

(19)



(11)

EP 1 785 723 A1

(12)

EUROPEAN PATENT APPLICATION

(43) Date of publication:

16.05.2007 Bulletin 2007/20

(51) Int Cl.:

G01N 25/20 (2006.01)

G01N 33/48 (2006.01)

G01K 17/00 (2006.01)

C12Q 1/04 (2006.01)

A61B 5/00 (2006.01)

(21) Application number: **05256977.9**

(22) Date of filing: **11.11.2005**

(84) Designated Contracting States:

**AT BE BG CH CY CZ DE DK EE ES FI FR GB GR
HU IE IS IT LI LT LU LV MC NL PL PT RO SE SI
SK TR**

Designated Extension States:

AL BA HR MK YU

(72) Inventor: **Kjeldsen, Naja Joslin**
4055 Basel (CH)

(74) Representative: **Benson, John Everett**
J.A. Kemp & Co.,
14 South Square,
Gray's Inn
London WC1R 5JJ (GB)

(71) Applicant: **Stratos Bio Ltd.**
1205 Geneva (CH)

(54) **Micro calorimetric analysis of mammalian cell susceptibility to agents that affect cell growth, metabolism and immune response, and use of micro calorimetric analysis in a transplantation test**

(57) Method of determining whether or not an organ for transplantation (from a human donor) or xenotransplantation (from an animal donor) is infected with a microorganism (which is optionally a virus) comprising
- measuring heat output from the organ to detect activity of said microorganism, or

- forcing expression of latent DNA by using agents that stimulate m-RNA synthesis,

to thereby determine the presence of a microorganism in the organ.

EP 1 785 723 A1

Description**Field of the Invention**

[0001] The invention relates to microcalorimetric analysis of agents used for either stimulating or repressing the immune system, such as anti-retroviral agents and agents used for transplantation. The present invention also relates to use of microcalorimetry to test for cell necrosis and rejection of organs subsequent to transplantation and elucidate the mode of action of already existing anti rejection agents such as cyclosporins both alone and in combinations. Further the invention relates to

- use of microcalorimetry to develop new agents either alone or in combination to prevent rejection
- use of microcalorimetry to assess organs for viral vectors prior to transplantation by stimulating eukaryotic mRNA synthesis and expression of latent DNA
- use of microcalorimetry to elucidate the mode of action of anti retroviral agents and their protective effect on DNA expression and stimulation of RNA synthesis.

Background of the Invention

[0002] There is a need for improved methods of testing organs for transplantation and to further investigate agents used to treat transplantation patients.

Summary of the Invention

[0003] Microcalorimetric studies of both growing and respiring bacteria and antibiotic action of those have concluded that agents that interfere with RNA synthesis may be responsible for both cell growth and expression of latent DNA, which is an important factor when attempting xenotransplantation.

[0004] These findings can also be applied to transplantation in that immuno suppressive agents such as cyclosporin can be tested in living cell systems both alone and in combinations and can give early warning on cell necrosis and potential rejection episodes as well as immunosuppressant effect.

[0005] They can also be applied for assessment of agents that stimulate the immune system such as anti-retroviral agents used for treatment of HIV positive patients

[0006] The invention concerns, amongst other aspects, use of micro/nano calorimetry to detect viruses in an organ for transplantation. Accordingly the invention provides a method of determining whether or not an organ for transplantation (from a human donor) or xenotransplantation (from an animal donor) is infected with a microorganism (which is optionally a virus) comprising

- measuring heat output from the organ to detect activity of said microorganism, and/or
- forcing expression of latent DNA by using agents that stimulate mRNA synthesis,

to thereby determine the presence of a microorganism in the organ.

Detailed description of the invention

[0007] Herein the term "microorganism" is understood to include viruses unless the context requires otherwise.

[0008] The invention provides a method of determining whether or not an organ for transplantation comprises (for example is infected with) a microorganism (for example a virus) comprising measuring heat output from the organ to detect presence of the microorganism and to thereby determine the presence of a virus in the organ. The heat output is typically due to an activity of the microorganism.

[0009] The organ may be from a human donor or from an animal donor. The organ may be from a human donor for transplantation into a human donor. The organ may be from an animal donor for transplantation into a human donor (i.e. for xenotransplantation). Such an animal is typically a mammal, such as a primate or pig.

[0010] The invention also provides a method of determining the suitability of a donor organ for transplantation to a patient comprising adding in vitro a portion of the organ to a fluid or tissue sample from the patient and measuring heat output from the sample using a calorimeter to thereby detect whether or not there is a rejection reaction against the organ.

[0011] The invention also includes a method of determining whether or not a candidate compound has anti-cancer activity comprising adding the compound to a cancer cell sample in vitro and measuring the heat output from the cancer cell using a calorimeter to thereby determine whether or not the candidate compound has anti-cancer activity. The cancer cell may be of any species mentioned herein, but is preferably a human cancer cell.

[0012] The invention also provides a method for determining whether an anti-microorganism compound affects the growth of a mammalian or avian animal comprising adding said compound to cells of said mammal and determining whether detecting the rate of growth of the cells by measuring the heat output from the cells using a calorimeter.

[0013] The invention also provides a method of determining a suitable administration regimens for a patient comprising taking a sample from a patient on a first administration regimen and detecting the effectiveness of the first administration regimen by measurement of the immune response, microorganism load or numbers of malignant cells using calorimetry, and then optionally if the first administration regimen is found to be of low effectiveness selecting a second administration regimen for the patient. The first and second administration regimen differ in respect to the drug or combination of drugs which are administered. The patient may be infected with HIV.

[0014] The invention also provides a method of determining whether an anti-microorganism or anti-infective compound is able to stimulate or suppress the immune system this can be tested by adding the compound to a respiring or growing cell culture optionally at a pre-determined low concentration threshold, such as $< 10 \times \text{MIC}$, $< 8 \times \text{MIC}$, $< 2 \times \text{MIC}$, cell culture $< 1 \times \text{MIC}$ to determine whether or not heat output of the microorganism/virus/mammalian cell occurs in response to the addition of the compound, wherein growth of the microorganism is optionally measured by detecting heat output from the microorganism/virus/mammalian cell using a calorimeter, and wherein the compound may be alone or in a combination with 2 or more other anti-microorganism or immuno suppressive or stimulating agents. The threshold may be less than 0.01ug/ml, 0.01 ug/ml to 10ug/ml, such as 0.1 ug/ml to 1ug/ml.

[0015] If the said compound is found to cause increased growth of microorganisms at the pre-determined low concentration, then the compound may be administered to an individual suffering from a condition caused by the microorganism or at risk of such a condition, at a high dosage, but optionally not at dosages causing increased respiration of the said compound.

The compounds investigated/used in the embodiments of the invention

[0016] One or more of the compounds may act at the cell wall or membrane, or at an intracellular location in the microorganism, such as at a ribosome, or directly on DNA synthesis. In the case where the microorganism is a virus one or more of the compounds may insert themselves into the virus. The mode of action of one or more of the compounds may not be (previously) known.

[0017] The compound may either activate or suppress the immune system either alone or in combination with other agents

[0018] The compound may be a biological agent, such as a protein or monoclonal or polyclonal antibody of any origin. The compound may be a nucleic acid. The compound may be a gene fragment. The compound (particularly proteins, peptides and antibodies) may differ in respect of their amino acid sequence, the type or extent of glycosylation (for example differing in the type and number of sugars which are attached) or the type or extent of lipid modification.

[0019] Generally the method comprises performing more than one analysis with the compounds, wherein during in each analysis the compounds are added to the sample in different ratios, which would normally be achieved by providing the compounds in different concentrations (i.e. different molarities).

[0020] In one embodiment more than one compound may be used. Thus for example in one analysis a first compound will be present at 0.1 to 1% of the molarity of a second compound, or at 1 to 10%, 20 to 30%, 30 to 40 %, 40 to 50 %, 50 to 60%, 60 to 70%, 70 to 80%, 90 to 100%, 100 to 150%, 150 to 250%, 250 to 500%, 500 to 1000%, 1000 to 10,000%. If a first compound is present at a molarity which is 100% of the molarity of the second compound then clearly the compounds are present in a ratio of 1:1 (i.e. an equimolar ratio). Preferably at least 3, such as at least 5, 10, 20, 30 or more analyses are performed with different ratios of the compounds. Typically in the method analyses are performed using at least 2, 3, 5, 10 or more of the specific ratios mentioned above.

Samples used in the methods of the invention

[0021] Any suitable sample from the individual may be used in the methods mentioned herein (for example a sample from a tissue or body fluid which is normally infected by the microorganism or a sample that shows cancerous growth).

The sample is typically urine, stools, whole blood, plasma, a body secretion (or a swab thereof) or a tissue sample. The sample may or may not be processed before it is analysed in the method of the invention, but in a preferred embodiment it is not processed before analysis. Such processing typically results in a removal of a component from the sample or may result in dilution of the sample (such as dilution of a swab sample). As discussed below substances such as nutrients may be added to the sample to aid growth of the microorganism. Such additions of substances are not understood herein as being "processing" of the sample.

[0022] Typically the sample (at the point of being taken from the individual) has a volume of less than 10 millilitres, such as less than 5, less than 3 or less than 3 millilitres, down to for example 25 ul or less. The sample which is analysed by calorimetry typically has a volume of less than 10 millilitres, such as less than 5, less than 3 or less than 3 millilitres,

down to for example 25 ul or less.

The donor, patient or individual from whom a sample is taken

- 5 **[0023]** The donor or patient may be from any species, and is typically a bird or mammal. The individual is preferably human or an animal. The individual may be one which is in a state that renders it susceptible to infection by the microorganism, or susceptible to cancer.

Microorganisms and cells which are used and/or detected in the methods of the invention

- 10 **[0024]** Any suitable microorganism may be used or detected in the methods of the invention. The microorganism is typically a bacterium, yeast, fungus or protozoa or virus. A tumor or malignant cell culture may be used or detected in the method of the invention.

- 15 **[0025]** The microorganism may be vancomycin resistant enterococci, pencillin resistant *Streptococcus pneumoniae*, methicillin resistant *Staphylococcus aureus*, multi-drug resistant gram-negative bacillus, multi-drug resistant *Mycobacterium tuberculosis*, multi-drug resistant gram-positive bacteria, *Achromobacter*, *acinetobacter*, *aerococcus*, *aeromonas*, *alcaligenes*, *bacillus*, *bacteroides*, *burkholderia*, *chryseobacterium*, *citrobacter*, *clostridium*, *corynebacterium*, *enterobacter*, *enterococcus*, *escherichia*, *fusobacterium*, *haemophilus*, *hafnia*, *klebsiella*, *listeria*, *micrococcus*, *moraxella*, *morganella*, *pantoaea*, *peptococcus*, *peptostreptococcus*, *proteus*, *providencia*, *pseudomonas*, *salmonella*, *serratia*, *staphylococcus*, *stenotrophomonas*, *streptococcus* multi-drug resistant malaria parasite or HIV, HSV, RSV or viruses which are members of the families Picomaviridae (e.g. polioviruses or rhinoviruses); Caliciviridae; Togaviridae (e.g. rubella virus or dengue virus); Flaviviridae; Coronaviridae; Reoviridae (e.g. rotavirus.); Bimaviridae; Rhabdoviridae (e.g. rabies virus); Orthomyxoviridae (e.g. influenza virus types A, B and C); Filoviridae; Paramyxoviridae (e.g. mumps virus, measles virus, respiratory syncytial virus or parainfluenza virus); Bunyaviridae; Arenaviridae; Retroviridae (e.g. HTLV-I ; or HTLV-II); Papillomavirus, or a tick-borne encephalitis virus.

- 25 **[0026]** The tissue could be from any organ such as liver, kidney , heart, lung etc and from any species

Conditions under which detection is performed

- 30 **[0027]** Typically detection of heat is carried out for at least 2 to 48 hours, such as 3 to 24 or 8 to 12 or 4 to 5 hours (typically after addition of the relevant substance to the calorimeter). Preferably continuous measurement of heat detection is performed, such as at least once every 10 minutes, such as over any of the above specified time periods. For example over a 24 or 48 hour period cell death can be demonstrated (and thus the bacteriocidal effect of antibiotic in vivo), or over a 2 hour period inhibition of growth (and return to baseline) or compatibility of compounds.

- 35 **[0028]** The sample may be placed in a temperature which allows growth/heat output of the cell or microorganism to be detected, such as in a temperature of 20 to 45 degrees C, typically in a temperature of 30 to 40 degrees C, such as at a temperature of 35 to 38 degrees C or at 37 degrees C.

[0029] The sample is typically placed in conditions allowing growth of the cell or microorganism for 3 to 12 hours, and typically for less than 8 hours, such as less than 6 or less than 4 hours.

- 40 **[0030]** As mentioned above in the method heat is detected (for example produced by the respiration, activity or growth of microorganisms may be detected). In one embodiment the heat produced is analysed over time, for example in the form of a graph of heat produced versus time (or power versus time). Such an analysis may allow detection of the quantity of microorganisms present in the original sample or may allow determination of the species of the microorganism. The heat data may be compared to control data to aid in ascertaining microorganism numbers and/or species.

- 45 **[0031]** The micro/nano calorimeter which is used in the method of the invention is normally one which is capable of continuously measuring heat effects (enthalpy changes) of a sample. In one embodiment the calorimeter is capable of detecting the heat output of 10^2 to 10^5 CFU/ml of microorganisms, such as of any of the species mentioned herein. Typically the calorimeter is capable of detecting a rate of heat change of 1uW or 1nW. The calorimeter may be capable of measuring heat output simultaneously from more than one sample, such as from at least 4, at least 20, at least 50 or at least 100 samples. During analysis of the heat output the sample may be in aerobic or anaerobic conditions.

- 50 **[0032]** The invention also provides a kit for carrying out a method according to any of the preceding claims comprising a calorimeter suitable for use in said method. The calorimeter may be specifically adapted so that any of the types of samples mentioned herein can be analysed and/or data concerning heat output from a sample can be provided to a computer or database for analysis of the data.

55

Detecting side effects or detrimental effects of anti-microorganism compounds

- [0033]** One aspect of the invention is based on the effect of low concentrations of anti-microorganism compounds on

the growth of cells. Such cells can be bacterial cells (such as any type or species of microorganism mentioned herein), preferably bacterial cells, or eukaryotic cells. It has been found that certain antimicroorganism compounds, especially compounds that have an effect on RNA synthesis, can cause cells to grow and thus may explain bacterial resistance (increased intracellular protein synthesis providing nutrients for the bacteria in vivo), viral infections (increased synthesis of mRNA and the expression of dormant DNA fragments, e.g. the appearance of shingles after administration of antibiotics) and also the emergence of HIV (a mixture of the two above mechanisms eventually rendering the immune system ineffective). The same effect in eukaryotic cells would give rise to tumour growth. Thus previously observed apparent resistance to anti-microorganism compounds may be due to stimulation of growth of the relevant microorganism at a concentration of anti-microorganism compound which is too low or alternatively due to stimulation of eukaryotic growth when administered in large excess or when no microorganism is present.

[0034] The effect of the anti-microorganism compound on cells is consistent with the known growth promoting effects of antibiotics. This also suggests a mechanism for anti-microorganism compounds to cause cancer.

[0035] The present work thus provides evidence of actions of anti-microorganism compounds which can cause detrimental effects in the individual being treated. Such actions will contribute for example to some of the side effects observed when using anti-infective compounds.

[0036] Accordingly the invention provides a method of determining whether an anti-microorganism compound (e.g. an anti-infective compound) is able to cause increased growth or respiration of the microorganism/virus/cell comprising adding the compound to either a respiring or growing microorganism, optionally at a pre-determined low concentration threshold, such as $< 10 \times \text{MIC}$, $< 8 \times \text{MIC}$, $< 2 \times \text{MIC}$, or $< 1 \times \text{MIC}$. A single anti-microorganism compound may be added or 2, 3, 4 or more anti-microorganism compounds may be added.

[0037] For neomycin (with a known mode of action of cause misreading during RNA synthesis) an increased heat output can be observed at concentrations of $< 1 \mu\text{g/ml}$ or in combinations with other antibiotics. For respiring E.coli all concentrations of Neomycin caused an increased heat output.

[0038] This can be used to determine whether or not growth of a microorganism/virus or cell occurs in response to the addition of the compound. Preferably growth of the microorganism/virus or cell is measured by detecting heat output from the microorganism/virus or cell using a calorimeter, such as in any of the ways discussed previously. The microorganism is generally any microorganism that causes disease, for example any such microorganism disclosed herein.

[0039] If the said compound is found to cause increased growth of microorganisms at the pre-determined low concentration, then appropriate action can be taken in the manner in which it is used, for example it can be administered to an individual suffering from a condition caused by the microorganism or at risk of such a condition, at a high dosage which would ensure killing of the microorganism. It can also be used to monitor infections in patients and thus administer anti-infective agents when necessary and stop treatment when the microorganism/virus or cell is dead (indicated by return to baseline of heat output, confirmed by viable count measurements).

Anti-microorganism compounds and agents that cause expression of latent DNA

[0040] The anti-microorganism compound may be Ampicillin, Chloramphenicol, Erythromycin, Nalidixic acid, Tetracycline, Aminoglycoside (such as streptomycin, kanamycin, gentamicin, tobramycin, amikacin, netilmicin and neomycin), tetracycline, minocycline and doxycycline, Spectinomycin, lincomycin, clindamycin, a macrolide (such as Erythromycin, clarithromycin, azithromycin or spiramycin), Fusidic acid, Rifampin, rifamycin, rifampicin, Quinolone (such as nalidixic acid, ciprofloxacin, oxolinic acid, Ofloxacin, norfloxacin, levofloxacin, lomefloxacin, sparfloxacin), sulfonamide, sulfone, Trimethoprim, methotrexate, pyrimethamine, Para-aminosalicylic acid (PSA), Dapsone, Isoniazid (INH), Spectinomycin, a Penicillin, flagyl metronidazole, Actinomycin, a sulphadiazine, vancomycin, adrenamycin, daquomycin, rifamycin, heparin, afo/3, streptogidin, or Streptovaricin.

[0041] Any of these specific compound may also be used to cause expression of latent DNA, although any suitable agent for causing DNA expression can be used, such as a polymerase enzyme (for example DNA polymerase or RNA polymerase).

Administration of agents

[0042] As mentioned above the invention includes administration of particular agents. Such an agent will generally need to be formulated into an appropriate composition to make it suitable for administration. The agent will typically be administered to an individual (human or animal) in need thereof.

[0043] The formulation will depend upon factors such as the nature of the substance and the individual to be treated. Any such substance may be administered in a variety of dosage forms. It may be administered orally, parenterally, subcutaneously, intravenously, intramuscularly, intrasternally, transdermally or by infusion techniques. The substance may also be administered as suppositories.

[0044] Typically the substance is formulated for use with a pharmaceutically acceptable carrier or diluent. The phar-

maceutical carrier or diluent may be, for example, an isotonic solution.

[0045] An effective amount of substance is administered. The dose may be determined according to various parameters, especially according to the substance used; the age, weight and condition of the individual to be treated; the route of administration; and the required regimen. A typical daily dose is from about 0.01 to 500 mg per kg, preferably from about 0.1mg/kg to 10mg/kg of body weight.

[0046] The following Examples illustrate the invention:

Examples

[0047] A micro/nano calorimeter is an apparatus that can measure the heat lost or gained during all chemical, physical and biological processes. The heat produced during these processes creates a temperature difference between the sample in the test chamber and its surroundings. The intensity of the process can thus be established through continuous monitoring of the temperature difference. The instrument used in the work is a heat conduction microcalorimeter and is designed to minimise temperature difference and encourage the most rapid heat flow away from the reaction site to a large heat sink ($\Delta T < 10^{-4}^{\circ}\text{C}$).

[0048] Micro/nano calorimetry is a non-destructive and non-invasive direct technique that does not require any prior sample treatment (like the lysis-filtration with impedance detection method). The analysis is also not limited to a physical state of a sample i.e. the calorimeter can deal with any sample form: solids, liquids, gases, transparent and non-transparent samples alike can be analysed. The colour of the sample is also not an issue using calorimetry. The calorimeter's indifference to colour and substrate nature makes it an ideal system for analysing the red, complex, suspended, cell composition of blood. Calorimetry provides a favourable advantage to the classical method of blood culturing as it can analyse very small samples of blood.

[0049] If an uninfected sample is placed in a calorimeter, one would expect there to be a clinically insignificant number of organisms present per ml (i.e. $< 10^4$ organisms/ml). It would thus take some time for the organisms in the sample to grow (to $> 10^4$ organisms/ml) and produce enough heat so that it can be detected in the calorimeter. Conversely, an infected sample will have $> 10^4$ organisms/ml. When this sample is placed in a calorimeter, it would have enough organisms to generate an immediate detectable level of heat.

[0050] The same principle can be used when assessing tissue culture and agents affecting the immune system. The sensitivity, reproducibility and ability to assess mode of action of agents has been demonstrated in an prokaryotic system both assessing growth and metabolism stimulating and repressive agents alone and in combinations

The Calorimetric Incubation Vessel

[0051] The bacterial cultures were grown in a fermenter vessel of approximately 200ml capacity which had a flat ground-glass rim. To the vertical face of the vessel close to its base, equidistantly spaced around the circumference, ports were fitted for an oxygen electrode and for sampling/inoculation. The fermenter was closed with a flat flanged glass lid which had five ports accommodating respectively, gas inlet tube, gas exhaust (via air condenser), inlet and outlet flow-lines to and from the calorimeter. The fermenter and lid was sealed with a p.t.f.e. gasket and clamped with a metal clip. The fermenter was placed on a magnetic stirring unit in a water jacket which allowed circulation of the thermostatically controlled water at 30°C or 37°C . The fermenter which contained a Teflon coated magnetic stirring bar, was sterilised by autoclaving.

The Calorimetric Incubation

[0052] For a typical calorimetric incubation the medium was passed through the calorimetric vessel at the same flow rate and temperature as those used in the experiment to establish a steady baseline deflection. The following operations then took place to a strict timetable. The liquid nitrogen stored ampoule was thawed at 38°C for 3 min and then stirred for 20 sec on a whirlimixer. The bacterial suspension (0.5-1.5ml) volumes was inoculated into the medium (100ml) two minutes after completion of thawing. A Gilson automatic pipette was used for inoculation. The calorimeter inlet flow-line was placed in the incubation medium 30 seconds after inoculation. The flowing incubation medium reached the calorimetric vessel two min after inlet flow-line insertion. At this time the recording of the p-t curves was started. Eight min after inoculation the outflow from the calorimeter was returned to the fermenter (after the "baseline" medium had run to waste but before the inoculated medium ran to waste) thus making a closed system. Aeration was started as soon as the inlet tube was inserted in the fermenter. Microbiological purity of the culture was checked by plating out a sample of the culture at the end of each incubation on nutrient agar (lab M).

[0053] The flow-microcalorimeter cannot be sterilised by conventional methods but can be decontaminated by successively pumping through solutions of 0.1 % w/v NaOH and finally sterilised deionised water. (For M. Flavus it was necessary also to use 70% w/v ethanol as well to remove any traces of peptone in the flow lines).

Presentation of Calorimetric Results

[0054] The output of calorimetric experiments is a plot of power (energy evolution per unit of time) as a function of time. The unit of power (p) is the Watt (W) and the unit of time is the hour (h). In some experiments seconds (s) were used as time unit.

Precision

[0055] The accuracy of the flow microcalorimetric assay is high. The parameter most likely to cause any variation in accuracy would be a variation in antibiotic standards. The sensitivity of this assay is also very high. This can be seen from the results of the EM's of the bacteria where no visible damage to the bacteria can be detected within the sensitivity ranges of the assay implying that very small metabolic changes can be detected in the calorimeter. This is important as conventional microbiological assays can not compete with the sensitivity which is obtained at concentrations close to those found "*in vivo*".

[0056] Flow - microcalorimetry is a very simple method to use for routine assay of antibiotics, when all the conditions discussed previously are observed. For this assay the standard deviation was $\pm 2.9\%$ which is a great improvement in reproducibilities from plate assays. (See table ref below).

Criteria	Calorimetry	Plate Method
Inter Assay Reproducibility %	3	5-10
Sensitivity u/ml	0.1	20
Range u/ml	0.1-100	20-100
Time per assay (h)	1	16
Capacity for automation	High	low
Convenience and simplicity	High	high

EXAMPLE TO DEMONSTRATE THE CAPABILITY OF THE TECHNIQUE TO ASSESS BOTH STIMULATION AND REPRESSION OF GROWTH AND METABOLISM ON RESPIRING OR GROWING CELLS.

ASSAY OF MIXTURE OF ANTIBIOTICS

[0057] Even in the earliest days of the antibiotic era, despite dramatic cures of what had previously been fatal infections, it was apparent that in certain situations the use of a single drug led to unacceptable failure rates.

[0058] The exact role of antibiotic combinations in current therapy of clinical infections often remains controversial or unclear, in part because few tightly controlled studies have been carried out in this area.

[0059] Compatibility of different anti-infectives or indeed non compatibility involves several mechanisms, including sequential blockage in a given metabolic pathway e.g. trimethoprim and sulfonamides or changes in permeability to the bacterial surface by one drug e.g. penicillin and aminoglycosides or inhibition of inactivating enzymes e.g. penicillin and cloxacillin.

[0060] Results have varied when combinations of aminoglycosides and antipseudomonas penicillins have been used against *Ps. aeruginosa* and when combinations of cephalosporins and aminoglycosides have been used against *Klebsiella* species. However it is clearly unusual for non compatibility to be suggested by one technique when another method shows compatibility of an antibiotic combination against the same organism.

[0061] In the interpretation of the results of all tests of *in vitro* antimicrobial activity, attention must be paid to the overwhelming importance of host factors in the outcome of treatment of any infectious process. *In vitro* tests do not usually take into account the role of phagocytic cells, humoral antimicrobial substances, pharmacokinetic factors and pathophysiological conditions at the site of infections. There are techniques to establish bactericidal effects in blood but these have serious microbiological limitations as regards performance and interpretation of results. The results depend on how the test was done e.g. on the salt concentration and other environmental parameters. However some correlation could be achieved between the test and the clinical outcome.

[0062] No clinical studies have been carried out in which an antibiotic combination consistently compatible with another against an infecting organism was compared to a single agent with *in vivo* activity against the infecting pathogen.

[0063] Biological compatibility and non compatibility of antibiotics can be demonstrated by micro calorimetry (see Figures 9 to 18). The antibiotics assayed in combination in this study were POLB, NEO and ZNB. These antibiotics were

EP 1 785 723 A1

available as an antibiotic spray "TRISEP" from ICI and contained, on average, NEO 500.00 u, POLB: 150.000 u and ZNB 40000 u per can. Converted to mg the content was NEO 7.5×10^2 mg, POLB 1.7×10^2 mg and ZNB 6.32×10^2 mg. When assaying antibiotics in combination one antibiotic is usually added at some multiple of its MIC, whereas the amount of the second or third antibiotic added is often empirically gauged. A simpler and more regular picture of the interaction of mixtures of antibiotics would be achieved by using equimolar concentrations or concentrations taking into account the mode of action of the antibiotics. In this study antibiotic mixtures have been assayed in the proportions found in the Trisep mixture and in equimolar concentrations. The relative molecular masses (RMM) for the three antibiotics were as follows: ZNB: RMM 1477, POLB: RMM 1300 and NEO RMM 700.

MIXTURES OF ANTIBIOTICS ASSAYED AGAINST BORDETELLA BRONCHISEPTICA (see Fig. 9)

[0064]

Bacterial Concentration <i>Bordetella Bronchiceptica</i>	Antibiotic Concentration	Observed effect on p-t curves	Comments
As % of p-t curve	Ug/ml MIC NEO 0.25 ug/ml ZNB 100ug/ml POL 2ug/ml		
50	0.25ug/ml Neo	No effect on p-t curve	
50	100ug/ml ZNB	Slightly delayed peak	
50	Pol 1 ug/ml, Neo 4 ug/ml (Trisep conc)	Sharp decline in curve 10 min after addition of antibiotics response polB 0.7 ug/ml	Non compatibility of antibiotics demonstrated
50	POL 1 ug/ml ZNB 3.57ug/ml	0.39 ug/ml POLB	Non compatibility of antibiotics demonstrated
50	All three antibiotics trisep combinations	0.47 ug/ml Pol B	2 hours after addition of antibiotic pt-power out put declined 10%, No return to baseline
50	All three antibiotics equimolar amounts	0.39 ug/ml PolB	2 hours after addition of antibiotic pt-power out put declined 20% No return to baseline
Fig 9			

MIXTURES OF ANTIBIOTICS ASSAYED AGAINST MICROCOCCUS FLAVUS

[0065] (see Fig. 10)

Bacterial Concentration <i>Micrococcus flavus</i>	Antibiotic Concentration	Observed effect on p-t curves	Comments
As % of p-t curve	Ug/ml MIC NEO 4 ug/ml ZNB 0.2ug/ml POL 2ug/ml		
0?	4 ug/ml Neo	No effect on p-t curve	
0?	2ug/ml POLB	Slightly delayed peak	
50	0.2 ug/ml ZNB 056 ug/ml POLB (trisep)	Delay in peak ZNB conc 0.11 ug/ml	Non compatibility of antibiotics demonstrated

EP 1 785 723 A1

(continued)

Bacterial Concentration Micrococcus flavus	Antibiotic Concentration	Observed effect on p-t curves	Comments
50	NEO 0.226 ug/ml ZNB 0.2ug/ml (trisep)	0.1 ug/ml ZNB	Non compatibility of antibiotics demonstrated
50	All three antibiotics trisep combinations	1.3 times standard cuve	Growth promoting effect
50	All three antibiotics equimolar amounts	0.1 ug/ml ZNB	Small delay in peak compared to standard curve
Fig 10			

MIXTURES OF ANTIBIOTICS ASSAYED AGAINST BACILLUS PUMILUS

[0066] (see Fig. 11)

Bacterial Concentration Bacillus Pumilus	Antibiotic Concentration	Observed effect on p-t curves	Comments
As % of p-t curve	Ug/ml MIC NEO 1 ug/ml ZNB 20 ug/ml POL 20 ug/ml		
0?	ZNB 20 ug/ml	No effect on p-t curve	
0?	20ug/ml POLB	Within 9 hours growth was inhibited completely	
0	0.5 µg/ml) NEO + 0.125 µg/ml (0.006 MIC) POLB (TRISEP)	Compatibility Effect 1ug/ml neomycin	compatibility of antibiotics demonstrated
0	And 0.25 MIC (0.5 µg/ml) NEO + 0.44 µg/ml 0.022 MIC ZNB (TRISEP). (trisep)	Compatibility Effect 1 ug/ml neomycin	compatibility of antibiotics demonstrated
0	All three antibiotics trisep combinations	1.0 times standard cuve	No effect
0	All three antibiotics equimolar amounts neomycin 1 ug/ml, POLB, ZNB	1.7ug/ml Neomycin	Compatibility demonstrated
Fig 11			

DISCUSSION

[0067] Compatibility was clearly demonstrated for all antibiotic combinations against *B. pumilus* except the Trisep mixture. These studies were all carried out at pH 8.00 as this was the most effective pH for neomycin. It seems likely that when using the Trisep mixture POLB and ZNB interfered with each other whereas an equimolar concentration no such effect was seen. The results suggest a critical range of concentration within which compatibility is observed.

[0068] When assaying the three antibiotics against *Bord. Bronch* and *M. flavus* no such effect could be observed. In fact an indication of non compatibility could be observed for all these mixtures, and even a growth promoting effect at lower concentrations of antibiotics. It seems likely that the compatible action against *Bacillus pumilus* was due to POLB and ZNB facilitating the transport of neomycin into the cytoplasm by disrupting the bacterial membrane. All the above

EP 1 785 723 A1

results have been verified by electron microscopy (see Figs. 12 to 14). It has been shown by EM that ZNB had some effect on *E. coli* as protrusions have been observed on the bacterial membrane after treatment with ZNB.

[0069] A possible assay of POLB and ZNB in the combinations used here could probably be achieved using *Bord. bronch* and *M. flavus* respectively as test organism. The assays would not be as sensitive as the assays for the individual antibiotics but a linear relationship between dose and response may be achievable. For the neomycin assay, using *B. pumilus* as test organism, a great variation in concentration of the antibiotics would have to be carried out to see at which concentrations, if any, an assay of all three antibiotics would be possible i.e. that an assay could be developed for neomycin with a constant response for ZNB and POLB.

RESPIRATION AND GROWTH OF *E. COLI*

[0070] *E. coli* was suspended in 10% DMSO prior to freezing and the viable counts were for batch 1: pre-freeze: $4.0 \pm 0.6 \times 10^{10}$ cfu/ml and post-freeze $2.0 \pm 0.44 \times 10^{10}$ cfu/ml and for batch 2: pre-freeze: $3.45 \pm 0.58 \times 10^{10}$ cfu/ml and post-freeze $2.93 \pm 0.43 \times 10^{10}$ cfu/ml. The freezing rate for both batches was 20°C/min the survival rate was 50% for batch 1 and 85% for batch 2. These differences may have been due to batch 1 being kept for 24h at 4°C before being frozen.

[0071] The respiration experiments were carried out in glucose buffer at different pH. The respiration curves obtained from batch 1 (inoculum 2×10^{10} cfu/ml) and batch 2 Inoculum 3.2×10^{10} cfu/ml) were essentially similar and a mean was used as a reference). The respiration experiments were carried out at pH 7 for polymyxin and Znbacitracin and at pH 8 for neomycin. No differences in p-t control curves could be observed at pH 7 or pH 8.

[0072] The p-t curves rose and reached a plateau as has been observed for respiration of yeasts and some 20 min later fell to a lower plateau value. Prior incubation of the cells in the buffered glucose for periods up to 20 min before presentation to the calorimeter could reduce or entirely remove this effect.

INTERACTION OF POLYMYXIN B, NEOMYCIN AND ZNBACITRACIN WITH RESPIRING

[0073] *E. COLI*

Bacterial Concentration <i>E. coli</i>	Antibiotic Concentration	Observed effect on p-t curves	Comments
As % of p-t curve	Ug/ml MIC&MBC NEO 5900 ug/ml ZNB > 1000 ug/ml POL 6.3ug/ml		
Respiration	POLB 6.3 ug/ml	Reduced height of p-t curve initially and returned to baseline within 1 hour	Bacteriocidal action demonstrated
Respiration	NEO 500 ug/ml	Increase in heat output Linear dose-response between 10-100ug/ml	Consistent with increased rate of RNA synthesis
Respiration	ZNB 300 ug/ml - 1000 ug/ml	No response	Resistance demonstrated
Respiration	: POLB: 20µg/ml NEO: 80.8µg/ml and ZNB 71.4µg/ml (TRISEP combination).	POLB-NEO longer plateau drop after 20 min to baseline POLB ZNB NEO ZNB NEO response curve	POLB/NEO interaction: delayed response compared to polymyxin alone but drop to baseline no increase in power NEO/ZNB no interference of ZNB with NEO action higher output but not as high as with NEO alone
Respiration	All three antibiotics trisep combinations	Polymyxin curve with delayed return to baseline 20 min	Effect not as good as polymyxin alone or Neomycin alone

(continued)

Bacterial Concentration E. coli	Antibiotic Concentration	Observed effect on p-t curves	Comments
Respiration	All three antibiotics equimolar amounts	Polymyxin curve with delayed return to baseline 9 min	Effect not as good as polymyxin alone or Neomycin alone

[0074] Fig 15,16 and 18

Interaction of Polymyxin B with E.coli

[0075] The MIC and MBC for E.coli was found to be 6.3 µg/ml. Polymyxin B reduced the initial peak height and thereafter the p-t curve returned to baseline (Fig. 15). This effect was in accordance with the mode of action of polymyxin as the bacteriocidal action was very rapid. This was also observed when using Bord. Branch as test organism.

Interaction of Neomycin with E.coli

[0076] The effect of neomycin on E.coli is given in Fig. 16. In contrast to polymyxin, neomycin caused an increase in power which confirms that Neomycin fundamentally stimulates RNA synthesis and only inhibits it when present in very large amounts when added to respiring E.coli. (MIC: 500 µg/ml). A linear relationship between dose and the reciprocal of increase in response from the control curve was found between 20-100 µg/ml neomycin.

Interaction of Znbacitracin with E.coli

[0077] No response could be observed on addition of Znbacitracin to respiring E.coli. The highest concentration added was 300 µg/ml (MIC > 1000 µg/ml). ZNB has been reported to be inactive against E.coli. However electron micrographs have shown that ZNB had an observable effect on E.coli i.e. did actually cause protrusions to occur on the bacterial membrane. This response was not lethal for the bacterium but small changes in heat could possibly have been expected to occur. This has in fact been observed when ZNB was added to growing cultures of Bord. branch. a small plateau was reached before growth reoccurred (see Fig. 17).

Interaction of Mixtures of Antibiotics

[0078] Fig. 18 shows the p-t curves for the effect of mixtures of antibiotics on E.coli. The concentrations were: POLB: 20 µg/ml NEO: 80.8 µg/ml and ZNB 71.4 µg/ml (TRISEP combination). Fig. 18 shows the p-t curve for ZNB + NEO; when compared with the p-t curves for neomycin alone it can be seen that this p-t curve was essentially similar to that for neomycin alone (20 µg/ml NEO). Fig. 18 shows the effect of POLB + NEO on E.coli. When comparing the effect of the mixture to that of POLB alone (Fig. 15) it can be seen that the effect of the two antibiotics was not comparable to the polymyxin interactions as the kinetics of the interactions seemed to have changed resulting in a plateau value for much longer periods than found for the polymyxin alone followed by a decline to a baseline value after 20 min. When compared to the neomycin interactions (Fig. 16) the p-t curve for NEO and POLB was very different.

[0079] Fig. 18 shows the effect of polymyxin and Znbacitracin in combination and when compared with that for polymyxin alone (Fig. 9) the p-t curve is seen to be similar in shape to that for polymyxin but shows a higher output. This response was equivalent to a response of 15 µg/ml polymyxin and suggests non compatibility.

[0080] Fig. 18 shows the effects of all three antibiotics added to respiring E.coli (A) as the TRISEP mixture and (B) as the equimolar mixture. The TRISEP mixture shows a response similar to that of POLB + NEO apart from showing a decline to baseline which was at a slower rate whereas the equimolar mixture at (20 µg/ml POLB) showed a polymyxin type curve with a plateau value very similar to that of the control curve, but then, after 9 min, a steady decline to baseline.

[0081] The heat effects observed with polymyxin B were those that would be expected of membrane active antibiotics; i.e. severe reduction in power output followed by a return to baseline signifying death or lysis of cells after a breakdown in membrane potential and leakage of cell constituents from the cytoplasm of the cell. The polymyxin type interactions showed kinetics which were essentially simply and which have been observed with antifungal antibiotics. Neomycin, in contrast, caused an increase in power output also observed in growth studies with *Basillus Pumilus* which would indicate an immediate effect on RNA synthesis and a very good reason not to use this antibiotic

[0082] That ZNB had no effect on power output was not surprising since the antibiotic has been reported to be active only against Gram positive organisms. The assays for POLB were not particularly sensitive (3 µg/ml) but had the advan-

tage of being rapid. They do compare favourably with the assay using Bord. Bronch. as test organism for this reason. Which assay to use would depend entirely on the sensitivity required. The reproducibilities for the E.coli assay were also smaller than those for the Bord. bronch. assay. The neomycin assay was much more sensitive when using B.pumilus as test organism. However the assay with E.coli was possible at concentrations much lower than the measured MIC (500 µg/ml). It can be concluded that these two assays would be suitable for screening antibiotics where sensitivity is not essential, since they are very rapid (1h).

[0083] It can be concluded from Fig. 18 that all the antibiotics assayed together show (two by two) possible antagonistic effects. The effect observed with neomycin and polymyxin was an initial increase in peak deflection followed by a gradual decrease to baseline. This suggested that the initial effect on E.coli was due to neomycin followed by the effect of polymyxin. For POLB + NEO assayed against E.coli it can be observed that the rate of decline of power was identical to that noted for the antibiotic on its own suggesting that these interactions followed the same order of kinetics at this point. That polymyxin and bacitracin assayed together showed non compatibility was not surprising since a possible explanation could be that competition for binding sites on the bacterial membrane would occur. This effect was also in agreement with the effect observed for this mixture when assayed against Bord. bronch. and M. flavus. This possible competitive effect could result in the inhibition of binding of the most active agent. When all three antibiotics were added together either as the TRISEP combination or as the equimolar combination, p-t curves resulted that were different from those observed either for the antibiotics added singly or in binary combinations.

[0084] These results form an excellent basis for rapid compatibility testing of anti-infective agents both during development and also as part of clinical practise in hospitals, doctors offices, dental clinics, veterinary clinics and in development countries

Claims

1. Method of determining whether or not an organ for transplantation (from a human donor) or xenotransplantation (from an animal donor) is infected with a microorganism (which is optionally a virus) comprising
 - measuring heat output from the organ to detect activity of said microorganism, or
 - forcing expression of latent DNA by using agents that stimulate m-RNA synthesis,
 to thereby determine the presence of a microorganism in the organ.
2. Method of determining the suitability of a donor organ for transplantation to a patient comprising adding in vitro a portion of the organ to a fluid or tissue sample from the patient and measuring heat output from the sample using a calorimeter to thereby detect whether or not there is a rejection reaction against the organ.
3. Method of determining whether or not a candidate compound has anti-cancer activity comprising adding the compound to a cancer cell sample in vitro and measuring the heat output from the cancer cell using a calorimeter to thereby determine whether or not the candidate compound has anti-cancer activity.
4. Method for determining whether an anti-microorganism compound affects the growth of a mammalian or avian animal comprising adding said compound to cells of said mammal and determining whether detecting the rate of growth of the cells by measuring the heat output from the cells using a calorimeter.
5. Method of determining a suitable administration regimen for a patient comprising taking a sample from a patient on a first administration regimen and detecting the effectiveness of the first administration regimen by measurement of the immune response, microorganism load or numbers of malignant cells in a sample from a patient on the first administration regimen, wherein measurement is performed by using calorimetry, and then optionally if the first administration regimen is found to be of low effectiveness selecting a second administration regimen for the patient
6. Method according to claim 5 wherein the first and second administration regimen differ in respect to the drug or combination of drugs which are administered.
7. Method according to claim 5 or 6 wherein the patient is diagnosed to be HIV positive and antiretroviral agents are used or being developed.
8. Method of determining whether an anti-microorganism or anti-infective compound is able to cause growth of the microorganism/virus/mammalian cell comprising adding the compound to the microorganism, optionally at a pre-

determined low concentration threshold, such as $< 10 \times \text{MIC}$, $< 8 \times \text{MIC}$, $< 2 \times \text{MIC}$, $< 1 \times \text{MIC}$ to determine whether or not growth or heat output of the microorganism/virus/mammalian cell occurs in response to the addition of the compound, wherein growth of the microorganism is optionally measured by detecting heat output from the microorganism/virus/mammalian cell using a calorimeter, and wherein the compound may be alone or in a combination with 2 or more other anti-microorganism or anti-infective agents.

9. Method according to claim 8 wherein if the said compound is found to cause increased growth of microorganisms at the pre-determined low concentration, then administering the compound to an individual suffering from a condition caused by the microorganism or at risk of such a condition, at a high dosage, but optionally not at dosages causing increased respiration of the said compound.
10. Method according to any one of the preceding claims wherein the donor or patient is a human, animal, mammal, primate, bird, domestic pet, agricultural animal, wild animal, sporting animal or fish or where the sample or cell is from an individual of any such organism.
11. Method according to any one of the preceding claims wherein the donor or patient is of any of the following species, or where the sample or cell is from an individual of any of the following species: horse, pig, cow, dog, cat, sheep, goat, turkey, chicken, camel, llama, deer, duck, fish, monkey, mouse, rat, guinea pig or hamster.
12. Method according to any one of the preceding claims wherein heat output is measured using calorimeter able to detect changes in heat flow of 25 nanocalories/sec and/or heat effects of 10 microcalories.
13. Kit for carrying out a method according to any of the preceding claims comprising a calorimeter suitable for use in said method.

Fig. 1

Interaction of Antibiotics with *Bacillus Pumilus*

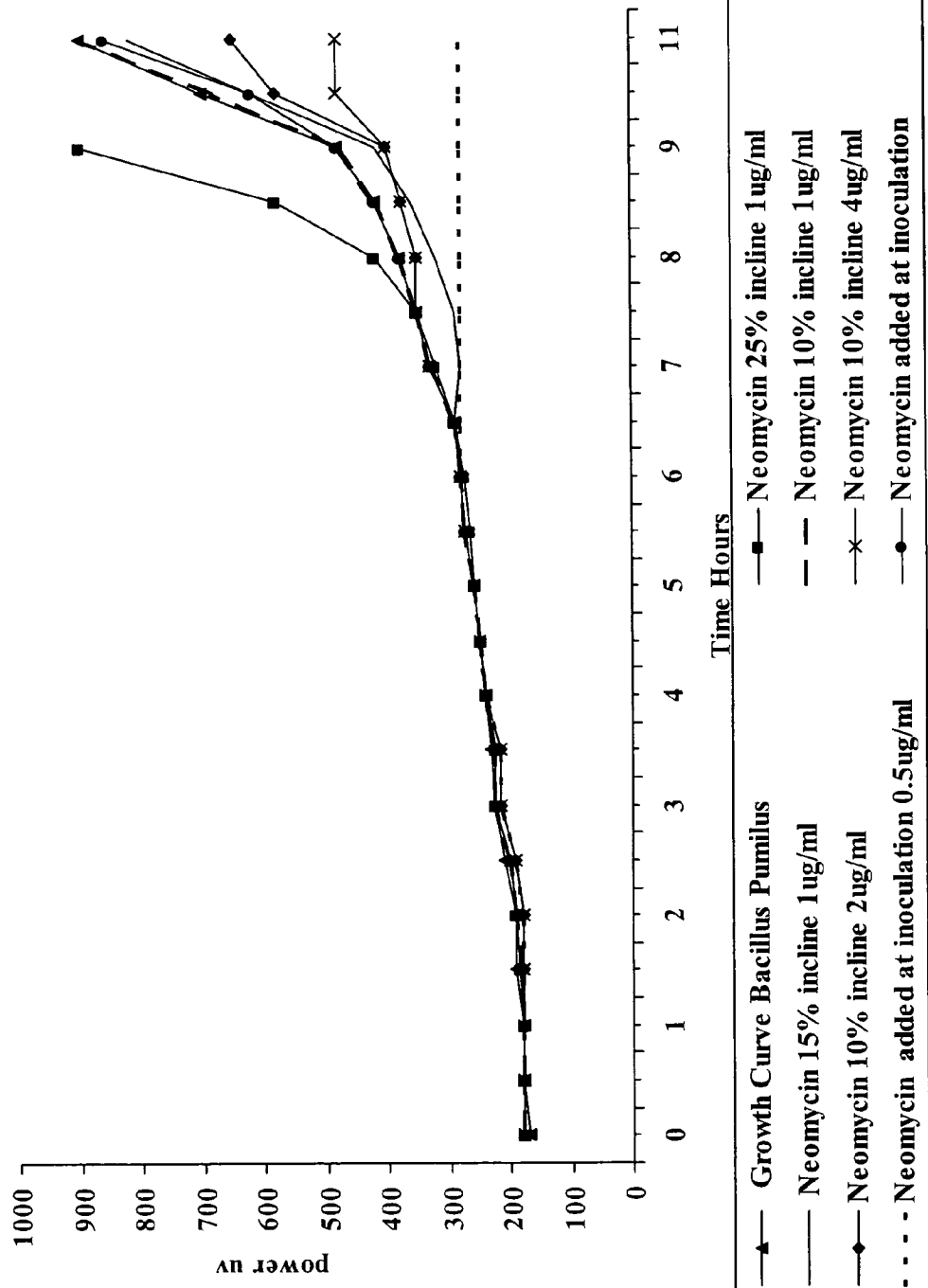
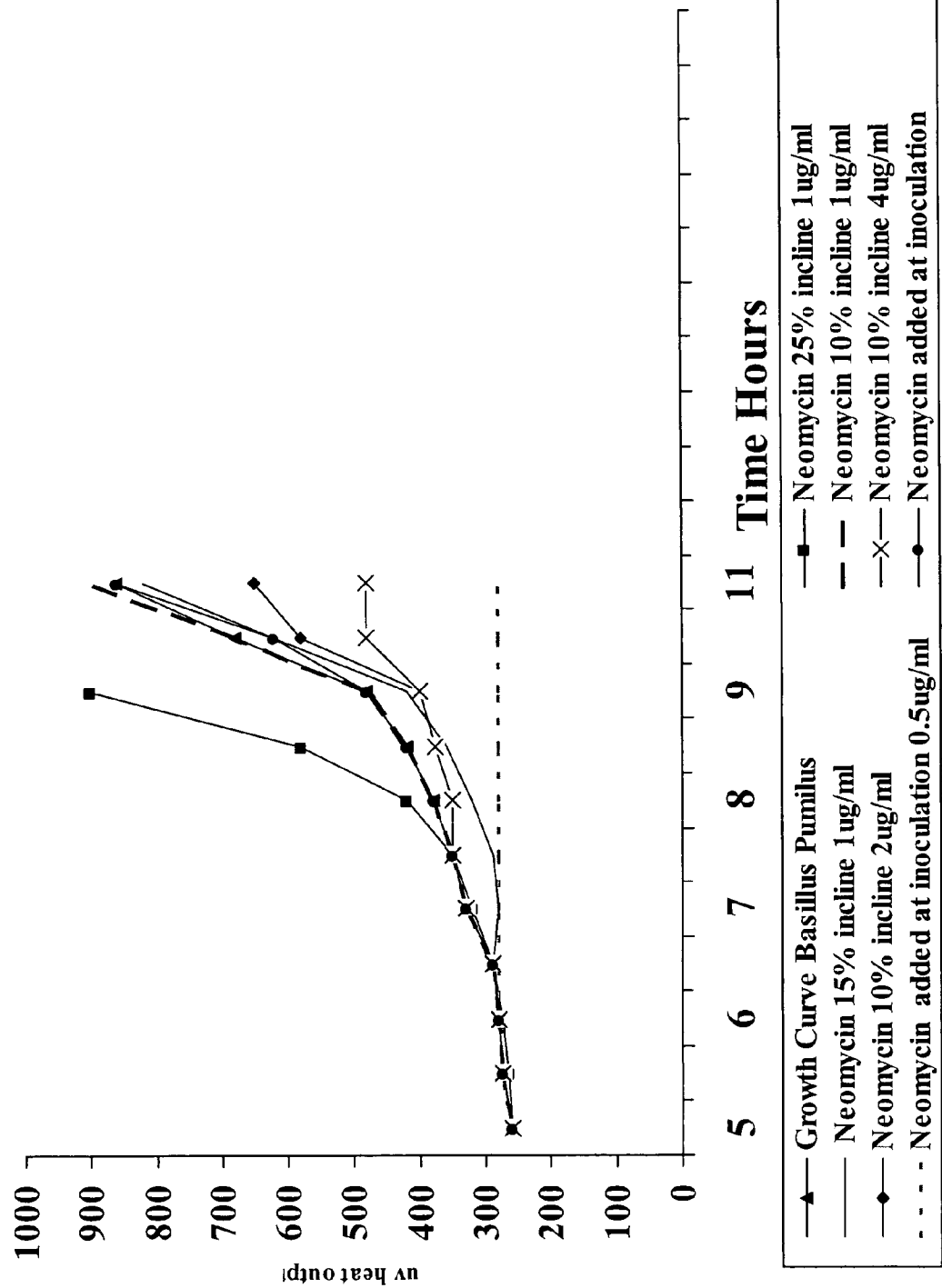


Fig. 2

Interaction of Antibiotics with Bacillus Pumilus 5 – 11 hours growth



Interaction of Antibiotics with *Micrococcus Flavus*

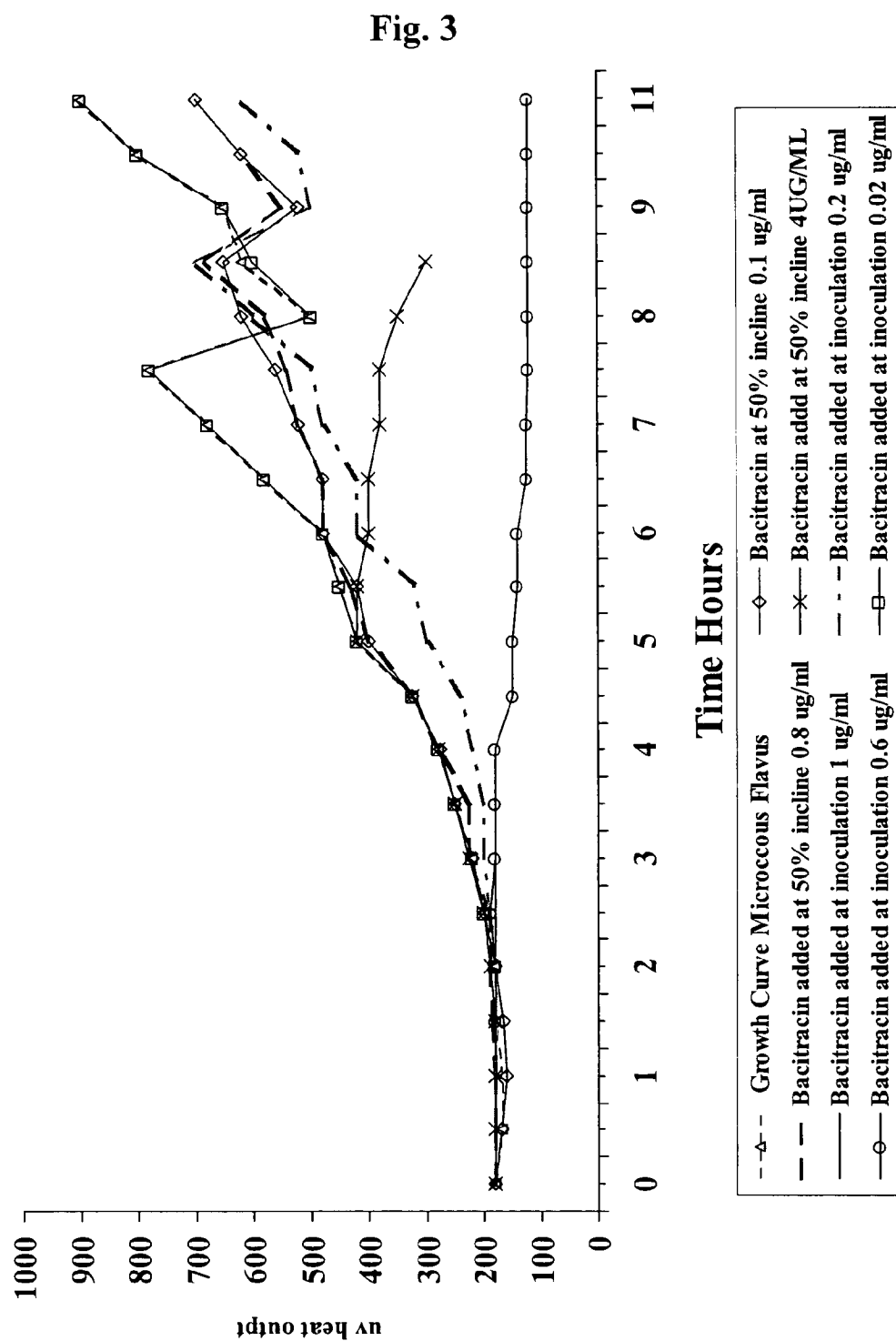
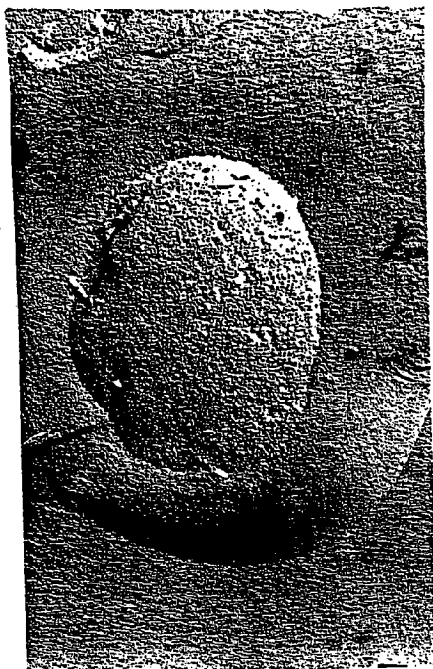


FIG 4

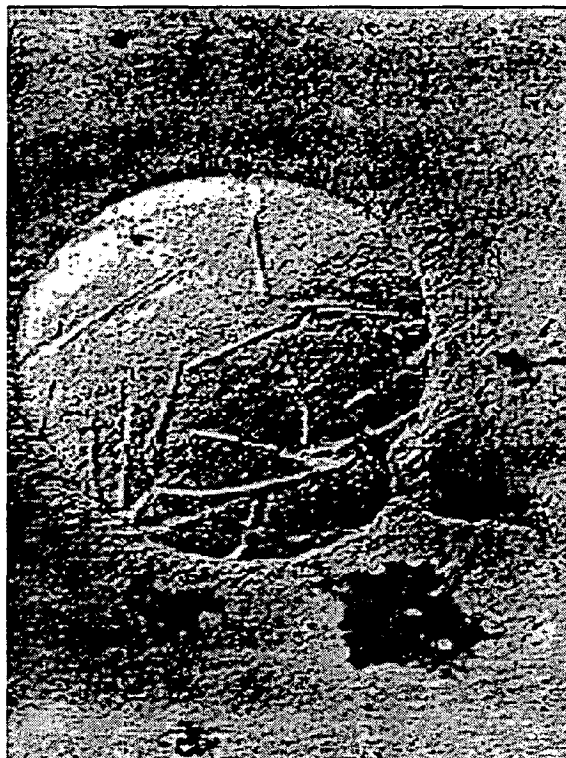


FROZEN
Magnification x 43,000

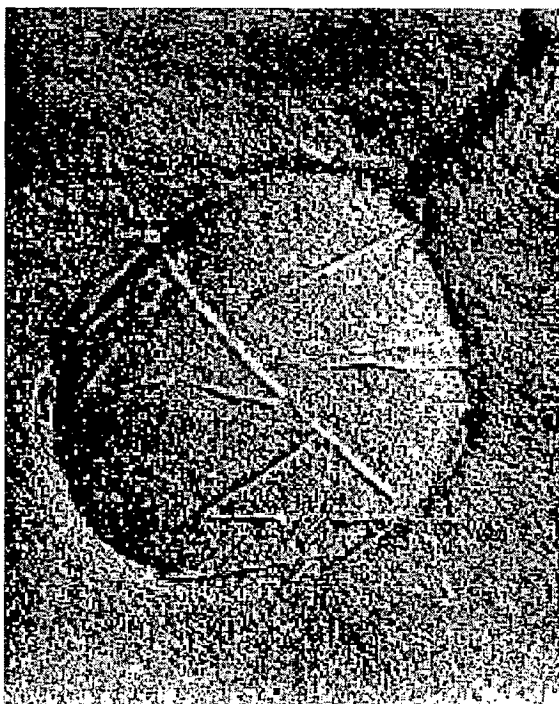


FRESH
Magnification x 43,000

Fig. 5



Bacitracin treated
Micrococcus flavus
(frozen)
0.025 x MIC



Bacitracin treated
Micrococcus flavus
(fresh)
0.025 x MIC

Fig. 6

Interactions of AntiBiotics with Bordetella bronchiceptica

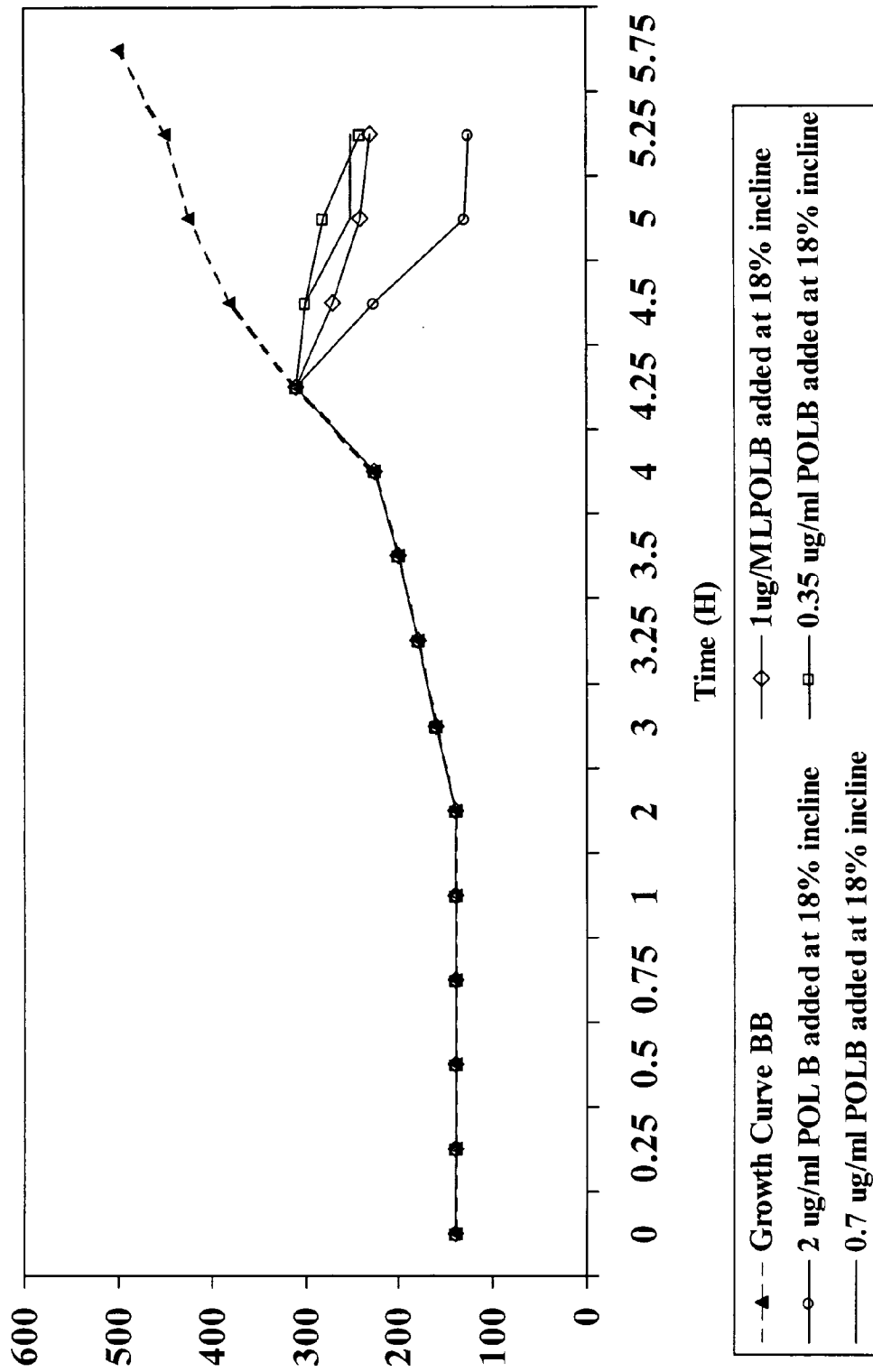
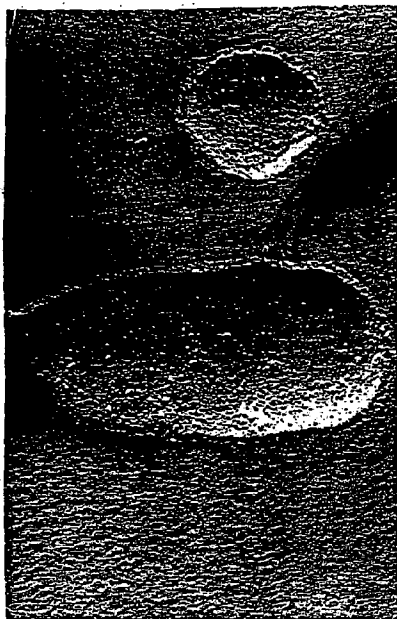


FIG 7



FROZEN
Magnification x 59000

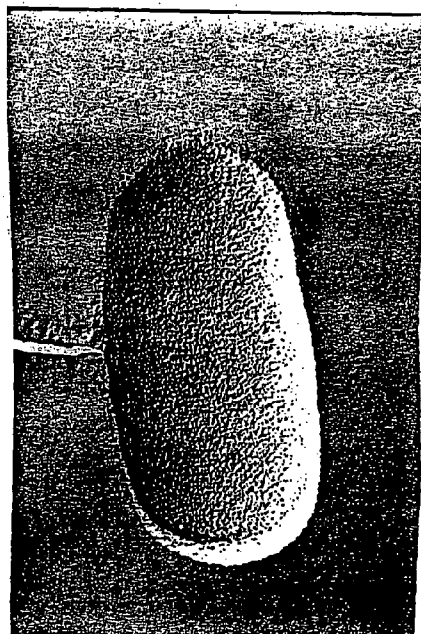


FRESH
Magnification x 75000

FIG 8



Polymyxin treated
Bord bronch (frozen)
1µg/ml
Magnification x 75000



Polymyxin treated
Bord bronch (fresh)
1µg/ml
Magnification x 59000

Fig. 9

Interactions of AntiBiotic mixtures with Bordetella bronchiceptica

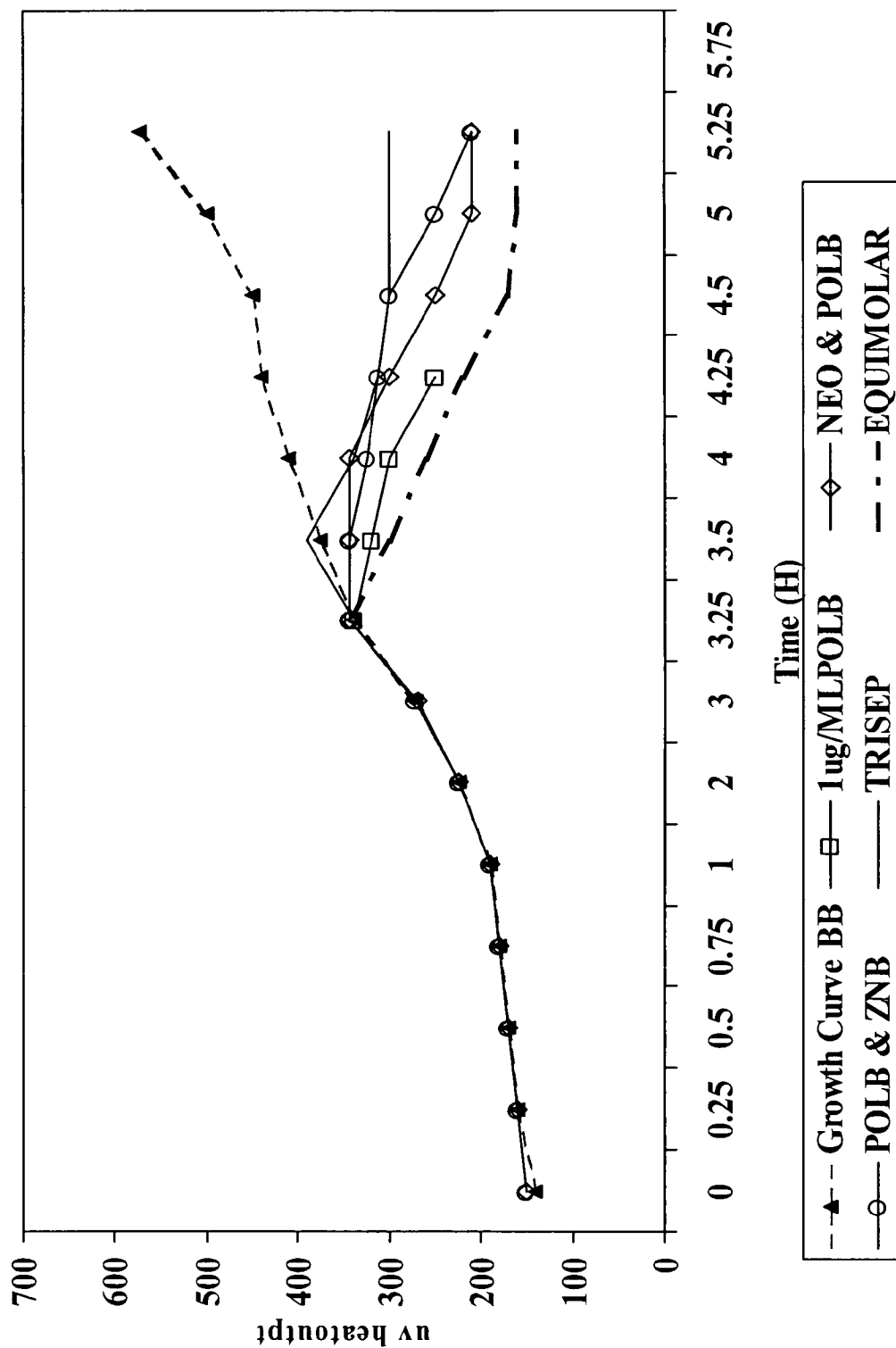


Fig. 10

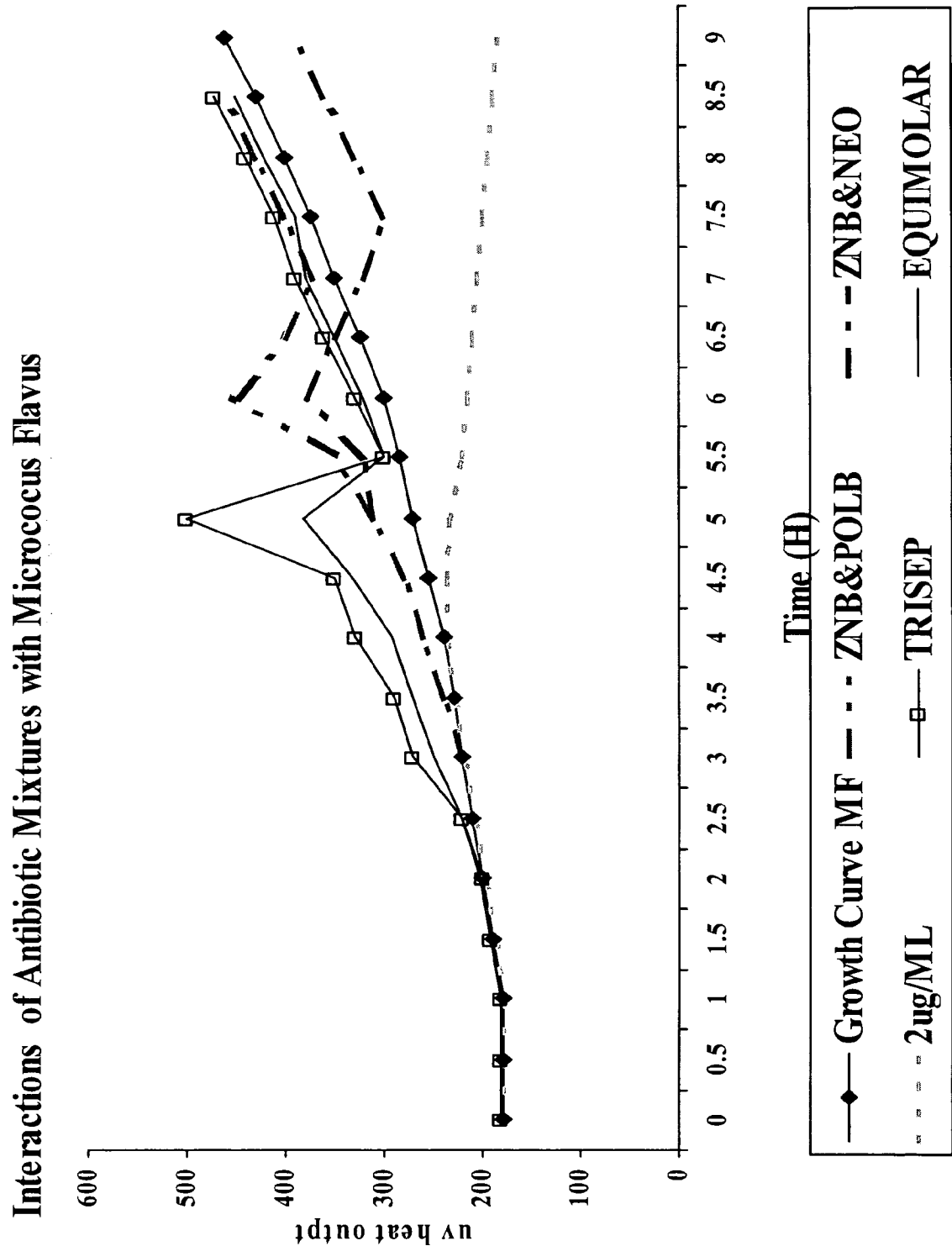


Fig. 11

Interaction of Antibiotic Mixtures with *Bacillus Pumilus*

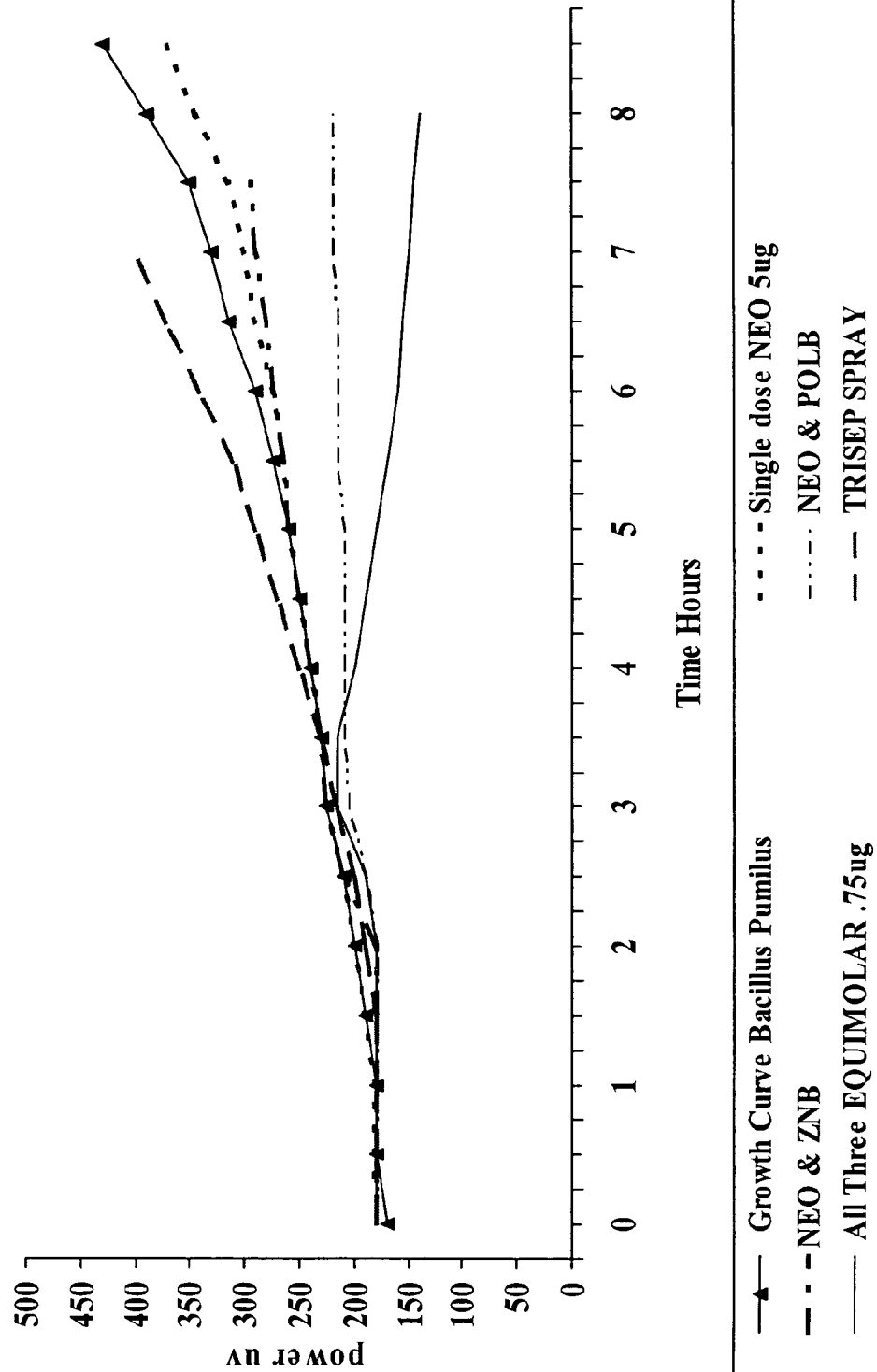
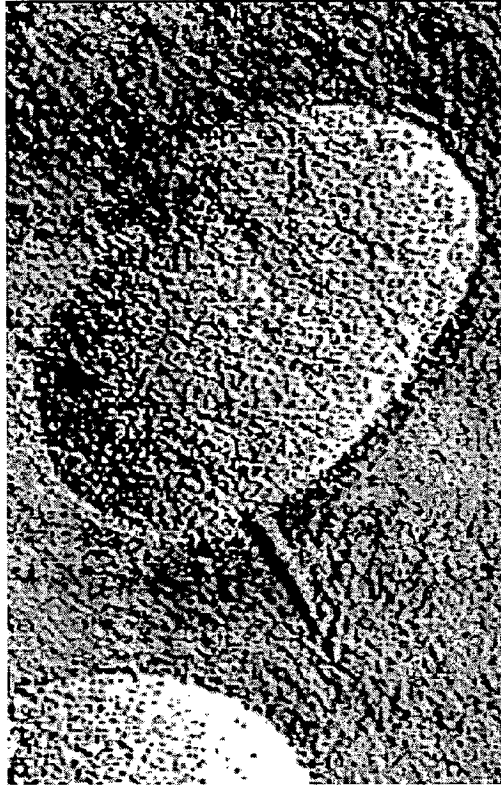


Fig. 12



**EM for Equimolar combinations
against Bordetella
bronchiceptica**



**EM for TRISEP combinations
against Bordetella
bronchicptica**

Fig. 13

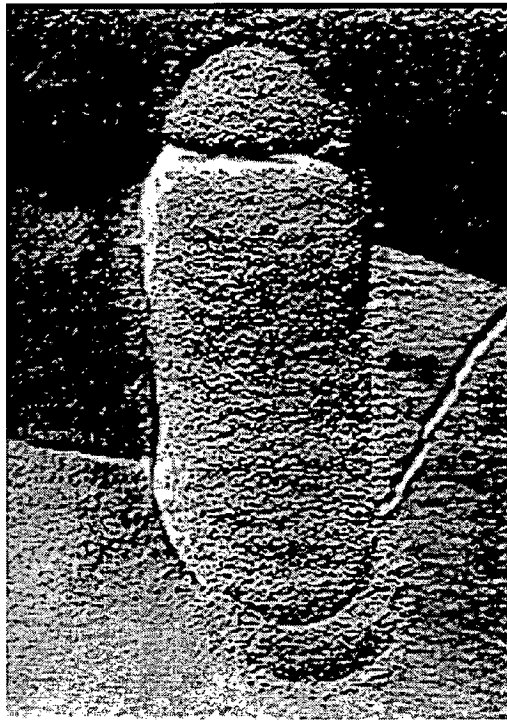


**EM for Equimolar
combinations against
Micrococcus flavus**



**EM for TRISEP
combinations against
Micrococcus flavus**

Fig. 14



**EM for Equimolar
combinations against
Bacillus pumilus**



**EM for TRISEP
combinations against
Bacillus pumilus**

Fig. 15

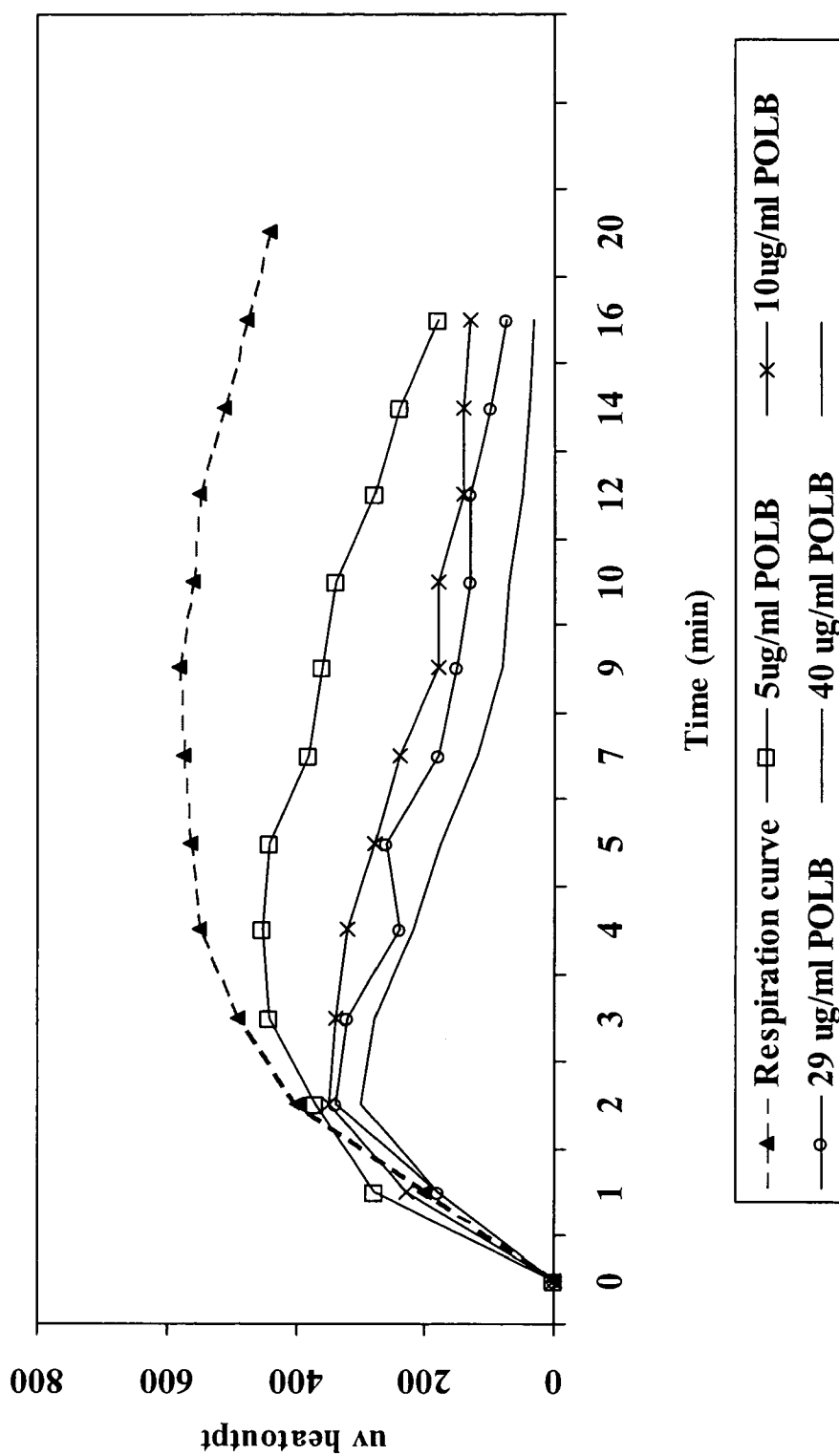
Interactions of Polymyxin B with *Escherichia coli*

Fig. 16

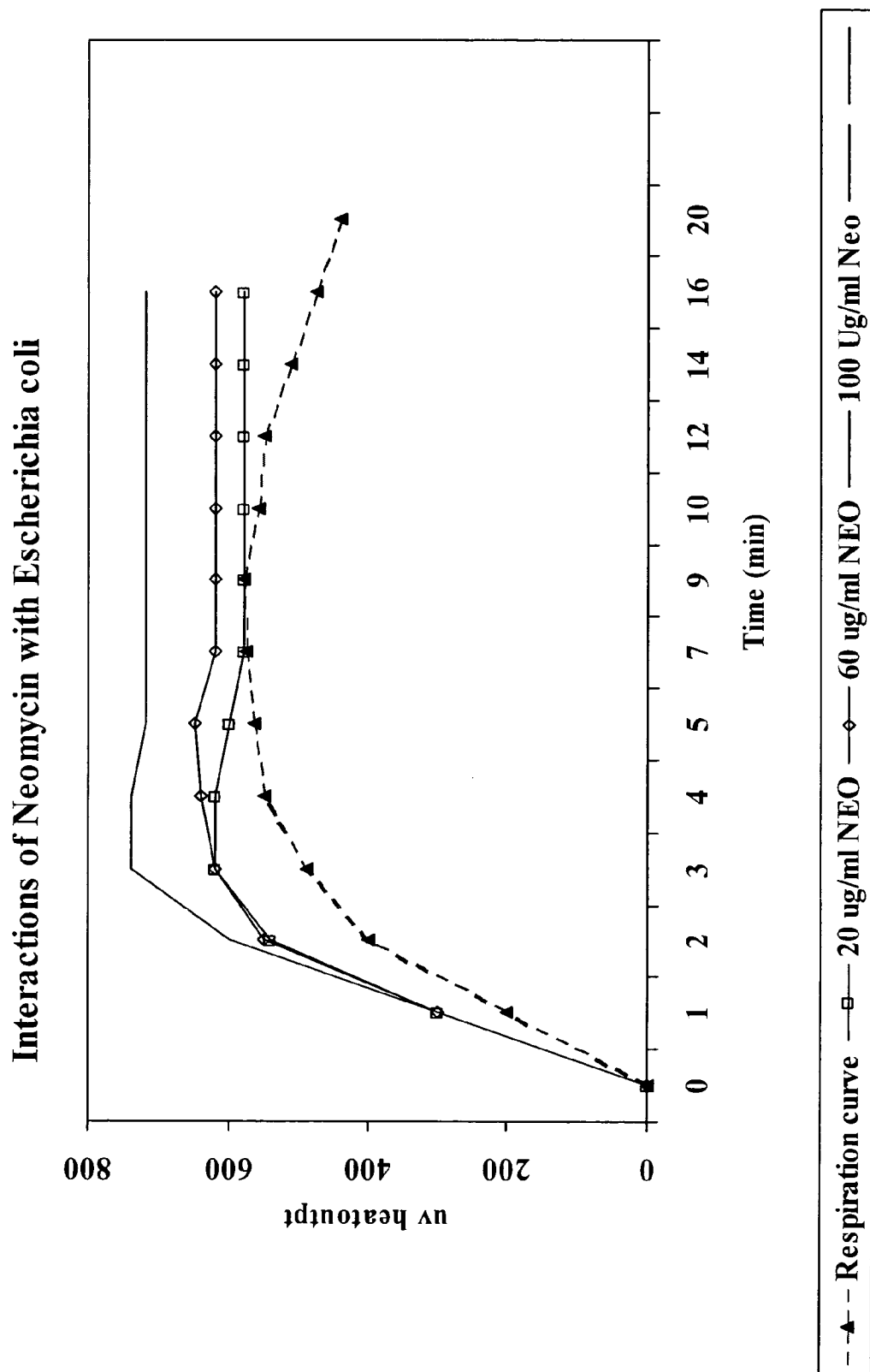


FIG 17

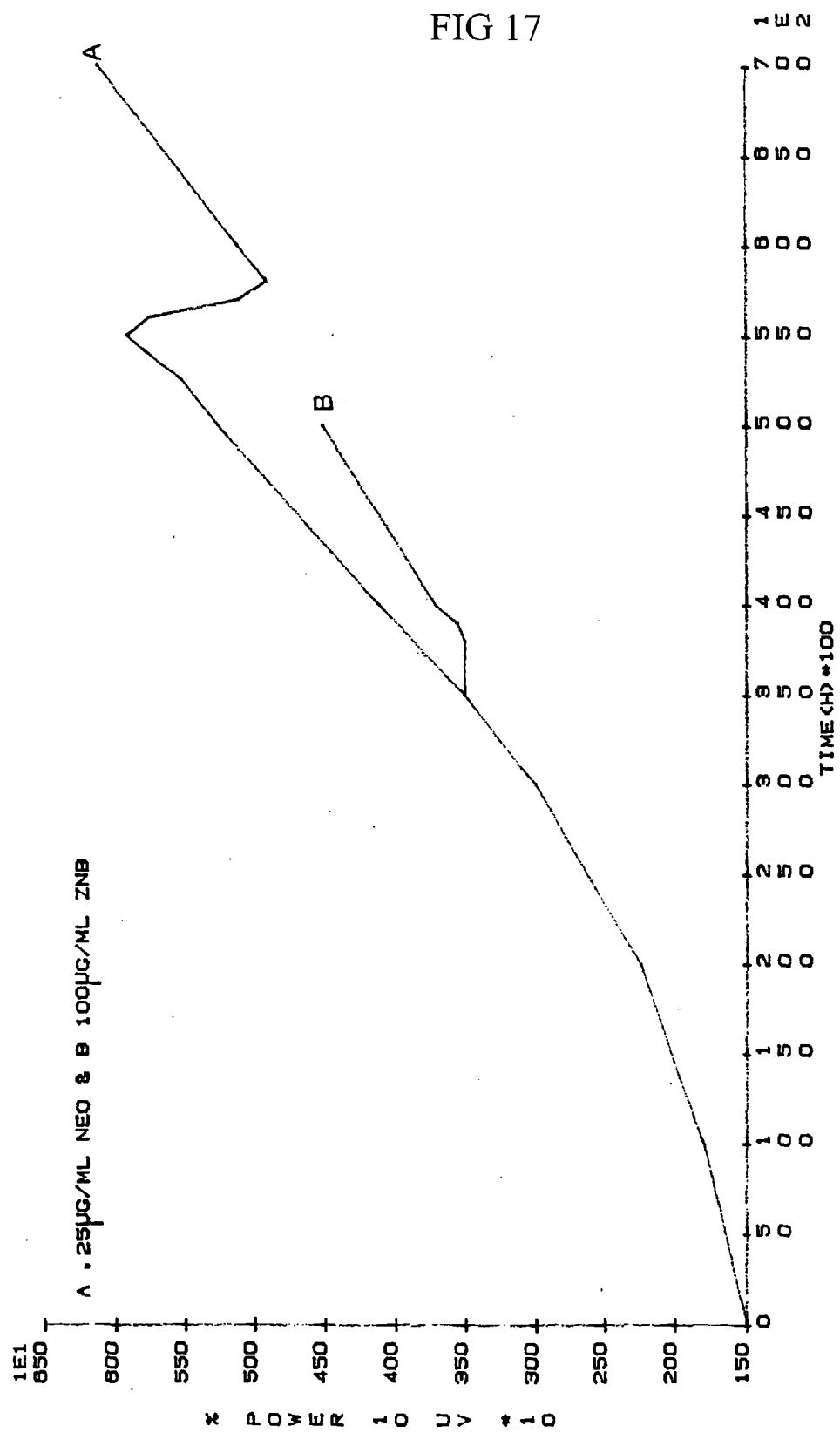
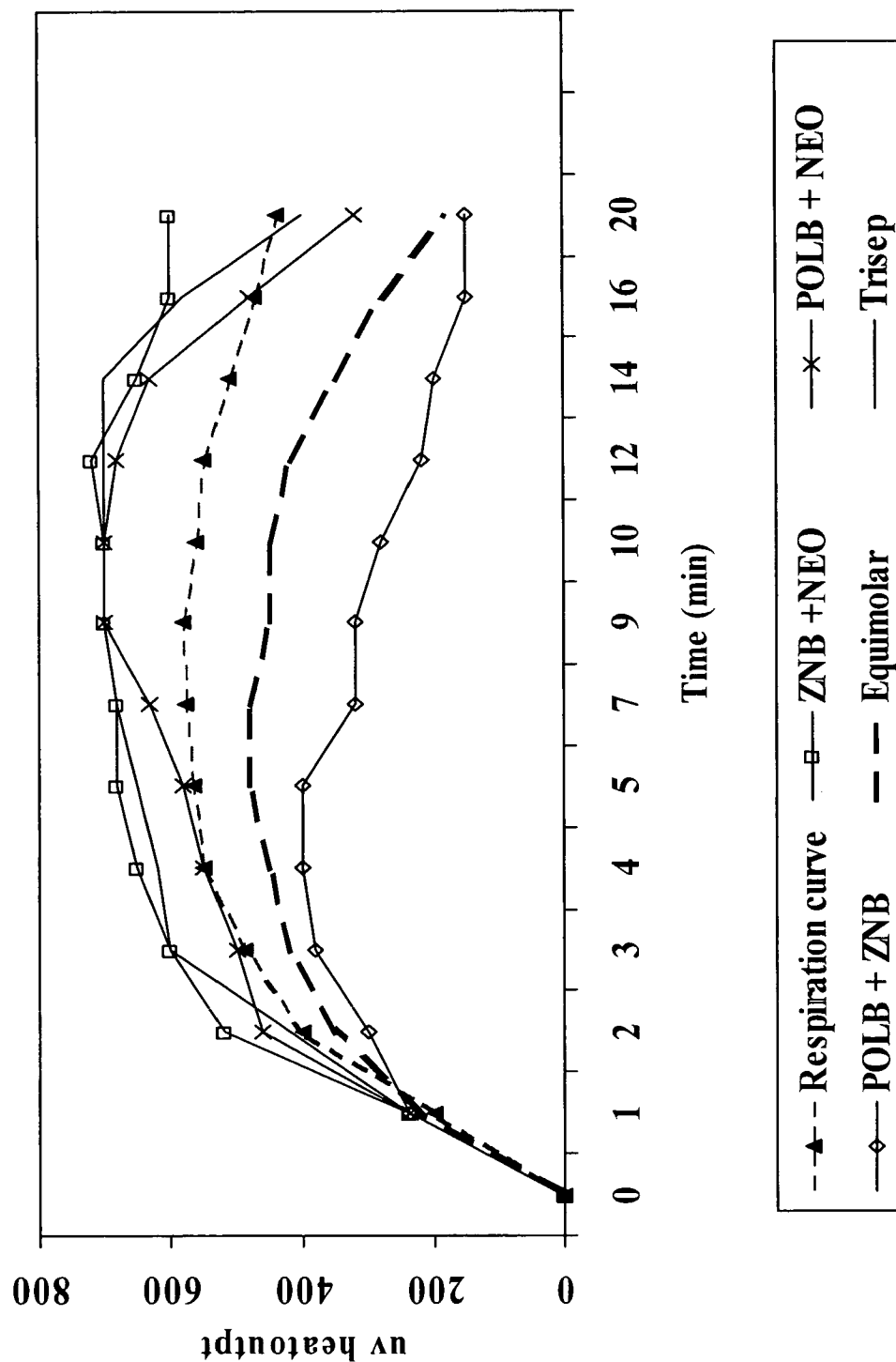


Fig. 18

Interactions of Mixtures of Antibiotics with *Escherichia coli*





European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 05 25 6977

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
Y	TUCKER ALEXANDER W ET AL: "Control of microbial contamination during surgical harvest of pig renal xenografts." XENOTRANSPLANTATION, vol. 11, no. 1, January 2004 (2004-01), pages 91-96, XP002368284 ISSN: 0908-665X * the whole document *	1,10-12	INV. G01N25/20 G01N33/48 G01K17/00 C12Q1/04 A61B5/00
X	US 3 765 237 A (BLACKMER D,US) 16 October 1973 (1973-10-16)	13	
Y	* column 1, lines 4-19 * * column 2, lines 34-39 * * column 5, line 64 - column 6, line 3 * * abstract *	1,10-12	
A	FISHMAN J A: "Xenotransplantation and infection: Opportunity or apocalypse?" ABSTRACTS OF THE INTERSCIENCE CONFERENCE ON ANTIMICROBIAL AGENTS AND CHEMOTHERAPY, vol. 41, 2001, page 532, XP008060134 & 41ST ANNUAL MEETING OF THE INTERSCIENCE CONFERENCE ON ANTIMICROBIAL AGENTS AND CHEMOTHERAPY; CHICAGO, ILLINOIS, USA; SEPTEMBER 22-25, 2001 * the whole document *	1,10-12	TECHNICAL FIELDS SEARCHED (IPC) G01N G01K C12Q A61B
X	US 6 436 346 B1 (DOKTYCZ MITCHEL J ET AL) 20 August 2002 (2002-08-20)	13	
A	* column 1, lines 22-25 * * column 2, lines 6,7,65-67 * * column 3, lines 61,62 * * column 8, line 54 - column 9, line 9 *	1,10-12	
X	US 3 298 790 A (BENZINGER THEODOR H) 17 January 1967 (1967-01-17) * column 1, lines 17-26 * * column 3, lines 57-63 *	13	
1 The present search report has been drawn up for all claims			
Place of search Munich		Date of completion of the search 17 February 2006	Examiner Filipas, A
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			

EPO FORM 1503 03 82 (P04C01)

**CLAIMS INCURRING FEES**

The present European patent application comprised at the time of filing more than ten claims.

- ☐ Only part of the claims have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claim(s):
- ☐ No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.

LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

see sheet B

- ☐ All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
- ☐ As all searchable claims could be searched without effort justifying an additional fee, the Search Division did not invite payment of any additional fee.
- ☐ Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims:
- ☒ None of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims:
- 1 (first alternative), 10-13



The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

1. claims: 1(first alternative), 10-13

Method of determining whether or not an organ for transplantation is infected with a microorganism, by measuring heat output from the organ,
and
Kit for carrying out said method, comprising a calorimeter.

2. claims: 1(second alternative), 10-13

Method of determining whether or not an organ for transplantation is infected with a microorganism, by forcing expression of latent DNA by using agents that simulate m-RNA synthesis,
and
Kit for carrying out said method, comprising a calorimeter.

3. claims: 2,10-13

Method of determining the suitability of an organ for transplantation to a patient, by adding in vitro a portion of the organ to a fluid/tissue sample from the patient and measuring heat output using a calorimeter to detect whether or not there is rejection against the organ,
and
Kit for carrying out said method, comprising a calorimeter.

4. claims: 3,10-13

Method of determining whether or not a compound has anti-cancer activity, by adding the compound to a cancer cell sample in vitro and measuring heat output using a calorimeter,
and
Kit for carrying out said method, comprising a calorimeter.

5. claims: 4,10-13

Method for determining whether an anti-microorganism compound affects the growth of a mammalian/avian animal, by adding said compound to cells of said animal and detecting the rate of growth of the cells by measuring heat output using a calorimeter,
and
Kit for carrying out said method, comprising a calorimeter.



The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

6. claims: 5-7, 10-13

Method of determining a suitable administration regimen for a patient, by measuring the immune response, microorganism load or number of malignant cells in a sample from the patient by using calorimetry,
and
Kit for carrying out said method, comprising a calorimeter.

7. claims: 8-13

Method of determining whether an anti-microorganism or anti-infective compound is able to cause growth of a microorganism/virus/mammalian cell, by adding the compound to the microorganism to determine whether or not growth or heat output of the microorganism/virus/mammalian cell occurs,
and
Kit for carrying out said method, comprising a calorimeter.

**ANNEX TO THE EUROPEAN SEARCH REPORT
ON EUROPEAN PATENT APPLICATION NO.**

EP 05 25 6977

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

17-02-2006

Patent document cited in search report		Publication date		Patent family member(s)		Publication date
US 3765237	A	16-10-1973	DE	2216411 A1		26-10-1972
			FR	2132484 A5		17-11-1972
			GB	1341944 A		25-12-1973
			IT	972421 B		20-05-1974
			JP	53025273 B		26-07-1978

US 6436346	B1	20-08-2002	AU	7480500 A		17-04-2001
			WO	0120283 A1		22-03-2001

US 3298790	A	17-01-1967	NONE			

专利名称(译)	微量热分析哺乳动物细胞对影响细胞生长，代谢和免疫反应的药剂的敏感性，以及在移植试验中使用微量热分析		
公开(公告)号	EP1785723A1	公开(公告)日	2007-05-16
申请号	EP2005256977	申请日	2005-11-11
[标]申请(专利权)人(译)	STRATOS BIO		
发明人	KJELDTSEN, NAJA JOSLIN		
IPC分类号	G01N25/20 G01N33/48 G01K17/00 C12Q1/04 A61B5/00		
CPC分类号	G01K17/006 C12Q1/04 C12Q1/18		
代理机构(译)	BENSON , JOHN EVERETT		
外部链接	Espacenet		

摘要(译)

确定用于移植的器官（来自人类供体）或异种移植（来自动物供体）是否被微生物（任选地是病毒）感染的方法包括 - 测量来自器官的热输出以检测所述微生物的活性。或者 - 通过使用刺激m-RNA合成的试剂强制潜伏DNA的表达，从而确定器官中微生物的存在。

