 and supervisory module 56, adjusts the magnitude of a neural modulation signal in response to the sensed neural response. Signal processor 71 extracts relevant information from the sensed condition signals, and control circuit 72 uses this extracted information in the calculation of an output neuromodulation signal (NMS) 998. Neuromodulation signal 998 subsequently travels along stimulator output path 111 to IC stimulating electrode array 37. In one embodiment, control circuit 72 is a state machine, utilizing current and past system behavior in the calculation of a control signal. In an alternative embodiment, control circuit 72 includes an embedded microprocessor to process nonlinear control laws. Alternative embodiments of the control circuit 72 appropriate for the particular application may be also be used.

Control circuit 72 receives control law selection information, control law parameter information, stimulation waveform parameter range information, stimulation modulation mode, output stage regulation mode, and medication dose and timing information from patient interface module 55 and supervisory module 56. The waveform parameter or parameters which are modulated by control law output signal U 997 are determined by the stimulation modulation mode; these parameters include but are not limited to pulse amplitude, pulse width, pulse frequency, pulses per burst, and burst frequency. Selection between regulation of pulse voltage or pulse current as the regulated pulse amplitude is determined by the output stage regulation mode.

Control circuit 72 provides stimulation waveform parameter history information, disease state history information, control law state variable history information, control law error history information, control law input variable history information, control law output variable history information, stimulating electrode impedance history information, sensory input history information, battery voltage history information, and power consumption history information to patient interface module 55 and supervisory module 56.

Provision of stimulating electrode impedance history information allows monitoring of stimulating electrode performance and functionality. If an electrode is determined to be fractured, shorted, or encapsulated by fibrotic tissue, any of various control law parameters, output stage parameters, and waveform range parameters may be adjusted to allow compensation for these changes. Additionally, the Neuromodulation Signal (NMS) 998 may be delivered to different sets of electrodes to insure that it reaches neural tissue 250.

Sensory input history information allows evaluation of validity of any given sensory input. This is useful in determining the functionality of a given sensor and serves as an indicator for sensor replacement or adjustment of the signal processing parameters or algorithm or the control law parameters or algorithm to continue to generate reliable disease state estimate signals X and control law outputs U despite the loss of any particular individual or set of sensory signals.

Signal processor 71 receives amplifier gain setting information, filter parameter information, weighting information, and disease state estimator parameter and algorithm information from patient interface module 55 and supervisory module 56. The function and operation of patient interface module 55 and supervisory module 56 are described above. As noted, patient interface module 55 may be used by the patient or home health care personnel to monitor disease state, stimulation parameters, and response to therapy. Limited adjustment of stimulation parameters and ranges is facilitated. Patient interface module 55 may be used by the patient or home health care personnel to provide information to the physician, avoiding the need for an office visit for the obtaining of said information.

Patient information module 55 queries signal processor 71 for present and time histories of monitored values. Time histories of selected variables in signal processor 71 and control circuit 72 are stored in memory module 240 for subsequent retrieval by patient interface module 55 and supervisory module 56. Selected variables include but are not limited to disease state, tremor frequency, tremor magnitude, EMG magnitude, EMG frequency spectra (EMG magnitude within frequency ranges), and acceleration of limb, head, mandible, or torso. Selected variables may also include disease state, frequency spectra of limb, torso, and head movements, as determined by EMG and accelerometer signals.

Stimulating and recording unit 26 also includes an output stage circuit 77. Output stage circuit 77 takes for an input the control law output signal U, which may be comprised of a single or multiplicity of channels or signals, from control circuit 72. This control law output signal U 997 modulates the magnitude of the sequence of waveforms comprising the desired output neuromodulation signal (NMS.sub.D) which is produced by output stage circuit 77 and delivered via intracranial stimulating electrode array 37 to neural tissue 250.

Output stage circuit 77 generates a neuromodulation signal (NMS.sub.D) 998 with a magnitude specified by control law output signal U 997 received from control circuit 72. In one preferred embodiment, the waveform parameter of the desired output neuromodulation signal (NMS.sub.D) which is modulated by control law output signal U is the stimulation current magnitude. The capability to specifically modulate the stimulation current confers efficacy resistance to perturbations or changes in electrode impedance. Presently implanted systems suffer from a decline in efficacy which results from an increase in electrode impedance which accompanies the normal tissue response to a foreign body, that is fibrotic encapsulation of the electrode. In this design taught in the present invention, a the magnitude of the current delivered to the neural tissue 250 will not vary as the electrode becomes encapsulated with fibrotic tissue or its impedance otherwise changes over time. A further advantage conferred by current modulation is the ability to monitor electrode impedance. If a current-modulated waveform, preferably a sinusoid, is delivered to the electrodes, and the resultant voltage potential waveform is concurrently monitored, the relative magnitudes and phase shifts of these waveforms may be computed. From these magnitudes and phases, the complex

impedance and hence the resistive and capacitive components of the electrode impedance may be calculated.

In an alternative embodiment, the waveform parameter of the desired output neuromodulation signal (NMS.sub.D) which is modulated by control law output signal U 997 is the stimulation voltage magnitude. This design would not enjoy the independence of the stimulation current and efficacy from impedance variation enjoyed by the embodiment described above. If fibrosis was uneven around the surface of the electrode, this embodiment would avoid potentially undesirably large current densities along narrow tracts of remaining low resistance unfibrosed regions of neural tissue 250.

Alternatively, regulation of stimulus pulse width may be desired. In certain circuit implementations, the available resolution or bits for specifying the magnitude of pulse width may be greater than that for specifying the pulse voltage or current. In such a case, if finer control of the magnitude of Neuro-modulation signal (NMS) 998 is desired than is provided by the control of pulse current or pulse voltage, then it may be desirable to modulate the pulse width. Furthermore, the spatial neuron recruitment characteristics of a pulse width modulated neuromodulation signal (NMS) 998 may provide a more linear, predictable, or controllable response than that obtained with current or voltage modulation. Selection between regulation of pulse voltage, pulse current, or pulse width as the regulated pulse amplitude parameter is determined by the output stage regulation mode, which may be set using supervisory module 56. In alternative embodiments, the modulation of pulse frequency and the modulation of the number of pulses per burst are regulated. As one of ordinary skill in the relevant art would find apparent. Other such characteristics may be regulated in addition to or instead of the ones noted above.

Output stage circuit 77 includes a pulse generator 73, an output amplifier 74 and a multiplexor 75. Pulse generator 73 generates one or more stimulus waveforms, each of which is characterized by several parameters, including but not limited to pulse amplitude, pulse width, pulse frequency, number of pulses per burst, and burst frequency. As noted above, pulse amplitude may comprise pulse voltage or pulse current. Preferably, each of these parameters may be independently varied, as specified by control law output signal U 997 generated by control circuit 72. As noted, the stimulus waveforms comprising the neuromodulation signal (NMS) generated by output stage circuit 77 are applied to patient through intracranial (IC) stimulating electrode array 37. Pulse generator 73 generates a single waveform when single channel stimulation is to be used, and a plurality of waveforms when multiple channel stimulation is to be used. It may generate monophasic or biphasic waveforms.

In one preferred embodiment, charge balanced biphasic waveforms are produced. Those skilled in the art are aware that the net charge contained in a given pulse is given by the time integral of the stimulus current over the duration of the pulse. In a biphasic configuration, a pair of pulses of opposite polarity is generated, and the pulse current amplitude and pulse width are chosen such that the charge amplitude is equal in magnitude and opposite in polarity. In some cases, it is desirable for the pulses comprising the biphasic pulse pair to have different amplitudes; in this case, the pulse widths are chosen to insure equal and opposite charges so the pulse pair introduces zero net charge to the neural tissue 250. The capability to deliver pulse pairs with balanced charges is yet a further advantage conferred by the current regulation mode described above.

Even though the waveform parameters of the pulse pairs are calculated to deliver a zero net charge, in practice, noise

and precision limitations in computation and resolution limitations and nonlinearities in the digital to analog conversion and amplification stages may result in slight imbalances in the pulse pair charges. Over time, this can result in the delivery of a substantial accumulated net charge to the neural tissue. To eliminate this potential for net charge delivery to neural tissue, a direct current (DC) blocking capacitor is employed. This is a technique that is well known to those of ordinary skill in the art. In one preferred embodiment, a DC blocking capacitor is included within multiplexor 75 in series with stimulator output path 111.

Typically, multi-channel stimulation is used in the case of bilateral stimulation. Since the disease progression is typically asymmetrical, and the normal motor control systems governing movement on the left and right side of the body are also highly independent of each other, the delivery of treatment to the left and right sides of the body should be controlled separately. This represents one need for a multiple channel neuromodulation signal (NMS) 998. Multichannel stimulation is also expected to be beneficial in treating patients with variable involvement of different limbs. For example, the magnitude neuromodulation of a portion of the globus pallidus required to achieve optimal controls of arm tremor may be different from the optimal level of neuromodulation of separate portion of the globus pallidus to achieve optimal control of leg tremor. In this case, separate electrodes or electrode pairs are required to deliver optimal levels of neuromodulation to control tremor in these two regions of the body. Correspondingly, these separate electrodes or electrode pairs will be driven by separate neuromodulation signal (NMS) channels, necessitating a multichannel system.

A further need for multichannel neuromodulation signal (NMS) is the control of multiple symptoms of the movement disorder and the side effects arising from pharmacologic treatment. Optimal control of tremor, dyskinesias, and rigidity are not achieved by modulation of the same site at the same intensity. For this reason, multiple and separately controlled channels of neuromodulation are required to simultaneously achieve optimal control of these multiple symptoms and side effects. Each of these symptoms and side effects may be considered to comprise one or more element in a multivariable disease state. A multivariable control system will be required to optimally drive each of these disease state elements to its desired value, ideally toward a target minimum level and thus achieve optimal control of this multiplicity of disease states. This multivariable control system may be implemented as multiple independent control laws each with separate though potentially overlapping sensory inputs or as a multivariable control law matrix.

Stimulation via each of the multiple channels comprising the neuromodulation signal (NMS) 998 is characterized by separate though possibly overlapping sets of one or more of the following parameters: stimulation voltage, stimulation current stimulation frequency of pulses within the same burst, frequency of bursts, pulse width, pulses per burst, duration of burst, and interpulse interval. The stimulus waveforms are amplified by output amplifier 74 to generate an amplified stimulus waveform. Specifically, pulse generator 73 transfers information to output amplifier 74 which includes information that uniquely specifies the desired stimulation waveform. In a preferred embodiment, the information is in the form of an analog signal which represents a scaled version of the voltage or current waveform to be delivered to the tissue. It should be understood that other forms of the signal generated by pulse generator 73 may be used, including combinations of at least one of analog and digital representations. Output amplifier 74 performs amplification and regulation of the

received stimulus waveform generated by the pulse generator 73. This may be regulation of electrical current to achieve desired voltage or regulation of electrical voltage to achieve desired current, depending on whether a voltage or current waveform is to be delivered to the nervous system component.

As one skilled in the relevant art would find apparent, voltage regulation is simpler to implement, and is a technique which is commonly used by many conventional stimulators. Current regulation, on the other hand, is more complex but allows for more precise control of the applied stimulation. Current regulation insures that a specified amount of current is delivered, regardless of the impedance of the electrode. Current regulation is advantageous in that it allows for precise control of stimulation level despite changes in electrode impedance which invariably occur over time. Since electrode impedances often change, typically increasing as they become encapsulated by fibrosis, current regulation is preferred to avoid the decrease in current which would occur if voltage regulation were to be used in such circumstances.

The amplified stimulus waveform generated by output amplifier 74 is conducted along stimulator amplifier output path 112 to multiplexor 75. Multiplexor 75 allows for delivery of a stimulating electrode output signal (SEOS) to the intracranial stimulating electrode array 37, multiplexed with sensing of a stimulating electrode input signal (SEIS). Specifically, multiplexor 75 serves to alternately connect intracranial stimulating electrode (ICSE) array 37 to output amplifier 74 and intracranial stimulating electrode amplifier 57. Connection of intracranial stimulating electrode (ICSE) array 37 to output amplifier 74 facilitates delivery of neural modulation signal to neural tissue, while connection of intracranial stimulating electrode (ICSE) array 37 to intracranial stimulating electrode amplifier 57 facilitates monitoring of neural activity in the region being stimulated.

Multiplexor 75 allows delivery of neural modulation signals to neural tissue concurrent with monitoring of activity of same neural tissue; this facilitates real-time monitoring of disease state and response to treatment. Stimulating electrode output signal (SEOS) from output amplifier 74 is conducted along stimulator amplifier output path 112 to multiplexor 75. Multiplexor 75 conducts output from output amplifier 74 to stimulator output path 111 which conducts the stimulating electrode output signal to intracranial stimulating electrode array 37. To facilitate periodic sampling of neural activity in tissue being stimulated, multiplexor 75 alternatively conducts signal arising from stimulated tissue via intracranial stimulating electrode array (ICSE) 37 and stimulator output path 111 to multiplexed stimulator recording input path 113 and intracranial stimulating electrode amplifier 57.

Multiplexor 75 selectively conducts the signal on multiplexed stimulator recording input path 113 to amplifier 57. Multiplexor 75 may alternate conduction between path 111 and path 112 or path 113 using temporal multiplexing, frequency multiplexing or other techniques to allow concurrent access to the intracranial stimulating electrode (ICSE) array 37 for modulation of tissue activity and monitoring of tissue activity. Temporal multiplexing is a well known technique and frequency multiplexing of stimulation and recording signals in known to those skilled in the art. In this embodiment, temporal multiplexing is accomplished by alternately connecting stimulator output path 111 to stimulator amplifier output path 112 and multiplexed stimulator recording input path 113. In one embodiment, frequency multiplexing is accomplished by passing a band-limited portion of stimulating electrode output signal SEOS via the stimulator output path 111 to intracranial stimulating electrode array 37 while

simultaneously monitoring activity on intracranial stimulating electrode array 37 within a separate frequency band, thereby generating a stimulating electrode input signal SEIS. Thus, stimulating electrode input signal SEIS is conducted from the intracranial stimulating electrode array 37 to stimulator output path 111 to multiplexor 75 and via multiplexed stimulator recording input path 113 to intracranial stimulating electrode array amplifier 57.

Multiplexor 75 facilitates conduction between stimulator amplifier output path 112 and multiplexed stimulator recording input path 113 to allow automated calibration. In this mode, a calibration signal of known amplitude is generated by pulse generator 73 and amplified by output amplifier 74 which, for calibration purposes, delivers a voltage regulated signal via stimulator amplifier output path 112 to multiplexor 75. Multiplexor 75 conducts amplified calibration signal to multiplexed stimulator recording input path 113 which conducts signal to intracranial stimulating electrode amplifier 57.

Although not included in the illustrative embodiment, multiplexed or intermittent connection of stimulator amplifier output path 112 to the inputs of at least one of the other amplifiers, including EMG amplifier 59, EEG amplifier 60, accelerometer amplifier 61, acoustic amplifier 62, peripheral nerve electrode amplifier 63, and intracranial recording electrode amplifier 58, may be implemented without departing from the present invention. The same multiplexed connections may be used to calibrate the pulse generator 73 and output amplifier 74.

Referring to FIG. 15, an analog switch may be used to connect one or an opposing polarity pair of Zener diodes across the noninverting and inverting inputs of intracranial recording electrode amplifier 58. In this configuration, the Zener diodes would limit the maximal amplitude of the calibration signal in one or both polarities to known values, allowing for accurate calibration of intracranial recording electrode amplifier 58. The analog switch may then be deactivated, removing the cathode of the single or pair of Zener diodes from the input of intracranial recording electrode amplifier 58 to allow measurement of stimulating electrode output signal (SEOS) for calibration of pulse generator 73 and output amplifier 74. This is described in greater detail below.

Multiplexor 75 also facilitates conduction between stimulator amplifier output path 112, multiplexed stimulator recording input path 113, and stimulator output path 111 to allow measurement of impedances of components of intracranial stimulating electrode array 37. In this electrode impedance measurement mode, a three way connection between stimulator amplifier output path 112, multiplexed stimulator recording input path 113, and stimulator output path 111 is created. When output amplifier 74 is operated in current regulated mode, it delivers an SEOS of known current via stimulator output path 111 to intracranial stimulating electrode array 37. The voltages generated across the elements of intracranial stimulating electrode array 37 generally are the products of the electrode impedances and the known stimulating currents. These voltages are sensed as the stimulating electrode input signal SEIS by the intracranial stimulating electrical amplifier 57.

Reference module 1116 contains memory registers in which control law reference values are stored. Such reference values include but are not limited to target disease state levels, target symptom levels, including target tremor level, and threshold levels. Threshold levels include but are not limited to disease and symptom levels, including tremor threshold levels. Neural modulation amplitude may be increased when at least one of disease state and symptom level exceed the

corresponding threshold. Similarly neural modulation amplitude may be decreased or reduced to zero when either the disease state or symptom level falls below the corresponding threshold.

Reference module 116 is connected to patient interface module 55, facilitating both monitoring and adjustment of reference values by patient. Reference module 116 is also connected to supervisory module 56, facilitating both monitoring and adjustment of reference values by physician or other health care provider. Supervisory module 56 may be used by the neurologist, neurosurgeon, or other health care professional, to adjust disease state reference R values for the one or more control laws implemented in control circuit 72. The disease state reference R values specify the target level at which the corresponding disease states are to be maintained, as quantified by the disease state estimate X values, providing reference values for control laws implemented in control law circuit block 231 (FIG. 11; discussed below) and contained within control circuit 72. Reference module 116 may also receive input from control circuit 72, facilitating the dynamic adjustment of reference disease state "r" (discussed below). Reference module 116 may additionally receive input from disease state estimator module array (DSEMA) 229 (FIG. 11; discussed below) and aggregate disease state estimator 195 (FIG. 11 discussed below) and components of signal processor 71, for use in dynamically determining reference disease state "r".

FIG. 10 is a schematic diagram of signal processor 71. In this illustrative embodiment, signal processor 71 includes a disease state estimator module array 229 that includes one or more signal processor modules that generate a quantitative estimate of at least one disease state or parameter thereof based upon its respective input. For example, magnitude of tremor in the 3 to 5 Hz range represents one possible representation of a disease state. This could be an absolute or normalized quantification of limb acceleration in meters per second squared. This component of the disease state would be calculated almost exclusively from sensory feedback from accelerometer array 52. Another possible disease state is the frequency of occurrence of episodes of tremor activity per hour. This element of the disease state may be estimated from any of several of the sensory feedback signals. In this case, the most accurate representation of this disease state element is obtained by applying a filter such as a Kalman filter to calculate this parameter based upon a weighted combination of the sensory feedback signals. Such weighting coefficients are calculated from quantified measures of the accuracy of and noise present upon each sensory feedback channel.

In the illustrative embodiment, disease state estimator module array 229 includes an EMG signal processor 233, EEG signal processor 234, accelerometer signal processor 235, acoustic signal processor 236, peripheral nerve electrode (PNE) signal processor 237, intracranial recording electrode (ICRE) signal processor 238, and intracranial stimulating electrode (ICSE) signal processor 239. It should be understood that other signal processors may also be included in the array 229. Inputs to these modules include conditioned EMG signal path 78, conditioned EEG signal path 79, conditioned accelerometer signal path 80, conditioned acoustic signal path 81, conditioned peripheral nerve electrode (PNE) signal path 82, conditioned intracranial recording electrode (ICRE) signal path 83, and conditioned intracranial stimulating electrode (ICSE) signal path 84, respectively. Communication between these modules is facilitated. The output(s) of each of the modules is connected to an aggregate disease state estimator 195. Aggregate disease state estimator 195 generates a

25

single or plurality of disease state estimates "X" indicative of state of disease and response to treatment.

In the preferred embodiment, the acceleration of at least one of the affected limb and the head, each of which is sensed as a sensory feedback channel by an element of the accelerometer array **52**, serves as respective elements in the disease state estimate X. These elements of disease state estimate X are inputs to respective control laws implemented in control circuit **72**. of input to the control law. A control law governing the function of a proportional controller using acceleration as its single sensory feedback channel is given by equation (1):

$$u_1 = 0.3166(V \cdot S^2/m) \cdot ACC \quad (1)$$

and if

$$u_2 = 0.6333(V \cdot S^2/m) \cdot ACC \quad (2)$$

where u_1 and u_2 are the stimulation voltage given in volts; and ACC is the limb, mandible, or head acceleration given in meters per second squared (m/s^2).

In equation (1), the stimulation site is the ventroposterolateral pallidum, the output stage mode is voltage regulated, the waveform is a continuous train of square waves, the amplitude u_1 is given in volts (typically approximately 1 volt), and the remaining stimulation parameters include a pulse width of 210 microseconds, and a stimulation frequency of 130 Hz. In equation (2), the stimulation site is the ventral intermediate thalamic nucleus (Vim), the output stage mode is voltage regulated, the waveform is an intermittent train of square waves with an on time of 5 minutes and an off time of 45 seconds, the amplitude u_2 is given in volts (typically approximately 3 volts), and the remaining stimulation parameters include a pulse width of 60 microseconds, and a stimulation frequency of 130 Hz.

In one preferred embodiment, the ACC signal represents the average acceleration over a finite time window, typically 15 to 60 seconds. This effective lowpass filtering provides a stable sensory feedback signal for which a proportional control law is appropriate. If stability and performance requirements dictate, as is familiar to those practiced in the art of feedback control, other components, including an integrator and a differentiator may be added to the control law to produce a proportional-integral-differential (PID) controller, as needed.

One preferred embodiment also includes electromyographic (EMG) signals as sensory feedback in the calculation of at least one element of the disease state estimate X which is an input to the control law. As discussed in the section describing EMG signal processor **233**, the EMG signals are rectified by full wave rectifier **123**, passed through envelope determiner **124**, passed through several bandpass filters **125**, **127**, **129**, **131**, **133** and associated threshold discriminators **126**, **128**, **130**, **132**, **134** and then passed in parallel to each of integrator **135** and counter **136**. Integrator **135** generates an output which is a weighted function of it inputs and represents the average magnitude of tremor activity over a given time window $-w/2$ to $+w/2$. A simplified representation of this is given by equation (3):

$$u_3 = \int_{-w/2}^{+w/2} X_{EMG} - dt \quad (3)$$

over a given time window $-w/2$ to $+w/2$. A simplified representation of this is given by the equation:

As is familiar to those skilled in the art of control theory, an integral controller is marginally stable. To confer stability to

26

this control law, the equivalent of a finite leak of the output magnitude $u_{sub.4}$ to zero is added to maintain stability. A more general form of this equation is given by equation (4):

$$-C_1 \partial u_4 / \partial t + C_2 u_4 = B_1 \cdot \partial X_{EMG} / \partial t + B_2 \cdot X_{EMG} \quad (4)$$

Shown as a system function, the control law output U is given as the product of a transfer function H(s) and the disease estimate X, the input to the control law:

$$u(s) / (C_1 \circ s + C_2) = X_{EMG}(s) / (B_1 \circ s + B_2) \quad (5)$$

$$u(s) / X_{EMG}(s) = (B_1 \circ s + B_2) / (C_1 \circ s + C_2) \quad (6)$$

$$H(s) = u(s) / X_{EMG}(s) = (B_1 \circ s + B_2) / (C_1 \circ s + C_2) \quad (7)$$

One such control law with an appropriate time response is given by:

$$H(s) = u(s) / X_{EMG}(s) = G_{VEMG} (0.1 \circ s + 1) / (2s + 1) \quad (8)$$

where G_{VEMG} is the gain in neuromodulation signal (NMS) (volts per volt of EMG signal).

For intramuscular EMG electrodes, signal amplitudes are on the order of 100 microvolts. For neuromodulation signal (NMS) parameters of 2 volts amplitude, 60 microseconds pulse width, 130 Hz stimulation frequency, the appropriate overall gain G'_{VEMG} is 20,000 volts_{NMS}/volts_{EMG}. Since the preamplifier stage performs amplification, 1000, in the preferred embodiment, the actual value for G_{VEMG} as implemented in the control law is 20 volts_{NMS}/volts_{PREAMPL EMG}.

Disease state estimator **195** determines estimates of disease state including but not limited to long-term, or baseline, components, circadian components, postprandial components, medication induced alleviation of components, medication induced components, and future predicted behavior of said components. Output of disease state estimator **195** includes output of observer **228**, depicted in FIG. **11**, which makes use of an adaptive model of disease behavior to estimate disease states which are not directly detectable from sensors. Such sensors provide input to the adaptive model to correct state estimates and model parameters. Each of the signal processor modules in disease state estimator module array **229** are described below.

FIG. **3** is a block diagram of intracranial recording electrode (ICRE) signal processor **238** and intracranial stimulating electrode (ICSE) signal processor **239**, each of which are included within signal processor **71** in the illustrative embodiment illustrated in FIGS. **2** and **10**. ICRE signal processor module **238** and ICSE signal processor module **239** process signals from one or more intracranial electrodes, including but not limited to those comprising intracranial recording electrode array **38** and intracranial stimulating electrode array **37**. As noted, intracranial stimulating electrode array **37** is comprised of one or more intracranial stimulating electrodes while intracranial recording electrode array **38** is comprised of one or more intracranial recording electrodes.

Input to ICRE signal processor **238** is conditioned intracranial recording electrode (ICRE) signal path **83** noted above. This input is connected to a spike detector **85** which identifies action potentials. Spike detection techniques are well known to those skilled in the art and generally employ low and high amplitude thresholds. Waveforms having amplitudes greater than the low threshold and lower than the high threshold are determined to be action potentials. These thresholds may be predetermined or adjusted manually using supervisory module **56** or may be adapted in real-time by an algorithm which sweeps the threshold through a range of values to search for values at which action potential spikes are consistently recorded. The low amplitude threshold is set above the amplitude of background noise and that of nearby cells not of

interest, and the high amplitude threshold is set above the amplitude of the desired action potentials to allow their passage while eliminating higher amplitude noise spikes, such as artifacts arising from electrical stimulation currents. Bandpass, notch, and other filtering techniques may also be used to improve signal to noise ratio and the sensitivity and specificity of spike detectors. Individual neuron action potentials are usually recorded using fine point high-impedance electrodes, with impedances typically ranging from 1 to 5 megohms. Alternatively, larger lower-impedance electrodes may be used for recording, in which case the signals obtained typically represent aggregate activity of populations of neurons rather than action potentials from individual neurons. Spike detector **85** passes the waveform(s) to a spike characterizer **86**. Spike characterizer **86** determines firing patterns of individual neurons. The patterns include, for example, tonic activity, episodic activity, and burst firing. Spike characterizer **86** calculates parameters that characterize the behavior of the individual and groups of neurons, the activity of which is sensed by intracranial recording electrode array **38**. In one embodiment, the characterization includes parameterization of recorded action potentials, also referred to as spikes, bursts of spikes, and overall neural activity patterns. This parameterization includes, but is not limited to, calculation of frequencies of spikes, frequencies of bursts of spikes, inter-spike intervals, spike amplitudes, peak-to-valley times, valley-to-peak times, spectral composition, positive phase amplitudes, negative phase amplitudes, and positive-negative phase differential amplitudes. These parameters are depicted in FIG. **14** and are discussed below. Based on these parameterization, spike characterizer **86** discriminates individual spikes and bursts originating from different neurons. This discrimination facilitates serial monitoring of activity of individual and groups of neurons and the assessment and quantification of activity change, reflective of change in disease state and of response to therapy.

A spike analyzer **87** receives as input the parameters from spike characterizer **86**. Spike analyzer **87** extracts higher level information, including but not limited to average spike frequencies, average interspike intervals, average amplitudes, standard deviations thereof, trends, and temporal patterning. By comparing current spike frequency rates to historical spike frequency data, spike analyzer **87** additionally calculates the rates of change of spike parameters. Prior trends and current rates of change may then be used to predict future behaviors. Rates of change of the parameters include but are not limited to autocorrelation and digital filtering.

Spike analyzer **87** may receive additional input from accelerometers, including but not limited to at least one of head mounted accelerometer **12**, proximal accelerometer **28**, enclosure mounted accelerometer **36**, and distal accelerometer **33**. Spike analyzer **87** may receive indirect input from accelerometers, such as from conditioned or processed signals arising therefrom. This may include, for example, the signal transmitted by conditioned accelerometer signal path **80**.

Spike analyzer **87** may also receive additional input from EMG arrays **50**, such as a proximal EMG electrode array **45**, enclosure-mounted EMG electrode array **46**, or distal EMG electrode array **47**. Spike analyzer **87** may receive indirect input from such EMG electrode arrays **50**, such as from conditioned or processed signals arising therefrom, including but not limited to the signal transmitted by conditioned EMG signal path **78**.

These additional inputs from accelerometers and EMG arrays facilitates the characterization of neuronal firing patterns relative to activity of muscle groups and movement

of joints, including but not limited to characterization of neuronal spike amplitudes and tuning of firing to movement, including but not limited to movement velocity and direction. The characterization may be used to assess functioning of the sensorimotor system, including but not limited to motor response time, and to measure the disease state and response to therapy.

Intracranial recording electrode (ICRE) single unit-based (SU) disease state estimator **88** receives input from spike characterizer **86** and/or spike analyzer **87**. Spike analyzer **87** provides higher level information, including but not limited to average spike frequencies, average interspike intervals, average amplitudes, standard deviations thereof, trends, and temporal patterning to disease state estimator **88**. These inputs are representative of the current neuronal activity in the tissue from which the intracranial recording electrodes (ICRE) are recording. ICRE SU disease state estimator **88** may also receive input representative of one or more signals, including desired neuronal activity, from control circuit **72**. The ICRE SU disease state estimate X_{ICRE_SU} calculated by ICRE SU disease state estimator **88**, may be comprised of a single or a plurality of signals, consistent with a representation of the disease state by a single or a multitude of state variables, respectively. The ICRE MU disease state estimate X_{ICRE_MU} calculated by ICRE MU disease state estimator **88**, may be comprised of a single or a plurality of signals, each representative of multiunit neurophysiological signals, i.e. reflective of concurrent activity of numerous neurons. Both ICRE SU disease state estimate X_{ICRE_SU} and ICRE MU disease state estimate X_{ICRE_MU} are output to aggregate disease state estimator **195**.

Referring to FIG. **3**, conditioned intracranial recording electrode (ICRE) signal path **83** additionally connects to filter **101**. Filter **101** is preferably of the bandpass type filter. In one embodiment, the bandpass filter **101** has a passband of 0.1 to 100 Hz, although other ranges may be used. Output of filter **101** connects to spectral energy characterizer **102**, which may be implemented in any of several hardware or software forms. For example, in one embodiment, the spectral energy characterizer **102** is implemented using real-time fast Fourier transform (FFT) techniques. Alternatively, other digital or analog techniques may also be used.

It should be understood that inputs and outputs from spike detector **85**, spike characterizer **86**, spike analyzer **87**, disease state estimator **88**, filter **101**, spectral energy characterizer **102**, spectral energy analyzer **103**, and disease state estimator **104** may be comprised of individual signals or a plurality of signals. Further, spike detector **85**, spike characterizer **86**, spike analyzer **87**, disease state estimator **88**, filter **101**, spectral energy characterizer **102**, spectral energy analyzer **103**, and disease state estimator **104** may each have different parameters and signal processing characteristics for each of the multiple signals processed. Because baseline neuronal firing rates differ among various anatomical and functional regions of the brain, and their involvement in disease states and susceptibility to change in firing patterns varies, the respective signal processing circuitry and logic will vary correspondingly. For example, baseline firing rates among neurons in the globus pallidus externus are approximately 43 Hz and those in the globus pallidus internus are 59 Hz.

The input to intracranial stimulating electrode ICSE signal processor **239**, referred to above as conditioned intracranial stimulating electrode (ICSE) signal path **84**, connects to spike detector **89**. Spike detector **89** identifies action potentials in a manner similar to that described above with reference to spike detector **85**. Intracranial stimulating electrode ICSE signal processor **239** performs a similar set of functions as intracra-

nial recording electrode ICRE signal processor **238** on a different set of sensory feedback signals. As noted above, spike detection techniques are well known to those skilled in the art.

Spike detector **89** passes waveforms to spike characterizer **90**, which uses well known techniques to calculate parameters than characterize the behavior of the individual and groups of neurons, the activity of which is sensed by intracranial stimulating electrode array **37**. As noted above with respect to spike characterizer **86**, this characterization may include parameterization of spikes, bursts of spikes, and overall neural activity patterns. Similarly, the parameterization may include calculation of spike frequencies, burst frequencies, inter-spike intervals, amplitudes, peak-to-valley times, valley-to-peak times, spectral composition, positive phase amplitudes, negative phase amplitudes, and positive-negative phase differential amplitudes. Such characterization of neural spikes is known to those skilled in the art of neurophysiology. Based on this parameterization, spike characterizer **90** discriminates individual spikes and bursts originating from different neurons. As noted, such discrimination facilitates serial monitoring of activity of individual and groups of neurons and the assessment and quantification of activity change, reflective of change in disease state and of response to therapy.

Spike analyzer **91** receives the parameters from spike characterizer **90**, and extracts higher level information, including average spike frequencies, average interspike intervals, average amplitudes, standard deviations thereof, trends, and temporal patterning. The function and operation of spike analyzer **91** is similar to that described herein with reference to spike analyzer **87**. Similarly, spike analyzer **91** may receive additional input directly or indirectly from accelerometers and/or EMG arrays to facilitate the characterization of neuronal firing patterns relative to activity of muscle groups and movement of joints. This may include, for example, characterization of neuronal spike amplitudes and tuning of firing to movement, including but not limited to movement velocity and direction. Such characterization may be used to assess functioning of the sensorimotor system, including but not limited to motor response time, and to measure the disease state and response to therapy.

Intracranial stimulating electrode (ICSE) single unit-based (SU) disease state estimator **92** receives input from either or both spike characterizer **90** and spike analyzer **91**. ICSE SU disease state estimator **92** receives input representative of the current neuronal activity from spike characterizer **90**. ICSE SU disease state estimator **92** may receive input representative of at least one of several signals, including desired neuronal activity, actual neuronal activity, and the difference between these quantities. The ICSE SU disease state estimate, calculated by ICSE SU disease state estimator **92**, may be comprised of a single or a plurality of signals, consistent with a representation of the disease state by a single or a multitude of state variables, respectively.

As with intracranial recording electrode signal processor **238**, inputs and outputs from spike detector **89**, spike characterizer **90**, spike analyzer **91**, disease state estimator **92**, filter **106**, spectral energy-characterizer **107**, spectral energy analyzer **108**, and disease state estimator **109** may include individual or a plurality of signals, and each may have different parameters and signal processing characteristics for each of the multiple signals processed. Because baseline neuronal firing rates differ among various anatomical and functional regions of the brain, and their involvement in disease states

and susceptibility to change in firing patterns varies, the respective signal processing circuitry and logic varies correspondingly.

FIG. **4** is a schematic diagram of a globus pallidus **119** implanted with stimulating and recording electrodes. Intracranial catheter **7** is shown in place with electrode of the intracranial stimulating electrode array **37** located within the globus pallidus internus (Gpi) **120**, including globus pallidus internus internal segment (GPI,i) **94** and globus pallidus internus external segment (GPI,e) **95**, and globus pallidus externus (GPe) **96**.

Intracranial stimulating electrodes **1** and **2** are shown implanted in the globus pallidus internus internal segment (GPI,i) **94**; and intracranial stimulating electrodes **3** and **4** are shown implanted in the globus pallidus internus external segment (GPI,e) **95** and globus pallidus externus (GPe) **96**, respectively. It should be understood that this arrangement is illustrative of one preferred embodiment, and other stimulating and recording electrode configurations may be employed without departing from the present invention.

The optic tract **97** is shown in its close anatomical relationship to the globus pallidus internus (Gpi) **120**. The risk inherent in treatment modalities involving irreversible tissue ablation should be apparent; stereotactic errors of only one to several millimeters during lesioning of the globus pallidus internus (Gpi) **120** may result in irreversible damage or complete destruction of the optic tract **97**. Furthermore, the advantage of a system which dynamically adjusts the amplitude of inhibitory electrical stimulus to the globus pallidus **119** to minimize said amplitude offers the potential advantage of minimization of side effects including interference with visual signals of the optic tract **97** and prevention of overtreatment.

Intracranial stimulating electrodes **1**, **2**, **3**, **4** are shown implanted in the GPI,i **94**, GPI,e **95**, GPe **96**, respectively. This is one preferred embodiment. Numerous permutations of electrode stimulation site configuration may be employed, including more or fewer electrodes in each of these said regions, without departing from the present invention. Electrodes may be implanted within or adjacent to other regions in addition to or instead of those listed above without departing from the present invention, said other reasons including but not limited to the ventral medial Vim thalamic nucleus, other portion of the thalamus, subthalamic nucleus (STN), caudate, putamen, other basal ganglia components, cingulate gyrus, other subcortical nuclei, nucleus locus ceruleus, pedunculo-pontine nuclei of the reticular formation, red nucleus, substantia nigra, other brainstem structure, cerebellum, internal capsule, external capsule, corticospinal tract, pyramidal tract, ansa lenticularis, white matter tracts, motor cortex, premotor cortex, supplementary motor cortex, other motor cortical regions, somatosensory cortex, other sensory cortical regions, Broca's area, Wernicke's area, other cortical regions, other central nervous system structure, other peripheral nervous system structure, other neural structure, sensory organs, muscle tissue, or other non-neural structure.

Referring to FIGS. **3** and **4**, a small percentage of cells in the globus pallidus internus internal segment **94** and globus pallidus internus external segment **95** exhibit tremor-synchronous discharges. As noted, at least one of single unit recordings from individual cells and multiple unit recordings from a plurality of cells are processed by signal processor **71**. The single and multiple unit recordings may be derived from signals arising from intracranial stimulating electrode array **37**, intracranial recording electrode array **38**, or other sources. The output from signal processor **71** is connected to control circuit **72** and the output may represent at least one of disease

state, magnitude of symptomatology, response to therapy, other parameter, and combination thereof.

Individual electrodes comprising intracranial stimulating electrode array 37 and intracranial recording electrode array 38 may each be of the microelectrode type for single unit recordings, macroelectrode type for multiple unit recordings, other electrode type, or a combination thereof, without departing from the spirit of the present invention. In one preferred embodiment, intracranial stimulating electrode array 37 consists of macroelectrodes. The macroelectrodes facilitate delivery of stimulation current at a lower charge density (coulombs per unit of electrode surface area) than microelectrodes of the same chemistry and surface treatment. The dimensions of intracranial stimulating electrodes 1-4 are selected such that the current density, or electrical current divided by electrode surface area, is below the threshold of reversible charge injection for the given electrode material.

Standard single cell recording technique, using an electrode with an impedance of typically 1-2 Megohms, involves bandpass filtering with -6 decibel (dB) points at 300 and 10,000 Hertz. This filtering, or a modification thereof, may be accomplished by ICRE filter 65 and ICSE filter 64; alternatively, it may be performed in spike detector 85 and spike detector 89, respectively, or other portion of stimulating and recording circuit 26.

FIG. 5 is a block diagram of one embodiment of an EMG signal processor 233 which is included in a preferred embodiment of signal processor 71. EMG signal processor 233 processes signals from EMG electrode array 50, performing functions including but not limited to full wave rectification, envelope determination, bandpass filtering, threshold discrimination, and others described in more detail below, to produce signals indicative of the overall magnitude of tremor as well as the frequency at which tremor episodes occur. As noted, EMG electrode array 50 includes, but is not limited to, proximal EMG electrode array 45, enclosure-mounted EMG electrode array 46, and distal EMG electrode array 47. EMG electrodes may be located in any implanted or external location without departing from the present invention. For example, electrodes may be located within or in proximity to the hand, forearm, arm foot, calf, leg, abdomen, torso, neck, head, jaw, lip, eyelid, larynx, vocal cords, and tongue.

Conditioned EMG signal path 78 is also connected to a well-known full wave rectifier 123 now or later developed. Output from the full wave rectifier 123 is coupled to an input of an envelope determiner 124. Determination of the envelope of a modulated signal is well known to those skilled in the art of electronics; this may be readily implemented in analog or digital hardware or in software. Output of envelope determiner 124 is connected to inputs of filters 125, 127, 129, 131 and 133. In one embodiment, filters 125, 127, 129, 131, 133 implement passbands of approximately 0.1-2 Hz, 2-3 Hz, 3-5 Hz, 7-8 Hz, and 8-13 Hz, respectively. Outputs of filters 125, 127, 129, 131 and 133 are connected to threshold discriminators 126, 128, 130, 132, 134, respectively.

Threshold discriminators 126, 128, 130, 132, and 134 generate outputs representing episodes of normal voluntary movement (Mv), low frequency intention tremor (Til) resting tremor (Tr), high frequency intention tremor (Tih), and physiologic tremor (Tp), respectively. These outputs are each connected to both of integrator 135 and counter 136. Integrator 135 generates outputs representative of the total activity of each of the above types of movement over at least one period of time. One such time period may be, for example, time since implantation, time since last visit to physician or health care provider, month interval, week interval, day interval, interval since last medication dose, interval since last change in stimu-

lation parameters, weighted average of multiple time windows, and convolution of said activity with arbitrary time window function.

Counter 136 generates outputs representative of the number of episodes of each of the above types of movement over at least one period of time. Such period of time may be, for example, time since implantation, time since last visit to physician or health care provider, month interval, week interval, day interval, interval since last medication dose, interval since last change in stimulation parameters, and weighted average of said number of episodes over multiple time windows. Outputs from integrator 135 and counter 136 are connect to EMG analyzer 137. EMG analyzer 137 performs a number of functions including, for example, calculation of proportions of tremor activity which are of the rest and the intention type, ratios of different types of tremor activity, the level of suppression of resting tremor activity with voluntary movement, assessment of temporal patterns of EMG activity. EMG disease state estimator 138 receives inputs from EMG analyzer 137 and generates output representative of disease state based upon said input. In one preferred embodiment, two disease states are calculated, including a signal representative of the overall magnitude of tremor activity and a signal representative of the frequency of occurrence of tremor events. It should be understood that all signals paths may transmit one or more signals without departing from the present invention.

EMG signals may be sensed from any individual or group of muscles and processed in a manner including but not limited to the determination of severity and frequency of occurrence of various tremor types. Normal or physiologic tremor includes movement in the 8-13 Hz range and may be used as a normalization for the other types of sensed tremor. The predominant pathological form of tremor exhibited in Parkinson's disease patients is the classical "resting" tremor which includes movements in the 3-5 Hz range which are present at rest and suppressed in the presence of voluntary movement. In the present invention, quantification of this tremor type serves as a heavily weighted sensory input in the assessment of disease state and response to therapy. Parkinson's disease patients may also exhibit intention tremor, of which there are two types. The first type of intention tremor is referred to as "low frequency intention tremor" (Til in the present invention) and consists of movements in the 2-3 Hz range. A second type of intention tremor is referred to as "high frequency intention tremor" (Tih in the present invention) and consists of irregular movements in the 7-8 Hz range which persist throughout voluntary movement. Other types of tremor having associated movement in other ranges may be sensed and represented by the EMG signals.

EMG signals from at least one of orbicularis oculi (effecting eye closure), levator palpebrae (effecting eye opening), and other muscles contributing to eyelid movement, may be sensed and processed to determine frequency of eye blinking. Patients with Parkinson's disease exhibit a reduction in eye-blinking frequency from the normal of 20 per minute to 5 to 10 per minute, and this parameter is sensed as a measure of disease severity and response to treatment. Additionally, said EMG signals may be sensed and processed for detection and quantification of blepharoclonus, or rhythmic fluttering of the eyelids, and used as a measure of disease state and response to therapy. EMG signals, including baseline levels thereof, may be used to quantify rigidity and hypertonus as measures of disease state and response to therapy. Discharge patterns of individual motor units, including but not limited to synchronization of multiple units and distribution of intervals preced-

ing and following discharge, may be used as measures of disease state and response to therapy.

FIG. 6 is a block diagram of one embodiment of an EEG signal processor module **234** which is included in embodiments of signal processor **71**. The EEG signal processor module **234** processes signals from EEG electrode array **51**. Conditioned EEG signal path **79** connects to an input of artifact rejecter **139** which rejects signals with amplitudes above a threshold. In one embodiment, this threshold is 0.1 mV. An output from artifact rejecter **139** connects to an input of each of supplementary motor area signal extractor **140** and filters **143, 146, 149, 152, 219**. Filters **143, 146, 149, 152, and 219** are preferably of the bandpass type with passbands of 13-30 Hz, 8-13 Hz, 4-7 Hz, 0.1-4 Hz, and 0.1-0.3 Hz, respectively. Each filter output is connected to an input of an associated full wave rectifier **141, 144, 147, 150, 153, 220**. Each full wave rectifier **141, 144, 147, 150, 153, 220** is connected to an input of an associated envelope determiner **142, 145, 148, 151, 154, and 221**, respectively. The envelope determiners generate a signal representative of the envelope of the input signal, typically performed by lowpass filtering with a time constant of 5 seconds. Finally, outputs of envelope determiners **142, 145, 148, 151, 154, and 221** are connected to EEG disease state estimator **155**.

Signal SMA generated by supplementary motor area signal extractor **140** represents activity in the supplementary motor area ipsilateral to the intracranial stimulating electrode array (ISEA) **37**. Supplementary motor area signal extractor **140** amplifies signals which are unique to elements of the EEG electrode array **51** which overlie the supplementary motor area. The supplementary motor area receives neural signals via neural projections from the basal ganglia and exhibits decreased activity in patients with Parkinson disease. The SMA is essential for sequential movements, which are often impaired in Parkinson's disease patients. The SMA signal provides a quantitative measure of disease state and response to therapy. The SMA signal is extracted from the anterior EEG leads, predominantly from those in the vicinity of the frontal cortex, and provides a quantitative measure of disease state and response to therapy. Signals beta, alpha, theta, and delta consist of 13-30 Hz, 8-13 Hz, 4-7 Hz, and 0.1-4 Hz activity, respectively.

Signal "resp" consists of 0.1-0.3 Hz activity and reflects respiration. Parkinson's disease patients exhibit irregular respiratory patterns characterized by pauses and by abnormally deep breathing while at rest and preceding speech. Assessment of respiratory irregularity as well as other parameters derived from such resp signal serve as quantitative measures of disease state and response to therapy.

Anterior EEG electrodes are also used to sense EMG signals, and the EMG signals are processed to determine activity of muscles including but not limited to those related to eye blinking activity. Processing of the EMG signals is included in the FIG. 6 circuit block diagram which contains the EEG signal processing component of signal processor **71**. However, the processing could be incorporated into EMG signal processing component of signal processor **71** without departing from scope of the present invention. Conditioned EEG signal path **79** is additionally connected to input of full wave rectifier **222**, the output of which is connected to the input of an envelope determiner **223**. Envelope determiner **223** includes an output connected to input of filter **224**. Filter **224** is preferably of the bandpass type with a passband range of 0.1 to 20 Hz. Filter **224** has an output connected to input of threshold discriminator **225**, the output of which is connected to EEG disease state estimator **155**.

Preferably, EMG signals arising from activity of at least one of orbicularis oculi (effecting eye closure), levator palpebrae (effecting eye opening), and other muscles the activity of which is associated with eyelid movement are sensed by anterior EEG electrodes. These EMG signals are processed to determine eye blink events, and the rates and regularity of eye blinking activity are calculated. Frequency and irregularity of eyeblinking as well as blepharoclonus, or rhythmic fluttering of the eyelids, are quantified as measures of disease state and response to therapy.

FIG. 7 is a block diagram of one embodiment of an accelerometer signal processor **235** which is incorporated into certain embodiments of signal processor **71**. The accelerometer signal processor **235** processes signals from accelerometer array **52**. Conditioned accelerometer signal path **80** is connected to an input of each of a plurality of filters **156, 160, 164, 168, 172**. The filters are preferably of the bandpass type with passbands of 0.1-2 Hz, 2-3 Hz, 3-5 Hz, 7-8 Hz, and 8-13 Hz, respectively. Other passband frequency ranges may also be used. The output of each filter **156, 160, 164, 168, 172** is connected to an associated full wave rectifiers **157, 161, 165, 169, and 173**, respectively. The output of each rectifier **157, 161, 165, 169, and 173** is connected to an associated envelope determiners **158, 162, 166, 170, and 174**, respectively. Outputs of envelope determiners **158, 162, 166, 170, and 174** are connected to inputs of an associated threshold discriminators **159, 163, 167, 171, and 175**, respectively.

Outputs of threshold discriminators **159, 163, 167, 171, 175** represent episodes of normal voluntary movement (Mv), low frequency intention tremor (Til), resting tremor (Tr), high frequency intention tremor (Tih), and physiologic tremor (Tp), respectively. These outputs are each connected to an integrator **176** and a counter **177**. Integrator **176** generates outputs representative of the total activity of each of the above types of movement over at least one period of time. As noted, such a time period may be, for example, time since implementation, time since last visit to physician or health care provider, or some other time interval, weighted average of multiple time windows, or convolution of selected activities with an arbitrary time window function.

Counter **177** generates outputs representative of the number of episodes of each of the above types of movements over at least one such period of time. Outputs from integrator **176** and counter **177** are connect to an acceleration analyzer **178**. Acceleration analyzer **178** calculates proportions of tremor types, such as the rest and intention types, ratios of different types of tremor activity, the level of suppression of resting tremor activity with voluntary movement, and assessment of temporal patterns of movement and acceleration. Acceleration analyzer **178** may perform some or all of these calculations, as well as other calculations, on alternative embodiments of the present invention. Acceleration-based disease state estimator **179** receives input from acceleration analyzer **178** and generates output representative of disease state based upon such input.

It should be understood that accelerometer signals may be sensed from any individual or group of body components. For example, such signals may be sensed from joints, bones, and muscles. Furthermore, such signals may be processed in any well known manner, including the determination of severity and frequency of occurrence of various tremor types. The types of tremor have been described above with respect to FIG. 5.

FIG. 8 is a block diagram of one embodiment of an acoustic signal processor **236** which is included in certain embodiments of signal processor **71**. Acoustic signal processor **236** processes signals from acoustic transducer array **53**. Condi-

tioned acoustic signal path **81** is connected to a full wave rectifier **180** and a spectral analyzer **185**. The output of full wave rectifier **180** is connected to an input of an envelope determiner **181**, an output of which is connected to an input of a low threshold discriminator **182** and a high threshold discriminator **183**. Low threshold discriminator **182** and high threshold discriminator **183** each have an output connected to an input of timer **184**. Timer **184** generates an output signal representing latency (Lat) and is connected to acoustic analyzer **186**. An output of acoustic analyzer **186** is connected to an input of acoustic-based disease state estimator **187**. Latency (Lat) represents the latency between initiation of vocal utterance and the subsequent achievement of a threshold level of vocal amplitude. Such a vocal amplitude level is set by high threshold discriminator **183** and may represent steady state vocal amplitude or a preset or dynamically varying threshold. Latency from voice onset to achievement of steady state volume may be delayed in patients with Parkinson's disease and is calculated as a measure of disease state and response to therapy.

Acoustic analyzer **186** receives input from spectral analyzer **185**. The respiratory pattern is determined from rhythmic modulation of voice and breathing sounds, sensed by elements of the acoustic transducer array **53**. Irregularity and pauses in respiration as well as abnormally deep breathing patterns at rest and preceding speech are exhibited in Parkinson's disease patients. Such parameters are quantified and used as estimates of disease state and response to therapy. Respiration durations are quantified; abnormally deep respiration both during rest and preceding speech are identified and used as indicators of disease state and response to therapy. Pauses in speech and decline in speech amplitude, or fading, are additionally monitored as indicators of disease state and response to therapy. Spectral composition of speech is monitored and the change in spectral composition, reflective of changes of pharyngeal and laryngeal geometry, are quantified. Additionally, the fundamental vocal frequency; that is, the frequency at which the epiglottis vibrates, is extracted and that standard deviation of the fundamental vocal frequency is calculated over various time intervals as a quantified measure of the monotonic quality of speech characteristic of Parkinson's disease. This serves as yet another indicator of disease state and response to therapy.

FIG. 9 is block diagram of one embodiment of a peripheral nerve electrode (PNE) signal processor **237** which is implemented in certain embodiments of signal processor **71**. PNE signal processor **237** processes signals from peripheral nerve electrode array **54**. These signals provided by peripheral nerve electrode array **54** are provided to PNE signal processor **237** via conditioned PNE signal path **82**. Conditioned PNE signal path **82** is connected to an input of a spike detector **188** and a filter **191**.

Spike detector **188** identifies action potentials. As noted, spike detection techniques are well known to those skilled in the art, and generally employ low and high amplitude thresholds. Waveforms with amplitudes greater than the low threshold and lower than the high threshold are determined to be action potentials. These thresholds may be adjusted in real-time, and the low amplitude threshold is set above the amplitude of background noise and that of nearby cells not of interest, and the high amplitude threshold is set above the amplitude of the desired action potentials to allow their passage while eliminating higher amplitude noise spikes, such as artifacts arising from electrical stimulation currents. It should be understood that bandpass, notch, and other filtering techniques may also be used to improve signal to noise ratio and the sensitivity and specificity of spike detectors. Individual neuron

action potentials are usually recorded using fine point high-impedance electrodes, with impedances typically ranging from 1 to 5 megohms. Alternatively, larger lower-impedance electrodes may be used for recording, in which case the signals obtained typically represent aggregate activity of populations of neurons rather than action potentials from individual neurons. As noted above, peripheral nerve electrode array **54** may include such electrodes as single unit recording microelectrodes, multiple unit recording microelectrodes, intrafascicular electrodes, other intraneural electrodes, epineural electrodes, and any combination thereof.

A spike characterizer **189** determines firing patterns of individual neurons, including, for example, tonic activity, episodic activity and burst firing. Spike characterizer **189** receives the signals passed by spike detector **188** and calculates parameters that characterize the behavior of the individual and groups of neurons, the activity of which is sensed by peripheral nerve electrode array **54**. Such characterization includes but is not limited to parameterization of spikes, bursts of spikes, and overall neural activity patterns. Parameterization includes but is not limited to calculation of frequencies of spikes, frequencies of bursts of spikes, inter-spike intervals, spike amplitudes, peak-to-valley times, valley-to-peak times, spectral composition, positive phase amplitudes, negative phase amplitudes, and positive-negative phase differential amplitudes. These parameters are described in further detail below with reference to FIG. 14. Based on this parameterization, spike characterizer **189** discriminates individual spikes and bursts originating from different neurons. The discrimination facilitates aerial monitoring of activity of individual and groups of neurons and the assessment and quantification of activity change, reflective of change in disease state and of response to therapy.

A spike analyzer **190** receives as input the parameters from spike characterizer **189**, and extracts higher level information, including but not limited to average spike frequencies, average frequencies of bursts of spikes, average interspike intervals, average spike amplitudes, standard deviations thereof, trends, and temporal patterning.

Preferably, spike analyzer **190** additionally calculates the rates of change of spike parameters. From prior and current rates of change, future behaviors may be predicted. Rates of change of the parameters include but are not limited to first, second, and third time derivatives. In alternative embodiments, spike analyzer **190** additionally calculates weighted combinations of spike characteristics and performs convolutions of spike waveforms with other spike waveforms, and other preset and varying waveforms. Such operations may be performed, for example, for purposes including but not limited to autocorrelation and digital filtering.

Spike analyzer **190** may receive additional input from accelerometers, such as those described above, including head mounted accelerometer **12**, proximal accelerometer **28**, enclosure mounted accelerometer **36**, and distal accelerometer **33**. Spike analyzer **190** may receive indirect input from these or other accelerometers, as well as from conditioned or processed signals arising therefrom. Such conditioned or processed signals include, for example, the signal transmitted by conditioned accelerometer signal path **80** (FIG. 7).

Spike analyzer **190** may receive additional input from EMG arrays. As noted, such EMG arrays may include, for example, proximal EMG electrode array **48**, enclosure-mounted EMG electrode array **46**, and distal EMG electrode array **47**. Spike analyzer **190** may also receive indirect input from these or other EMG electrode arrays, as well as from conditioned or processed signals arising therefrom. Such conditioned or processed signals include but are not limited to

the signal transmitted by conditioned EMG signal path 78 (FIG. 5). These additional inputs from accelerometers and EMG arrays facilitates the characterization of neuronal firing patterns relative to activity of muscle groups and movement of joints. Such characterization may include, for example, characterization of neuronal spike amplitudes and tuning of neuronal spike frequencies to movement, including but not limited to the signal transmitted by conditioned EMG signal path 78.

The additional input from accelerometers and EMG arrays also facilitates the characterization of neuronal firing patterns relative to activity of muscle groups and movement of joints, including but not limited to characterization of neuronal spike amplitudes and tuning of neuronal spike frequencies to movement, including but not limited to movement velocity and direction. These characterizations may be used to assess functioning of the sensorimotor system, including but not limited to motor response time, and to measure the disease state and response to therapy.

Peripheral nerve electrode (PNE)-based single unit (SU) disease state estimator 194 receives an input representative of the current neuronal activity from spike characterizer 189. PNE-based single unit disease state estimator 194 may receive input representative of at least one of several signals, including desired neuronal activity, actual neuronal activity, and the difference between these quantities. The output from estimator 194 may carry a single or a plurality of signals, consistent with a representation of the disease state by a single or a multitude of state variables, respectively.

Filter 191 has an output connected to an input of spectral energy characterizer 192. Spectral energy characterizer 192 calculates the spectral composition of the signals sensed by the peripheral nerve electrode array 54. Spectral energy characterizer 192 provides outputs to each of spectral energy analyzer 193 and peripheral nerve electrode (PNE)-based multiple unit disease state estimator 232. Output of spectral energy analyzer 193 is connected to an input of PNE-based multiple unit (MU) disease state estimator 232. PNE SU disease state estimator 194 both receives input from and provides output to PNE MU disease state estimator 232.

PNE MU disease state estimator 232 receives as an input signals representative of the current neuronal activity from spectral energy characterizer 192. PNE MU disease state estimator 232 may receive input representative of at least one of several signals, including desired neuronal activity, actual neuronal activity, and the difference between these quantities. The output from PNE MU disease state estimator 232 may carry a single or a plurality of signals, consistent with a representation of the disease state by a single or a multitude of state variables, respectively.

It should be understood that inputs and outputs from each spike detector 188, spike characterizer 189, spike analyzer 190, filter 191, spectral energy characterizer 192, spectral energy analyzer 193, and PNE-based single unit disease state estimator 194, and PNE-based multiple unit disease state estimator 232 may each be comprised of individual signals or a plurality of signals. It should also be understood that each of these units, spike detector 188, spike characterizer 189, spike analyzer 190, filter 191, spectral energy characterizer 192, spectral energy analyzer 193, and PNE-based single unit disease state estimator 194, and PNE MU disease state estimator 232 may each have different parameters and signal processing characteristics for each of the multiple signals processed. Modifications of this processing circuitry may be made to accommodate various combinations of intraneural electrodes, used for single and multiple unit recordings, and

epineural electrodes, used for compound action potential recordings, without departing from the present invention.

FIG. 11 is a schematic diagram of one embodiment of a patient-neural modulator system 999 illustrated in FIG. 2 with feedback control. Patient-neural modulator system 999 primarily includes an observer 228 and a controller 229. An observer is a component of a control system that is known to those of ordinary skill in the art of control systems. An observer is a functional block in which variables, typically represented in software as parameter values or in hardware as electrical signal amplitudes, represent states of the controlled system. Such a component is used in controlling systems in which one or more of the state variables are not directly observable from the sensed signals. An observer essentially includes a simulated version of the controlled system. Its input are the same control law output signals delivered to the controlled system, and its outputs are desired to match those sensed outputs of the controlled system. The difference between the outputs of the observer and the measured outputs of the controlled system, that is, the outputs of a motor control portion of the patient's nervous system in this case, are used to calculate an observer error signal which may then be used to correct the observer error. Since the observer is implemented in software or hardware, all of its signals, including all state variables, are accessible. In a system such as the complex neural circuitry of the patient, one or more of the state variables may not be "observable", that is directly measurable or calculatable based on measured values. In such a case, the state variables present in the observer may be used as "estimates" of the actual state variables and included in the control law. The general use of "observers" for estimation of "unobservable" state variables is known to those skilled in the art of control theory. The use of observers for the estimation of neural state variables, disease states, and responses to therapy is one of the teachings of the present invention.

Observer 228 includes signal conditioning circuit 76 (FIG. 2) and signal processor 71 (FIGS. 2, 10). Signal processor 71, as noted, includes disease state estimator module array (DSEMA) 229 and aggregate disease state estimator 195. Observer 228 receives patient output "y" from patient 227. Patient output "y" is comprised of one or more signals arising from patient 227. In one preferred embodiment patient output "y" includes one or more signals from EMG electrode array 50, EEG electrode array 51, accelerometer array 52, acoustic transducer array 53, peripheral nerve electrode array 54, intracranial recording electrode array 38, and intracranial stimulating electrode array 37. It should be understood that additional signals of the same or different type may also be included.

Control circuit 72 (FIG. 2) includes summator 226 which receives an input from reference module 116, and a control law circuit block 231. Controller 229 includes the control law circuit block 231 and output stage circuit 77. Controller 229 generates a neural modulation waveform "u", described in detail below with reference to FIG. 13. The function and operation of each of these modules is described in detail below.

Reference disease state "r", generated by reference module 116, is a non-inverting input to summator 226, providing disease state and target reference values for the single or plurality of control laws implemented in control law circuit block 231 introduced above with reference to FIG. 2. Reference module 116 may also receive input from control circuit 72, facilitating the dynamic adjustment of reference values. Reference disease state "r" may comprise a single or plurality of signals, each of which may be zero, constant, or time-varying independent of the other. Disease state error "e" is

output from summator **226** and input to controller **229**. Disease state error “e”, which may comprise a single or plurality of signals, represents a difference between a desired disease state (represented by reference disease state “r”) and an actual disease state (represented by disease state estimate “x”). Other methods of calculating disease state estimate “x”, including but not limited to linear or nonlinear combinations of reference disease state “r” and disease state estimate “x”, may be employed without departing from the present invention. Controller **229** is comprised of control law circuit block **231** and output stage circuit **77**.

Disease state error “e” is input to control law circuit block **231** which generates a control circuit output “uc.” Control law circuit block **231** is connected to an input of output stage circuit **77**. The output of the controller **229**, which is generated by the output stage circuit **77**, “u”, is delivered to patient **227** in the form of neural modulation waveforms, described in detail below with reference to FIG. **13**.

Patient output “y” is input to signal conditioning circuit **76**, the output of which is connected to the input of DSEMA **229**. The output of DSEMA **229** is provided to an aggregate disease state estimator **195**, the output of which is the disease state estimate x. Disease state estimate x, which may be comprised of a single or plurality of signals, is an inverting input to summator **226**.

Control law circuit block **231** receives disease state estimate x as an additional input, for use in nonlinear, adaptive and other control laws. Reference module **116** receives input from DSEMA **229** and aggregate disease state estimator **195** for use in dynamically determining reference disease state r. Other modifications, including substitutions, additions, and deletions, may be made to the control loop without departing from the present invention.

Control law circuit block **231** has an autocalibration mode in which multivariable sweeps through stimulation parameters and stimulating electrode configurations are performed to automate and expedite parameter and configuration optimization. This autocalibration feature enables rapid optimization of treatment, eliminating months of iterations of trial and error in optimizing stimulation parameters and electrode configuration necessitated by the prior technique of constant parameter stimulation. Additionally, this autocalibration feature permits real-time adjustment and optimization of stimulation parameters and electrode configuration. This is particularly useful to overcome increases in electrode impedance which result from the body’s normal response to implanted foreign bodies in which a fibrotic capsule is commonly formed around the electrodes. Effects of shifts in electrode position relative to a target structures may be minimized by said autocalibration feature. Detection of changes in electrode impedance and position are facilitated by autocalibration feature. The autocalibration feature facilitates detection of changes in electrode impedance and position. Notification of patient and health care provider allows proactive action, including automated or manual adjustment of treatment parameters and advance knowledge of impending electrode replacement needs.

FIG. **12** is a schematic diagram of control circuit **72**. As noted, control circuit **72** comprises control laws circuit block **231** and summator **226**. Disease state error “e” is input to gain stages of control laws, including but not limited to at least one of proportional gain **197**, differential gain **198**, integral gain **199**, nonlinear gain **200**, adaptive gain **201**, sliding gain **202**, and model reference gain **203**.

An output of each of these gain stages is connected to what is referred to herein as control law stages. In the illustrative embodiment, control law stages includes proportional con-

troller **230**, differential controller **204**, integral controller **205**, nonlinear controller **206**, adaptive controller **207**, sliding controller **208**, and model reference controller **209**, respectively.

Outputs of these control law stages are connected to weight stages, including proportional controller weight **210**, differential controller weight **211**, integral controller weight **212**, nonlinear controller weight **213**, adaptive controller weight **214**, sliding controller weight **215**, and model reference controller weight **216**. Outputs of the weight stages are noninverting inputs to summator **217**, the output of which is control circuit output “uc”. The weight stages may be any combination of at least one of constant, time varying, and nonlinear without departing from the present invention.

Disease state estimate x is input to nonlinear controller **206**, adaptive controller **207**, sliding controller **208**, and model reference controller **209**. The control laws depicted are representative of one possible implementation; numerous variations, including substitutions, additions, and deletions, may be made without departing from the present invention.

The present invention optimizes the efficiency of energy used in the treatment given to the patient by minimizing to a satisfactory level the stimulation intensity to provide the level of treatment magnitude necessary to control disease symptoms without extending additional energy delivering unnecessary overtreatment. In the definition of the control law, a command input or reference input (denoted as r in FIGS. **11** and **12**) specifies the target disease state. In the preferred embodiment, r specifies the target amplitude of tremor. The control law generates an electrical stimulation magnitude just sufficient to reduce the patient’s tremor to the target value. With this apparatus and method, the precise amount of electrical energy required is delivered, and overstimulation is avoided. In present stimulation systems, a constant level of stimulation is delivered, resulting in either of two undesirable scenarios when disease state and symptoms fluctuate: (1) undertreatment, i.e. tremor amplitude exceeds desirable level or (2) overtreatment or excess stimulation, in which more electrical energy is delivered than is actually needed. In the overtreatment case, battery life is unnecessarily reduced. The energy delivered to the tissue in the form of a stimulation signal represents a substantial portion of the energy consumed by the implanted device; minimization of this energy substantially extends battery life, with a consequent extension of time in between reoperations to replace expended batteries.

FIG. **13** is a schematic diagram of electrical stimulation waveforms for neural modulation. The illustrated ideal stimulus waveform is a charge balanced biphasic current controlled electrical pulse train. Two cycles of this waveform are depicted, each of which is made of a smaller cathodic phase followed, after a short delay, by a larger anodic phase. In one preferred embodiment, a current controlled stimulus is delivered; and the “Stimulus Amplitude” represents stimulation current. A voltage controlled or other stimulus may be used without departing from the present invention. Similarly, other waveforms, including an anodic phase preceding a cathodic phase, a monophasic pulse, a triphasic pulse, or the waveform may be used without departing from the present invention.

The amplitude of the first phase, depicted here as cathodic, is given by pulse amplitude **1** PA1; the amplitude of the second phase, depicted here as anodic, is given by pulse amplitude **2** PA2. The durations of the first and second phases are pulse width **1** PW1 and pulse width **1** PW2, respectively. Phase **1** and phase **2** are separated by a brief delay d. Waveforms repeat with a stimulation period T, defining the stimulation frequency as $f=1/T$.

The area under the curve for each phase represents the charge Q transferred, and in the preferred embodiment, these quantities are equal and opposite for the cathodic (Q1) and anodic (Q2) pulses, i.e. $Q=Q1=Q2$. For rectangular pulses, the charge transferred per pulse is given by $Q1=PA1*PW1$ and $Q2=PA2*PW2$. The charge balancing constraint given by $-Q1=Q2$ imposes the relation $PA1*PW1=-PA2*PW2$. Departure from the charge balancing constraint, as is desired for optimal function of certain electrode materials, is included in the present invention.

The stimulus amplitudes PA1 and PA2, durations PW1 and PW2, frequency f, or a combination thereof may be varied to modulate the intensity of the said stimulus. A series of stimulus waveforms may be delivered as a burst, in which case the number of stimuli per burst, the frequency of waveforms within the said burst, the frequency at which the bursts are repeated, or a combination thereof may additionally be varied to modulate the stimulus intensity.

Typical values for stimulation parameters include $f=100$ -300 Hz, PA1 and PA2 range from 10 microamps to 10 milliamps, PW1 and PW2 range from 50 microseconds to 100 milliseconds. These values are representative, and departure from these ranges is included in the apparatus and method of the present invention.

FIG. 14 is a schematic diagram of one example of the recorded waveforms. This represents an individual action potential from a single cell recorded from intracranial microelectrodes. Aggregates of multiple such waveforms are recorded from larger intracranial electrodes. The action potentials may be characterized according to a set of parameters including but not limited to time to valley 1 TV1, time to peak 1 TP1, time to valley 2 TV2, amplitude of valley 1 AV1, amplitude of peak 1 AP1, amplitude of valley 2 AV2, and algebraic combinations and polarity reversals thereof.

When recording activity from more than one cell, said characterization facilitates discrimination of waveforms by individual recorded cell. The discrimination allows activity of a plurality of cells to be individually followed over time. The parameterization may be performed separately on signals recorded from different electrodes. Alternatively, said parameterization may be performed on signals pooled from multiple electrodes.

Following is a description of a general form for representing disease state.

Disease State DS is a vector of individual disease states, including intrinsic disease states DSI and extrinsic disease states DSE:

$$DS=[DS_I, DS_E]$$

Intrinsic disease states and extrinsic disease states are, themselves vectors of individual disease states:

$$DS_I=[DS_{I1}, DS_{I2}, DS_{I3}, \dots, DS_{IN}]$$

$$DS_E=[DS_{E1}, DS_{E2}, DS_{E3}, \dots, DS_{EM}]$$

Intrinsic Disease States include those disease states which characterize the state of disease at a given point in time. Extrinsic Disease States include variations of intrinsic disease states, including but not limited to cyclical variations in Intrinsic Disease States, variations in Intrinsic Disease States which occur in response to external events, and variations in Intrinsic Disease States which occur in response to levels of and changes in levels of electrical stimulation. Said external events include but are not limited to pharmacologic dosing, consumption of meals, awakening, falling asleep, transitioning from Parkinsonian "on" state to Parkinsonian "off" state, transitioning from Parkinsonian "off" state to Parkinsonian "on" state.

Each of Intrinsic Disease States and Extrinsic Disease States include but are not limited to those defined herein;

additional disease states and definitions thereof may be added without departing from the present invention.

The first intrinsic disease state DS_{I1} represents the level of resting tremor

$$DS_{I1}=RT_N$$

Where Normalized Resting Tremor Magnitude $RT_{sub.N}$ is given by:

$$RT_N = T_{A,3-5} * W_{TA,3-5} + T_{E,3-5} * W_{TE,3-5} + T_{P,3-5} * W_{PE,3-5} + T_{C,3-5} * W_{TC,3-5} + T_{N,3-5} * W_{TN,3-5} + T_{S,3-5} * W_{TS,3-5} + T_{E,3-5} * W_{TE,3-5}$$

Where the factors from which the Resting Tremor Magnitude RT_N is determined, representing estimates of the magnitude of 3-5 Hertz movement of selected body segments, including but not limited to limbs, torso, and head are:

$T_{A,3-5}$ =Tremor level determined by acceleration monitoring

$W_{TA,3-5}$ =Weighting factor for tremor $T_{A,3-5}$

$T_{E,3-5}$ =Tremor level determined by electromyographic (EMG) monitoring

$W_{TE,3-5}$ =Weighting factor for tremor $T_{E,3-5}$

$T_{P,3-5}$ =Tremor level determined by peripheral nerve electrode monitoring

$W_{TP,3-5}$ =Weighting factor for tremor $T_{P,3-5}$

$T_{C,3-5}$ =Tremor level determined by cortical electrode monitoring

$W_{TC,3-5}$ =Weighting factor for tremor $T_{C,3-5}$

$T_{N,3-5}$ =Tremor level determined by neural monitoring, including subcortical nuclei, white matter tracts, and spinal cord neurons

$W_{TN,3-5}$ =Weighting factor for tremor $T_{N,3-5}$

$T_{S,3-5}$ =Tremor level determined by acoustic sensor monitoring

$W_{TS,3-5}$ =Weighting factor for tremor $T_{S,3-5}$

Weighting factors are adjusted after implantation to achieve normalization of RT_N and to allow for selective weighting of tremor levels as determined from signals arising from various sensors, including but not limited to those listed.

These calculations may be implemented in analog hardware, digital hardware, software, or other form. In the preferred embodiment, values are implemented as 16-bit variables; ranges for said weighting factors and tremor levels are 0 to 65535. These ranges may be changed or implemented in analog form without departing from the present invention.

The second intrinsic disease state DS_{I2} represents the level of dyskinesia:

$$DS_{I2}=D_N$$

Where Normalized Dyskinesia Magnitude D_N is given by:

$$D_N = D_A * W_{DA} + T_E * W_{TE} + T_P * W_{PE} + T_C * W_{TC} + T_N * W_{TN} + T_S * W_{TS} + T_E * W_{TE}$$

Where

$D_{A,3-5}$ =Dyskinesia level determined by acceleration monitoring

$W_{DA,3-5}$ =Weighting factor for Dyskinesia $D_{A,3-5}$

$D_{E,3-5}$ =Dyskinesia level determined by electromyographic (EMG) monitoring

$W_{DE,3-5}$ =Weighting factor for Dyskinesia $D_{E,3-5}$

$D_{P,3-5}$ =Dyskinesia level determined by peripheral nerve electrode monitoring

$W_{DP,3-5}$ =Weighting factor for Dyskinesia $D_{P,3-5}$

$D_{C,3-5}$ =Dyskinesia level determined by cortical electrode monitoring

$W_{DC,3-5}$ =Weighting factor for Dyskinesia $D_{C,3-5}$

$D_{N,3-5}$ =Dyskinesia level determined by neural monitoring, including subcortical nuclei, white matter tracts, and spinal cord neurons

$W_{DN,3-5}$ =Weighting factor for Dyskinesia $D_{N,3-5}$

$D_{S,3-5}$ =Dyskinesia level determined by acoustic sensor monitoring

$W_{DS,3-5}$ =Weighting factor for Dyskinesia $D_{S,3-5}$

The third intrinsic disease state DS_{13} represents the level of rigidity.

$$DS_{13}=R_N$$

Where Normalized Rigidity Magnitude R_N is given by:

$$R_N = \frac{R_A * W_{RA} + R_E * W_{RE} + R_P * W_{RP} + R_C * W_{RC} + R_N * W_{RN} + R_S * W_{RS} + R_E * W_{RE}}{R_S * W_{RS} + R_E * W_{RE}}$$

Where

$R_{A,3-5}$ =Rigidity level determined by acceleration monitoring

$W_{RA,3-5}$ =Weighting factor for Rigidity $R_{A,3-5}$

$R_{E,3-5}$ =Rigidity level determined by electromyographic (EMG) monitoring

$W_{RE,3-5}$ =Weighting factor for Rigidity $R_{E,3-5}$

$R_{P,3-5}$ =Rigidity level determined by peripheral nerve electrode monitoring

$W_{RP,3-5}$ =Weighting factor for Rigidity $R_{P,3-5}$

$R_{C,3-5}$ =Rigidity level determined by cortical electrode monitoring

$W_{RC,3-5}$ =Weighting factor for Rigidity $R_{C,3-5}$

$R_{N,3-5}$ =Rigidity level determined by neural monitoring, including subcortical nuclei, white matter tracts, and spinal cord neurons

$W_{RN,3-5}$ =Weighting factor for Rigidity $R_{N,3-5}$

$R_{S,3-5}$ =Rigidity level determined by acoustic sensor monitoring

$W_{RS,3-5}$ =Weighting factor for Rigidity $R_{S,3-5}$

The fourth intrinsic disease state DS_{14} represents the level of bradykinesia.

$$DS_{14}=B_N$$

Where Normalized Bradykinesia Magnitude B_N is given by:

$$B_N = \frac{B_A * W_{BA} + B_E * W_{BE} + B_P * W_{PE} + B_C * W_{BC} + B_N * W_{BN} + B_S * W_{BS} + B_E * W_{BE}}{B_S * W_{BS} + B_E * W_{BE}}$$

Where

R_A =Bradykinesia level determined by acceleration monitoring

W_{RA} =Weighting factor for Bradykinesia R_A

R_E =Bradykinesia level determined by electromyographic (EMG) monitoring

W_{RE} =Weighting factor for Bradykinesia R_E

R_P =Bradykinesia level determined by peripheral nerve electrode monitoring

W_{RP} =Weighting factor for Bradykinesia R_P

R_C =Bradykinesia level determined by cortical electrode monitoring

W_{RC} =Weighting factor for Bradykinesia R_C

R_N =Bradykinesia level determined by neural monitoring, including subcortical nuclei, white matter tracts, and spinal cord neurons

W_{RN} =Weighting factor for Bradykinesia R_N

R_S =Bradykinesia level determined by acoustic sensor monitoring

W_{RS} =Weighting factor for Bradykinesia R_S

The control law drives these disease states toward their reference values, nominally 0, according to a vector of weights, establishing a prioritization.

Side effects and other parameters, such as power consumption and current magnitude, are also quantified and minimized according to a cost function.

One advantage of the present invention is that it provides prediction of future symptomatology, cognitive and neuro-

motor functionality, and treatment magnitude requirements. Such predictions may be based on preset, learned and real-time sensed parameters as well as input from the patient, physician or other person or system. The prediction of future symptomatology is based upon any of several weighted combination of parameters. Based upon prior characterization of the circadian fluctuation in symptomatology (that is, tremor magnitude for deep brain stimulation or level of depression for stimulation of other sites including locus ceruleus), future fluctuations may be predicted. An estimate, or model, of fluctuation may be based upon a combination of preset, learned, and real-time sensed parameters. Preset parameters are derived from clinical studies designed specifically for the purpose of gathering such data, or from estimates extracted from data gleaned from published literature. Real-time sensed parameters are derived from the current states (and changes, i.e. derivatives and other processed signals, thereof) of sensed and processed signals. Learned parameters are based upon the time histories of previously sensed signals. For example, the circadian fluctuation in tremor amplitude may be sensed; a weighted average of this data collected over numerous prior days provides as estimate of the expected tremor amplitude as well as a standard deviation and other statistical parameters to characterize the anticipated tremor amplitude. Similarly, in the presence of closed-loop feedback, the level of stimulation required to reduce or eliminate tremor may be used as an estimate of the "amplitude" or state of the underlying disease.

Another advantage of the present invention is that it performs automated determination of the optimum magnitude of treatment—by sensing and quantifying the magnitude and frequency of tremor activity in the patient, a quantitative representation of the level or "state" of the disease is determined. The disease state is monitored as treatment parameters are automatically varied, and the local or absolute minimum in disease state is achieved as the optimal set of stimulation parameters is converged upon. The disease state may be represented as a single value or a vector or matrix of values; in the latter two cases, a multivariable optimization algorithm is employed with appropriate weighting factors.

Having now described several embodiments of the invention, it should be apparent to those skilled in the art that the foregoing is merely illustrative and not limiting, having been presented by way of example only. For example, all signal paths may transmit a single or plurality of signals without departing from the present invention. Numerous modifications and other embodiments are within the scope of one of ordinary skill in the art and are contemplated as falling within the scope of the invention as defined by the appended claims.

Current implanted neurostimulators suffer from limited battery life necessitating replacement. The original filing of this invention teaches a closed-loop technique which allows lower average power delivery, providing extended batter life. A further improvement is taught in the present invention. The present invention teaches a novel form of power delivery, in which electromagnetic power is provided externally and radiated transcutaneously through the skin 382 to the implanted circuit.

A multiplicity of electromagnetic coils is used, to provide electromagnetic fields of multiple non-collinear orientations. The purpose of this design is to overcome fluctuations and interruptions in power that otherwise occur when the implanted circuit moves along with the body parts relative to electromagnetic coils.

An additional improvement is taught in the application of time multiplexing of signals delivered by said multiplicity of electromagnetic coils. Multiplexing signals in time from a

multiplicity of coils allows electromagnetic energy to be delivered at a multiplicity of spatial orientations without mutual interference between fields. In one embodiment, three coils are positioned such that they are mutually orthogonal. Regardless of the orientation of the implanted circuit, mutual coupling between the implanted circuit and at least one element of the coil array will facilitate power transmission.

The present invention further includes an embodiment in which the coils are embedded in a flexible cloth assembly, including but not limited to a pillow, bandana, hat, or other accessory or piece of apparel.

Additionally, low profile mounting of the neurological control system 999 is taught, including a design in which the device is implanted entirely adjacent to the head, avoiding the subcutaneous cables plaguing current designs with high failure and fracture rates. Additionally, a low-profile design is taught in which the neurological control system is implanted beneath the skin and recessed in the caldarium.

FIG. 16 and FIG. 18 show two coil and three coil embodiments, respectively, of power delivery unit 413. Different numbers of coils may be used without departing from the present invention. Each of electromagnetic coil 372, 373, 374 radiates electromagnetic power, represented by magnetic flux 375, 376, 377, which radiates through the skin 382 and is then converted to electrical energy by power conversion unit 378. Power is then transmitted from power conversion unit 378 via power cable 379 to stimulating and recording circuit 26, a component of the neurological control system 999.

A single or multiplicity of coil holder 380 holds electromagnetic coil 372, 373, 374 in place. Coil holder 380 may be placed atop bedding 381, which may include a pillow, blanket, or mattress. Alternatively, coil holder 380 may be placed below bedding 381, which may include a pillow, blanket, mattress, or other element.

FIG. 17 shows this configuration at greater magnification, in which magnetic flux 375 is seen to penetrate skin 382.

FIG. 19 depicts power delivery unit 413 with coil holder 380 shown holding electromagnetic coil 372, 373, 374 in place, as seen from above. In this design, all electromagnetic coils 372, 373, 374 are contained within coil holder 380. For illustrative purposes, coil holder 380 is shown in this embodiment as a convex shape. Other shapes may be used without departing from the present invention.

FIG. 20 shows coil holder 380 with electromagnetic coil 372, 373, 374 in place, as seen from above. In this design, all electromagnetic coils 372, 373, 374 are contained within coil holder 380. For illustrative purposes, coil holder 380 is shown in this embodiment as a multi-lobed shape, with electromagnetic coil 372, 373, 374 contained within coil pockets 383, 384, 385, respectively. Other shapes may be used without departing from the present invention.

Electromagnetic coils 372, 373, 374 may be permanently affixed within coil holder 380. Alternatively, electromagnetic coils 372, 373, 374 may be removable from coil holder 380 to facilitate cleaning or laundering of coil holder 380.

Further, use of electromagnetic fields, as direct modulators of neural activity is taught in the present invention. [This also references provisional application filed Jul. 29, 2002.]

As shown in FIG. 21, electromagnetic coils 372, 373, 374 may be positioned by coil holder 380 to lie in close proximity to the head 398 of a patient. Magnetic flux 375, 376, 377, which radiate through the skin 382 penetrate into underlying brain 251, or other neural tissue 354, and generate electrical currents. Said electrical currents serve to stimulate or inhibit neural activity. Other neural tissue includes but is not limited to brain 251, brainstem, spinal cord, peripheral nerves, cranial nerves, neurosensory organs, and other structures.

FIG. 22 shows multiple coil embodiments. Different numbers of coils may be used without departing from the present invention. FIG. 22 shows a distributed arrangement of coils, to facilitate strong signal coupling with a minimum of tissue heating or sensory stimulation. Brain 251 is shown underlying skin 382 in head 398. Temporal cortex 255 and thalamus 270 are shown.

Headband coil holder 386 is in mechanical connection with each of electromagnetic coils 387, 388, 389, 390, 391, 392, 393, which radiate electromagnetic power through the skin 382. This is then converted to electrical energy by power conversion unit 378, which is implanted in any of several locations, shown in FIG. 17 and FIG. 18, omitted here for diagram clarity. As shown in FIG. 17, power is then transmitted from power conversion unit 378 via power cable 379 to stimulating and recording circuit 26, a component of the neurological control system 999.

Power source 397 provides power to power modulator 396, which modulates power on a carrier frequency and is electrically connected via electromagnetic coil cable 394 to electromagnetic coils 387, 388, 389, 390 which radiate power through skin 382. Power modulator 396 is also electrically connected via electromagnetic coil cable 395 to electromagnetic coils 391, 392, 393 which radiate power through skin 382. Electromagnetic coils 387, 388, 389, 390, 391, 392, 393 can be arranged in a different geometrical or anatomical configuration, with the same, a higher or a lower number of electromagnetic coils, without departing from the present invention. Electromagnetic coils 387, 388, 389, 390, 391, 392, 393 may alternatively be affixed to a different form of apparel, clothing, or fixture without departing from the present invention. Such embodiments include a hat, cap, bandana, or other device.

FIG. 23 shows the paracranial design, with implanted components in close proximity to the head 398. This design eliminates the pervasive problem of moving parts, inherent in all current technology devices. All cabling is in close approximation to the caldarium, a rigid structure; therefore all implanted parts remain stationary. Since no implanted parts are subjected to movement, they are at greatly reduced risk for mechanical fatigue, the single highest failure mode for current implanted neuromodulators. This innovation dramatically reduces the frequency and likelihood for re-operation to repair broken implanted components, a major concern in all patients and particularly the elderly, who are often poor operative candidates, and who are at highest risk for many of the neurological and neurodegenerative diseases amenable to treatment by neuromodulation.

FIG. 23 depicts an embodiment in which the head 398 of the patient lies atop coil holder 380, placing the power conversion unit 378 within at least one of magnetic flux 375, 376, 377. Power conversion unit 378 converts power transmitted as magnetic flux 375, 376, 377 into electrical energy, which is transmitted by power cable 379 to stimulating and recording circuit 26, a component of the neurological control system 999. Stimulating and recording circuit 26 generates neuromodulation signal that is transmitted via connecting cable 8 to intracranial stimulating electrode array 37, depicted in more detail in FIG. 1, which deliver neuromodulation signal to brain 251, or other neural tissue 354. Connecting cable 8 furthermore provides bidirectional electrical connection between intracranial electrodes 246 and stimulating and recording circuit 26, a component of neurological control system 999.

Catheter anchor **29** is in mechanical communication with intracranial catheter **7**, and secures it to caldarium **9**. Intracranial electrodes **246** are mounted on or incorporated into intracranial catheter **7**.

FIG. **24** shows power conversion unit **378** includes electromagnetic coupling element **399**, such as a wire coil, circuit board tracing coil, or equivalent implementation. In one embodiment, power conversion unit **378** also includes power conversion circuit **400**, which converts electrical signal induced by any of magnetic flux **375**, **376**, **377**, on electromagnetic coupling element **399**, into at least one of electrical power and electrical signal, which are transmitted via power cable **379** to stimulating and recording circuit **26**, which generates neuromodulation signal **412**, which is transmitted on connecting cable **8** to intracranial catheter **7**.

Power conversion circuit **400** includes rectifier **401**, energy storage element **402**, regulator **403**, filter **404**, demodulator **405**, and amplifier **406**. Electromagnetic coupling element **399** is electrically connected via electromagnetic coupling element cable **407** to power conversion circuit **400**.

Either of magnetic flux **375**, **376**, **377** passing through electromagnetic coupling element **399** induces induced current **408**, which is transmitted via electromagnetic coupling element cable **407** to power conversion circuit **400**. Induced current **408** is rectified by rectifier **401** to a direct current form and is stored or lowpass filtered by energy storage element **402** and regulated by regulator **403** and then transmitted as regulated power **409** by power cable **379** to stimulating and recording circuit **26**, a component of neurological control system **999**.

FIG. **25** shows the communication circuit. This circuit also provides bi-directional communication with external devices, including power delivery unit **413** as well as with patient interface module **55** and supervisory module **56**, shown in FIG. **1**.

Either of magnetic flux **375**, **376**, **377** passing through electromagnetic coupling element **399** induces induced current **408**, which is transmitted via electromagnetic coupling element cable **407** to power conversion circuit **400**. Induced current **408** is filtered by filter **404**, then demodulated by **405**, then amplified by amplifier **406**, then transmitted as incoming data stream **410** via power cable **379** to stimulating and recording circuit **26**. At least one of digital to analog or analog to digital conversion may also be performed without departing from the present invention.

Outgoing data stream **411** is transmitted from stimulating and recording circuit **26** via power cable **379** to power conversion circuit **400** where it is modulated by modulator **414** and then amplified by amplifier **415** and transmitted as inducing current **416** along electromagnetic coupling element cable **407** to electromagnetic coupling element **399** where it generated magnetic flux that is detected and decoded by external devices, including power delivery unit **413** as well as with patient interface module **55** and supervisory module **56**, shown in FIG. **1**. At least one of digital to analog or analog to digital conversion may also be performed without departing from the present invention.

FIG. **26** depicts the overall system, with power conversion unit **378** connected via power cable **379** to power management unit **417**, a component of stimulating recording and power circuit **419**. Power management unit **417** is in electrical connection with energy storage unit **418** and in electrical connection with stimulating and recording circuit **26**.

In one preferred embodiment, energy storage unit **418** is implemented as a capacitor. Power management unit **417** converts regulated power **409** into charge, which is stored in energy storage unit **418**.

In another preferred embodiment, energy storage unit **418** is implemented as a rechargeable battery. Power management unit **417** converts regulated power **409** into current, which recharges energy storage unit **418**.

When regulated power **409** is insufficient to power stimulating and recording circuit **26**, power management unit **417** withdraws energy from energy storage unit **418** and delivers energy to stimulating and recording circuit **26**.

An embodiment of this invention further includes novel geometrical features to enhance form and function of this neuromodulation system as well as any neural stimulation system. One invention taught is the refinement of the implanted case such that one side is concave. An additional invention taught is the refinement of the implanted case such that one side is convex. A further extension of this invention includes the use of at least one concave side and at least one convex side. Advantages of this design include the close positioning of the casing against the intact outer table of the skull of the patient.

FIG. **27** shows stimulating and recording unit **43** enclosed within system enclosure **434**, implanted beneath the scalp **10**, overlying the caldarium **9**. System enclosure **434** is designed with an enclosure inner surface **427** and enclosure outer surface **426**. In one preferred embodiment, enclosure inner surface **427** and enclosure outer surface **426** are concave and convex, respectively.

By designing enclosure inner surface **427** to be concave, system enclosure **434** may be placed tightly and securely against caldarium **9**. This represents a substantial innovation over flat machined enclosures, the designs currently implanted elsewhere and that are based upon established pacemaker technology. This innovation reduces or eliminates movement of the implant that would otherwise occur with existing devices.

By designing enclosure outer surface **426** to be convex, system enclosure **434** may be placed against calvarium **9** with a substantially improved cosmetic result and less displacement of scalp **10** or other skin **382**. This represents a substantial innovation over flat machined enclosures, the designs currently implanted elsewhere and that are based upon established pacemaker technology. This innovation reduces or eliminates movement of the implant that would otherwise occur with existing devices.

Furthermore, each of enclosure inner surface **427** and enclosure outer surface **426** serve to reduce the profile of system enclosure **434** and to improve the cosmetic appearance of the overlying skin **382** or scalp **10**.

In this embodiment, system enclosure **434** overlies the intact pericranium **420**. This facilitates attachment of System enclosure **434** to the pericranium **420** by means of mechanical attachment **424**, providing mechanical attachment between mechanical attachment mount **425** and at least one of pericranium **420**, skin **382**, scalp **10**, or other tissue of the patient. Mechanical attachment **424**, may be implemented as sutures, wires, clips, or other attachment means. In this embodiment, the caldarium outer table **421**, caldarium marrow layer **422**, and caldarium inner table **423** remain intact.

Alternatively, System enclosure **434** is implanted beneath the scalp **10** and beneath the pericranium **420**, directly overlying the caldarium **9**. In this embodiment, the pericranium **420** serves to secure System enclosure **434** in place against the caldarium **9**.

FIG. **28** depicts system enclosure **434** secured to the calvarium **9** using screws **429**, **431** which are shown inserted through screw mounts **428**, **430**, respectively, into calvarium **9**. Additional or fewer screws and screw mounts may be used without departing from the present invention. Screws **429**,

431 may penetrate in depth to the level of the calvarium outer table 421, calvarium marrow layer 422, or calvarium inner table 423.

FIG. 29 depicts a lower profile design in which the system enclosure 434 is partially recessed into the calvarium 9. Protruding component 432 lies above the outer surface of calvarium 9, and recessed component 433 lies below the outer surface of calvarium 9. Recessed component 433 is shown occupying volume previously occupied by calvarium outer table 421. Recessed component 433 may also occupy volume previously occupied by calvarium marrow layer without departing from the present invention.

FIG. 30 depicts a lower profile design in which the system enclosure 434 is partially recessed into the calvarium 9. Protruding component 432 lies above the outer surface of calvarium 9, and recessed component 433 lies below the outer surface of calvarium 9. Recessed component 433 is shown occupying volume previously occupied by calvarium outer table 421, calvarium marrow 422, and calvarium inner table 423. System enclosure 434 is shown secured to the calvarium outer table 421 portion of calvarium 9 using screws 429, 431 which are shown inserted through screw mounts 428, 430, respectively. Additional or fewer screws and screw mounts may be used without departing from the present invention. Screws 429, 431 may penetrate in depth to the level of the calvarium outer table 421, calvarium marrow layer 422, or calvarium inner table 423.

FIG. 31 depicts a lower profile design in which the system enclosure 434 is fully recessed into the calvarium 9. Recessed component 433 lies below the outer surface of calvarium 9. Recessed component 433 is shown occupying volume previously occupied by calvarium outer table 421, calvarium marrow 422, and calvarium inner table 423. System enclosure 434 is shown secured to the calvarium outer table 421 portion of calvarium 9 using screws 429, 431 which are shown inserted through screw mounts 428, 430, respectively. Additional or fewer screws and screw mounts may be used without departing from the present invention. Screws 429, 431 may penetrate in depth to the level of the calvarium outer table 421, calvarium marrow layer 422, or calvarium inner table 423.

FIG. 32 depicts a lower profile design in which the system enclosure 434 is fully recessed into the calvarium 9. Recessed component 433 lies below the outer surface of calvarium 9. Recessed component 433 is shown occupying volume previously occupied by calvarium outer table 421, calvarium marrow 422, and calvarium inner table 423. In one embodiment shown, enclosure inner surface 427 is shown extending below the level of calvarium inner table 423. This design facilitates close proximity of stimulating and recording unit 43 to underlying neural tissue 354 and brain 251, valuable in sensing neural activity and in delivering an output neuromodulation signal (NMS) 998 via intracranial stimulating electrode array 37, shown in FIG. 1 and FIG. 2.

System enclosure 434 is shown secured to the calvarium inner table 423 portion of calvarium 9 using screws 429, 431 which are shown inserted through screw mounts 428, 430, respectively. Additional or fewer screws and screw mounts may be used without departing from the present invention. Alternate means of mechanically attaching system enclosure 434 to calvarium 9 may be employed without departing from the present invention.

FIG. 33 depicts the neurological control system 999 employing a lower profile design in which the system enclosure 434 is recessed into the calvarium 9. Power delivery unit 413, via generated magnetic flux, transmits power to power conversion unit 378, which transmits power via power cable 379 to stimulating and recording circuit 26, part of stimulat-

ing and recording unit 43, contained within system enclosure 434. Stimulating and recording circuit 26 is electrically connected via connecting cable 8 to intracranial catheter 7 which provides electrical connection to intracranial stimulating electrode array 37 and intracranial recording electrode array 38. Intracranial catheter is mechanically secured via catheter anchor 29 to calvarium 9.

FIG. 34 depicts the neurological control system 999 employing a lower profile and more compact design in which the system enclosure 434 is recessed into the calvarium 9. Power delivery unit 413, via generated magnetic flux, transmits power to power conversion unit 378, which transmits power via power cable 379 to stimulating and recording circuit 26, part of stimulating and recording unit 43, contained within system enclosure 434.

Stimulating and recording circuit 26 is in close proximity to intracranial catheter 7, this includes but is not limited to a preferred embodiment in which stimulating and recording circuit 26 is recessed into calvarium at the site surrounding the entry point at which intracranial catheter 7 passes from outside the calvarium to inside the calvarium. Stimulating and recording circuit 26 is electrically connected directly or via connecting cable 8 to intracranial catheter 7 which provides electrical connection to intracranial stimulating electrode array 37 and intracranial recording electrode array 38. Intracranial catheter is mechanically secured via catheter anchor 29 to calvarium 9.

An advantage of the closed-loop system includes the natural variability of the output signal. The nervous system exhibits a natural tendency to attenuate responses to continuous stimuli, such as continuous background noises. A natural example includes imperceptibility of stationary visual images on frog retina. A related example is electrical stimulation of peripheral nerves, such as stimulation of the tibial or peroneal nerves for use in eliciting the flexion withdrawal reflex to facilitate gait restoration in paraplegics. With time, these reflexes attenuate or habituate, reducing their effectiveness.

The time-varying nature of a closed-loop signal, responding to environmental and system fluctuations and noise, the natural neural process of habituation to constant signals will be reduced.

Furthermore, by specifically and intentionally altering the output signal, such as intermittently reducing the amplitude of the stimulus, and allowing the closed-loop system to compensate in between these intentional amplitude restrictions, further reduction and prevention of habituation is achieved.

As shown previously, ventricular electrode catheter 1 is shown positioned within the lateral ventricle 2, with ventricular electrode 17 in contact with the CSF fluid 18 in lateral ventricle 2. Cortical electrode 3 is in contact with the CSF fluid overlying the cerebral cortex 19. Cortical electrode 3 may be placed over any portion of the cerebral or cerebellar cortex without departing from the present invention. Hippocampal electrode 4 is shown underlying the hippocampal regions of the cerebrum.

The parenchyma of the nervous system is used as a dielectric to establish a voltage potential gradient between any of the CSF reservoirs, including the lateral ventricles, third ventricle, cerebral aqueduct, fourth ventricle, foramen of Lushke, foramen of magendie, basal cisterns, CSF overlying the cerebellar hemispheres, CSF overlying the cerebral hemispheres, central spinal canal, CSF overlying the spinal cord, and CSF overlying the spinal nerves and roots.

Patients currently implanted with neuromodulation devices must undergo repeat surgery to replace implanted pulse generators every 3-5 years to replace units when batteries fail. Tradeoffs between acceptable implanted device size

and limited energy density with current battery technology are responsible for this pervasive problem. Many of these patients are older, and the risks of general anesthesia and surgery are not insignificant. Subjecting these patients to these large risks is a pervasive problem in the field of neuromodulation. The present invention overcomes these limitations with a clever design employing a novel external radio-frequency power delivery system.

The present embodiment of the invention includes an external coil use of Limited energy density because technology is limited.

The present embodiment of this invention teaches a device, method of implantation, and surgical tools for the rapid implantation of a neuromodulation system. Current devices suffer from the need to implant a pulse generator in the subclavicular pocket or other site remote from the site of electrode implantation. Because of this, subcutaneous cables must be implanted to connect the implanted circuit to the implanted stimulating electrode. The present invention teaches a device and method for implanting the circuit in close proximity to the site of stimulation. The design taught herein obviates the need for any subcutaneous cables. Additionally, it teaches a compact design that allows placement of the implanted circuit and the implantation of the intracranial electrode catheter through a single hole. This is a substantial improvement, facilitating much more rapid implantation and eliminating the need for subcutaneous cable implantation. As a result, surgical procedures are much faster and may be performed under local anesthesia, no longer requiring general anesthesia. This substantially increases the market size, allowing implantation in older and frail patients who might otherwise not benefit from neuromodulation technology because of their being poor surgical candidates, due to their inability to safely tolerate general anesthesia and long surgical procedures.

FIGS. 35 and 36 depict the neurological control system 999 employing a lower profile and more compact design in which the system enclosure 434 is recessed into the calvarium 9, shown in the anteroposterior and lateral projections, respectively. Power delivery unit 413, is included within system enclosure 434; however, power delivery unit 413 may also be external to system enclosure 434 without departing from the present invention.

Stimulating and recording circuit 26 is in close proximity to intracranial catheter 7, this includes but is not limited to a preferred embodiment in which stimulating and recording circuit 26 is recessed into calvarium at the site surrounding the entry point at which intracranial catheter 7 passes from outside the calvarium to inside the calvarium. Stimulating and recording circuit 26 is electrically connected to intracranial catheter 7 which provides electrical connection to intracranial stimulating electrode array 37 and intracranial recording electrode array 38. Intracranial catheter is mechanically secured to system enclosure 434. System enclosure 434 is secured to calvarium 9 via mechanical attachment 424, which is attached to system enclosure 434 via machine screw 439 or equivalent means and to calvarium 9 via bone screw 438 or equivalent means. Catheter recess 441 provides space for establishment of electrical and mechanical connection of stimulating and recording circuit 26 to intracranial catheter 7. Catheter mount socket 437 and catheter mount ball 436.

FIG. 37 shows an expanded view of the neurological control system 999 also shown in FIGS. 35 and 36.

FIGS. 38 and 39 depict a Calvarium drill bit is used to create a circular hole in the calvarium. The present invention teaches a major advance in the expeditiousness of the implantation procedure for creating a craniotomy, with particular

relevance to the implantation of a neuromodulation device recessed in the calvarium. In a single pass, a drill bit creates a hole of a diameter similar to that of the implanted device. The outer diameter portion is created by calvarium bit outer diameter segment 443. Attached to and deep to calvarium bit outer diameter segment 443 is the calvarium bit inner diameter segment 442, which creates a hole in the calvarium of a smaller diameter. The resulting geometry, as seen in FIG. 38 and FIG. 39, produces bone ledge 447. Bone ledge 447 serves to provide mechanical support to system enclosure 434, preventing system enclosure 434 from becoming displaced and impinging upon brain 251.

Intracranial catheter 7 is secured to system enclosure 434 by means of catheter mount ball 436. A compressible material is either adjacent to or comprises catheter mount ball 436.

FIG. 40 depicts the path of the intracranial catheter 7 and its connection to the electrical elements of stimulating and recording circuit 26, which is contained within system enclosure 434 and in close proximity to intracranial catheter 7. Intracranial Catheter Proximal End 451 is largely contained within or at least has a component that occupies a portion of the catheter recess 441. Intracranial Catheter Proximal Electrode 452 and Intracranial Catheter Proximal Electrode 453 are shown. Additional electrodes or alternate connection means may be employed without departing from the present invention.

In one embodiment, set screws are used to electrically connect electrodes on intracranial catheter 7 to contacts mounted on system enclosure 434. Electrode contacts and set screw are recessed in catheter recess 441 in system enclosure 434.

System enclosure is constructed from a castable material, such as potted epoxy, methylmethacrylate, or other plastic, silicone, or polymer. RF electrodes are included within the system enclosure. A titanium or stainless steel shield may be included into center of the system enclosure 434. The outer surface of system enclosure 434 may have a convex shape, to better approximate the shape of the calvarium removed and replaced by system enclosure 434, and the convex shape of outer surface of system enclosure 434 minimized stress concentrations on the overlying scalp 10.

Disease state is estimated using a measure or estimate of chaos of neural activity. Such measures include Lyapunov functions and other measures of chaos or system synchronicity or correlation.

Bottom surface of system enclosure 434 may include electrodes used for at least one of recording, stimulation, ground, or reference electrode functions.

An array of at least one EEG electrode is used for recording electroencephalographic signals. Said EEG electrode may be placed in at least one of a subgaleal location, subcutaneous location, epidural location, subdural location, intracerebral location, or other location enabling EEG electrode to sense neural activity.

An advancement in intracranial catheter design is taught in the present invention. Intracranial catheter 7 includes a microelectrode channel 448. Microelectrode 449 is inserted through microelectrode channel 448, with microelectrode tip 450 protruding beyond tip of intracranial catheter 7. Microelectrode tip 450 is used to record single cell activity to identify neural structures during advancement of intracranial catheter 7 through brain 251. At the completion of the insertion of intracranial catheter 7 through brain 251, microelectrode 449 may be removed from intracranial catheter 7.

This allows microelectrode recording and implantation of intracranial catheter 7 during a single pass, saving substantial time during the implantation procedure.

53

During and Following implantation, closed-loop optimization of electrical field shaping is performed by control circuit 72.

Disease state is characterized by a measure of correlation between neural signals measured from neural tissue. Disease State DS is a vector of individual disease states, including intrinsic disease states DSI and extrinsic disease states DSE:

$$DS=[DS_I, DS_E]$$

Intrinsic disease states and extrinsic disease states are, themselves vectors of individual disease states:

$$DS_I=[DS_{I1}, DS_{I2}, DS_{I3}, \dots, DS_{IN}]$$

$$DS_E=[DS_{E1}, DS_{E2}, DS_{E3}, \dots, DS_{EM}]$$

Intrinsic Disease States include those disease states which characterize the state of disease at a given point in time. Extrinsic Disease States include variations of intrinsic disease states, including but not limited to cyclical variations in Intrinsic Disease States, variations in Intrinsic The fifth intrinsic disease state DS_{I5} represents the level of correlation between neural activity in multiple areas of the nervous system.

$$DS_{I5}=C_N$$

Where Normalized Correlation Magnitude matrix CN is given by:

$$C_N = \begin{bmatrix} C_{1,1} & C_{1,2} & C_{1,3} & \dots & C_{1,M} \\ C_{2,1} & C_{2,2} & C_{2,3} & \dots & C_{2,M} \\ C_{3,1} & C_{3,2} & C_{3,3} & \dots & C_{3,M} \\ \dots & \dots & \dots & \dots & \dots \\ C_{M,1} & C_{M,2} & C_{M,3} & \dots & C_{M,M} \end{bmatrix}$$

Which becomes:

$$C_N = \begin{bmatrix} 1 & C_{1,2} & C_{1,3} & \dots & C_{1,M} \\ C_{2,1} & 1 & C_{2,3} & \dots & C_{2,M} \\ C_{3,1} & C_{3,2} & 1 & \dots & C_{3,M} \\ \dots & \dots & \dots & \dots & \dots \\ C_{M,1} & C_{M,2} & C_{M,3} & \dots & 1 \end{bmatrix}$$

This matrix or a weighted sum of its components represents an additional measure of disease state. This has broad applications in neurological disease quantification and also has particular relevance to measurement of tremor and assessment of seizure activity as well as prediction of likelihood and onset of seizure activity.

$$\begin{aligned} C_N &= C_{1,2} * W_{1,2} + C_{1,3} * W_{1,3} + \dots + C_{1,M} * W_{1,M} + \\ &C_{2,1} * W_{2,1} + C_{2,3} * W_{2,3} + \dots + C_{2,M} * W_{2,M} + \\ &C_{3,1} * W_{3,1} + C_{3,2} * W_{3,2} + \dots + C_{3,M} * W_{3,M} + \\ &\dots \\ &C_{M,1} * W_{M,1} + C_{M,2} * W_{M,2} + C_{M,3} * W_{M,3} + \dots \end{aligned}$$

Where

$C_{I,J}$ =Correlation between signal I and signal J

$W_{I,J}$ =Weighting factor for Correlation between signal I and signal J

The sixth intrinsic disease state DS_{I6} represents the level of chaos in neural activity in a single or multiplicity of areas of the nervous system. This may be implemented as any measure of entropy or chaos, including variance, standard deviation,

54

Lyapunov exponent values, maximum Lyapunov exponent value, or other measure of entropy or chaos without departing from the present invention.

$$DS_{I6}=S_N$$

$$DS_{I6}=S_1 * W_{S1} + S_2 * W_{S2} + S_3 * W_{S3} + \dots + S_N * W_{SN}$$

Where

S_I =Entropy measure for signal I, which may be implemented in any of several ways outlined above, some of which are detailed below.

W_{S_I} =Weighting factor for Entropy measure for signal Chaos measurement implemented and quantified as Entropy:

$$S_I = \text{Entropy measure for signal } I$$

$$= -k \int [dX] P[X_I] \log P[X_I]$$

Where

k =a constant, i.e. Boltzmann's constant as in thermodynamics, or by convention a dimensionless constant in information theory;

X_I =Signal I, such as EEG voltage signal I or implanted electrode signal I or microelectrode voltage signal I

$P[X_I]$ =Probability distribution of Signal I

dX =integration variable

In a typical implementation in digital hardware, based upon a base 2 digitization scheme, chaos measurement implemented and quantified as Entropy becomes:

$$S_I = -\sum p_i \log_2 p_i \text{ bits}$$

Where

S_I =Entropy measure for signal I

p_i =probability of outcome i, for example of sensed voltage being within a particular discretization bin window or value range.

$\log_2 p_i$ =Log base 2 of p_i , alternative bases could be used, but 2 is typically chosen to be consistent with digital hardware, which is implemented using the binary (base 2) system.

Chaos measurement implemented and quantified as Lyapunov exponent, which is defined as:

$$S_I = L = \left(1 / \sum_{k=1}^M D_{t_k} \right) * \sum_{k=1}^M L_k \text{ bits per second}$$

Where

L =Lyapunov Exponent

L_k Satisfies the condition: $L(t_k) = L(t_{k-1}) * 2^{L_k * D_{t_k}}$

$D_{t_k} = t_k - t_{k-1}$ is the evolution time of $L(t_k)$

$L(t_k)$ is the distance between two close points in phase space at time t_k

S_I =Entropy measure for signal I, calculated with Lyapunov Exponent

Other measures of chaos, including variations of these and other measures or estimates for chaos, may be used without departing from the present invention.

FIGS. 37 and 40 show system enclosure in cross section, in which intracranial catheter 7 traverses catheter mount ball 436, which rotates within catheter mount socket 437, providing a swivel mechanism to facilitate the selection of a continuum of potential intracranial target sites for intracranial electrode array and intracranial catheter using a single mounted position for system enclosure 434 on caldarium 9.

The reader is requested to note the following labels on FIG. 37:

Catheter mount ball 436

Catheter mount socket 437

stimulating and recording unit 43

FIG. 41 depicts one design for the system enclosure for implantation in the calvarium. Mechanical attachment 424 is shown in plurality, facilitating mechanical attachment of system enclosure 434 to calvarium 9, including calvarium outer table 421 or calvarium inner table 423, calvarium stabilization lip 501, or other portion of calvarium without departing from the present invention. Alternatively, other attachment means may be fashioned to perform the equivalent function of attaching system enclosure 434 to calvarium 9 without departing from the present invention. A single or plurality of screw 429 is used to perform the attachment of mechanical attachment 424 to calvarium 9 and to system enclosure 434. Mechanical attachment may be implemented as a cranial plate, craniofacial plate, or other form well known to neurosurgeons, or it may be implemented in another equivalent fashion without departing from the present invention.

Electrode 452, 453, 454, 455, 456, 457, 458, 459, are shown on the end of intracranial catheter 7, said electrode facilitate contact between intracranial electrodes 246, and circuitry enclosed within system enclosure 434, including but not limited to output stage, signal conditioning circuit, signal processing circuit, control circuit, and other circuit components.

FIGS. 42 and 43 depict a dual intracranial catheter design, shown from above and shown in cross section profile implanted in a patient, respectively. Mechanical attachment 424 are as described in FIG. 41. Intracranial catheter 7 and Electrode 452, 453, 454, 455, 456, 457, 458, 459 are as described in FIG. 41. Electrode 476, 477, 478, 479, 480, 481, 482, 483 are shown on the end of intracranial catheter 500. A multiplicity of intracranial catheter 7 or intracranial catheter 500 may be employed without departing from the spirit of the present invention. In FIG. 43, a plurality of bone screw 438 is shown facilitating mechanical attachment of mechanical attachment 424 to calvarium 9. In FIG. 43, a plurality of machine screw 439 is shown facilitating mechanical attachment of mechanical attachment 424 to system enclosure 434. In FIG. 43, catheter recess 441 is seen in cross section, providing space for intracranial catheter 7 and intracranial catheter 500 to minimize the profile or height of system enclosure and avoid surface protuberances that could cause ulceration of overlying portions of scalp 10.

FIG. 44 is a schematic diagram of one embodiment of the intracranial stimulator of the present invention implanted bilaterally in a human patient. In the embodiment illustrated in FIG. 44, two neurological control systems 999 are shown implanted bilaterally. Each system 999 includes a stimulating and recording unit 26 and one or more intracranial components described below. As described in this illustrative embodiment, the intracranial components preferably include a stimulating electrode array 37. However, it should become apparent to those of ordinary skill in the relevant art after reading the present disclosure that the stimulating electrodes may also be extracranial; that is, attached to a peripheral nerve in addition to or in place of being located within the cranium.

FIG. 45 is a schematic diagram of one embodiment of the neurological control system 999 implanted unilaterally in a human patient. A multiplicity of components or multiplicity of entire systems may be implanted unilaterally or bilaterally without departing from the present invention. In the embodiment illustrated in FIG. 45, one neurological control systems 999 is shown implanted unilaterally in the temporal lobe,

shown in an orientation intersecting the hippocampus 277. Each system 999 includes a stimulating and recording unit 26 and one or more intracranial components described in detail above in parent cases. As described in this illustrative embodiment, the intracranial components preferably include an intracranial catheter 7 with at least one of a sensor array 560 or neuromodulator array 561. Sensor array 560 may be implemented as a intracranial recording electrode array 38, EEG electrode array 51, or other form of sensory input including sensory input modalities 247 shown in FIG. 1 or other form, including optical, metabolic, chemical, or other sensor, without departing from the present invention. Neuromodulator array 561, shown on intracranial catheter 7, serve to stimulate, inhibit, or both, neural activity. Neuromodulator array 561 may be on the same intracranial catheter 7 as sensor array 560 or on a second intracranial catheter 500, as shown in FIG. 46, on a multiplicity of intracranial catheter 7, or on other structure, without departing from the present invention. There may be partial or complete overlap in elements that comprise neuromodulator array 561 and sensor array 560. For example, neural interface array 562 may be implemented as electrode array 563, some or all of elements of intracranial recording electrode array 38 and elements of intracranial stimulating electrode array 37 may be shared. Various forms of multiplexing, in various dimensions, including time, space, frequency, phase, or other dimension, may be employed to facilitate dual or multiple function of elements of sensor array 560 and neuromodulator array 561, to one skilled in the art, without departing from the present invention. It should become apparent to those of ordinary skill in the relevant art after reading the present disclosure that the stimulating electrodes may also be extracranial; that is, attached to a peripheral nerve or other location such as on the surface of the skin in addition to or in place of being located within the cranium.

FIG. 46 is a schematic diagram of one embodiment of the neurological control system 999 implanted unilaterally in a human patient. A multiplicity of components or multiplicity of entire systems may be implanted unilaterally or bilaterally without departing from the present invention. In the embodiment illustrated in the sagittal view diagram in FIG. 46, one neurological control systems 999 is shown implanted unilaterally, with two intracranial catheters, intracranial catheter 7 and intracranial catheter 500, in the right thalamus 335 and the right hippocampus 564, respectively. The same or a multiplicity of neurological control systems 999 may be placed in contralateral structures, including left thalamus 336 and left hippocampus 565, and other structures including the right anterior nucleus of the thalamus, left anterior nucleus of the thalamus, right subthalamic nucleus, left subthalamic nucleus, right substantia nigra, left substantia nigra, other single structures, or other multiplicity of structures.

Modulation of right hippocampus 564 is also accomplished via hippocampal modulator 534 and hippocampal modulator 535. Modulation of left hippocampus 565 is also accomplished via hippocampal modulator 536 and hippocampal modulator 357. Cortex 252 is modulated via cortical modulator 530, cortical modulator 531, cortical modulator 532, and cortical modulator 533.

FIG. 46 also shows cortical modulator 530, cortical modulator 531, cortical modulator 532, cortical modulator 533, hippocampal modulator 534, hippocampal modulator 535, hippocampal modulator 536, and hippocampal modulator 537, which facilitate modulation of multiple regions of cortex 252, right hippocampus 564, and left hippocampus 565.

FIG. 46 and FIG. 47 are sagittal and lateral view diagrams, respectively, depicting a multiplicity of neural interfacing

nodes. Neural interfacing is accomplished with olfactory nerve **525** via at least one of olfactory nerve modulator **527** and with trigeminal nerve **368** via at least one of trigeminal nerve modulator **528**.

FIG. **47** depicts neural interfacing with vagus nerve **369** via at least one of vagus nerve modulator **529**, with baroreceptor **370** via baroreceptor modulator **566**, with sympathetic ganglion **371** via sympathetic modulator **567**. Modulation of these structures is used for control of neurological state in the treatment of neurological disease as well as hypertension and hypotension.

FIG. **47** also depicts several noninvasive techniques for the treatment of neurological disease. Orbitofrontal modulator **279**, prefrontal modulator **280**, precentral modulator **281**, postcentral modulator **282**, parietal modulator **283**, parietooccipital modulator **284**, occipital modulator **285**, cerebellar modulator **286** are shown overlying the scalp **10**. These may be implemented as electromagnetic coils or as electrodes, to produce electromagnetic waves for the induction of current or to directly produce currents, respectively. There may also be implemented optically or using other modality for the modulation of neurological activity. These neuromodulators may be located at any anatomical depth, including superficial to the scalp, implanted at any level including but not limited to within the scalp, under the scalp, recessed in the caldarium, deep to the caldarium, in the epidural space, in the subdural space, overlying the cortex, within the cortex, or deep to the cortex, without departing from the present invention. Neuromodulators may take the form of macroelectrodes, macroelectrode arrays, microelectrodes, microelectrode arrays, nerve cuffs, other designs described in the present invention, other electrode designs known in the art, and other neural interfaces to be developed, without departing from the present invention.

Neural chaos is modulated by the controlled introduction of energy into a single or multiplicity of neural structures including but not limited to olfactory nerve **525**, trigeminal nerve **368**, vagus nerve **369**, cutaneous nerves, other cranial nerves, subcutaneous nerve endings, cortex **252**, cerebral cortex, cerebellar cortex **257**, deep brain structures **349**, thalamus **121**, subthalamic nucleus **122**, basal ganglia, locus ceruleus, any portion or portions of Papez' circuit, hippocampus, amygdala, fornix, subthalamic nucleus, anterior nucleus of the thalamus, prepyriform cortex, solitary nucleus, dorsal column nucleus, cerebellar nuclei, caudate, putamen, corpus callosum, other nuclei, other tracts, other nerves, other nuclei, other neural structures, and other non-neural structures.

FIG. **48** shows dermatomal zones, revealing specific areas of innervation of skin **382** by underlying nerves. Dermatoma zone **538**, dermatoma zone **539**, and dermatoma zone **540** are innervated by trigeminal nerve branches **1**, **2**, and **3**, respectively. Dermatoma zone **541**, dermatoma zone **542**, and dermatoma zone **543** are innervated by cervical nerve roots **C2**, **C2**, and **C3**, respectively.

FIG. **49** depicts two implementations of noninvasive versions of neurological control system **999**; these are shown as neurological control system **994** which is preferably mounted on the head and neurological control system **995** which is preferably mounted on the neck.

Neurological control system **994** comprises head band **568**, recording and stimulating unit **555**, connecting cable **556**, connecting cable **557**, and neuromodulator **545**, neuromodulator **546**, neuromodulator **547**, neuromodulator **548**, neuromodulator **549**, neuromodulator **550**, neuromodulator **551**, neuromodulator **552**, and an additional set, which may be symmetrical or asymmetrical, on the contralateral side.

Neurological control system **995** comprises head band **569**, recording and stimulating unit **570**, connecting cable **558**, connecting cable **559**, and neuromodulator **553**, neuromodulator **554**, and an additional set, which may be symmetrical or asymmetrical, on the contralateral side.

Any single or plurality of said dermatomal zone **538**, **539**, **540**, **541**, **542**, **543** may be modulated using implanted nerve cuff electrode disclosed in pending patent application Ser. No. 10/198,871 (GISTIM) and the cited provisional application No. 60/307,124. Other techniques for modulating innervation to these regions may be employed without departing from the present invention. These include surface electrical stimulation, magnetic stimulation, transcranial magnetic stimulation, vibrotactile stimulation, thermal stimulation, pressure stimulation, optical stimulation, or other stimulation modality.

FIG. **50** depicts a functional block diagram of the neurological control system **999**, connected to intracranial catheter **7**, shown implanted in the temporal lobe **571**, intersecting the hippocampus **277**. In this diagram, signal from sensor array **560** is conditioned, conducted along intracranial recording electrode (ICRE) signal path **83** to signal processor **71**, which generates control input (Y) **996**, which is conducted to control circuit **72**, which generates control output (U) **997**, which is conducted to Output stage circuit **77**, which generates neuromodulating signal (NMS) **998**, which is conducted along stimulator output path **111** to neuromodulator array **561**. Neuromodulator array **561** and sensor array **560** are implemented as intracranial recording electrode array (ICREA) **38** and intracranial stimulating electrode array (ICSEA) **37**, in one preferred embodiment. In the present invention, control input (Y) **996** is a function of at least one of neural chaos, T-index, neural signal correlation, neural signal cross-correlation, and neural synchronization. Control output (U) **997** and neuromodulatory signal (NMS) **998** are selected to modulate at least one of neural chaos, neural signal correlation, neural signal cross-correlation, and neural synchronization.

FIG. **51** depicts the same functional block diagram as in FIG. **50** with the addition of a second intracranial catheter **500** positioned to intersect at least one of the thalamus **121**, subthalamic nucleus **122**, or other anatomical target, including at least one of the centromedian nucleus, substantia nigra, locus ceruleus, reticular activating center, nucleus solitarius, or other neural structure or non-neural structure.

FIG. **52** depicts the same functional block diagram as in FIG. **51** with the addition of a time diagram of the control input (Y) **996** and control output (U) **997** shown versus time. The control input (Y) **996** is shown to vary with time and to remain within the bounds delimited by threshold **572** and threshold **575**. Control output (U) **997** is shown to vary with time in a manner designed to maintain control input (Y) **996** within the range specified, below threshold **572** and above threshold **575**. Any of the control laws specified in this patent as well as any others may be used without departing from the present invention. In FIG. **52**, signals form a nonlinear control law is shown; as the disease state varies and approaches threshold **575** toward the center of the diagram, control output (U) **997** is seen to increase and drive control input (Y) **996** back toward the center of the range delimited by threshold **572** and threshold **575**.

FIG. **53** depicts the same functional block diagram and time diagram as in FIG. **52** with the addition of a threshold **573**, threshold **574**, and target value **576**. In this time diagram, control output (U) **997** is shown to vary in accordance with a more complex nonlinear control law. The variation is representative of a piecewise linear control law, which response differently when the control input (Y) is in different regions as

defined by threshold 572, threshold 573, threshold 574, threshold 575, and target value 576. In this embodiment, as control input (Y) 996 diverges farther from target value 576, the absolute value of the magnitude of control output (U) 997 becomes larger. The vertical center of the time diagram for control output (U) may be interpreted as being zero, though other offsets and baseline values may be used without departing from the present invention. As disease state enters the range delimited by threshold 572 and threshold 573, the gain of control law is seen to increase, as reflected by the incremental increase in magnitude of control output (U) 997. As disease state enters the range delimited by threshold 574 and threshold 575, the gain of control law is seen to increase, as reflected by the incremental increase, in the opposite polarity, in magnitude of control output (U) 997. This behavior demonstrates a control law designed to maintain control input (Y) 996 within the operating range defined by threshold 573 and threshold 574, shown as target range 577 and target range 578, which are above and below target value 576, respectively. As is shown subsequently, state (X) 991 is a scalar or vector of value representative of neurological state and disease state. Components of state (X) 991 may be identical to or functions of control input (Y). Furthermore, the same, similar, or different control laws and systems may be employed to control the behavior of state (X) 991. fcontrol input (Y) 996 diverges outside this range and into critical range 579 or critical range 580, a different control scheme is used. This may be implemented as the same control law with different gains or as a different control law.

In FIG. 53, the signal path is shown along with the system block diagram, detailed in FIG. 2, which is from the original filing of the parent case of this patent. Neural recording signal (NRS) 993 is sensed by sensor array 560, and is transmitted to signal conditioning circuit 76. Feedback signal (F) 992 is generated by signal conditioning circuit 76 from neural recording signal (NRS) 993 and transmitted to signal processor 71. Feedback signal (F) is shown transmitted on Conditioned Intracranial recording electrode (ICRE) signal path 83, though for a non-implanted neurological control system 999, a different transmission means would be used, without departing from the present invention. Signal conditioning circuit 76, may have a unity gain, and equivalently be omitted from neurological control system 999, without departing from the present or parent case invention.

In FIG. 54, a time diagram of the control input (Y) 996 and control output (U) 997 shown versus time. In addition, a time diagram of state vector (X) 991, with components state 1 (X1) 991-1, state 2 (X2) 991-2, state 3 (X3) 991-3, state 4 (X4) 991-4, and state 5 (X5) 991-5, are shown. More or fewer states, comprising the range of state 1 (X1) 991-1 to state N (XN) 991-N, as shown in FIG. 56, may be defined without departing from the present invention.

In state time diagram shown, state 1 (X1) 991-1 is identical to control input (Y) 996. In the application of seizure control, one embodiment for such a state as represented by state 1 (X1) 991-1 is a function of neural chaos. This may be used to facilitate a proportional control law component in the control of neural chaos. In one preferred embodiment the function of neural chaos includes the disentrainment of neural chaos, inversely related to the synchronization of neural chaos among at least 1 recording electrode or other neural signal transducer.

There are a multiplicity of methods and designs in which this can be implemented in the present invention. In one such preferred embodiment, one state may be allocated to represent the measure of neural chaos for each element of neural recording signal (NRS) 993, as recorded from each recording

electrode or signal transducer, including at least one of intracranial recording electrode 5, intracranial recording electrode 6, elements of intracranial recording electrode array 38, elements of neural interface array 562 (shown in FIG. 45), or other neural signal transducer. At least one additional state is defined as a function of these states, specifically the disentrainment of neural chaos, inversely related to the correlation or synchronization of neural chaos, which is a measure of the correlation between these measures of neural chaos derived from elements of neural recording signal (NRS) 993. These measures of neural chaos and the correlation between neural chaos measurements may be alternatively calculated within signal processor 71 as a neural state or disease state estimate, without departing from the present invention.

In state time diagram shown, state 2 (X2) 991-2 is a time derivative of control input (Y) 996. In the application of seizure control, one embodiment for such a state as represented by state 2 (X2) 991-2 is a measure of the first derivative with respect to time of neural chaos or function thereof, which is used to facilitate a differential control law component in the control of said function of neural chaos.

In state time diagram shown, state 3 (X3) 991-3 is a time integral of control input (Y) 996. In the application of seizure control, one embodiment for such a state as represented by state 3 (X3) 991-3 is a measure of the first integral with respect to time of neural chaos or function thereof, which is used to facilitate an integral control law component in the control of said function of neural chaos.

In state time diagram shown, state 4 (X4) 991-4 is a nonlinear function of control input (Y) 996. In the application of seizure control, one embodiment for such a state as represented by state 4 (X4) 991-3 is a measure of a nonlinear function of neural chaos, which is used to facilitate a nonlinear control law component in the control of neural chaos or function thereof, including disentrainment of neural chaos or of synchronization of neural chaos. As shown, state 4 (X4) 991-4 depicts a nonlinear function which is approximately zero during the condition in which control input (Y) 996 is within target range 577 and target range 578 and increases in magnitude as control input (Y) deviates from the region defined by the union of target range 577 and target range 578 and migrates into at least one of critical range 579 and critical range 580. State 4 (X4) 991-4 is shown to represent "target range deviation", a continuous function which is shown as the amount by which state 1 (X1) deviates outside the limits defined by the union of target range 577 and target range 578.

In state time diagram shown, state 5 (X5) 991-5 is an error signal representing the difference between a constant or time varying reference value and control input (Y) 996. As such it represents an error signal. It may also be processed to generate additional states, such as a combination of first, second, and higher order derivatives and integrals with respect to time, to produce control laws of arbitrary complexity to facilitate tracking of any or all components of state (X) 991 to a desired reference signal. In the application of seizure control, one embodiment for such a state as represented by state 5 (X5) 991-5 is a measure of the difference between the actual level of neural chaos and the desired level of neural chaos, such as target value 576 or other signal.

These components of state (X) 991, and other signals and functions thereof, are used to drive the neural chaos to a desired level, such as within the normal level seen during inter-ictal periods, and to prevent the neural chaos from entering critical range 579 or critical range 580. When neural chaos decreases to a value within critical range 580, the probability of a seizure is increased. Generation of neuromodulatory signal (NMS) 998 is performed to drive neural chaos out of

this region into either of target range 577 or target range 578, to reduce the probability of a seizure and thereby prevent the occurrence of a seizure.

This control system is also used to prevent occurrence of other neurological events, including mania, depression, psychosis, rage, narcolepsy, desire for addicting agents (i.e. opiates, cocaine, nicotine, alcohol, or other drug), or other undesirable neurological state, condition, perception, or symptom.

Neuromodulatory signal (NMS) 998 is transmitted to neuromodulator array 561. Neuromodulator array 561, encompasses any single or plurality of neuromodulator elements, including neuromodulator 545, neuromodulator 546, neuromodulator 547, neuromodulator 548, neuromodulator 549, neuromodulator 550, neuromodulator 551, neuromodulator 552, neuromodulator 553, neuromodulator 554, intracranial stimulating electrode 1, intracranial stimulating electrode 2, intracranial stimulating electrode 3, and intracranial stimulating electrode 4. Any of said neuromodulator elements may be implemented as a singularity or combination of an electrode for the delivery of electrical energy, a drug delivery catheter for the delivery of drug or chemical agent, a micro-catheter for the intraparenchymal delivery of drug or agent, an optical element for the delivery of optical energy, a coil or other transceiver for the delivery of electromagnetic energy, ultrasonic transducer for the delivery of ultrasound energy, thermal source for the introduction of thermal energy, thermal sink for the removal of thermal energy, microdialysis device for the introduction or control of chemical concentrations, or other device for the modulation of neural activity.

Neuromodulatory signal (NMS) 998 is transmitted to neuromodulator array 561, according to closed-loop control laws described above, to control state (X) 991 in any of several manners or combinations thereof, including control laws to: (1) maintain state (X) 991 at or near a target value (scalar or vector), (2) maintain state (X) 991 within a single or plurality of target ranges, including but not limited to target range 577 and target range 578, (3) prevent state (X) 991 from entering any critical range, including but not limited to critical range 579 and critical range 580, (4) to maintain state (X) 991 at or near any constant or time-varying reference value, (scalar or vector), (5) maintain state (X) 991 within a single or plurality of constant or time-varying target ranges, (6) prevent state (X) 991 from entering any constant or time-varying critical range, and (7) other form of closed-loop control.

As is typical of control systems, the farther any single or plurality of component of state (X) 991 is from a critical range, such as critical range 579 and critical range 580, the lower the probability of entering said critical range by said single or plurality of component of state (X) 991. The larger the difference between a single or plurality of component of state (X) 991 from said critical range, the larger the system dynamic effect, external perturbation, or noise magnitude would be required to drive said single or plurality of component of state (X) 991 into said critical range. For a neurological condition that has an elevated probability of occurring when said single or plurality of component of state (X) 991 are in critical range, the probability of said neurological condition occurring can be minimized if said single or plurality of component of state (X) 991 is kept outside or as far in value as possible from said critical range.

When the present invention is applied to the treatment of epilepsy, at least one component, i.e. state 1 (X1) 991-1, of state (X) 991, is decreases as the probability of a seizure occurring increases. Through the introduction of chaos into the nervous system, the neural chaos is increased, thereby reducing the probability of a seizure occurring. With suffi-

cient control gain, the probability of a seizure occurring can be driven to small values approaching zero; resulting in the prevention of a seizure.

State (X) 991 comprises disease state estimate as shown in FIG. 12. As taught in the parent case, and is shown in FIG. 12, a control error (e) is calculated as the difference between the disease reference state (r) and the disease state estimate (X). Control circuit 72 operates to drive this control error (e) toward zero, such that the disease state estimate (X) follows the disease state reference (r). This same invention, taught in the parent case, is used to control state (X) as also shown in FIG. 52, to follow a desired disease reference state. This same invention, taught in the parent case, is also used to control state (X) as also shown in FIG. 52, to remain within a desired target range. This same invention, taught in the parent case, is also used to control state (X) as also shown in FIG. 52, to remain outside of a critical range. Control law circuit block 231 teaches a configuration for accomplishing each of these control schemes.

In the control of neurological state to prevent seizures, the control circuit 72 shown in the parent invention may be employed to perform at least one of (1) maintenance of neurological chaos or function thereof at a reference value, typically defined within the normal inter-ictal range; (2) maintenance of neurological chaos or function thereof within a target range, typically representative of normal inter-ictal chaos range; (3) maintenance of neurological chaos or function thereof outside of a critical range, typically outside of the normal inter-ictal chaos range, (4) maintenance of neurological chaos or function thereof as far as possible from a critical range which is associated with an increased probability of seizure; (5) maintenance of neurological chaos or function thereof at a constant value, (6), maintenance of neurological chaos at a time varying value. Functions of neurological chaos included in the present invention comprise disentrainment of neural chaos, entrainment of neural chaos, synchronization of neural chaos, correlation of neural chaos, differences between actual and reference values of neural chaos, and functions thereof.

In FIG. 55, a time diagram of the control input (Y) 996 and control output (U) 997 shown versus time. In addition, a time diagram of state vector (X) 991, with components state 1 (X1) 991-1, state 2 (X2) 991-2, state 3 (X3) 991-3, state 4 (X4) 991-4, and state N (XN) 991-N, are shown. More or fewer states may be defined without departing from the present invention.

In this figure, state 1 (X1) 991-1 is shown to be the same as control input 1 (Y1) 996-1, which represents disentrainment of neural chaos among elements of neural recording signal (NRS) 993, calculated in this preferred embodiment as a disease state estimate by signal processor 71 and transmitted to control circuit 72. The calculations may be allocated differently among signal processor 71 and control circuit 72 without departing from the present invention. State 2 (X2) 991-2 is shown as the first derivative with respect to time of state 1 (X1) 991-1. State 3 (X3) 991-3 is shown as the first integral with respect to time of state 1 (X1) 991-1.

As seen in FIG. 55, control input 1 (Y1) 996-1 remains within target range 577 and target range 578. State 4 (X4) 991-4 represents a function of the amount by which state 1 (X1) deviates outside the limits defined by the union of target range 577 and target range 578. In state time diagram shown, state 4 (X4) 991-4 is a nonlinear function of control input (Y) 996. In the application of seizure control, one embodiment for such a state as represented by state 4 (X4) 991-3 is a measure of a nonlinear function of the disentrainment of neural chaos. As seen in related FIG. 54, state 4 (X4) 991-3 depicts a

nonlinear function which is approximately zero during the condition in which control input (Y) 996 is within target range 577 and target range 578 and increases in magnitude as control input (Y) deviates from the region defined by the union of target range 577 and target range 578 and migrates into at least one of critical range 579 and critical range 580. State 4 (X4) 991-4 is shown to represent "target range deviation", a continuous function which is shown as the amount by which state 1 (X1) deviates outside the limits defined by the union of target range 577 and target range 578. In FIG. 55, state 4 (X4) 99-14 remains approximately zero during the time span shown, since control input 1 (Y1) 996-1 remains within the union of union of target range 577 and target range 578, suggesting satisfactory closed-loop control performance, specifically in the regulation of the disentrainment of neural chaos.

In FIG. 56, a time diagram of the unperturbed control input (Y') 988 and perturbed control input (Y'') 989, unperturbed control output (U') 983 and perturbed control output (U'') 984, unperturbed state (X') 986, and perturbed state (X'') 987 in response to perturbation (P) 990. This figure shows the effect of perturbation (P) 990 on the neurophysiology of patient 227 and signals processed by neurological control system 999.

Unperturbed control input 1 (Y-1') 988, unperturbed control output 1 (U-1') 983, and unperturbed state 1 (X-1') 986 represent the neurophysiology of patient 227 and signals processed by neurological control system 999 in the absence of perturbation (P) 990. Additional elements of any values, including but not limited to unperturbed control input (Y') 988, unperturbed control output (U') 983, and unperturbed state (X') 986 are included in the present invention, in a multivariable implementation of a preferred embodiment of the present invention.

Perturbed control input 1 (Y-1'') 989-1, perturbed control output 1 (U-1'') 984-1, and perturbed state 1 (X-1'') 987-1 represent the neurophysiology of patient 227 and signals processed by neurological control system 999 in the presence of perturbation (P) 990, such as the application of flashing lights. Additional elements of any values, including but not limited to perturbed control input (Y'') 989, perturbed control output (U'') 984, and perturbed state (X'') 987 are included in the present invention, in a multivariable implementation of a preferred embodiment of the present invention.

Independence of neural chaos and disentrainment of neural chaos are seen to decrease, reflective of an increase in synchronization of neural chaos, in response to perturbation (P) 990; and other state variables, including perturbed state 1 (X1'') 991-1, through perturbed state N (XN'') 991-N, as described in detail in FIG. 54 and FIG. 55, are shown to respond accordingly.

Perturbed control output 1 (U-1'') 984-1 is seen to increase relative to unperturbed control output 1 (U-1') 983-1, as determined by control law implemented in control circuit 72, in response to the increase in perturbed state 1 (X1'') 987-1. This incremental increase in perturbed control output 1 (U-1'') 984-1 causes a corresponding incremental increase in neuro-modulating signal (NMS) 998, which causes an incremental increase in the level of neural disentrainment to compensate for the effects of perturbation (P) 990, driving the level of disentrainment of neural chaos back into the desired range, comprising the region defined by the union of target range 577 and target range 578.

In FIG. 57, a time diagram of the EEG signal (E) 985, unperturbed control input (Y') 988 and perturbed control input (Y'') 989, unperturbed control output (U') 983 and perturbed control output (U'') 984, unperturbed state (X') 986

and perturbed state (X'') 987 in response to perturbation (P) 990. This figure shows the effect of perturbation (P) 990 on the neurophysiology of patient 227 and signals processed by neurological control system 999.

In addition to the values shown in FIG. 56, FIG. 57 further shows EEG signal 985, including EEG signal 1 (E1) 985-1 and EEG signal 2 (E2) 985-2, during which time span there are no EEG abnormalities 599. Neurological control system 999 maintains perturbed control input 1 (Y1) 989-1 within the desired range, comprising the region defined by the union of target range 577 and target range 578, preventing any neurological signs or symptoms and preventing any EEG abnormalities which may precede or be concurrent with such neurological signs or symptoms.

In FIG. 58, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the normal unperturbed state. This diagram shows normal EEG signals, (E1) 985-1 and (E2) 985-2, with no EEG abnormalities 599 nor neurological signs or symptoms 600 during this time span.

In one preferred embodiment, control input 1 (Y1) 996-1 calculated by signal processor 71 is representative of disease state, which is implemented as state 1 (X1) 991-1 in control circuit 72. Additional states (X) 991 may be defined or used without departing from the present invention. In one embodiment, for the treatment of seizures, state 1 (X1) 991-1 represent the level of correlation of neural chaos between EEG signals (E1 to E-N) 985-1 to 985-N.

Control input 1 (Y1) 996-1 may occupy a range of values that are subdivided into specific ranges shown in FIG. 58.

Under normal baseline conditions, as shown for baseline nonperturbed nontreated period 592, control input 1 (Y1) 996-1 is shown to vary within normal range 587. This demonstrated variation of control input 1 (Y1) 996-1 occurs throughout the day under normal conditions and in the absence of any signs or symptoms of disease. This represents a normal range of disease state. In one preferred embodiment for the treatment of epilepsy, this represents the normal range of a metric of disentrainment of neural chaos and during which there are no signs and no symptoms of seizure.

Under normal conditions and during which time the present invention is in operation ("Closed-Loop Neuromodulator Control ON"), shown as nonperturbed treated period 593, control input 1 (Y1) 996-1 is shown to vary within control range 586. Control range 586 is a subset of normal range 587. This demonstrates variation of control input 1 (Y1) 996-1 that occurs throughout the day under conditions in which the present invention is active ("Closed-Loop Neuromodulator Control ON") and in the absence of any signs or symptoms of disease. This represents a normal range of disease state. In one preferred embodiment for the treatment of epilepsy, this represents the normal range of a metric of neural chaos during which there are no signs and no symptoms of seizure. During nonperturbed treated period 593, the Closed-Loop Neuromodulator Control is ON and control output (U1) 997-1 is active. Due to the action of control output (U1) 997-1, implemented as electrical current or voltage pulses, flow of pharmacological or chemical agent, emission of light, delivery of vibratory or ultrasound energy, production of electromagnetic energy, application of pressure or other neuro-modulating energy form, control input 1 (Y1) 996-1 is maintained within control range 586.

Under normal conditions and during which time the present invention is inactive ("Closed-Loop Neuromodulator Control OFF"), shown as nonperturbed nontreated period 594, control input 1 (Y1) 996-1 is shown to vary within

normal range 587. This demonstrated variation of control input 1 (Y1) 996-1 occurs throughout the day under normal conditions and in the absence of any signs or symptoms of disease. This represents a normal range of disease state. In one preferred embodiment for the treatment of epilepsy, this represents the normal range of a metric of neural chaos during which there are no signs and no symptoms of seizure. There may be some aftereffects that persist following the deactivation of the present invention (Closed-Loop Neuromodulator Control OFF); these are anticipated in the present invention. The persistence of beneficial effects for some period following the use of the present invention allows the duty cycle of operation to be reduced, thereby minimizing tissue stimulation, drug or other agent delivery.

There are no EEG abnormalities 599 and no neurological signs or symptoms 600, including seizures, during the time periods depicted in FIG. 58. As shown during nonperturbed nontreated period 594, the action of control input 1 (Y1) 996-1 provides a continuous stabilizing influence on control input 1 (Y1) 996-1, preventing it from deviating outside of control range 586. At no point in time in FIG. 58 does control input 1 (Y1) 996-1 indicate any adverse neurological conditions that would be reflective of a seizure, and aura, or associated event. Control input 1 (Y1) 996-1 does not enter borderline range 588, borderline range 589, critical range 590, or critical range 591.

In FIG. 59, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the normal unperturbed state. This diagram shows normal EEG signals, (E1) 985-1 and (E2) 985-2, during this time span, occupying interictal period 603. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, outside of normal range 587 and into borderline range 589 and critical range 591 without the occurrence of any EEG abnormalities 599 or signs or symptoms of neurological disease 600.

In one preferred embodiment, control input 1 (Y1) 996-1 calculated by signal processor 71 is representative of disease state, which is implemented as state 1 (X1) 991-1 in control circuit 72. Additional states (X) 991, including but not limited to the time derivative of state 1 (X1) 991-1, the integral with respect to time of state 1 (X1) 991-1, and other functions of state 1 (X1) 991-1, other elements of state (X) 991 and functions thereof, other elements of control input (Y) 996 and functions thereof, and other values without departing from the present invention.

Control input 1 (Y1) 996-1 is shown to vary first within the normal range 587 and to have multiple excursions into borderline range 589 and critical range 591. During this time, there are no episodes of abnormal EEG nor episodes of neurological signs or symptoms. Control input 1 (Y1) 996-1, as well as corresponding elements of state (X) 991, exhibit excursions outside of normal range 587, and corresponding ranges likewise for elements of state (X) 991, and return to normal range 587 without event.

In FIG. 60, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the unperturbed state in which the present invention is not activated and a seizure spontaneously develops. This diagram shows the development of EEG abnormalities 599, shown in (E1) 985-1 and (E2) 985-2, during this time span, followed by the progression of the seizure to be manifest by neurological signs or symptoms 600. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, outside of normal range 587 and into borderline

range 589 and critical range 591, during which time EEG abnormalities 599 subsequently develop, followed by neurological signs or symptoms 600.

In one preferred embodiment, control input 1 (Y1) 996-1 calculated by signal processor 71 is representative of disease state, which is implemented as state 1 (X1) 991-1 in control circuit 72. Additional states (X) 991, including but not limited to the time derivative of state 1 (X1) 991-1, the integral with respect to time of state 1 (X1) 991-1, and other functions of state 1 (X1) 991-1, other elements of state (X) 991 and functions thereof, other elements of control input (Y) 996 and functions thereof, and other values without departing from the present invention.

Control input 1 (Y1) 996-1 is shown to vary first within the normal range 587 and to have multiple excursions into borderline range 589 and critical range 591. During the first such excursion, which occurs in interictal period 595, there are no episodes of EEG abnormalities 599 nor episodes of neurological signs or symptoms 600. Sometime during the second such excursion into critical range 591, EEG abnormalities 599 develop, as seen during sustained organized paroxysmal discharge period 596. The EEG abnormalities 599 that appears during sustained organized paroxysmal discharge period 596 then progresses to involve a larger portion of the brain and is manifest as neurological signs or symptoms 600 which define the clinical seizure 597. Following clinical seizure 597, neurological signs or symptoms 600 cease and EEG abnormalities 599 cease. During and Following clinical seizure 597, Control input 1 (Y1) 996-1 progressively moves back to normal range 587.

In FIG. 61, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the unperturbed state in which the present invention is activated and providing a continuous stabilizing influence on the nervous system, during controlled interictal period 604. This diagram shows the persistence of normal EEG signals, shown in (E1) 985-1 and (E2) 985-2, during this time span, occupying controlled interictal period 604, accompanied by no neurological signs or symptoms 600. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, within the control range 586. A single or multiplicity of additional states may be used without departing from the present invention.

In one preferred embodiment, control input 1 (Y1) 996-1 calculated by signal processor 71 is representative of disease state, which is implemented as state 1 (X1) 991-1 in control circuit 72. Additional states (X) 991, including but not limited to the time derivative of state 1 (X1) 991-1, the integral with respect to time of state 1 (X1) 991-1, and other functions of state 1 (X1) 991-1, other elements of state (X) 991 and functions thereof, other elements of control input (Y) 996 and functions thereof, and other values without departing from the present invention.

In one preferred embodiment for the treatment of epilepsy, control input 1 (Y1) 996-1 is calculated as a measure of the disentrainment of neural chaos. Other measures of neural chaos, functions of neural chaos, or other function of at least one of neural chaos or neural entropy may be used without departing from the present invention. Neural chaos may be measured using a Lyapunov exponent, Kolmogorov entropy, or other measure of chaos. These measures of neural chaos, disentrainment of neural chaos, entrainment of neural chaos, synchronization of neural chaos, and other functions of neural chaos may be calculated using any of the methods taught in the present invention or using other measures, including those in which information rate, neural signal correlations or cross-

correlations, or other measure of entropy, chaos, or equivalent measure, or other function of neural or physiological signals are employed. Other functions include metrics of neural signal overall energy levels, energy levels within a single or multiplicity frequency bands or ratios thereof, changes in spike and wave frequencies, or other parameters.

Control output 1 (U1) 997-1 is used to generate Neuromodulatory signal (NMS) 998, which is transmitted to at least one of the implementations neural modulator described herein or to other neuromodulator; these include but are not limited to neuromodulator array 561, hippocampal modulator 534 and hippocampal modulator 535, hippocampal modulator 536 and hippocampal modulator 357, cortical modulator 530, cortical modulator 531, cortical modulator 532, and cortical modulator 533, olfactory nerve modulator 527, trigeminal nerve modulator 528, vagus nerve modulator 529, sympathetic modulator 567, orbitofrontal modulator 279, prefrontal modulator 280, precentral modulator 281, postcentral modulator 282, parietal modulator 283, parietooccipital modulator 284, occipital modulator 285, cerebellar modulator 286, neuromodulator 545, neuromodulator 546, neuromodulator 547, neuromodulator 548, neuromodulator 549, neuromodulator 550, neuromodulator 551, neuromodulator 552, neuromodulator 553, neuromodulator 554, neuromodulator array 561, contralateral implementations of any of these, and other neuromodulators. Other variations of these neuromodulator designs or locations may be envisioned by one skilled in the art, and these are included in the present invention. Other anatomical locations or electrode designs or configurations may be used without departing from the present invention.

Control input 1 (Y1) 996-1 is shown to remain within control range 586, while responding to the continuous stabilizing influence provided by control output 1 (U1) 997-1. There are no EEG abnormalities 599 and no neurological signs or symptoms 600 during the action provided by the feedback driven controller embodied in the present invention. The present invention encompasses the use of any single or multiplicity of elements of control input Y 996, representing any single or multiplicity of disease state. One preferred state used in the treatment of epilepsy is the use of at least one measure of disentrainment of neural chaos. These and other measures of neural activity, chaos, disentrainment of neural chaos, entrainment of neural chaos, synchronization of chaos are also used in the treatment of psychosis, depression, schizophrenia, mania, bipolar disorder, rage, anxiety, and other neurological and psychiatric conditions.

In FIG. 62, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the unperturbed state in which the control output 1 (U1) is initially OFF during uncontrolled interictal period 605 and is subsequently activated and begins providing a continuous stabilizing influence on the nervous system, during controlled interictal period 606.

The transition from control output 1 (U1) in the OFF state during uncontrolled interictal period 605 to the ON state during controlled interictal period 606 occurs at an arbitrary time and under normal conditions. In the diagram shown, control input 1 (Y1) 996-1 has deviated from arbitrarily defined control range 586 and is still within normal range 587. As has been shown in FIG. 59, this does not represent a precursor to a seizure, and even if control input 1 (Y1) 996-1 were to enter orderline range 588, borderline range 589, critical range 590, or critical range 591, this condition would not represent a precursor to a seizure. At the point shown, control input 1 (Y1) 996-1 has deviated by an arbitrary amount from

target value 576, at which point control output 1 (U1) 997-1 is activated to apply closed-loop feedback control to stabilize control input 1 (Y1) 996-1 and bring control input 1 (Y1) 996-1 into control range 586.

This diagram shows the persistence of normal EEG signals, shown in (E1) 985-1 and (E2) 985-2, during this time span, occupying controlled interictal period 604, accompanied by no neurological signs or symptoms 600. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, within the control range 586. A single or multiplicity of additional states may be used without departing from the present invention.

In FIG. 63, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time in the unperturbed state in which the control output 1 (U1) is initially OFF during uncontrolled interictal period 607, is subsequently activated and begins providing a continuous stabilizing influence on the nervous system during controlled interictal period 608, and control output 1 (U1) is subsequently OFF in uncontrolled interictal period 609.

The transition from control output 1 (U1) in the OFF state during uncontrolled interictal period 605 to the ON state during controlled interictal period 606 occurs at an arbitrary time and under normal conditions. In the diagram shown, control input 1 (Y1) 996-1 has deviated from arbitrarily defined control range 586 and is still within normal range 587. As has been shown in FIG. 59, this does not represent a precursor to a seizure, and even if control input 1 (Y1) 996-1 were to enter borderline range 588, borderline range 589, critical range 590, or critical range 591, this condition would not represent a precursor to a seizure. At the point shown, control input 1 (Y1) 996-1 has deviated by an arbitrary amount from target value 576, at which point control output 1 (U1) 997-1 is activated to apply closed-loop feedback control to stabilize control input 1 (Y1) 996-1 and bring control input 1 (Y1) 996-1 into control range 586. Once control input 1 (Y1) 996-1 has been stabilized into an arbitrary range, such as control range 586 or other range, control output 1 (U1) is turned OFF, as shown in uncontrolled interictal period 609. The turning of control output 1 (U1) OFF is performed to conserve battery power, minimize electrical current induced tissue damage, minimize pH changes at the tissue-electrode interface, to further minimize neural habituation, which is itself a benefit of closed-loop feedback driven neuromodulation, and other benefits.

This diagram shows the persistence of normal EEG signals, shown in (E1) 985-1 and (E2) 985-2, during this time span, occupying controlled interictal period 604, accompanied by no neurological signs or symptoms 600. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, within the control range 586. A single or multiplicity of additional states may be used without departing from the present invention.

In FIG. 64, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time before, during, and after the application of a perturbation (P) 990, such as flashing lights. This diagram shows normal EEG signals, (E1) 985-1 and (E2) 985-2, during this time span, occupying uncontrolled baseline period 610 preceding the application of perturbation (P) 990, uncontrolled perturbation period 611 during the application of perturbation (P) 990, and uncontrolled post-perturbation period 612 following the application of perturbation (P) 990. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, within normal range 587, and

without the occurrence of any EEG abnormalities **599** or neurological signs or symptoms **600**. Despite the application of perturbation (P) **990**, control input **1** (Y1) **996-1**, a measure of disease state, remains in normal range **587**; and corresponding elements of state (X) **991** remain in their corresponding normal ranges.

In one preferred embodiment, control input **1** (Y1) **996-1** calculated by signal processor **71** is representative of disease state, which is implemented as state **1** (X1) **991-1** in control circuit **72**. In a preferred embodiment for the treatment of seizures, control input **1** (Y1) **996-1** is a function of disen- 10 trainment of neural chaos, for which neural chaos is calculated using Lyapunov exponents, Kolmogorov entropy or other measure of chaos. Additional elements of state (X) **991**, including but not limited to the time derivative of state **1** (X1) **991-1**, the integral with respect to time of state **1** (X1) **991-1**, and other functions of state **1** (X1) **991-1**, other elements of state (X) **991** and functions thereof, other elements of control input (Y) **996** and functions thereof, and other values without departing from the present invention.

In FIG. **65**, a time diagram of the EEG tracings, (E1) **985-1** and (E2) **985-2**, control input **1** (Y1) **996-1**, state **1** (X1) **991-1**, and control output **1** (U1) **997-1** are shown versus time before, during, and after the application of a perturbation (P) **990**, such as flashing lights. This diagram shows normal EEG signals, (E1) **985-1** and (E2) **985-2**, during this time span, occupying uncontrolled baseline period **610** preceding the application of perturbation (P) **990**, uncontrolled perturbation period **611** during the application of perturbation (P) **990**, and uncontrolled post-perturbation period **612** following the application of perturbation (P) **990**. This figure demonstrates fluctuation of control input **1** (Y1) **996-1**, and correspond- 25 ingly of state **1** (X1) **991-1**, outside of normal range **587** and into borderline range **589** and critical range **591** without the occurrence of any EEG abnormalities **599** or neurological signs or symptoms **600**.

In one preferred embodiment, control input **1** (Y1) **996-1** calculated by signal processor **71** is representative of disease state, which is implemented as state **1** (X1) **991-1** in control circuit **72**. In a preferred embodiment for the treatment of seizures, control input **1** (Y1) **996-1** is a function of disen- 40 trainment of neural chaos, for which neural chaos is calculated using Lyapunov exponents, Kolmogorov entropy or other measure of chaos. Additional elements of state (X) **991** may be used, including but not limited to the time derivative of state **1** (X1) **991-1**, the integral with respect to time of state **1** (X1) **991-1**, and other functions of state **1** (X1) **991-1**, other variables or constants and functions thereof, other elements of control input (Y) **996** and functions thereof, and other values without departing from the present invention.

Control input **1** (Y1) **996-1** is shown to vary first within the normal range **587** and to have multiple excursions into border- 45 line range **589** and critical range **591**. During this time, there are no episodes of EEG abnormalities **599** nor episodes of neurological signs or symptoms **600**. Control input **1** (Y1) **996-1**, as well as corresponding elements of state (X) **991**, exhibit excursions outside of normal range **587**, and corresponding ranges likewise for elements of state (X) **991**, and return to normal range **587** without event.

In FIG. **66**, a time diagram of the EEG tracings, (E1) **985-1** and (E2) **985-2**, control input **1** (Y) **996-1**, state **1** (X1) **991-1**, and control output **1** (U1) **997-1** are shown versus time before, during, and after the application of a perturbation (P) **990**, such as flashing lights, during which time the present inven- 60 tion is not activated and a seizure subsequently develops.

This diagram shows EEG signals, (E1) **985-1** and (E2) **985-2**, which are initially normal during uncontrolled base-

line period **613**, preceding the application of perturbation (P) **990**, and which remain normal during uncontrolled perturbation period **614** during the application of perturbation (P) **990**, and remain normal for some time thereafter during uncontrolled post-perturbation period **615** following the application of perturbation (P) **990**. This figure demonstrates fluctuation of control input **1** (Y1) **996-1**, and correspondingly of state **1** (X1) **991-1**, outside of normal range **587** and into borderline range **589** and critical range **591** without the occurrence of 10 any EEG abnormalities **599** or neurological signs or symptoms **600** during uncontrolled baseline period **613**, uncontrolled perturbation period **614**, and uncontrolled post-perturbation period **615**.

During the second excursion of control input **1** (Y1) **996-1** into critical range **591**, EEG abnormalities **599** develop, shown in (E1) **985-1** and (E2) **985-2**, defining the beginning of sustained organized paroxysmal discharge period **616**, which is followed by the progression of EEG abnormalities **599** into clinical seizure **617**, which is manifest by neurologi- 20 cal signs or symptoms **600**.

In one preferred embodiment, control input **1** (Y1) **996-1** calculated by signal processor **71** is representative of disease state, which is implemented as state **1** (X1) **991-1** in control circuit **72**. In a preferred embodiment for the treatment of seizures, control input **1** (Y1) **996-1** is a function of disen- 25 trainment of neural chaos, for which neural chaos is calculated using Lyapunov exponents, Kolmogorov entropy or other measure of chaos. Additional elements of state (X) **991** may be used, including but not limited to the time derivative of state **1** (X1) **991-1**, the integral with respect to time of state **1** (X1) **991-1**, and other functions of state **1** (X1) **991-1**, other variables or constants and functions thereof, other elements of control input (Y) **996** and functions thereof, and other values without departing from the present invention.

Control input **1** (Y1) **996-1** is shown to vary first within the normal range **587** and to have multiple excursions into border- 35 line range **589** and critical range **591**. During the first such excursion, which occurs in uncontrolled post-perturbation period **615**, there are no episodes of EEG abnormalities **599** nor episodes of neurological signs or symptoms **600**. Sometime during the second such excursion into critical range **591**, EEG abnormalities **599** develop, as seen during sustained organized paroxysmal discharge period **596**. The EEG abnormalities **599** that appears during sustained organized paroxysmal discharge period **596** then progresses to involve a larger portion of the brain and is manifest as neurological signs or symptoms **600** which define the clinical seizure **597**. Following clinical seizure **597**, neurological signs or symptoms **600** cease and EEG abnormalities **599** cease; and this remains the case during uncontrolled interictal period **618**. During and Following clinical seizure **597**, Control input **1** (Y1) **996-1**, reflective of disease state, progressively moves back to normal range **587**.

In FIG. **67**, a time diagram of the EEG tracings, (E1) **985-1** and (E2) **985-2**, control input **1** (Y1) **996-1**, state **1** (X1) **991-1**, and control output **1** (U1) **997-1** are shown versus time before, during, and after the application of a perturbation (P) **990**, such as flashing lights. The present invention is shown continuously activated, producing control output **1** (U1) **997-1** to provide a continuous stabilizing influence, and the potential for a neurological signs or symptoms **600** is prevented. Furthermore, the very potential for even a precursor, i.e. EEG abnormalities **599**, is prevented, because the nervous system is prevented from entering a state in which such neuro- 65 logical signs or symptoms **600** or their precursors may develop and progress. Said neurological signs or symptoms **600** include but are not limited to a seizure, status epilepticus,

headache, manic episode, depressive episode, anxiety episode, panic attack, rage episode, psychotic episode.

In FIG. 68, a time diagram of the EEG tracings, (E1) 985-1 and (E2) 985-2, control input 1 (Y1) 996-1, state 1 (X1) 991-1, and control output 1 (U1) 997-1 are shown versus time before, during, and after the application of a perturbation (P) 990, such as flashing lights. The present invention is shown conditionally activated, when control input 1 (Y1) 996-1 moves outside of control range, to resume the production of control output 1 (U1) 997-1 to provide a continuous stabilizing influence and prevent the potential for a neurological signs or symptoms 600 or their precursors. There are no EEG abnormalities 599 or neurological signs or symptoms 600 present and no potential for them to occur at any point in this figure.

This diagram shows EEG signals, (E1) 985-1 and (E2) 985-2, which remain normal during uncontrolled baseline period 622, preceding the application of perturbation (P) 990, and which remain normal during uncontrolled perturbation period 623 during which time perturbation (P) 990 is applied, and remain normal thereafter during uncontrolled post-perturbation period 624 and controlled post-perturbation period 625. This figure demonstrates fluctuation of control input 1 (Y1) 996-1, and correspondingly of state 1 (X1) 991-1, outside of control range 586 without the occurrence of any EEG abnormalities 599 or neurological signs or symptoms 600.

When control input 1 (Y1) 996-1 moves outside of control range 586 but remains within normal range 587, then control output 1 (U1) 997-1 is delivered to maintain the nervous system in a stable state and thereby prevent even the possibility of undesirable neurological signs or symptoms 600 or their precursors, i.e. EEG abnormalities 599, from developing. The precise time of initiation of delivery of control output 1 (U1) 997-1 is somewhat arbitrary, as the action is taken not in response to any event but as a measure to prevent a neural state, as quantified by control input 1 (Y1) 996-1 and correspondingly of state 1 (X1) 991-1, from exiting normal range 587 and progressing into a critical range 591 in which neurological signs or symptoms 600 may subsequently develop.

By this action of control output 1 (U1) 997-1, the very potential for even a precursor, i.e. EEG abnormalities 599, is prevented, because the nervous system is prevented from entering a state in which such neurological signs or symptoms 600 or their precursors may develop and progress. Said neurological signs or symptoms 600 include but are not limited to a seizure, status epilepticus, headache, manic episode, depressive episode, anxiety episode, panic attack, rage episode, psychotic episode.

Summary

In one preferred embodiment, the control input 1 (Y1) 996-1 is calculated as a measure of disentrainment of neural chaos among regions of the nervous system. This representation of control input 1 (Y1) 996-1 then reflects degree of neural entrainment, which is inversely related to entrainment of neural chaos and synchronization of neural chaos. During normal interictal states, when no seizure or other neurological signs or symptoms 600 are present, the nervous system possesses a normal degree of neural entrainment, that is, the various neural regions exhibit some correlation in their level of chaos. A seizure occurs as a significant portion of neural tissue exhibits synchronization of activity. By minimizing neural entrainment and maximizing disentrainment, seizures and their precursors may be prevented.

This is fundamentally different from and a dramatic improvement over the system described by but not enabled by Fischell U.S. Pat. No. 6,016,449. Fischell describes a system that detects and terminates a seizure. The patent then claims

but does not enable a system to detect and terminate a precursor to a seizure, by detecting EEG thresholds. Precursors are not defined and their detection not described nor enabled.

In either of these two cases of terminating seizures or precursors to seizures, the aberrant neural condition has already occurred and its progression to seizure is imminent and inevitable without intervention. At this point, terminating the process, which has already begun, is difficult and may not be possible. In the Fischell designs, an event detection signal is generated upon detection of a seizure or its precursor, in either case of which, progression to clinical seizure has begun or is imminent, as manifest by EEG abnormalities 599.

In contrast, the neurological control system 999 taught in the present invention controls fundamental neural states, thereby preventing the development of neural states in which a seizure is even possible. By maintaining neural disentrainment within the normal range, seizures do not occur or are extremely unlikely. Certain pharmacological agents may have this effect as well, by preventing the nervous system from being able to initiate the process that culminates in a seizure. The present invention can continuously monitor and maintain a desired level of therapy, controlling desired neural states to remain within stable regions and out of regions in which neurological signs and symptoms may develop. Through the use of feedback control, the present invention insures that the appropriate neural states, including but not limited to the level of disentrainment of neural chaos, which is inversely related to synchronization of neural chaos and entrainment of neural chaos, remains within the normal range, preventing the nervous system from entering a state, which itself is free of any abnormalities or signs or symptoms, in which it is even possible to develop a seizure or other neurological signs or symptoms 600.

Not only is a seizure prevented, the potential for a seizure is prevented. Furthermore, the potential for a precursor to a seizure is prevented. The present invention controls the degree of disentrainment of neural chaos as measured in at least one region of the brain. This degree of disentrainment, the quantification of which is embodied in control input 1 (Y1) 996-1, normally varies throughout the day. Certain external inputs, such as perturbations as well as medications, can alter this degree of disentrainment. If control input 1 (Y1) 996-1 is maintained in normal range 587, the development of a precursor to a seizure, specifically EEG abnormalities 599 as manifest during sustained organized paroxysmal discharge period 596, are prevented. Neural state, as quantified in control input 1 (Y1) 996-1, varies overtime. Under certain conditions, i.e. when control input 1 (Y1) 996-1 is in critical range 591, a seizure may possibly occur, but it is not predetermined to occur under this condition. Maintaining control input 1 (Y1) 996-1 outside of critical range 591 and preferably in normal range 587 and more preferably in a subset of normal range 587 labeled control range 586 prevents even the precursor, i.e. EEG abnormalities 599, of a neurological signs or symptoms 600 from developing. The present invention prevents the nervous system from entering a state in which it is possible to have a seizure. The present invention monitors the degree of response to therapy and modulates therapy to maintain effect within the desired therapeutic range, i.e. control input 1 (Y1) 996-1 within control range 586. Under such conditions, no neurological signs or symptoms 600 occur. No precursors, i.e. EEG abnormalities 599, occur, since the present invention applies a continuous stabilizing influence to modulate the nervous system and thereby maintain it in a stable state.

73

What is claimed is:

1. A method of monitoring and storing EEG signals from a patient, the method comprising:

establishing a wireless communication link between an implanted unit and a communication module that is external to the patient's body, wherein the implanted unit is coupled to an intracranial catheter having a distal end positioned inside the patient's calvarium, the intracranial catheter containing one or more wires coupling the implanted unit to one or more intracranial recording electrodes and one or more wires coupling the implanted unit to one or more intracranial stimulating electrodes; monitoring EEG signals from the patient with the intracranial recording electrodes and the implanted unit substantially continuously while the wireless communication link is established;

wirelessly transmitting in substantially real-time the monitored EEG signals from the implanted unit to the communication module that is external to the patient's body substantially continuously while the wireless communication link is established;

storing the EEG signals in a memory in the communication module;

monitoring high frequency head vibration, muscle vibration, or speech production using an acoustic sensor mounted in the patient's head and coupled to the implanted unit; and

monitoring head movement and position with respect to gravity using an accelerometer mounted in the patient's head and coupled to the implanted unit.

2. The method of claim 1 wherein the communication module is a portable patient interface module, and wherein the memory is within the portable patient interface module.

3. The method of claim 1 wherein the communication module is a supervisory module that is used by a healthcare provider, and wherein the memory is within the supervisory module.

74

4. The method of claim 1 wherein storing the transmitted EEG signals creates a time history record of the EEG signals for a time period when the wireless communication link is established between the implanted unit and the communication module.

5. The method of claim 1 further comprising wirelessly transmitting a stored time history of the EEG signals that was previously stored in a memory of the implanted unit.

6. The method of claim 1 further comprising analyzing the EEG signals to estimate the patient's condition.

7. The method of claim 6 wherein analyzing the EEG signals from the patient to estimate the patient's condition comprises predicting future symptomatology of the patient's condition.

8. The method of claim 1 further comprising using the communication module to provide an output to the patient that is indicative of the patient's estimated condition.

9. The method of claim 7 further comprising initiating delivery of a therapy to the patient when the future symptomatology is predicted.

10. The method of claim 1, further comprising: monitoring electromyographic signals using an electromyography electrode array positioned in the patient's neck.

11. The method of claim 1, further comprising: storing stimulation electrode impedance history in the implanted unit.

12. The method of claim 1, further comprising: monitoring eyelid movement using EMG sensors to determine frequency of eye blinking.

* * * * *

专利名称(译)	用于连续EEG监测的方法和系统		
公开(公告)号	US9113801	公开(公告)日	2015-08-25
申请号	US11/647785	申请日	2006-12-29
[标]申请(专利权)人(译)	BIONEURONICS CORP		
申请(专利权)人(译)	BIONEURONICS CORPORATION		
当前申请(专利权)人(译)	Cyberonics公司，INC.		
[标]发明人	DILorenzo DANIEL JOHN		
发明人	DILorenzo, DANIEL JOHN		
IPC分类号	A61B5/04 A61B5/0476 A61B5/0488 A61N1/372 A61B5/11 A61B5/00 A61N1/36 A61N1/05 A61N1/378		
CPC分类号	A61B5/04001 A61B5/04012 A61B5/0476 A61B5/0488 A61B5/1101 A61B5/4094 A61B5/4839 A61N1/36082 A61N1/372 A61B5/6864 A61B5/6868 A61B5/6882 A61B5/7203 A61N1/0529 A61N1/0531 A61N1/0539 A61N1/36114 A61N1/3787 A61B5/6852 A61B5/0816 A61B5/4082 A61N1/36064 A61N1/36067 A61N1/36139		
审查员(译)	德安杰洛，MICHAEL		
优先权	60/095413 1998-08-05 US 60/460140 2003-04-03 US 60/427699 2002-11-20 US 60/438286 2003-01-06 US 60/562487 2004-04-14 US 60/436792 2002-12-27 US		
其他公开文献	US20070161919A1		
外部链接	Espacenet USPTO		

摘要(译)

一种神经控制系统，用于调节任何组件或结构的的活动，所述组件或结构包括神经系统的整体或部分，或与其连接的任何结构，在本文中通常称为“神经系统组件”。神经系统控制系统产生传递给神经系统的神经调制信号。神经系统组件通过一个或多个神经调节剂来控制神经状态并预防神经系统症状和体征。这样的治疗参数可以源自对一个或多个传感器感测的先前传递的神经调制信号的神经反应，每个传感器配置成感测指示神经或精神病症的特定特征。

